

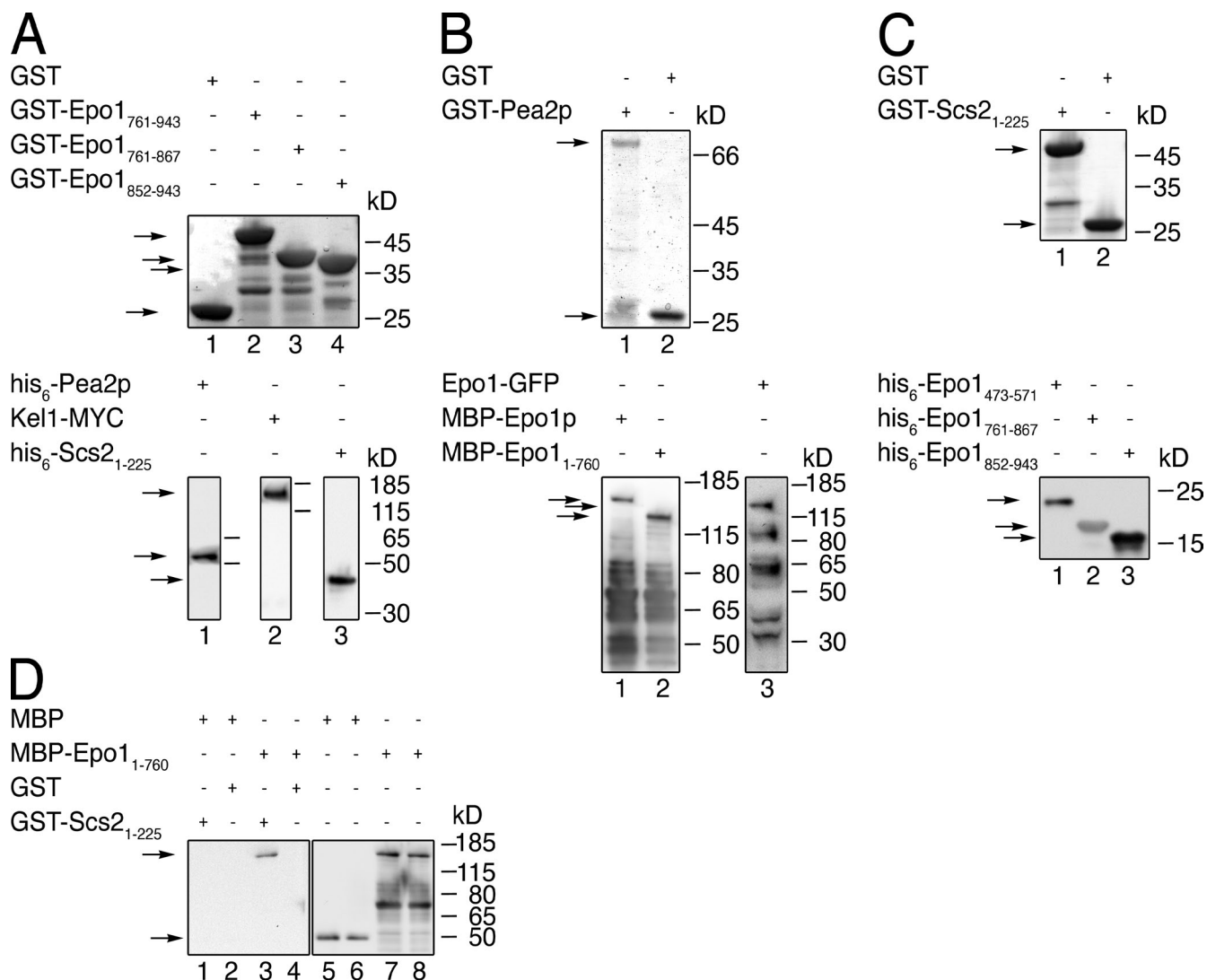
Neller et al., <http://www.jcb.org/cgi/content/full/jcb.201407126/DC1>

Figure S1. **Expression of fusion proteins used for pull-down analysis.** (A, top) SDS-PAGE of glutathione eluates of glutathione-coupled Sepharose beads incubated with extracts of bacterial cells expressing GST (lane 1), GST-Epo1₇₆₁₋₉₄₃ (lane 2), GST-Epo1₇₆₁₋₈₆₇ (lane 3), or GST-Epo1₈₅₂₋₉₄₃ (lane 4). The proteins were stained with Coomassie. (bottom) SDS-PAGE of yeast (lane 2) or *E. coli* extracts (lanes 1 and 3) containing his₆-Pea2p (lane 1), Kel1-MYC (lane 2), or his₆-Scs2₁₋₂₂₅ (lane 3). After transfer, the fusion proteins were detected by Western blotting using anti-His (lanes 1 and 3) or anti-MYC (lane 2) antibodies. (B, top) As in A (top), but with extracts from *E. coli* expressing GST-Pea2p (lane 1) or GST (lane 2). (bottom) SDS-PAGE of *E. coli* (lanes 1 and 2) or yeast extracts (lane 3) containing MBP-Epo1p (lane 1), MBP-Epo1₁₋₇₆₀ (lane 2), or Epo1-GFP (lane 3). After transfer, the fusion proteins were detected by Western blotting using anti-MBP (lanes 1 and 2) or anti-GFP (lane 3) antibodies. (C, top) As in A (top), but with extracts from *E. coli* expressing GST-Scs2₁₋₂₂₅ (lane 1) or GST (lane 2). (bottom) SDS-PAGE of *E. coli* extracts containing his₆-Epo1₄₇₃₋₅₇₁ (lane 1), his₆-Epo1₇₆₁₋₈₆₇ (lane 2), or his₆-Epo1₈₅₂₋₉₄₃ (lane 3). After transfer, the fusion proteins were detected by Western blotting using anti-His antibodies. (D) Eluates of Sepharose bead-coupled GST-Scs2₁₋₂₂₅ (lanes 1 and 3) or GST (lanes 2 and 4) incubated with *E. coli* extracts containing MBP (lanes 1, 2, 5, and 6) or MBP-Epo1₁₋₇₆₀ (3, 4, 7, and 8) were separated by SDS-PAGE. Western blots were probed with anti-MBP antibodies. Lanes 5–8 show the unbound fractions of the extracts. The arrows indicate the running positions of the fusion proteins on the gel.

