

# Supporting Information

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## SI Methods

**Practical Considerations and Limitations of the Proposed Sensing Strategy.** Nine practical considerations and limitations should be taken into account before attempting to implement this sensing strategy. (i) Not all materials are rf-transparent. Therefore, the technique can be compromised by the presence of materials that are rf-opaque or reflect rf radiation. (ii) CARDS cannot be stacked on top of one another (see the discussion in *Methods, Binary Logic for Chemical Discrimination Using Arrays of CARDS*). (iii) NFC relies on inductive coupling and therefore the technique is sensitive to its magnetic environment. (iv) The technique, as described in *Methods, Method for Determining Reflection Coefficient and Readability of CARDS with a Smartphone*, is sensitive to the relative orientation of and distance between the smartphone and CARD. (v) Based on our findings, the on/off threshold is dictated by the amplitude of power transfer between the smartphone and the CARD. Therefore, the make and model of the smartphone may influence the on/off threshold. (vi) Based on our findings, the on/off threshold is dependent on the PENCIL material. (vii) The chemiresponsive materials used in this study are unprotected from the atmosphere of the laboratory and their performance may degrade over time. (viii) Because the sensing element is exposed, the behavior of the chemiresistor may change abruptly if touched or otherwise disrupted. (ix) This technique is demonstrated in the controlled setting of a laboratory. In a non-laboratory setting, human and environmental exposure to nanomaterials would have to be addressed with packaging around the sensing element.

**General Materials and Methods.** SWCNTs (purified  $\geq 95\%$  as SWCNT) were kindly provided by Nano-C, Inc. HFIPN (CAS 2092-87-7) was purchased from SynQuest.  $\text{NH}_3$  (1% in  $\text{N}_2$ ) was custom-ordered from Airgas. All NFC tags used in this study (hereafter referred to generically as “NFC tag”) were Texas Instruments HF-I Tag-It 13.56-MHz rf identification transponder square inlays (RI-I11-114A-01), purchased from DigiKey.

**Choice of Tags.** This study uses commercially available Texas Instruments HF-I Tag-It Plus Transponder Inlays (TI-Tag) to demonstrate the concept of converting a commercially available NFC tag into a chemical sensor. These tags were chosen based on their chemically robust substrate, absence of protective polymeric coating over the circuitry, commercial availability, and low cost. The electronic circuitry of the unmodified tags is supported via polyurethane glue on both sides of a thin (47- $\mu\text{m}$ ), flexible sheet of polyethylene terephthalate, which also serves as a dielectric layer for the capacitor. The circuit comprises an aluminum antenna that serves as an inductor ( $L$ ), a parallel-plate aluminum capacitor ( $C$ ), and a silicon-based IC chip ( $R$ ), all connected in parallel, forming an  $LCR$  resonant circuit (Fig. 1).

**Choice of Analytes.** The selective detection of a target chemical analyte is a necessary requirement for any functional ultra-low-cost distributed chemical sensor. We achieve this requirement in a manner that does not use extensive data analysis or computationally intensive interpretation and achieves selectivity toward analytes by harnessing established properties of chemiresponsive materials (1, 2). We targeted detection of ammonia ( $\text{NH}_3$ ) gas and vapors of cyclohexanone ( $\text{C}_6\text{H}_{10}\text{O}$ ), hydrogen peroxide ( $\text{H}_2\text{O}_2$ ), and water ( $\text{H}_2\text{O}$ ) as model analytes for the detection of industrial, agricultural, and safety hazards.  $\text{NH}_3$  is commonly emitted in industrial and agricultural settings and is toxic at

relatively low levels (3). Cyclohexanone is a volatile organic compound, commonly used for recrystallization of explosives, such as RDX (4), that can also aid their detection (5).  $\text{H}_2\text{O}_2$  can be used in improvised explosive devices, as a commonly used industrial reagent, and is routinely used for sanitization in hospitals.

