

## **Fine-Scale Mapping of the 5q11.2**

### **Breast Cancer Locus Reveals at Least Three**

### **Independent Risk Variants Regulating *MAP3K1***

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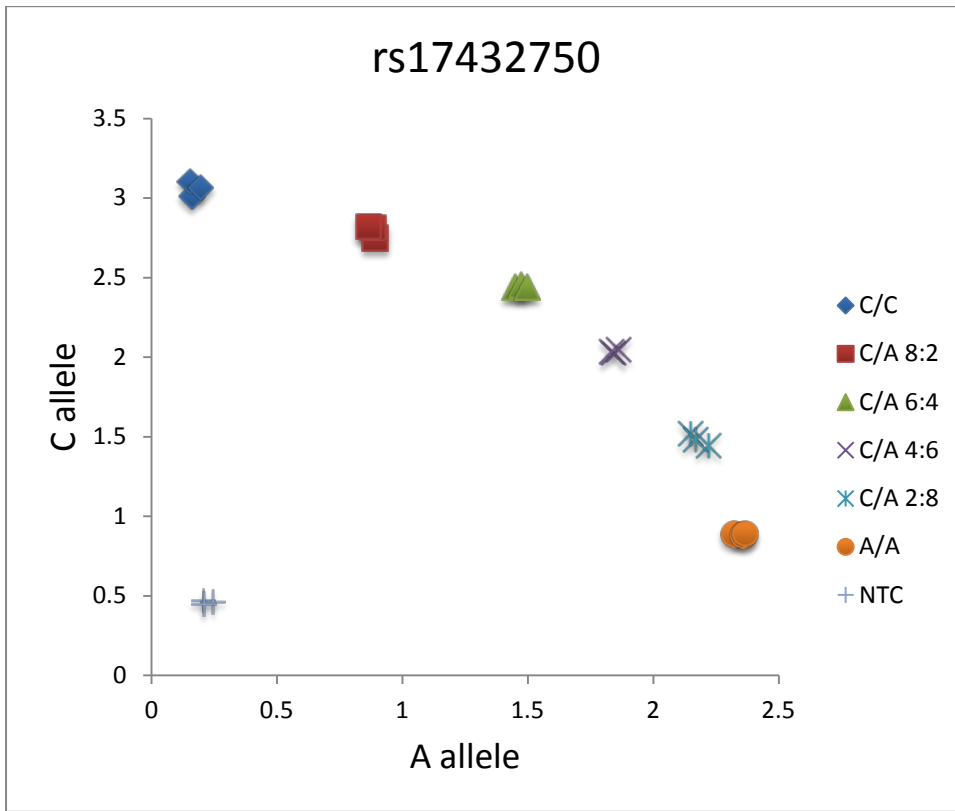