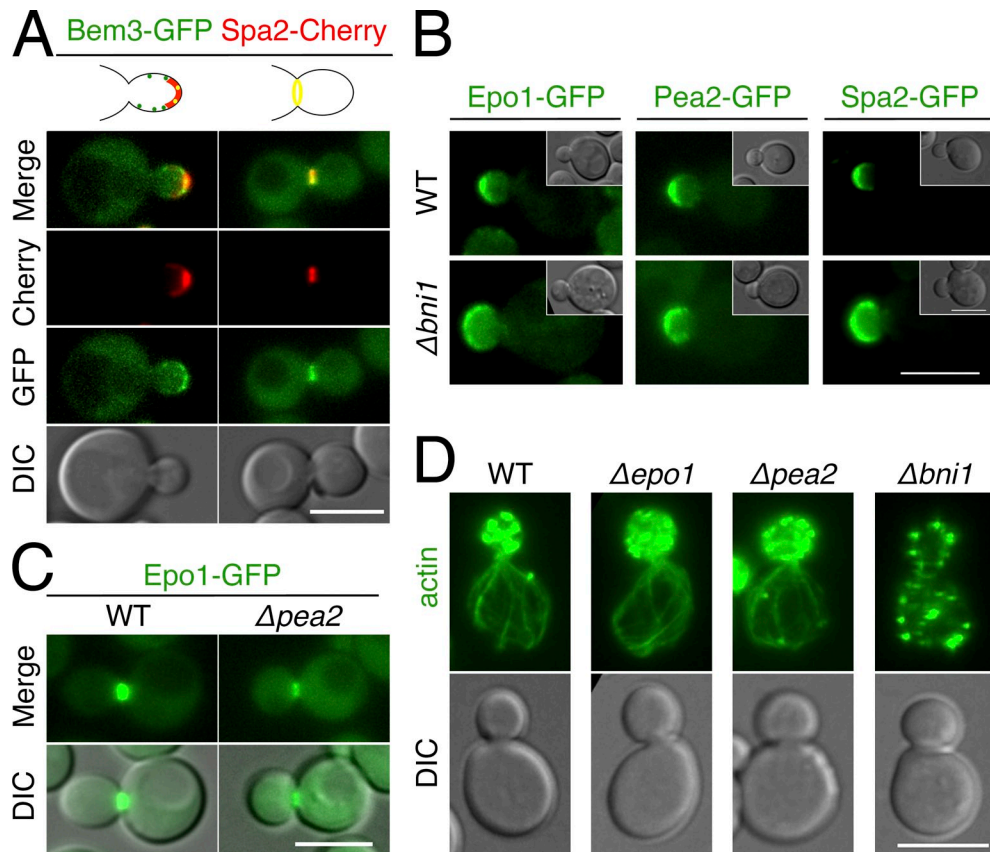


Figure S2. **Localizations of Bem3-GFP in WT cells and of Epo1-, Pea2-, and Spa2-GFP in $\Delta bni1$ or $\Delta pea2$ cells and the actin staining of WT and of $\Delta epo1$ cells.** (A) Bud tip and neck staining of Bem3-GFP. Two-channel microscopy of cells coexpressing Bem3-GFP and Spa2-Cherry. Shown is one representative cell during tip growth or mitosis. (B) Epo1-, Pea2-, and Spa2-GFP lose their focused distribution upon deletion of *BNI1*. Shown are the GFP images of one representative cell each during tip growth in WT (top) or $\Delta bni1$ cells (bottom). Insets show a reduction of the DIC image of the same cell. (C) Representative images of WT and $\Delta pea2$ cells displaying Epo1-GFP at the bud neck. (D) Alexa Fluor 488 Phalloidin staining of WT cells and cells lacking *EPO1*, *PEA2*, or *BNI1*. Note the WT-like actin cytoskeleton in $\Delta pea2$ and $\Delta epo1$ cells as compared with $\Delta bni1$ cells. Bars, 5 μ m.

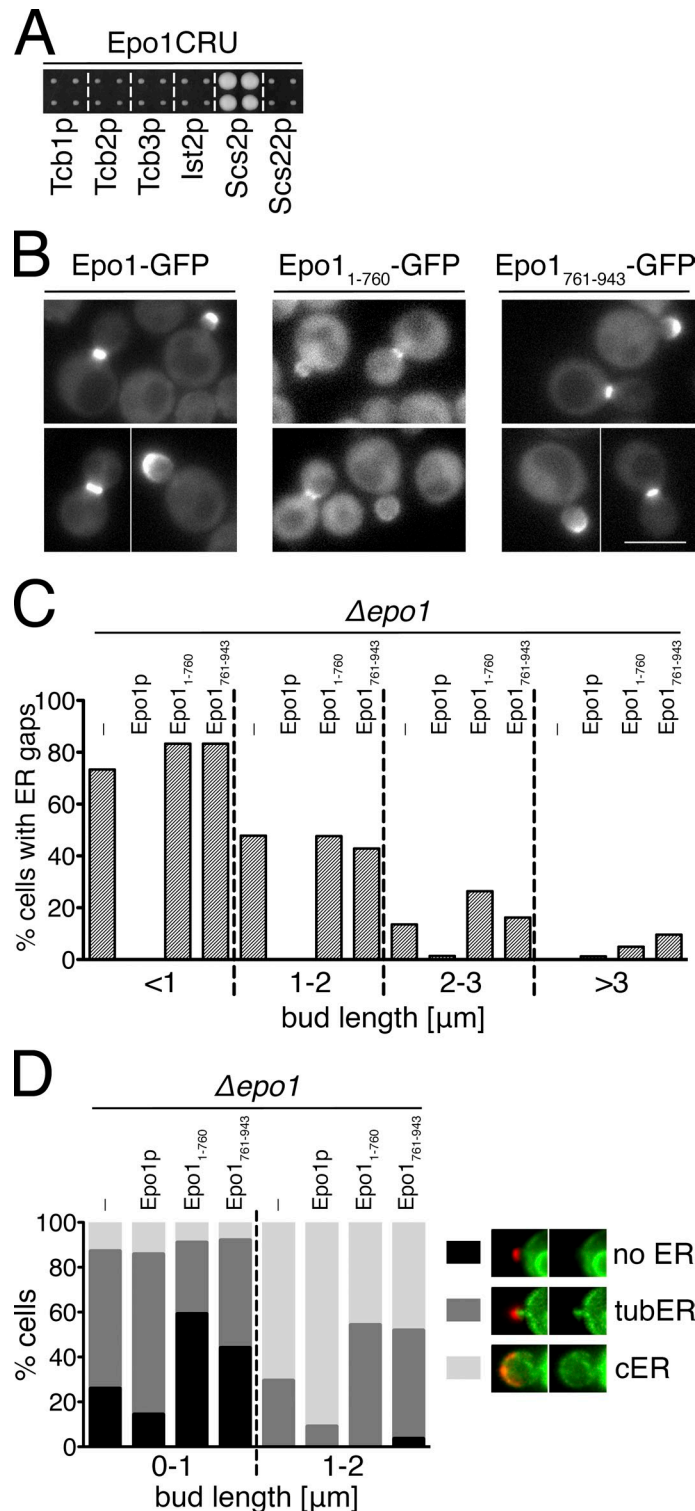


Figure S3. **Array-based Split-Ub interaction assay between Epo1CRU and the N_{ub} fusions of all ER-PM tethers and the complementation of Δ epo1 cells by the expression of Epo1p fragments.** (A) Epo1p interacts exclusively with the ER-PM tether protein Scs2p. Split-Ub interaction assay of six yeast strains each coexpressing Epo1CRU with either N_{ub} -fused Tcb1p, Tcb2p, Tcb3p, Ist2p, Scs2p, or Scs22p. Shown are quadruplets of each strain after 4 d of growth on medium containing 5-FOA. Dashed lines separate the yeast strains expressing the different N_{ub} fusion proteins. (B) The isolated Scs2p binding sites on Epo1p do not complement the loss of *EPO1*. Fluorescence images of cells expressing Epo1-GFP (left) or Epo1₁₋₇₆₀-GFP (middle), both from the native promoter, or Epo1₇₆₁₋₉₄₃-GFP (right) expressed from the *P_{MET17}* promoter. Bar, 5 μ m. (C) Δ epo1 cells expressing GFP-Sec2p and an empty plasmid (–) or a plasmid carrying the Cherry fusion to Epo1p, Epo1₁₋₇₆₀, or Epo1₇₆₁₋₉₄₃ were quantified for gaps of ER staining at the bud tip as in Fig. 7 B (42 < *n* < 300). (D) Δ epo1 cells expressing GFP-Sec2p and an empty plasmid (–) or a plasmid carrying the Cherry-fusion to Epo1p, Epo1₁₋₇₆₀, or Epo1₇₆₁₋₉₄₃ were scored for the three stages of cER inheritance (no, tubular [tubER], or cER). The same representative images were shown previously in Fig. 7 C. Quantification was as described in Fig. 7 C (105 < *n* < 225).

