

The *rv1184c* locus encodes Chp2, an acyltransferase in *Mycobacterium tuberculosis* polyacyltrehalose lipid biosynthesis

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SUPPLEMENTAL MATERIAL

Supplemental Table S1. Bacterial strains and plasmids used in this study

Supplemental Table S2. Primers used in this study

Supplemental Table S1. Bacterial strains and plasmids used in this study

Strain	Name	Genotype	Source
<i>M. tuberculosis</i> Erdman			
<i>M. tuberculosis</i> Erdman		$\Delta chp2$	This work
<i>M. tuberculosis</i> Erdman		$\Delta chp2::chp2$	This work
<i>M. tuberculosis</i> Erdman		$\Delta papA3$	Ref [1]
<i>M. tuberculosis</i> Erdman		$\Delta papA3::papA3$	Ref [1]
<i>M. tuberculosis</i> Erdman	jcm110	$\Delta mmpL10$	J. Cox
<i>M. smegmatis</i> mc ² 155			
Plasmids			
Reference name	Name	Description	Source
	pMV261	Kn ^r , pAL5000 origin, ColE1 origin, multiple cloning site, P _{<i>hsp60</i>}	Ref [2]
	pMV306	Kn ^r , <i>int</i> , attA5, ColE1 origin, multiple cloning site	Ref [3]
	pJSC407	oriE, cosA; HygR flanked by loxP sites	Ref [4]
$\Delta chp2$ KO construct		pJSC407; HindIII-804bp (first 156bp of <i>chp1</i>)-Hyg ^r -813bp (last 157bp of <i>chp1</i>)-KpnI	This work
	pRibo-BsaHind	pMV261 derivative; P _{<i>hsp60</i>} -theophylline riboswitch; BsaI for cloning	Ref [5]
Chp2-3xFLAG	pRibo Chp2-3xFLAG	pRibo-BsaHind derivative; contains <i>chp2</i> fused to 3xFLAG	This work
AP cytosolic control	pMB111	pMV261 derivative; contains <i>phoA</i>	M. Braunstein
AP secreted control	pMB124	pMV261 derivative; contains <i>fbpB(aa2-40)-phoA</i>	M. Braunstein
β Gal cytosolic control	pHsp- <i>lacZ</i>	pMWS114 derivative; P _{<i>hsp60</i>} - <i>lacZ</i>	Ref [5]
β Gal secreted control	sec- <i>lacZ</i>	pMB124 derivative; P _{<i>hsp60</i>} - <i>fbpB(aa2-40)-lacZ</i>	Ref [5]
full-length Chp2- β Gal		pRibo-BsaHind derivative; contains full-length <i>chp2</i> fused to <i>lacZ</i>	This work
N-term Chp2- β Gal		pRibo-BsaHind derivative; contains <i>chp2</i> (aa1-39) fused to <i>lacZ</i>	This work
cat Chp2- β Gal		pRibo-BsaHind derivative; contains <i>chp2</i> (aa26-359) fused to <i>lacZ</i>	This work

Reference name	Name	Description	Source
full-length Chp2-AP		pRibo-BsaHind derivative; contains full-length <i>chp2</i> fused to <i>phoA</i>	This work
N-term Chp2-AP		pRibo-BsaHind derivative; contains <i>chp2</i> (aa1-39) fused to <i>phoA</i>	This work
cat Chp2-AP		pRibo-BsaHind derivative; contains <i>chp2</i> (26-359) fused to <i>phoA</i>	This work
<i>chp2</i>		pMV306 derivative; contains 1kb upstream of <i>fadD21</i> ; <i>chp2</i>	This work
<i>mmpL10</i>		pMV306 derivative; contains 1kb upstream of <i>pks3/4</i> ; <i>mmpL10</i>	This work
$P_{native2kb}$ - <i>mmpL10</i>		pMV306 derivative; contains 2kb upstream of <i>pks3/4</i>	This work
P_{hsp60} - <i>mmpL10</i>		pMV306 derivative; P_{hsp60} - <i>mmpL10</i>	This work
P_{gs} - <i>mmpL10</i>		pMV306 derivative; P_{gs} - <i>mmpL10</i>	This work
	2BT	LIC vector with N-terminal 6xHis, TEV protease site, SspI site	S. Gradia
Chp2-cat		2BT derivative; 6XHis-TEV- <i>chp2</i> (aa26-359)	This work