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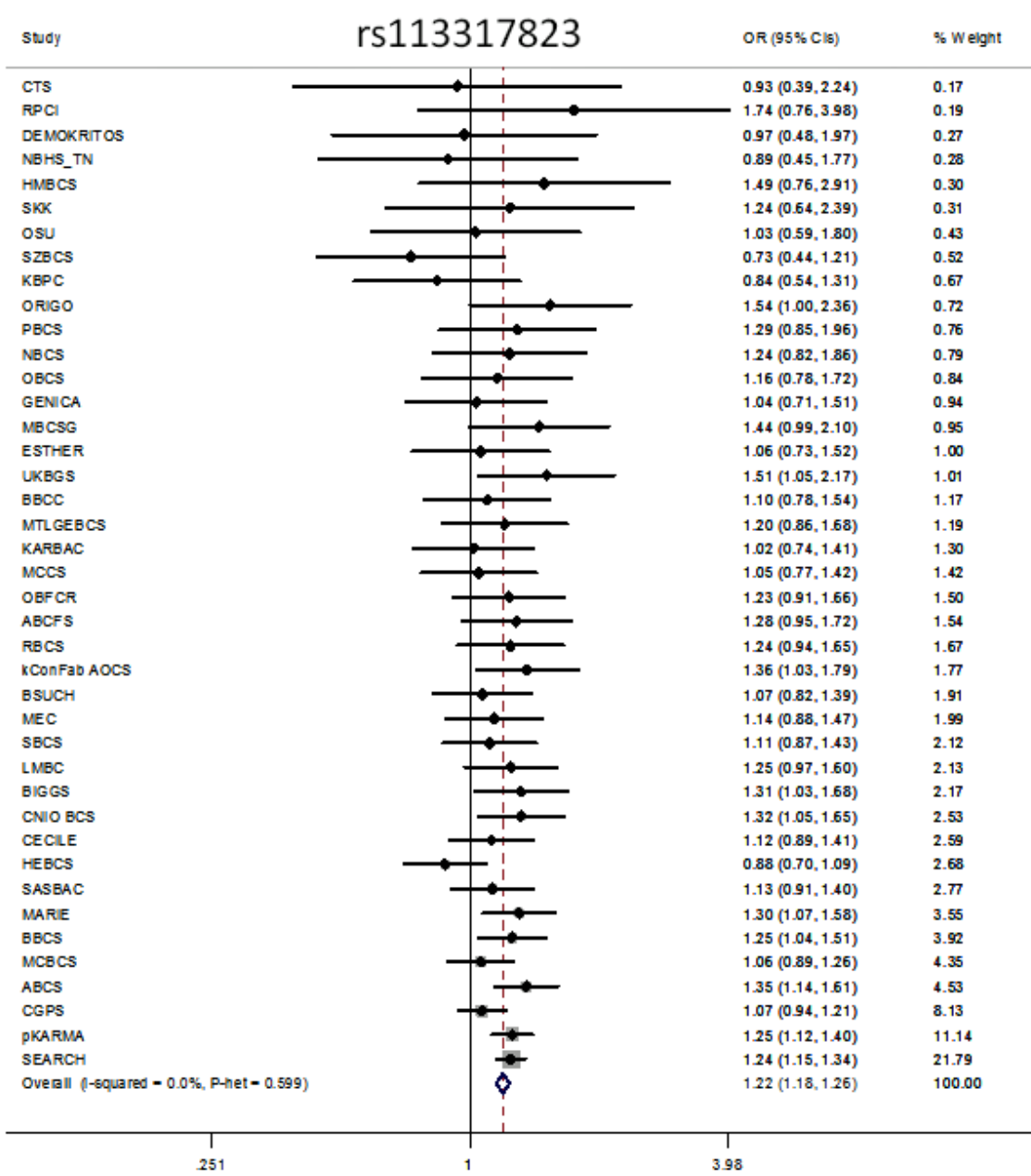
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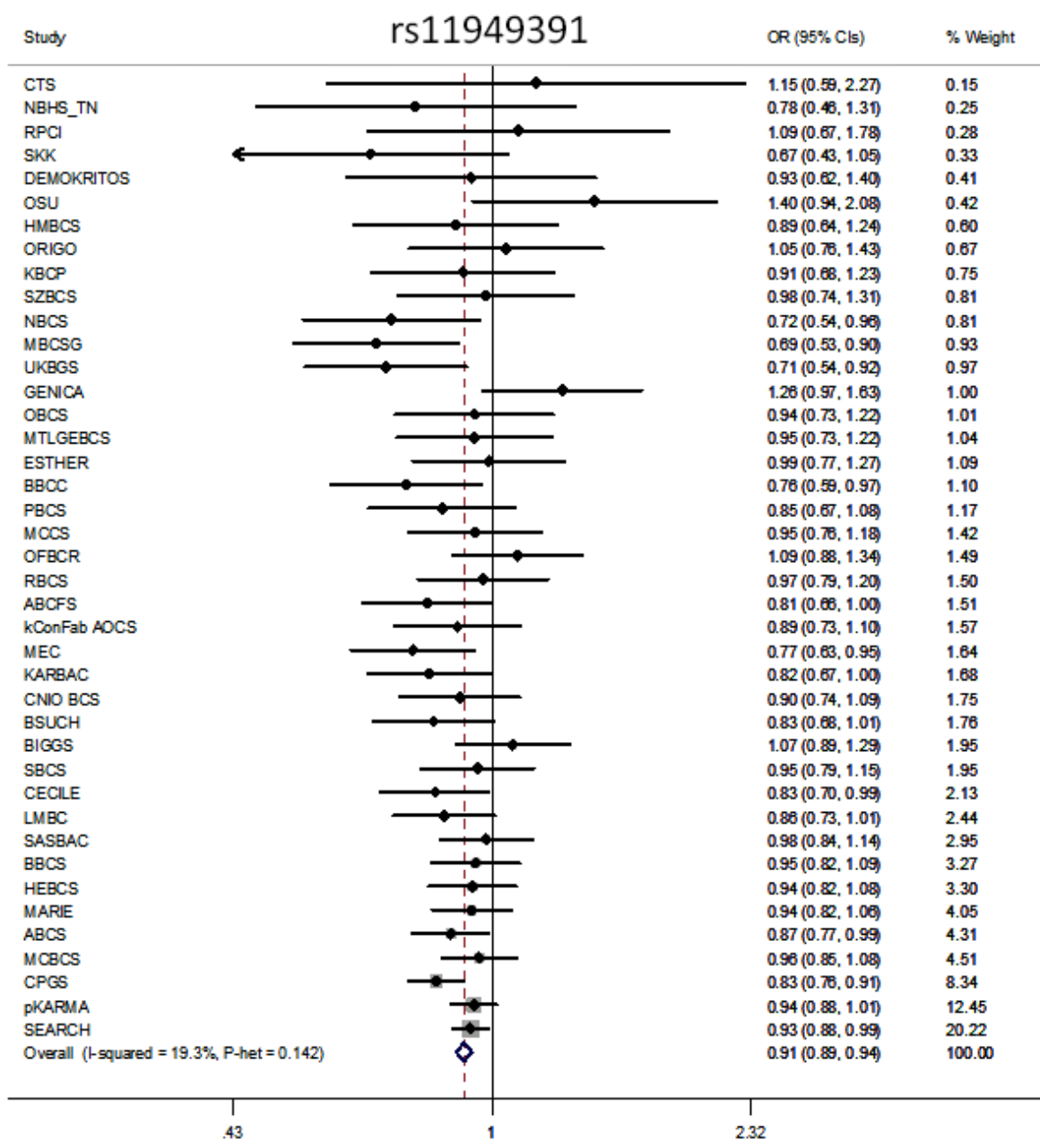