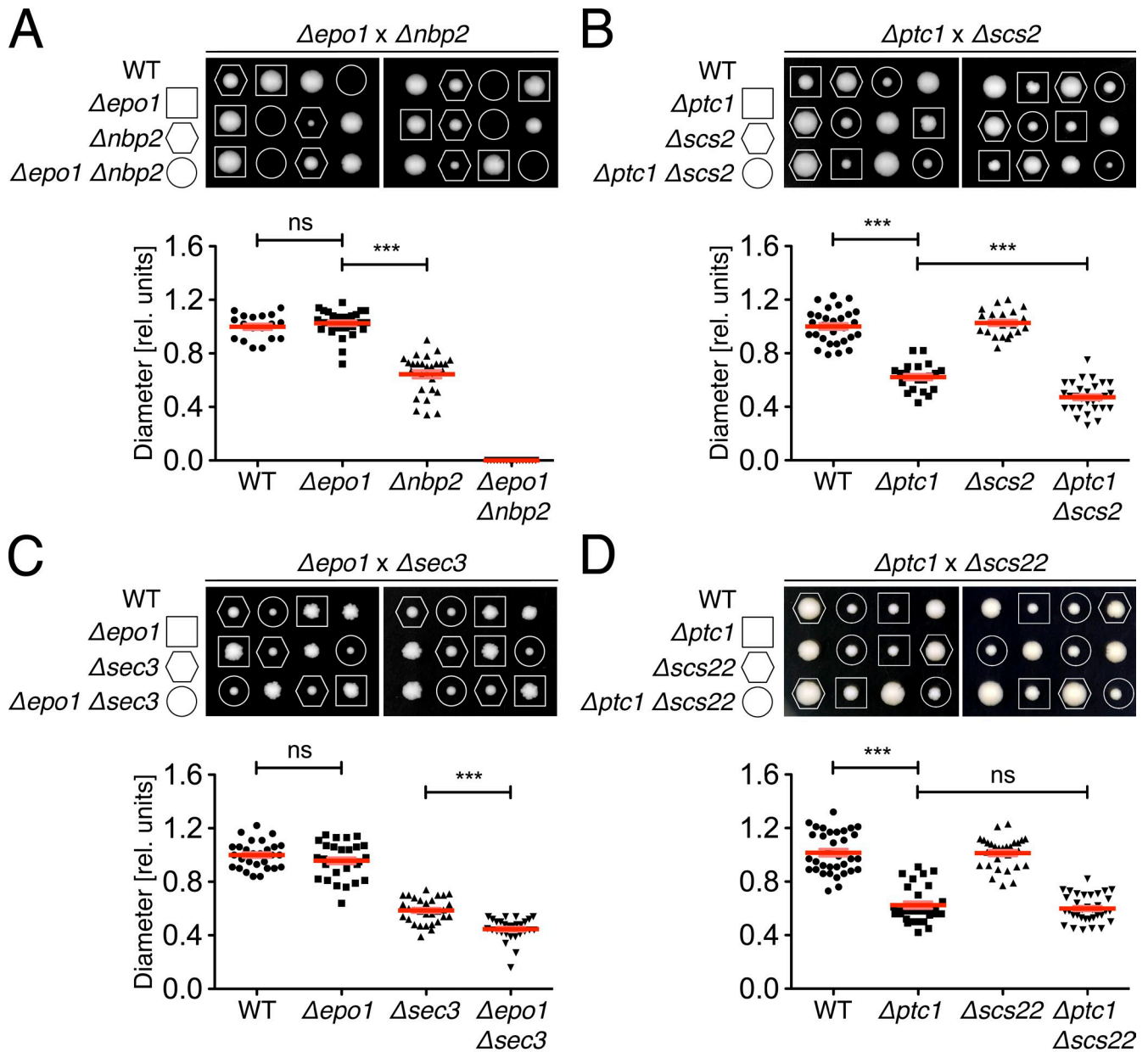


Figure S4. **Tetrad analyses of heterozygous double-deletion strains after sporulation.** (A, top) Tetrad analysis of the meiotic progeny derived from diploid cells heterozygous for $\Delta epo1$ and $\Delta nbp2$. (bottom) Quantification of the diameters of the spores ($17 < n < 31$; error bars show SEM). ***, $P < 0.001$. (B) As in A, but with $\Delta ptc1/PTC1$; $\Delta scs2/SCS2$ cells. (C) As in A, but with $\Delta epo1/EPO1$; $\Delta sec3/SEC3$ cells. (D) As in A, but with $\Delta ptc1/PTC1$; $\Delta scs22/SCS22$ cells. Spores in A, B, and D were grown at 30°C for 4 d and, in C, at 23°C for 6 d. rel., relative.

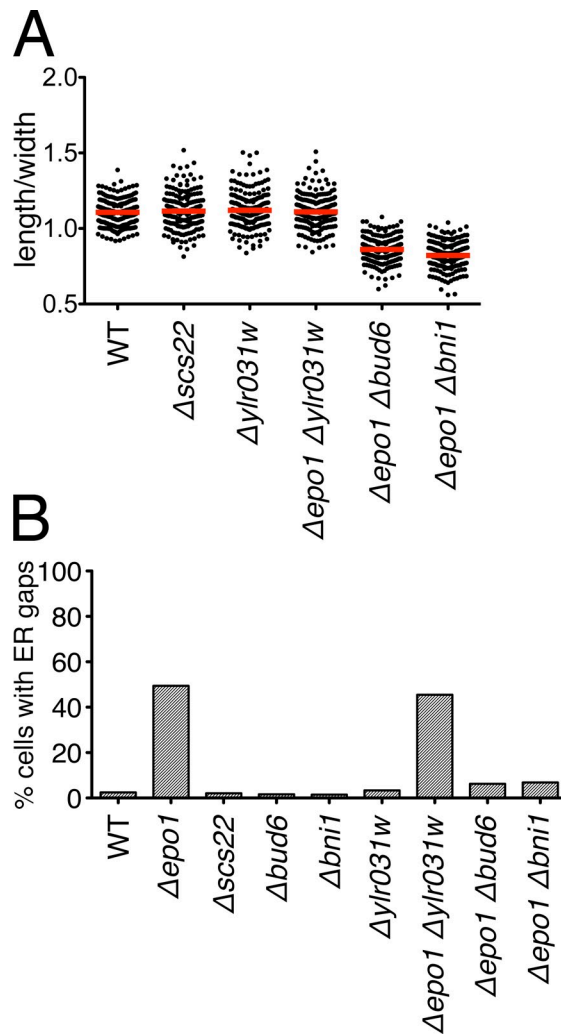


Figure S5. **Quantification of length/width ratios and ER gaps.** Quantification of length/width ratios ($200 < n < 206$, error bars show SEM; the value for the WT is taken from Fig. 3 D) and ER gaps ($80 < n < 139$) of different yeast deletion strains. Analysis was performed as in Fig. 8 (A and B).