**Choice of Smartphone.** We use an off-the-shelf smartphone to demonstrate the capability for wireless chemical sensing. This type of detector would be compatible with a highly distributed network of sensors accessible to a large number of people. In this context, the SGS4 was chosen as the primary NFC-enabled smartphone as a result of two factors: (i) At the time of this study, the Samsung Galaxy series are among the most widely distributed “smart” mobile devices in history and (ii) the SGS4 runs on Android, one of the most widely distributed operating systems that supports NFC applications. The demonstrated wireless chemical sensing via NFC is applicable to other NFC-enabled devices (Fig. S1). The NFC chip comprises an antenna for inductive coupling with NFC tags, a transmission module with microcontroller for 13.56-MHz carrier signal generation and tag signal demodulation, as well as embedded and external [subscriber identity module (SIM) card] security elements. When used with unmodified TI-tags, the SGS4 can read tags at  $\sim 5$  cm standoff distance through solid, nonmetallic objects such as paper, plastic, and liquids (Fig. S1).

**Choice of Smartphone Application.** The NFC Reader (Adam Nybäck) and NFC TagInfo (NFC Research Lab) applications were used to read the tags and were freely available from the Google Play Store at the time of this study. These applications were chosen because they display the tag’s UID without invoking other time- or energy-intensive functions of the smartphone. For the purposes of this study, the tag is considered on or readable if the UID can be retrieved within 5 s or less of holding the smartphone at  $\sim 2.5$  cm distance away from the tag. Conversely, the tag is considered off or unreadable if the UID cannot be retrieved under the same conditions.

**Instrumental Analysis.** We monitored the rf signal response of the modified TI-tags and smartphone antennas from 10 to 20 MHz, as well as the smartphone-transmitted rf signal, with a custom-made loop probe connected via a BNC cable to a vector network analyzer (VNA) (Agilent E5061B) by measuring reflection coefficient ( $S_{11}$ ) at 50  $\Omega$  port impedance and 0 dBm input power (Fig. S4).

**Ball Milling.** Cyclohexanone sensing material was generated by solvent-free ball milling of SWCNTs with HFIPN using an oscillating mixer mill (MM400; Retsch GmbH) within a stainless steel milling vial (5 mL) equipped with a single stainless steel ball (7 mm in diameter). The milling vial was filled with HFIPN (96 mg) and SWCNTs (24 mg) and the mixture was ball-milled for 5 min at 30 Hz.

**Fabrication of PENCILs.** PENCILs were fabricated by loading powdered sensing material into a steel pellet press (6-mm internal diameter) (SDS6; Across International) and compressing the powder by applying a constant pressure of 10 MPa for 1 min using a hydraulic press (MP24A; Across International).

**Fabrication of Loop Probe.** Hollow copper tubing covered in heat-shrink wrap was shaped into a square (5  $\times$  5 cm) shape and soldered to a BNC adapter. Heat-shrink wrap was placed over

the connection point and was shrunk using a heat gun in a fume hood.

**Dilution of Ammonia.** Delivery of controlled concentrations of  $\text{NH}_3$  to the sensing devices placed within a gas chamber was performed using a Smart-Trak Series 100 (Sierra Instruments) gas mixing system at total flow rates between 0.50 and 10.00 L/min.  $\text{NH}_3$  was diluted with  $\text{N}_2$ .

**Dilution of Vapors.** Delivery of controlled concentrations of cyclohexanone vapors to the sensing devices placed within the gas chamber was carried out using Precision Gas Standards Generator Model 491M-B (Kin-Tek Laboratories). Cyclohexanone was diluted with  $\text{N}_2$  at total flow rates of 0.25–0.50 L/min.

**Gas Chamber.** A custom gas chamber was fabricated by inserting two plastic syringes (1 mL; NORM-JECT), one on either side, in the bottom corners of a zippered plastic bag (1 L) and sealing it with electrical tape.

**Detection of  $\text{NH}_3$ .** Sensor tag data were collected according to the method described above. The sensor tag was kept on the benchtop of a fume hood for 10 min, followed by exposure to  $\text{NH}_3$  in  $\text{N}_2$  (35 ppm) in a gas chamber for 5 min, followed by removal and placement on a benchtop of a fume hood for 10 min. This procedure was repeated three more times; after the fourth cycle, the sensor tag was allowed to sit on the fume hood benchtop for an additional 10 min.