**Figure S1. Allelic Discrimination plot of known ratios of genomic DNA from breast cell lines carrying the C/C (ZR751) or A/A (MDA-MB-468) at rs17432750.**

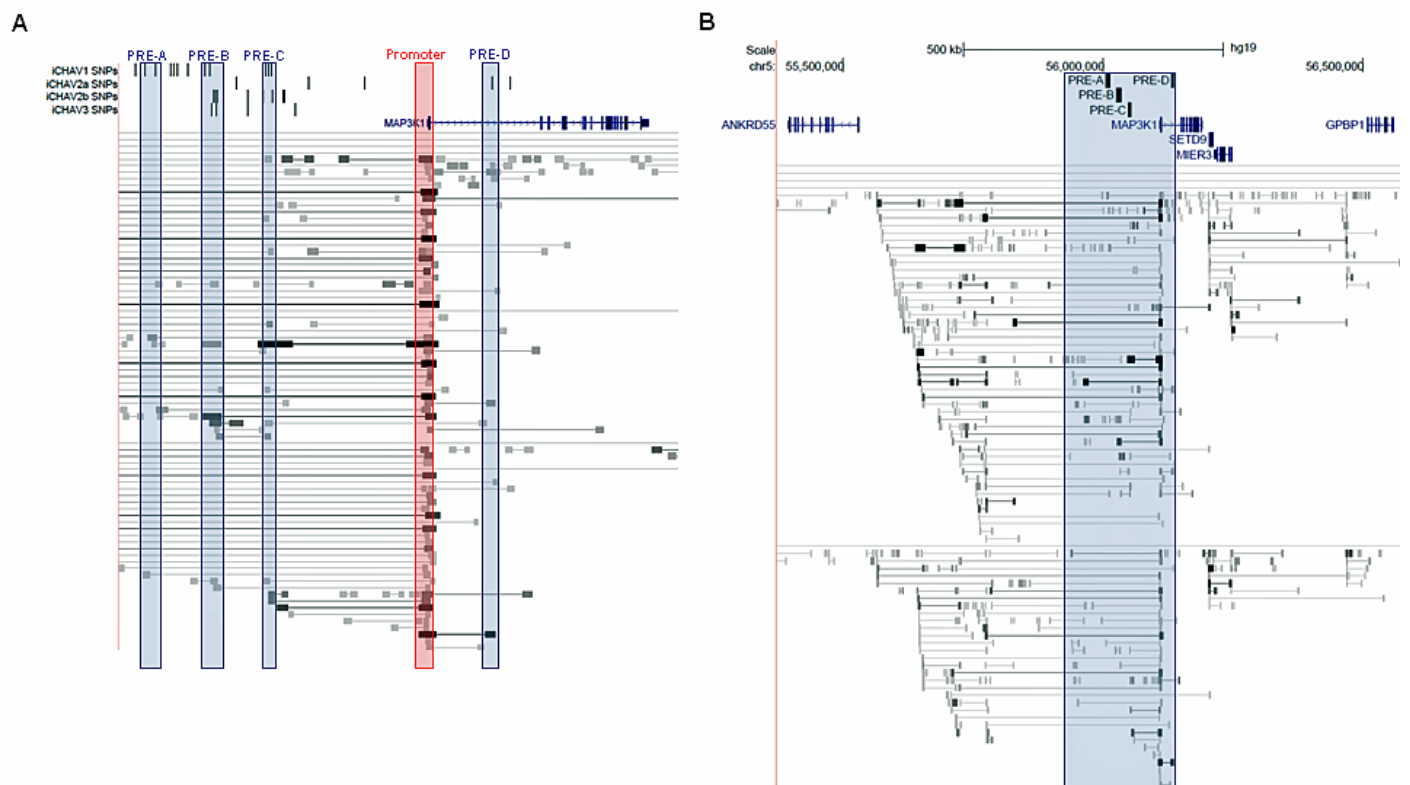




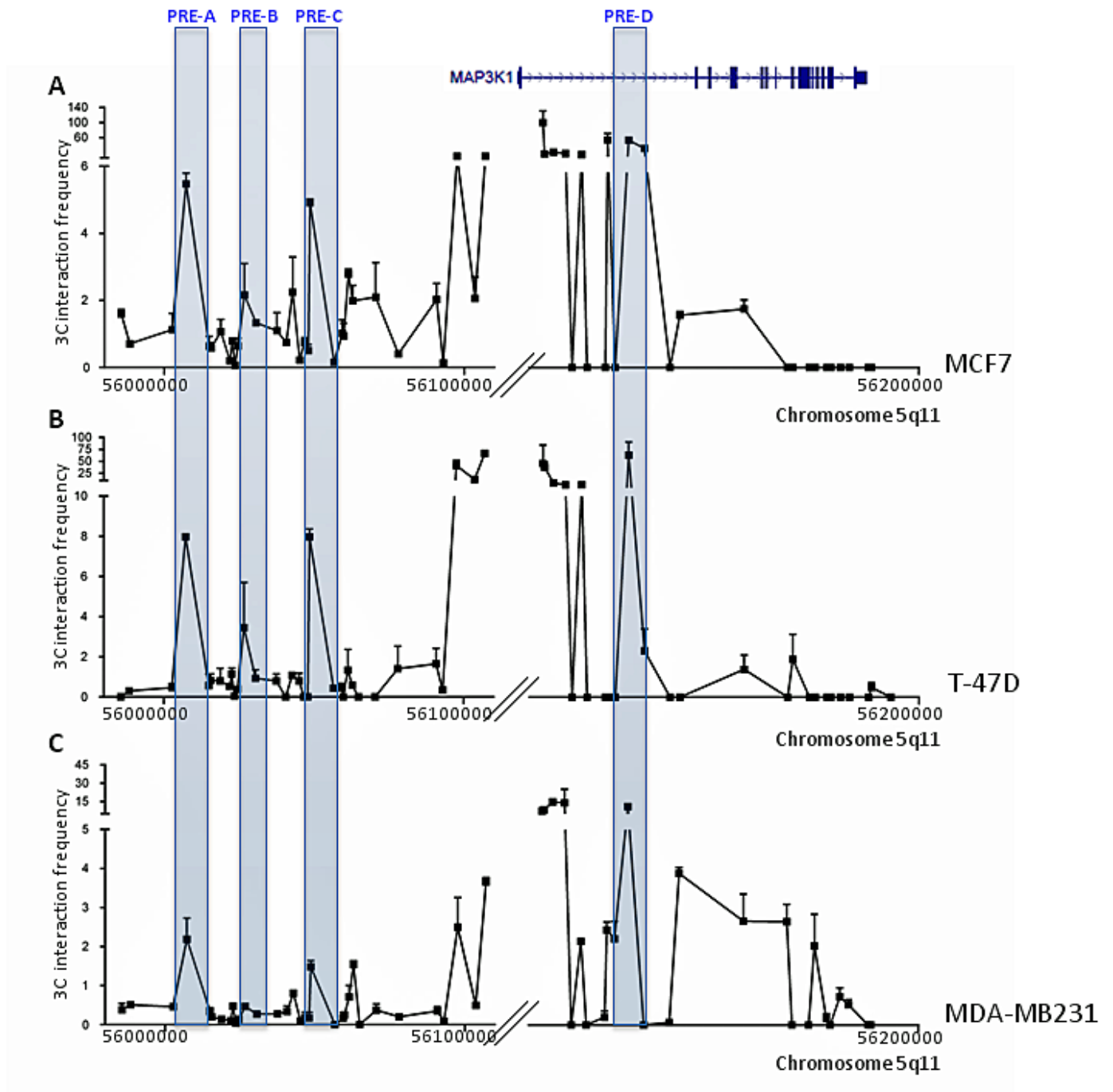




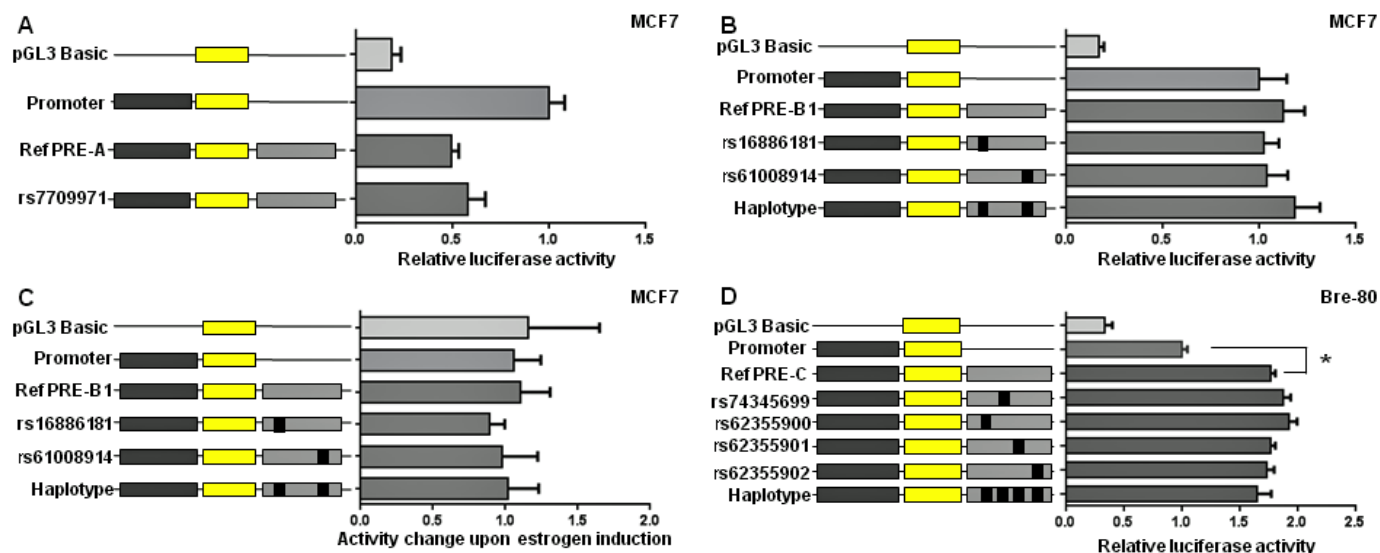