Table S1. List of all tested N_{ub} fusions

ABP1	BUD20	EDE1	KEL1	PAN1	SEC4	STE11
ACF2	CAP1	ELG1	KEL2	PBS2	SEC5	STE20
ACT1(P)	CBK1	ELM1	KIC1	PDE1	SEC6	STE50
ACT1 ^{V152G;T318I}	CDC3	ELO1	KIN1	PDS1	SEC8	STT4
AFR1	CDC5	END3	KIN2	PEA2	SEC9	STU1
AIP1	CDC6	ENT2	KIN4	PEX4(P)	SEC10	STU2
AIR1	CDC10	EPO1	KIP2	PEX19(P)	SEC14	SVL3
AMN1	CDC11	ESP1	KIP3	PFK1	SEC15	SWE1
APC2	CDC12	EXO70	KOG1	PFY1	SEC62(P)	SWI4
APG17	CDC14	EXO84	KSS1	PHO85	SFH1	SWI6
APP1	CDC20	FAB1	LAC1	PIK1	SFH2	SYP1
ARC1	CDC24	FAR1	LAG1	PIL1	SFH3	TAO3
ARC40	CDC25	FKH1	LAS17	PKC1	SFH4	TCB1
ARP1	CDC28	FKH2	LCB4	PKH1	SFH5	TCB2
ARP6	CDC37	FKS1	LCB5	PKH2	SFK1	TCB3
ASE1	CDC42(P)	FKS2	LDB17	PKH3	SHE4	TCO89
ASK10	CHK1	FUS2	LRE1	PLC1	SHO1(P)	TDH1
AUR1	CHS1	FUS3	LRG1	PNC1	SHO1 ^{Cterm}	TEF1
AVO1	CHS2	GIC1	LSP1	PPH3	SHS1	TEL1
AVO2(P)	CHS3	GIM4	LST8	PRK1	SIT4	TEM1(P)
AXL2 ^{Cterm}	CIK1	GIN4	LTE1	PSY4	SIZ1	TLG2(P)
BBC1	CIN1	GLG1	MAD2	PTC1	SKG6	TOK1
BCK1	CIN8	GPA2	MAD3	PTC2	SKM1	TOR1
BCK2	CLA4	GPB1	MCD1	PUF3	SKN7	TOR2
BCY1	CLB1	GPB2	MCK1	RAD9	SKO1	TOS2
BEM1	CLB2	GSY1	MEC1	RAS1	SKT5	TPK1
BEM2	CLB3	GUK1(P)	MID2 ^{Cterm}	RAS2	SLA1	TPM1
BEM3	CLB4	GUS1	MKK1	RAX1	SLA2	TPM2
BEM4	CLN1	HCM1	MLC1(P)	RGA2	SLI15	TSC11
BFA1	CLN2	HLR1	MLC2	RGD1	SLM1	TSC13
BIK1	CLN3	HOF1	MOB1	RGS2	SIT2	TUS1
BIM1	CMD1	HOF1 ⁹⁸⁻⁶⁹⁹	MPS1	RHO1	SMC1	UBC6(P)
BIR1	CMK1	HOG1	MSB1	RHO2	SMI1	UBC9
BIT61	CMP2	HOT1	MSB3	RHO4	SMP1	UBP7
BMH1	CNA1	HSK3	MSB4	RHO5	SMT3(P)	VAC8
BMH2	CNB1	HSL1	MSN1	RIM21	SMY1	VAC17
BNI1	CPR6	HSL7	MSN2	RLM1	SNC1(P)	VHS1
BNI1 ^{Cterm}	CRN1	HSP82	MSN4	ROM1	SOG2	VHS2
BNI4	CRZ1	HUA1	MSO1	ROM2	SPA2	VMA13
BNI5	CSE4	HYM1	MSS4	RSR1	SPC110	VRP1
BNR1	CYK3	IME2	MYO1	RVS161	SPC19	WSC1 ^{Cterm}
BNR1 ^{Cterm}	CYK3 ⁶⁹⁻⁸⁸⁵	INN1	MYO2	RVS167	SPC24	YBR053C
BOI1	CYK3 ¹⁹⁶⁻⁸⁸⁵	INP51	MYO5	SAC1	SPC34	YBR238C
BOI1 ¹⁻⁹⁵ (P)	CYR1	INP52	NAP1	SAC6	SPC72	YDL173W
BOI1 ⁷⁵⁸⁻⁹⁸⁰ (P)	DAD1	IPL1	NAS2	SAC7	SPH1	YDL203C
BOI2	DAD2	IPP1	NBA1	SAK1	SPR28	YDR348C
BUB2	DAD3	IQG1	NBP2	SCC2	SPT5	YEL023C
BUB3	DAM1	IQG1 ^{Cterm}	NET1	SCC4	SRO7	YGR058W
BUD2	DBF2	IRA1	NHA1	SCP1	SRO77	YKL161C
BUD3	DBF20	IRA2	NIP100	SCP160	SRV2	YOR304C-A
BUD4	DOP1	IRR1	NIS1	SCS2	SSD1	YPD1
BUD5	DSE1	IRS4	NUM1	SCS22	SSK1	YPK1
BUD6	DUN1	IST2	ORC6	SDP1	SSK2	ZDS1
BUD8	DUO1	KAR3	PAC1	SEC1	SSO1	ZDS2
BUD9	DYN1	KAR9	PAC10	SEC2	STE5	ZEO1
BUD14	ECM25	KCC4	PAM1	SEC3	STE7	

(P), plasmid encoded; Cterm, N_{ub} fused to the C terminus of the protein.

Table S2. Identities and positions of the N_{ub} fusions on the N_{ub} array shown in Fig. 1 A

Aip1	Bem1	Bem2	Bem3	Bem4	Boi1	Boi2	Scs2	Scs2	Bud6	Bud14	Cdc42	Ist2	Exo70	Exo84	Hof1	Kel1	Kel2	Kin1	Kin2	Mso1	Tcb1	Nbp2	Pec2
Ptc1	Rga2	Rgd1	Rho1	Rho4	Sac6	Sec1	Sec2	Sec3	Sec4	Sec5	Sec6	Sec8	Sec9	Sec10	Sec15	Snc1	Spa2	Sro7	Sro77	Sso1	Epo1	Yor304c	N_{ub}^-