References

1. Hatzios, S. K., et al. (2009) PapA3 is an acyltransferase required for polyacyltrehalose biosynthesis in *Mycobacterium tuberculosis*, *J Biol Chem* 284, 12745-12751.
2. Stover, C. K., et al. (1991) New use of BCG for recombinant vaccines, *Nature* 351, 456-460.
3. Stover, C. K., et al. (1993) Protective immunity elicited by recombinant bacille Calmette-Guerin (BCG) expressing outer surface protein A (OspA) lipoprotein: a candidate Lyme disease vaccine, *J Exp Med* 178, 197-209.
4. Stanley, S. A., et al. (2003) Acute infection and macrophage subversion by *Mycobacterium tuberculosis* require a specialized secretion system, *PNAS* 100, 13001-13006.
5. Seeliger, J. C., et al. (2012) A Riboswitch-Based Inducible Gene Expression System for Mycobacteria, *PLoS ONE* 7, e29266.

Supplemental Table S2. Primers used in this study

Name (purpose)	Sequence (cloning strategy)
pJSC407 Δchp2 (knockout)	(by assembly PCR across HygR cassette)
<u>HindIII</u> 5' flank	ggatccacgaagcttCGAGACACGGCACACCTTTG
5' flank- <u>loxP</u>	ACGCTTCAACCGCCAAAGTCggtggcccgtataacttcg
5' flank- <u>loxP</u> revcomp	cgaagttataccggggccaccGACTTTGGCGGTTGAAGCGT
<u>loxP</u> -3' flank	GAACTGTACCGGATTTTCGGACGCTGCAACCACAGATCGACG
<u>loxP</u> -3' flank revcomp	CGTCGATCTGTGGTTGCAGCGTCCGAAATCCGGTACAGTTC
3' flank <u>KpnI</u> revcomp	gactagagggtagaccGTTACCCGGTCATGCTGAGG
Δchp2 PCR (knockout confirmation)	
chp2 KO confirm REV	CGTCATGTTGTCTTGGTGGCTC
fadD21 FOR	GGCCACATAAAGAGTCGCTTCC
hygR confirm REV	CAGGCTCGCGTAGGAATCATC
ΔmmpL10 PCR (knockout confirm.)	
papA3 FOR	GCTGAAGTCTGTGTTCCAACGAGTC
mmpL10 internal REV	GTCTTCGTGCGCGCTGACATC
chp2 REV*	GTCGATCGTACGCTAGTTAACCGTGGATCCGGTGCGAG
hygR confirm REV	<i>same as above</i>
pMV306 chp2 (complement)	(in two cloning steps via XbaI-HindIII and HindIII-ClaI)
5' fadD21 promoter <u>XbaI</u>	GCTCTAGAgggtgctgcggtgctg
fadD21 promoter 3' <u>HindIII</u>	CTTCATAAGCTTCATtggtgctacattaccgtttc
chp2 5' <u>HindIII</u> for pMV306	ccaATGAAGCTTAAGCGAGTGATTGCGGGAG
chp2 3' <u>ClaI</u> for pMV306	GTCGACATCGATTTAGCCGCCGAAGGCGG
2BT chp2-cat (<i>E. coli</i> expression)	(by InFusion into SspI-cleaved 2BT)
5' chp2-cat FOR	GTACTIONCAATCCAATGCAAGCGAACCCGCGTACC
3' chp2-cat REV	TTATCCACTTCCAATtcattaGCCGCCGAAGGCGG
chp2 <u>S141A</u> QC FOR	GTCGGCCTAgCCCAGGGTTCCTCGTGCTC
chp2 S141A QC REV	GGAACCCCTGGGcTAGGCCGACGGCCGCGG
chp2 SA (Ser141Ala mutagenesis)	(by site-directed mutagenesis)
S141A FOR	GTCGGCCTAgCCCAGGGTTCCTCGTGCTC
S141A REV	GGAACCCCTGGGcTAGGCCGACGGCCGCGG

* Originally designed as a cloning primer. Only underlined portion is complementary to *chp2*.