**Figure S2. Forest plots for the three independently-associated SNPs marking iCHAVs 1-3.**



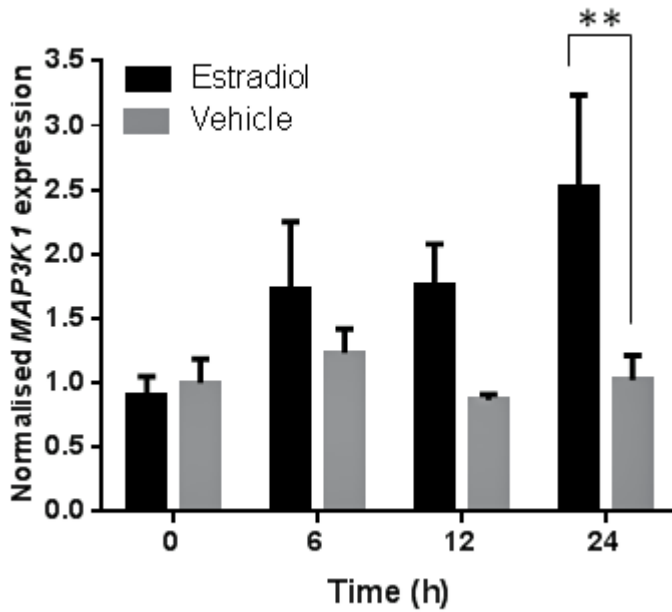
**Figure S3. RNA polymerase II ChIA-PET at the 5q11.2 breast cancer risk locus in MCF7 cells shows chromatin interactions between discrete genomic regions and the *MAP3K1* promoter.** ENCODE data (GEO sample accession GSM970209) was accessed using the UCSC genome browser. iCHAV variants and PREs are shown. Panel (A) shows the zoomed in region containing the PREs and *MAP3K1*. Panel (B) shows the 5q11.2 risk locus and flanking genes.