Table S3. List of constructs used and created in this study

Name	Description	Origin
Epo1-CRU pRS303	<i>EPO1-C_{ub}-R-URA3, Amp^R, HIS3</i>	This work
Pea2-CRU pRS303	<i>PEA2-C_{ub}-R-URA3, Amp^R, HIS3</i>	This work
Istf2-CRU pRS303	<i>IST2-C_{ub}-R-URA3, Amp^R, HIS3</i>	This work
Tcb1-CRU pRS303	<i>TCB1-C_{ub}-R-URA3, Amp^R, HIS3</i>	This work
Tcb2-CRU pRS303	<i>TCB2-C_{ub}-R-URA3, Amp^R, HIS3</i>	This work
Tcb3-CRU pRS303	<i>TCB3-C_{ub}-R-URA3, Amp^R, HIS3</i>	This work
<i>P_{MET17}-Kel1-CRU pRS313</i>	<i>P_{MET17}-KEL1-C_{ub}-R-URA3, Amp^R, HIS3</i>	This work
<i>P_{MET17}-Epo1-CRU pRS313</i>	<i>P_{MET17}-EPO1-C_{ub}-R-URA3, Amp^R, HIS3</i>	This work
<i>P_{MET17}-Epo1₇₆₁₋₉₄₃-CRU pRS313</i>	<i>P_{MET17}-EPO1₇₆₁₋₉₄₃-C_{ub}-R-URA3, Amp^R, HIS3</i>	This work
<i>Epo1₁₋₇₆₀-CRU pRS303</i>	<i>EPO1₁₋₇₆₀-C_{ub}-R-URA3, Amp^R, HIS3</i>	This work
<i>P_{MET17}-Epo1₁₋₈₇₆-CRU pRS313</i>	<i>P_{MET17}-EPO1₁₋₈₇₆-C_{ub}-R-URA3, Amp^R, HIS3</i>	This work
<i>P_{CUP1}-Nub-HA kanMX4</i>	<i>P_{CUP1}-Nub-HA, Amp^R, kanMX4</i>	Dünkler et al., 2012
<i>P_{CUP1}-Nub-HA-kanMX4 CEN</i>	<i>P_{CUP1}-Nub-HA kanMX4 CEN, Amp^R</i>	Dünkler et al., 2012
<i>P_{CUP1}-Nub-HA-Guk1 kanMX4 CEN</i>	<i>P_{CUP1}-Nub-HA-GUK1 kanMX4 CEN, Amp^R</i>	This work
<i>P_{CUP1}-Nub-HA-Bem3₁₋₆₃₃ kanMX4 CEN</i>	<i>P_{CUP1}-Nub-HA-BEM3₁₋₆₃₃ kanMX4 CEN, Amp^R</i>	This work
<i>P_{CUP1}-Nub-HA-Bem3₁₋₃₀₀ kanMX4 CEN</i>	<i>P_{CUP1}-Nub-HA-BEM3₁₋₃₀₀ kanMX4 CEN, Amp^R</i>	This work
<i>P_{CUP1}-Nub-HA-Bem3₁₋₁₀₀ kanMX4 CEN</i>	<i>P_{CUP1}-Nub-HA-BEM3₁₋₁₀₀ kanMX4 CEN, Amp^R</i>	This work
<i>P_{CUP1}-Nub-HA-Bem3₃₀₀₋₆₃₃ kanMX4 CEN</i>	<i>P_{CUP1}-Nub-HA-BEM3₃₀₀₋₆₃₃ kanMX4 CEN, Amp^R</i>	This work
<i>P_{CUP1}-Nub-HA-Scs2₁₋₂₂₅ kanMX4 CEN</i>	<i>P_{CUP1}-Nub-HA-SCS2₁₋₂₂₅ kanMX4 CEN, Amp^R</i>	This work
<i>Epo1-CCG pRS306</i>	<i>EPO1-Cherry-C_{ub}-R-GFP, Amp^R, URA3</i>	This work
<i>Bem3-GFP pRS304</i>	<i>BEM3-GFP, Amp^R, TRP1</i>	This work
<i>Pea2-GFP pRS304</i>	<i>PEA2-GFP, Amp^R, TRP1</i>	This work
<i>Spa2-GFP pRS304</i>	<i>SPA2-GFP, Amp^R, TRP1</i>	This work
<i>Epo1-GFP pRS304</i>	<i>EPO1-GFP, Amp^R, TRP1</i>	This work
<i>pRH475</i>	<i>HMG1-GFP, Amp^R, URA3</i>	Wilhovskiy et al., 2000
<i>P_{MET17}-GFP-Sec62 pRS313</i>	<i>P_{MET17}-GFP-SEC62, Amp^R, HIS3</i>	This work
<i>P_{MET17}-GFP-Scs2₁₋₂₂₅ pRS313</i>	<i>P_{MET17}-GFP-SCS2₁₋₂₂₅, Amp^R, HIS3</i>	This work
<i>P_{MET17}-Epo1₁₋₇₆₀-GFP pRS304</i>	<i>P_{MET17}-EPO1₁₋₇₆₀-GFP, Amp^R, TRP1</i>	This work
<i>P_{MET17}-Epo1₇₆₁₋₉₄₃-GFP pRS313</i>	<i>P_{MET17}-EPO1₇₆₁₋₉₄₃-GFP, Amp^R, HIS3</i>	This work
<i>Spa2-Cherry pRS305</i>	<i>SPA2-Cherry, Amp^R, LEU2</i>	This work
<i>Spa2-Cherry pRS306</i>	<i>SPA2-Cherry, Amp^R, URA3</i>	This work
<i>Kel1-Cherry pRS305</i>	<i>KEL1-Cherry, Amp^R, LEU2</i>	This work
<i>Shs1-Cherry pRS306</i>	<i>SHS1-Cherry, Amp^R, URA3</i>	Schneider et al., 2013
<i>P_{MET17}-Cherry pRS315</i>	<i>P_{MET17}-Cherry, Amp^R, LEU2</i>	This work
<i>P_{MET17}-Hhf2-Cherry pRS315</i>	<i>P_{MET17}-HHF2-Cherry, Amp^R, LEU2</i>	This work
<i>P_{MET17}-Epo1-Cherry pRS315</i>	<i>P_{MET17}-EPO1-Cherry, Amp^R, LEU2</i>	This work
<i>P_{MET17}-Epo1₁₋₇₆₀-Cherry pRS315</i>	<i>P_{MET17}-EPO1₁₋₇₆₀-Cherry, Amp^R, LEU2</i>	This work
<i>P_{MET17}-Epo1₇₆₁₋₉₄₃-Cherry pRS315</i>	<i>P_{MET17}-EPO1₇₆₁₋₉₄₃-Cherry, Amp^R, LEU2</i>	This work
<i>Kel1-9xMyc pRS304</i>	<i>KEL1-9xMYC, Amp^R, TRP1</i>	This work
<i>pGEX-2T</i>	<i>P_{10c}-GST, Amp^R</i>	GE Healthcare
<i>Pea2 pGEX-2T</i>	<i>P_{10c}-GST-PEA2, Amp^R</i>	This work
<i>Epo1₇₆₁₋₉₄₃ pGEX-2T</i>	<i>P_{10c}-GST-EPO1₇₆₁₋₉₄₃, Amp^R</i>	This work
<i>Epo1₇₆₁₋₈₆₇ pGEX-2T</i>	<i>P_{10c}-GST-EPO1₇₆₁₋₈₆₇, Amp^R</i>	This work
<i>Epo1₈₅₂₋₉₄₃ pGEX-2T</i>	<i>P_{10c}-GST-EPO1₈₅₂₋₉₄₃, Amp^R</i>	This work
<i>Scs2₁₋₂₂₅ pGEX-2T</i>	<i>P_{10c}-GST-SCS2₁₋₂₂₅, Amp^R</i>	This work
<i>Bem3₁₋₁₀₀ pGEX-2T</i>	<i>P_{10c}-GST-BEM3₁₋₁₀₀, Amp^R</i>	This work
<i>pAC</i>	<i>P₁₇-6HIS, kanMX^R</i>	Iffland et al., 2000
<i>Pea2 pAC</i>	<i>P₁₇-6HIS-PEA2, kanMX^R</i>	This work
<i>Epo1₄₇₃₋₅₇₁ pAC</i>	<i>P₁₇-6HIS-EPO1₄₇₃₋₅₇₁, kanMX^R</i>	This work
<i>Epo1₇₆₁₋₈₆₇ pAC</i>	<i>P₁₇-6HIS-EPO1₇₆₁₋₈₆₇, kanMX^R</i>	This work
<i>Epo1₈₅₂₋₉₄₃ pAC</i>	<i>P₁₇-6HIS-EPO1₈₅₂₋₉₄₃, kanMX^R</i>	This work
<i>Scs2₁₋₂₂₅ pAC</i>	<i>P₁₇-6HIS-SCS2₁₋₂₂₅, kanMX^R</i>	This work
<i>pAGT-Xpress</i>	<i>P₁₇-6HIS-AGT, Amp^R</i>	This work
<i>Spa2₁₋₄₈₈ pAGT-Xpress</i>	<i>P₁₇-6HIS-Spa2₁₋₄₈₈-AGT, kanMX^R</i>	This work
<i>Spa2₁₋₅₃₅ pAGT-Xpress</i>	<i>P₁₇-6HIS-Spa2₁₋₅₃₅-AGT, kanMX^R</i>	This work
<i>pMal-c5x</i>	<i>P_{10c}-MBP, Amp^R</i>	New England Biolabs, Inc.
<i>Epo1 pMal-c5x</i>	<i>P_{10c}-MBP-EPO1, Amp^R</i>	This work
<i>Epo1₁₋₇₆₀ pMal-c5x</i>	<i>P_{10c}-MBP-EPO1₁₋₇₆₀, Amp^R</i>	This work

Table S3. List of constructs used and created in this study (Continued)

Name	Description	Origin
pFA6α-natNT2	<i>P_{AgTEF}-natNT2, Amp^R</i>	Janke et al., 2004
pFA6α-hphNT1	<i>P_{AgTEF}-hphNT1, Amp^R</i>	Janke et al., 2004
pFA6α-kanMX6	<i>P_{AgTEF}-kanMX6, Amp^R</i>	Bähler et al., 1998
pFA6α-CmLEU2	<i>P_{CmLEU2}-CmLEU2, Amp^R</i>	Schaub et al., 2006