**Detection of a Single Exposure of  $\text{N}_2$  (Negative Control).** Sensor tag data  $R_s$  and readability by SGS4 was determined according to the method described above. The sensor tag was kept on the benchtop of a fume hood for 10 min, followed by exposure to  $\text{N}_2$  in a gas chamber for 5 min, followed by removal and placement on the fume hood benchtop for 20 min.

**Detection of a Single Exposure of  $\text{NH}_3$ .** Sensor tag data were collected according to the method described above. The sensor tag was kept on the benchtop of a fume hood for 10 min, followed by exposure to  $\text{NH}_3$  in  $\text{N}_2$  (4 ppm or 35 ppm) in a gas chamber for 5 min, followed by removal and placement on the fume hood benchtop for 20 min.

**Detection of a Single Exposure of  $\text{C}_6\text{H}_{10}\text{O}$ .** Sensor tag data were collected according to the method described above. The sensor tag was kept on a benchtop underneath a ventilation snorkel for 10 min, followed by exposure to cyclohexanone ( $\text{C}_6\text{H}_{10}\text{O}$ ) in  $\text{N}_2$  (335 ppm) in a gas chamber for 5 min, followed by removal and placement on a benchtop underneath a ventilation snorkel for 20 min.

**Detection of a Single Exposure of  $\text{H}_2\text{O}_2$ .** Sensor tag data were collected according to the method described above. The sensor tag was kept on the benchtop of a fume hood for 10 min, followed by exposure to  $\text{H}_2\text{O}_2/\text{H}_2\text{O}$  ( $P_{eq}$ ) in a plastic zippered bag containing an open jar of  $\text{H}_2\text{O}_2/\text{H}_2\text{O}$  (35%) for 5 min, followed by removal and placement on the fume hood benchtop for 20 min.

**Detection of a Single Exposure of  $\text{H}_2\text{O}$ .** Sensor tag data were collected according to the method described above. The sensor tag was kept on the benchtop of a fume hood for 10 min, followed by exposure to  $\text{H}_2\text{O}$  (100% humidity in air) in a plastic zippered bag

containing an open jar of water for 5 min, followed by removal and placement on the fume hood benchtop for 20 min.

**Semiquantitative Detection of  $\text{NH}_3$ .** A sensor tag for 4 ppm  $\text{NH}_3$  (**CARD-1B**) was fabricated with  $R_s = 19.2 \pm 0.2 \text{ k}\Omega$  and a sensor tag for 35 ppm  $\text{NH}_3$  (**CARD-1A**) with  $R_s = 16.3 \pm 0.5 \text{ k}\Omega$ . Before exposure to  $\text{NH}_3$  both types of tags were on and readable by the phone (Fig. 3B and Fig. S8). Upon exposure to 4 ppm  $\text{NH}_3$ , **CARD-1B** turned off within 1 min of experiencing a change to its local environment, whereas **CARD-1A** remained on. After 5 min of exposure to 4 ppm  $\text{NH}_3$ , **CARD-1B** had  $R_s = 21.9 \pm 0.4 \text{ k}\Omega$  ( $\Delta R_s = 2.8 \pm 0.4 \text{ k}\Omega$ ); **CARD-1A** displayed  $R_s = 18.8 \pm 0.3 \text{ k}\Omega$  ( $\Delta R_s = 2.6 \pm 0.1 \text{ k}\Omega$ ). The same type of experiment, with a new batch of **CARD-1A** and **CARD-1B**, each fabricated in triplicate, was repeated for 35 ppm  $\text{NH}_3$  (Fig. 3B). Under these conditions, CARDS turned off ( $\Delta R_s = 6.0 \pm 0.5 \text{ k}\Omega$ ); **CARD-1B**  $R_s$  increased to  $25.8 \pm 0.6 \text{ k}\Omega$  ( $\Delta R_s = 6.3 \pm 0.1 \text{ k}\Omega$ ), and **CARD-1A**  $R_s$  increased to  $21.9 \pm 0.8 \text{ k}\Omega$  ( $\Delta R_s = 5.4 \pm 0.8 \text{ k}\Omega$ ), both above the readability threshold.