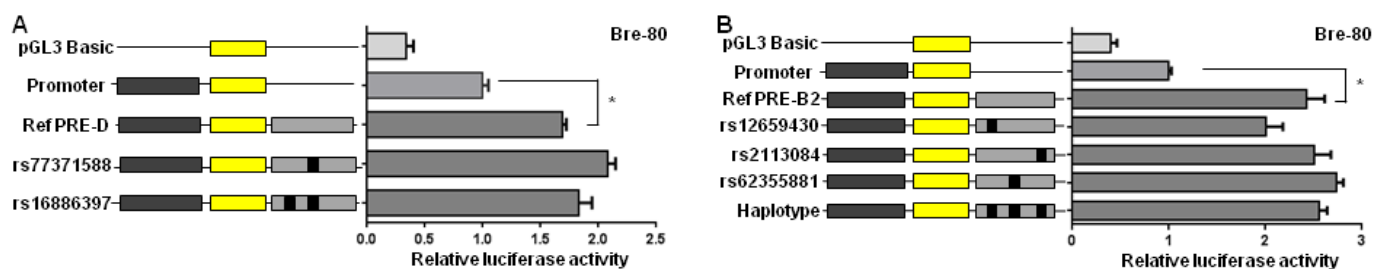
**Figure S4. Chromatin interactions with the *MAP3K1* promoter in breast cancer cell lines at the 5q11.2 risk locus.** 3C libraries were generated with EcoRI. Chromatin interaction frequencies were plotted at the chromosomal position of the corresponding EcoRI fragments for MCF7 (A), T-47D (B) and MDA-MB231 (C) libraries, respectively. PREs are highlighted at their chromosomal locations. A representative graph of at least two biological replicates is shown and error bars represent SD.



**Figure S5. Luciferase reporter assays of PRE constructs containing iCHAV1 SNPs.** MCF7 cells were transiently transfected with PRE-A (A) or PRE-B1 (B) and Bre-80 cells with PRE-C (D) constructs under basal conditions and assayed for luciferase activity after 24 h. Panel (C) shows results from luciferase assays after estrogen induction of MCF7 cells transfected with PRE-B1 constructs. For each reporter construct, the luciferase activity of estrogen treated cells was normalized to the activity of the corresponding vehicle treated cells. Error bars denote SEM from three experiments performed in triplicate. *P*-values were determined by repeated-measures ANOVA followed by Dunnett's multiple comparisons test (\**P*<0.05).