Table S4. List of yeast strains used and created in this study

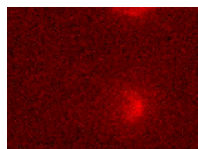
Strain	Name	Relevant genotype	Origin
JD47	JD47	MAT α , <i>his3-Δ200</i> , <i>leu2-3</i> , 112 <i>lys2-801</i> , <i>trp1-Δ63</i> , <i>ura3-52</i>	Dohmen et al., 1995
JD53	JD53	MAT α , <i>his3-Δ200</i> , <i>leu2-3</i> , 112 <i>lys2-801</i> , <i>trp1-Δ63</i> , <i>ura3-52</i>	Dohmen et al., 1995
JD51	JD51	MAT α / α , <i>his3-Δ200</i> / <i>his3-Δ200</i> , <i>leu2-3</i> / <i>leu2-3</i> , 112 <i>lys2-801</i> / <i>112 lys2-801</i> , <i>trp1-Δ63</i> / <i>trp1-Δ63</i> , <i>ura3-52</i> / <i>ura3-52</i>	Dohmen et al., 1995
YAD1222	Δ bni1	JD47, <i>BNI1::hphNT1</i>	This work
YAD1294	Δ bud6	JD47, <i>BUD6::natNT2</i>	This work
YAD249	Δ pea2	JD47, <i>PEA2::hphNT1</i>	This work
YAD248	Δ spa2	JD47, <i>SPA2::hphNT1</i>	This work
YAD250	Δ kel1	JD47, <i>KEL1::hphNT1</i>	This work
YAD251	Δ bem3	JD47, <i>BEM3::hphNT1</i>	This work
YAD1384	Δ ptc1	JD53, <i>PTC1::kanMX6</i>	This work
AHY140	Δ nbp2	JD47, <i>NBP2::natNT2</i>	Hruby et al., 2011
YWY473	Δ sec3	JD53, <i>SEC3::hphNT1</i>	This work
YAD1772	Δ scs22	JD47, <i>SCS22::hphNT1</i>	This work
YAD253	Δ ylr031w	JD47, <i>YLR031W::hphNT1</i>	This work
YAD398a	Δ epo1_hph	JD47, <i>EPO1::hphNT1</i>	This work
YAD400a	Δ epo1_kan	JD53, <i>EPO1::kanMX6</i>	This work
YAD399a	Δ epo1_nat	JD47, <i>EPO1::natNT2</i>	This work
YAD1153	Δ scs2_hph	JD47, <i>SCS2::hphNT1</i>	This work
YAD1385	Δ scs2_kan	JD53, <i>SCS2::kanMX6</i>	This work
YAD1405	Δ epo1 Δ scs2	<i>EPO1::hphNT1 SCS2::kanMX6</i>	This work
YAD1407	Δ epo1 Δ ptc1	<i>EPO1::hphNT1 PTC1::kanMX6</i>	This work
YAD1476	Δ epo1 Δ nbp2	<i>EPO1::kanMX6 NBP2::natNT2</i>	This work
YAD1482	Δ epo1 Δ sec3	<i>EPO1::natNT2 SEC3::hphNT1</i>	This work
YAD1478	Δ scs2 Δ ptc1	<i>SCS2::hphNT1 PTC1::kanMX6</i>	This work
YAD1801	Δ scs22 Δ ptc1	<i>SCS2::hphNT1 PTC1::kanMX6</i>	This work
YAD302.1	Δ epo1 Δ pea2	<i>PEA2::hphNT1 EPO1::kanMX6</i>	This work
YAD301.1	Δ epo1 Δ spa2	<i>SPA2::hphNT1 EPO1::kanMX6</i>	This work
YAD417.1	Δ epo1 Δ bem3	<i>BEM3::hphNT1 EPO1::kanMX6</i>	This work
YAD285	Δ epo1 Δ ylr031w	<i>YLR031W::hphNT1 EPO1::kanMX6</i>	This work
YAD1794	Δ epo1 Δ bud6	<i>EPO1::hphNT1 BUD6::natNT2</i>	This work
YAD1790	Δ epo1 Δ bni1	<i>EPO1::natNT2 BNI1::hphNT1</i>	This work
YAD416.1	Δ pea2 Δ bem3	<i>BEM3::hphNT1 PEA2::kanMX6</i>	This work
YAD213.1	Pea2-CRU	JD47, <i>PEA2::PEA2-Cub-R-URA3 HIS3</i>	This work
YAD347	Pea2-CRU Δ epo1	JD47, <i>PEA2::PEA2-Cub-R-URA3 HIS3 EPO1::CmLEU2</i>	This work
YAD346.1	Epo1-CRU	JD47, <i>EPO1::EPO1-Cub-R-URA3 HIS3</i>	This work
YAD288	P _{MET17} -Epo1-CRU	JD47, <i>P_{MET17}-EPO1-Cub-R-URA3 HIS3 pRS313</i>	This work
YAD967	P _{MET17} -Epo1-CRU Δ pea2	JD47, <i>P_{MET17}-EPO1-Cub-R-URA3 pRS313 PEA2::CmLEU2</i>	This work
YAD363	P _{MET17} -Epo1-CRU Δ spa2	JD47, <i>P_{MET17}-EPO1-Cub-R-URA3 pRS313 SPA2::CmLEU2</i>	This work
YAD341	P _{MET17} -Epo1-CRU Δ kel1	JD47, <i>P_{MET17}-EPO1-Cub-R-URA3 pRS313 KEL1::hphNT1</i>	This work
YAD410	P _{MET17} -Epo1-CRU Δ bem3	JD47, <i>P_{MET17}-EPO1-Cub-R-URA3 pRS313 BEM3::hphNT1</i>	This work
YAD1394	P _{MET17} -Epo1 ₇₆₁₋₉₄₃ -CRU	JD47, <i>P_{MET17}-EPO1₇₆₁₋₉₄₃-Cub-R-URA3 pRS313</i>	This work
YAD233	P _{MET17} -Epo1 ₁₋₇₆₀ -CRU	JD47, <i>P_{MET17}-EPO1₁₋₇₆₀-Cub-R-URA3 pRS313</i>	This work
YAD580	P _{MET17} -Epo1 ₁₋₈₇₆ -CRU	JD47, <i>P_{MET17}-EPO1₁₋₈₇₆-Cub-R-URA3 pRS313</i>	This work
YAD329	P _{MET17} -Kel1-CRU	JD47, <i>P_{MET17}-KEL1-Cub-R-URA3 HIS3 pRS313</i>	This work
YAD1175	Ist2-CRU	JD47, <i>IST2::IST2-Cub-R-URA3 HIS3</i>	This work
YAD1176	Tcb1-CRU	JD47, <i>TCB1::TCB1-Cub-R-URA3 HIS3</i>	This work
YAD1177	Tcb2-CRU	JD47, <i>TCB2::TCB2-Cub-R-URA3 HIS3</i>	This work
YAD1178	Tcb3-CRU	JD47, <i>TCB3::TCB3-Cub-R-URA3 HIS3</i>	This work
YAD305	N _{ub} -Spa2 Δ pea2	JD53, <i>P_{SPA2}::kanMX6 P_{CUP1}N_{ub}-HA PEA2::CmLEU2</i>	This work
YAD307	N _{ub} -Kel1 Δ pea2	JD53, <i>P_{KEL1}::kanMX6 P_{CUP1}N_{ub}-HA PEA2::CmLEU2</i>	This work
YAD311	N _{ub} -Bem3 Δ pea2	JD53, <i>P_{BEM3}::kanMX6 P_{CUP1}N_{ub}-HA PEA2::CmLEU2</i>	This work
YAD306	N _{ub} Δ pea2	JD53, <i>P_{CUP1}N_{ub}-HA kanMX4-CEN PEA2::CmLEU2</i>	This work
YAD310	N _{ub} -Bud6 Δ pea2	JD53, <i>P_{BUD6}::kanMX6 P_{CUP1}N_{ub}-HA PEA2::CmLEU2</i>	This work
YAD1358	N _{ub} -Scs2 Δ pea2	JD53, <i>P_{SCS2}::kanMX6 P_{CUP1}N_{ub}-HA PEA2::CmLEU2</i>	This work
YAD384	N _{ub} -Pea2 Δ spa2	JD53, <i>P_{PEA2}::kanMX6 P_{CUP1}N_{ub}-HA SPA2::natNT2</i>	This work
YAD380	N _{ub} -Kel1 Δ spa2	JD53, <i>P_{KEL1}::kanMX6 P_{CUP1}N_{ub}-HA SPA2::natNT2</i>	This work
YAD365	N _{ub} -Bem3 Δ spa2	JD53, <i>P_{BEM3}::kanMX6 P_{CUP1}N_{ub}-HA SPA2::natNT2</i>	This work

Table S4. List of yeast strains used and created in this study (Continued)