**Determination of Estimated Power Transfer from SGS4 to CARDS.** The power transferred from SGS4 to CARD-2 at each stage of fabrication was determined according to a seven-step procedure. (i) We collected  $S_{11}$  spectra ( $n = 5$ ) (10–20 MHz) of the SGS4-generated signal and averaged them into a single SGS4-signal spectrum. (ii) We collected  $S_{11}$  spectra ( $n = 5$ ) (10–20 MHz) at each stage of modification of a tag leading to the formation of CARD-2. Additionally we collected  $S_{11}$  spectra ( $n = 5$ ) (10–20 MHz) of CARD-2 before and after exposure to saturated cyclohexanone vapor, as described in Fig. 2A. (iii) We averaged the spectra collected in step ii into a single spectrum for each tag modification stage and for the gas exposure scenario. (iv) The SGS4-signal spectrum and each spectrum from step iii was zeroed according to their response at 20 MHz. (v) The zeroed SGS4-signal spectrum from step iv was added to each zeroed tag and CARD-2 spectrum from step iv to yield SGS4-tag composite spectra (Fig. S6A). (vi) The power reflected back to the network analyzer,  $P_{re}$ , was determined according to Eq. S1:

$$S_{11} = 10 \log \left( \frac{P_{re}}{P_{in}} \right), \quad [\text{S1}]$$

where incident power ( $P_{in}$ ) is 0 dBm (1  $\mu\text{W}$ ) (Fig. S6 B and C). (vii) The percent power transferred in each case ( $P_t$ ) (Fig. 2B) was estimated by Eq. S2 (Fig. S6C):

$$P_t(\%) = \left[ \frac{\left( \int_{13.53 \text{ MHz}}^{13.58 \text{ MHz}} P_{re}^{SGS4} df - \int_{13.53 \text{ MHz}}^{13.58 \text{ MHz}} P_{re}^x df \right)}{\int_{13.53 \text{ MHz}}^{13.58 \text{ MHz}} P_{re}^{SGS4} df} \right] \times 100\%, \quad [\text{S2}]$$

where  $x$  corresponds to scenarios 1–6 described in Fig. 2A of the main text.

**Determination of  $R_t$ .** The on/off threshold,  $R_t$ , was estimated (Table S1) by taking the average of the median  $R_s$  values found between the “last”  $R_s$  correlated with an unreadable CARD and the “first”  $R_s$  correlated with a readable CARD, during recovery from a given exposure to analyte.

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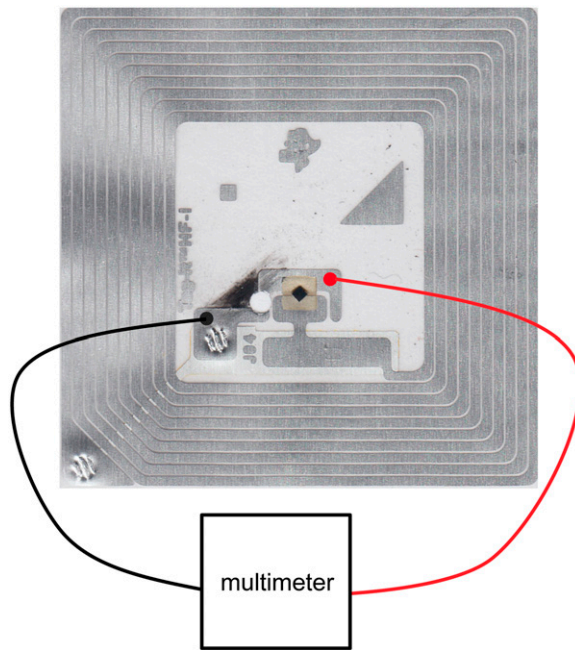


Fig. S5.  $R_s$  was measured using a multimeter by contacting the CARD at the locations depicted above.