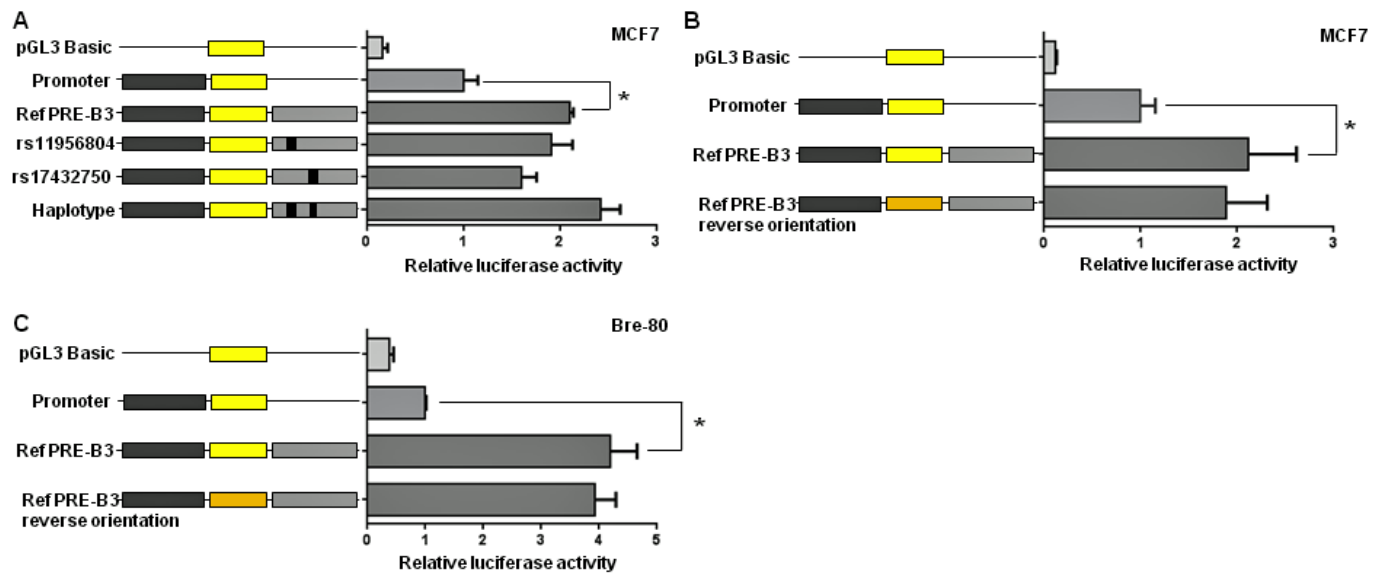


**Figure S6. *MAP3K1* expression is induced in MCF7 cells after estrogen treatment.** MCF7 cells were treated with either 100 nM estradiol or vehicle for 24 h. *MAP3K1* expression was measured at baseline and after 6, 12 and 24 h by qPCR and normalized using *GUS* expression as an internal control. Error bars denote SEM from three experiments performed in triplicate. Differences in expression between vehicle and estradiol treated cells were tested using a two-way repeated measures ANOVA followed by Sidak's multiple comparisons test (\*\* $P < 0.01$ ).

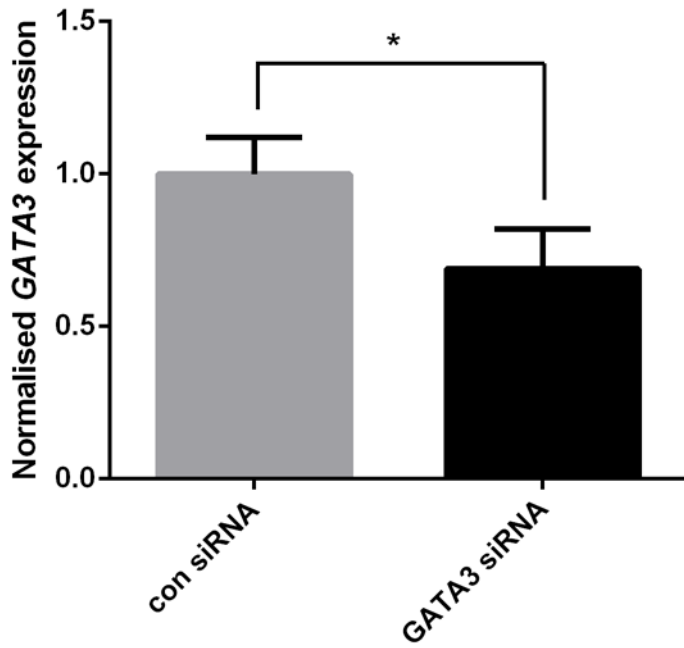


**Figure S7. Luciferase reporter assays of PRE-D constructs containing iCHAV2a SNPs and PRE-B2 constructs containing iCHAV2b variants in Bre-80 cells.** Bre-80 cells were transfected with PRE-D and PRE-B2 constructs. Cells were assayed for luciferase activity after 24 h. Error bars denote SEM from three experiments performed with triplicates. Statistical significance was determined by repeated-measures ANOVA followed by Dunnett's multiple comparisons test ( $*P<0.05$ ).





**Figure S8. Luciferase reporter assays of PRE-B3 constructs containing iCHAV3 SNPs in MCF7 and Bre-80 cells.** MCF7 cells were transiently transfected the PRE-B3 reference and variant luciferase constructs (A) or the PRE-B3 reference construct in the reverse orientation (B). Bre-80 cells were also transiently transfected with the reference PRE-B3 construct in the reverse orientation (C). Cells were assayed for luciferase activity after 24 h. Error bars denote SEM from three experiments performed with triplicates. *P*-values were determined by ANOVA followed by Dunnett's multiple comparisons test (\**P*<0.05).



**Figure S9. TaqMan qPCR assay confirming knockdown of *GATA3* in Bre80 cells.** *GATA3* expression from cells transfected with non-targeting siRNA and *GATA3* is denoted by con siRNA and *GATA3* siRNA, respectively. Error bars denote SEM from three experiments performed with triplicates. Statistical significance was determined by a paired t-test ( $*P < 0.05$ ).