Strain	Name	Relevant genotype	Origin
YAD382	N _{ub} Δspa2	JD53, P _{CUP1} N _{ub} -HA kanMX4-CEN SPA2::natNT2	This work
YAD351	N _{ub} -Pea2 Δkel1	JD53, P _{PEA2} ::kanMX6 P _{CUP1} N _{ub} -HA KEL1::CmLEU2	This work
YAD344	N _{ub} -Spa2 Δkel1	JD53, P _{SPA2} ::kanMX6 P _{CUP1} N _{ub} -HA KEL1::CmLEU2	This work
YAD352	N _{ub} -Bem3 Δkel1	JD53, P _{BEM3} ::kanMX6 P _{CUP1} N _{ub} -HA KEL1::CmLEU2	This work
YAD346	N _{ub} Δkel1	JD53, P _{CUP1} N _{ub} -HA kanMX4-CEN KEL1::CmLEU2	This work
YAD356	N _{ub} -Bud6 Δkel1	JD53, P _{BUD6} ::kanMX6 P _{CUP1} N _{ub} -HA KEL1::CmLEU2	This work
YAD402	N _{ub} -Pea2 Δbem3	JD53, P _{PEA2} ::kanMX6 P _{CUP1} N _{ub} -HA BEM3::natNT2	This work
YAD401	N _{ub} -Spa2 Δbem3	JD53, P _{SPA2} ::kanMX6 P _{CUP1} N _{ub} -HA BEM3::natNT2	This work
YAD403	N _{ub} -Kel1 Δbem3	JD53, P _{KEL1} ::kanMX6 P _{CUP1} N _{ub} -HA BEM3::natNT2	This work
YAD408	N _{ub} Δbem3	JD53, P _{CUP1} N _{ub} -HA kanMX4-CEN BEM3::natNT2	This work
YAD312	N _{ub} -Spa2 Δepo1	JD53, P _{SPA2} ::kanMX6 P _{CUP1} N _{ub} -HA EPO1::natNT2	This work
YAD313	N _{ub} -Kel1 Δepo1	JD53, P _{KEL1} ::kanMX6 P _{CUP1} N _{ub} -HA EPO1::natNT2	This work
YAD316	N _{ub} -Bem3 Δepo1	JD53, P _{BEM3} ::kanMX6 P _{CUP1} N _{ub} -HA EPO1::natNT2	This work
YAD315	N _{ub} Δepo1	JD53, P _{CUP1} N _{ub} -HA kanMX4-CEN EPO1::natNT2	This work
YAD1446	N _{ub} -Scs2 Δepo1	JD53, P _{SCS2} ::kanMX6 P _{CUP1} N _{ub} -HA EPO1::natNT2	This work
YAD1111	pN _{ub} -Scs2 ₁₋₂₂₅	JD53, P _{CUP1} N _{ub} -HA-SCS2 ₁₋₂₂₅ kanMX4-CEN	This work
YAD1814	pN _{ub} -Bem3 ₁₋₆₃₃	JD53, P _{CUP1} N _{ub} -HA-BEM3 ₁₋₆₃₃ kanMX4-CEN	This work
YAD1815	pN _{ub} -Bem3 ₁₋₃₀₀	JD53, P _{CUP1} N _{ub} -HA-BEM3 ₁₋₃₀₀ kanMX4-CEN	This work
YAD1816	pN _{ub} -Bem3 ₁₋₁₀₀	JD53, P _{CUP1} N _{ub} -HA-BEM3 ₁₋₁₀₀ kanMX4-CEN	This work
YAD1817	pN _{ub} -Bem3 ₃₀₀₋₆₃₃	JD53, P _{CUP1} N _{ub} -HA-BEM3 ₃₀₀₋₆₃₃ kanMX4-CEN	This work
YAD974	P _{MET17} -EPO1-CCG	JD47, P _{EPO1} ::P _{MET17} natNT2 EPO1::EPO1-Cherry-C _{ub} -R-GFP URA3	This work
YAD225	Epo1-GFP	JD47, EPO1::EPO1-GFP TRP1	This work
YAD239	Epo1 ₁₋₇₆₀ -GFP	JD47, EPO1::EPO1 ₁₋₇₆₀ -GFP TRP1	This work
YAD1321	P _{MET17} -Epo1 ₇₆₁₋₉₄₃ -GFP	JD47, P _{MET17} -EPO1 ₇₆₁₋₉₄₄ -GFP HIS3 pRS313	This work
YAD1195	Epo1-GFP Δpea2	JD47, EPO1::EPO1-GFP TRP1 PEA2::hphNT1	This work
YAD1223	Epo1-GFP Δbni1	JD47, EPO1::EPO1-GFP TRP1 BNI1::hphNT1	This work
YAD241	Pea2-GFP	JD47, PEA2::PEA2-GFP TRP1	This work
YAD1288	Pea2-GFP Δepo1	JD47, PEA2::PEA2-GFP TRP1 EPO1::hphNT1	This work
YAD1292	Pea2-GFP Δbni1	JD47, PEA2::PEA2-GFP TRP1 BNI1::hphNT1	This work
YAD959	Spa2-GFP	JD47, SPA2::SPA2-GFP TRP1	This work
YAD1293	Spa2-GFP Δbni1	JD47, SPA2::SPA2-GFP TRP1 BNI1::hphNT1	This work
YAD1112	P _{MET17} -GFP-Scs2 ₁₋₂₂₅	JD47, P _{MET17} -GFP-SCS2 ₁₋₂₂₅ HIS3 pRS313	This work
YAD1137	P _{MET17} -GFP-Scs2 ₁₋₂₂₅ Δepo1	JD47, P _{MET17} -GFP-SCS2 ₁₋₂₂₅ HIS3 pRS313 EPO1::hphNT1	This work
YAD1135	P _{MET17} -GFP-Scs2 ₁₋₂₂₅ Δpea2	JD47, P _{MET17} -GFP-SCS2 ₁₋₂₂₅ HIS3 pRS313 PEA2::hphNT1	This work
YAD1136	P _{MET17} -GFP-Scs2 ₁₋₂₂₅ Δspa2	JD47, P _{MET17} -GFP-SCS2 ₁₋₂₂₅ HIS3 pRS313 SPA2::hphNT1	This work
YAD1134	P _{MET17} -GFP-Scs2 ₁₋₂₂₅ Δkel1	JD47, P _{MET17} -GFP-SCS2 ₁₋₂₂₅ HIS3 pRS313 KEL1::hphNT1	This work
YAD1133	P _{MET17} -GFP-Scs2 ₁₋₂₂₅ Δbem3	JD47, P _{MET17} -GFP-SCS2 ₁₋₂₂₅ HIS3 pRS313 BEM3::hphNT1	This work
YAD519	P _{MET17} -GFP-SEC62	JD47, P _{MET17} -GFP-SEC62 HIS3 pRS313	This work
YAD520	P _{MET17} -GFP-SEC62 Δepo1	JD47, P _{MET17} -GFP-SEC62 HIS3 pRS313 EPO1::hphNT1	This work
YAD1297	Spa2-GFP Kel1-Cherry	JD47, SPA2::SPA2-GFP TRP1 KEL1::KEL1-Cherry LEU2	This work
YAD1309	Spa2-GFP Kel1-Cherry	JD53, SPA2::SPA2-GFP TRP1 KEL1::KEL1-Cherry LEU2	This work
YAD816	Epo1-GFP Kel1-Cherry	JD47, EPO1::EPO1-GFP TRP1 KEL1::KEL1-Cherry LEU2	This work
YAD1311	Epo1-GFP Kel1-Cherry	JD53, EPO1::EPO1-GFP TRP1 KEL1::KEL1-Cherry LEU2	This work
YAD230	Epo1-GFP Spa2-Cherry	JD47, EPO1::EPO1-GFP TRP1 SPA2::SPA2-Cherry URA3	This work
YAD1313	Epo1-GFP Spa2-Cherry	JD53, EPO1::EPO1-GFP TRP1 SPA2::SPA2-Cherry LEU2	This work
YAD794	Bem3-GFP Spa2-Cherry306 47	JD47, BEM3::BEM3-GFP TRP1 SPA2::SPA2-Cherry URA3	This work
YAD1312	Bem3-GFP304 Spa2-Cherry	JD53, BEM3::BEM3-GFP TRP1 SPA2::SPA2-Cherry LEU2	This work
YAD1536	Spa2-GFP Shs1-Cherry	JD47, SPA2::SPA2-GFP TRP1 SHS1::SHS1-Cherry URA3	This work
YAD1766	Epo1-GFP Shs1-Cherry	JD47, EPO1::EPO1-GFP TRP1 SHS1::SHS1-Cherry URA3	This work
YAD966	P _{MET17} -Hhf2-Cherry Spa2-GFP	JD47, P _{MET17} -HHF2-Cherry LEU2 pRS315 SPA2::SPA2-GFP TRP1	This work
YAD1199	P _{MET17} -GFP-SEC62 Spa2-Cherry	JD47, P _{MET17} -GFP-SEC62 HIS3 pRS313 SPA2::SPA2-Cherry LEU2	This work
YAD1200	P _{MET17} -GFP-SEC62 Spa2-Cherry	JD47, P _{MET17} -GFP-SEC62 HIS3 pRS313 SPA2::SPA2-Cherry LEU2	This work
	Δepo1	EPO1::hphNT1	
YAD1457	P _{MET17} -Cherry P _{MET17} -GFP-SEC62 Δepo1	JD47, P _{MET17} -Cherry LEU2 pRS315 P _{MET17} -GFP-SEC62 HIS3 pRS313 EPO1::hphNT1	This work
YAD1323	P _{MET17} -Epo1-Cherry P _{MET17} -GFP-SEC62 Δepo1	JD47, P _{MET17} -EPO1-Cherry LEU2 pRS315 P _{MET17} -GFP-SEC62 HIS3 pRS313 EPO1::hphNT1	This work
YAD1324	P _{MET17} -Epo1 ₁₋₇₆₀ -Cherry P _{MET17} -GFP-SEC62 Δepo1	JD47, P _{MET17} -EPO1 ₁₋₇₆₀ -Cherry LEU2 pRS315 P _{MET17} -GFP-SEC62 HIS3 pRS313 EPO1::hphNT1	This work

Table S4. List of yeast strains used and created in this study (Continued)

Strain	Name	Relevant genotype	Origin
YAD1325	P_{MET17} -Epo1 ₇₆₁₋₉₄₃ -Cherry P_{MET17} -GFP-Sec62 Δ epo1	JD47, P_{MET17} -EPO1 ₇₆₁₋₉₄₃ -Cherry LEU2 pRS315 P_{MET17} -GFP-SEC62 HIS3 pRS313 EPO1::hphNT1	This work
YAD1282	Hmg1-GFP	JD47, URA3::HMG1-GFP URA3	This work
YAD1283	Hmg1-GFP Δ epo1	JD47, URA3::HMG1-GFP URA3 EPO1::hphNT1	This work
YAD1449	Hmg1-GFP Δ scs2	JD47, URA3::HMG1-GFP URA3 SCS2::hphNT1	This work
YAD1448	Hmg1-GFP Δ ptc1	JD53, URA3::HMG1-GFP URA3 PTC1::kanMX6	This work
YAD1547	Hmg1-GFP Δ bem3	JD47, URA3::HMG1-GFP URA3 BEM3::hphNT1	This work
YAD1545	Hmg1-GFP Δ pea2	JD47, URA3::HMG1-GFP URA3 PEA2::hphNT1	This work
YAD1546	Hmg1-GFP Δ spa2	JD47, URA3::HMG1-GFP URA3 SPA2::hphNT1	This work
YAD1805	Hmg1-GFP Δ scs22	JD47, URA3::HMG1-GFP URA3 SCS22::hphNT1	This work
YAD1770	Hmg1-GFP Δ ylr031w	JD47, URA3::HMG1-GFP URA3 YLR031W::hphNT1	This work
YAD1798	Hmg1-GFP Δ bud6	JD47, URA3::HMG1-GFP URA3 BUD6::natNT2	This work
YAD1797	Hmg1-GFP Δ bni1	JD47, URA3::HMG1-GFP URA3 BNI1::natNT2	This work
YAD1451	Hmg1-GFP Δ epo1 Δ scs2	URA3::HMG1-GFP URA3 EPO1::kanMX6 SCS2::hphNT1	This work
YAD1453	Hmg1-GFP Δ epo1 Δ ptc1	URA3::HMG1-GFP URA3 PTC1::kanMX6 EPO1::hphNT1	This work
YAD1548	Hmg1-GFP Δ epo1 Δ pea2	URA3::HMG1-GFP URA3 EPO1::kanMX6 PEA2::hphNT1	This work
YAD1549	Hmg1-GFP Δ epo1 Δ spa2	URA3::HMG1-GFP URA3 EPO1::kanMX6 SPA2::hphNT1	This work
YAD1551	Hmg1-GFP Δ epo1 Δ bem3	URA3::HMG1-GFP URA3 EPO1::kanMX6 BEM3::hphNT1	This work
YAD1771	Hmg1-GFP Δ epo1 Δ ylr031w	URA3::HMG1-GFP URA3 EPO1::kanMX6 YLR031W::hphNT1	This work
YAD1800	Hmg1-GFP Δ epo1 Δ bud6	URA3::HMG1-GFP URA3 EPO1::hphNT1 BUD6::natNT2	This work
YAD1799	Hmg1-GFP Δ epo1 Δ bni1	URA3::HMG1-GFP URA3 EPO1::natNT2 BNI1::hphNT1	This work
YAD1550	Hmg1-GFP Δ pea2 Δ bem3	URA3::HMG1-GFP URA3 PEA2::kanMX6 BEM3::hphNT1	This work
YAD512	Kel1-MYC	JD47, KEL1::KEL1-9xMYC TRP1	This work



Video 1. **Time-lapse analysis of yeast cells coexpressing Spa2-Cherry with Epo1-GFP.** Cells were incubated at 30°C on SD complete. Frames were taken every 3 min for 120 min using a DeltaVision life cell imaging unit (Applied Precision). Selected stills are shown in Fig. 2 B.



Video 2. **Time-lapse analysis of yeast cells coexpressing Shs1-Cherry with Spa2-GFP.** Cells were incubated at 30°C on SD complete. Frames were taken every minute for 85 min using a DeltaVision life cell imaging unit. Selected stills are shown in Fig. 2 C.



Video 3. **Time-lapse analysis of yeast cells coexpressing Shs1-Cherry with Epo1-GFP.** Cells were incubated at 30°C on SD complete. Frames were taken every minute for 65 min using a DeltaVision life cell imaging unit. Selected stills are shown in Fig. 2 C.

References

- Bähler, J., J.Q. Wu, M.S. Longtine, N.G. Shah, A. McKenzie III, A.B. Steever, A. Wach, P. Philippsen, and J.R. Pringle. 1998. Heterologous modules for efficient and versatile PCR-based gene targeting in *Schizosaccharomyces pombe*. *Yeast*. 14:943–951. [http://dx.doi.org/10.1002/\(SICI\)1097-0061\(199807\)14:10<943::AID-YEA292>3.0.CO;2-Y](http://dx.doi.org/10.1002/(SICI)1097-0061(199807)14:10<943::AID-YEA292>3.0.CO;2-Y)
- Dohmen, R.J., R. Stappen, J.P. McGrath, H. Forrová, J. Kolarov, A. Goffeau, and A. Varshavsky. 1995. An essential yeast gene encoding a homolog of ubiquitin-activating enzyme. *J. Biol. Chem.* 270:18099–18109. <http://dx.doi.org/10.1074/jbc.270.30.18099>
- Dünkler, A., J. Müller, and N. Johnsson. 2012. Detecting protein-protein interactions with the Split-Ubiquitin sensor. *Methods Mol. Biol.* 786:115–130. http://dx.doi.org/10.1007/978-1-61779-292-2_7
- Hruby, A., M. Zapatka, S. Heucke, L. Rieger, Y. Wu, U. Nussbaumer, S. Timmermann, A. Dünkler, and N. Johnsson. 2011. A constraint network of interactions: protein-protein interaction analysis of the yeast type II phosphatase Ptc1p and its adaptor protein Nbp2p. *J. Cell Sci.* 124:35–46. <http://dx.doi.org/10.1242/jcs.077065>
- Iffland, A., P. Tafelmeyer, C. Saudan, and K. Johnsson. 2000. Directed molecular evolution of cytochrome c peroxidase. *Biochemistry.* 39:10790–10798. <http://dx.doi.org/10.1021/bi001121e>
- Janke, C., M.M. Magiera, N. Rathfelder, C. Taxis, S. Reber, H. Maekawa, A. Moreno-Borchart, G. Doenges, E. Schwob, E. Schiebel, and M. Knop. 2004. A versatile toolbox for PCR-based tagging of yeast genes: new fluorescent proteins, more markers and promoter substitution cassettes. *Yeast*. 21:947–962. <http://dx.doi.org/10.1002/yea.1142>
- Schaub, Y., A. Dünkler, A. Walther, and J. Wendland. 2006. New pFA-cassettes for PCR-based gene manipulation in *Candida albicans*. *J. Basic Microbiol.* 46:416–429. <http://dx.doi.org/10.1002/jobm.200510133>
- Schneider, C., J. Grois, C. Renz, T. Gronemeyer, and N. Johnsson. 2013. Septin rings act as a template for myosin higher-order structures and inhibit redundant polarity establishment. *J. Cell Sci.* 126:3390–3400. <http://dx.doi.org/10.1242/jcs.125302>
- Wilhovsky, S., R. Gardner, and R. Hampton. 2000. HRD gene dependence of endoplasmic reticulum-associated degradation. *Mol. Biol. Cell.* 11:1697–1708. <http://dx.doi.org/10.1091/mbc.11.5.1697>