**Table S1. 3C qPCR primers**

<b>3C primers</b>	<b>EcoRI site (GRCh37 chr 5 coordinates)</b>	<b>Sequence</b>
Fragment 1	55,988,536	aacctttcctgttctgcttaaggtgggtg
Fragment 2	56,001,687	aagacagccaaagactctccaattccttctgc
Fragment 3	56,002,653	cactggctcagtcacctcattgattcagtc
Fragment 4	56,015,010	caaagaaacgagctctaagaggtgggcagc
Fragment 5	56,015,005	gaggcagattccctggaaccttctttcc
Fragment 6	56,015,605	cctgagtctcattccctcttctgcaagagacc
Fragment 7	56,021,916	aagagtctcgtctgtcagacaaaatgtcacgg
Fragment 8	56,022,688	aagaatcactgggtgttattgctggaaagg
Fragment 9	56,023,585	ctcatggcctatgtacctcaaaggctcc
Fragment 10	56,024,547	catgtaactgctgggttccattccagttcagg
Fragment 11	56,026,749	ggttctatgtgaaggtgccctccaaaacc
Fragment 12	56,030,623	aatccagcacctgctatgaacacctcatcc
Fragment 13	56,037,547	cctaagtcttctccagagcattttgaccttgc
Fragment 14	56,040,674	atcaggaggactacaatctaggggtatgctgagg
Fragment 15	56,042,804	gcaggactctttgtcagcatctttctctttcc
Fragment 16	56,045,182	tgggactgtgggagacacacagttttgc
Fragment 17	56,046,719	agcaatcagtaatgggaaaagatgccatgc
Fragment 18	56,047,998	ctcttctgcctgttctctgacctgtgtcc
Fragment 19	56,048,601	tctaggtgtgtctgcaagggcatttctgg
Fragment 20	56,056,381	gccagtgtgtttgattccaaacctctgagc
Fragment 21	56,056,646	tggaacatgcttctactaaagccttctaggaagc
Fragment 22	56,059,823	gcagtgaatgaggtgacacagatctcc
Fragment 23	56,061,410	gcgtttgaactatccagagttaacacgcatcc
Fragment 24	56,062,862	aaggtgctgtcttatgaaagaggaaaccgagg
Fragment 25	56,064,941	ggaggaaagagtaggagacggctgttcagc
Fragment 26	56,070,521	agagtaagtaatgacctgcccacggcagg
Fragment 27	56,078,074	gacgaggtagggataattagtggaaccttaggc
Fragment 28	56,090,803	tcatagtgggtatggaatggtattgcattgtgc
Fragment 29	56,093,049	gtgaacacaagccgagatttccagaaggtagc
Fragment 30	56,097,598	ctatcctaccaatttgccagctaagtgattggg
Fragment 31	56,103,646	cagtttccctgatcccctattcccgc
Fragment 32	56,107,073	ttacaggaggatcttgggaaaatgtatgaacctc
Fragment 33	56,108,495	tctagccccagcactcctgcaagtattcc
Fragment 34	56,109,036	gcatgctgaaatcattgcaaggtttacg
Bait (promoter fragment)	56,112,866	gcttaactgaaaaggggttctccctctgc
Fragment 37	56,113,523	gcactgtacctggcactactgagaccaaagg
Fragment 38	56,115,580	ttgtactccagtcattcttgggttgagg
Fragment 39	56,118,301	cggccagaccattgcttgatgttaatagc
Fragment 40	56,119,851	aatgagagaaggcctaggaccaccttg
Fragment 41	56,121,912	cgtcatttcagcatcgtgctcagtttagtcc
Fragment 42	56,123,317	gatttacatcagagctgggacaagacagtctgg
Fragment 43	56,127,536	ttcgagttaatcagaccaactatcaggcaggc
Fragment 44	56,128,078	gaccgtccccattactgtcctaacactttgg
Fragment 45	56,129,808	aactaggatgattgaggctggatcccagacc
Fragment 46	56,132,919	cataagcagttcacataaaagacttatgggccagc

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Fragment 47	56,136,559	gaagcaaccaagaggaagaaaggaactagc
Fragment 48	56,142,493	cttcacctcctgcttttctcctcacctgc
Fragment 49	56,144,772	gtaaattggagttattgtgcatgacatgtgg
Fragment 50	56,159,585	ggacatttgagctgctcctgtttggtgg
Fragment 51	56,169,636	gaagctgacaaaaacgctcctagggtgagg
Fragment 52	56,170,833	gatgggtcatgaaagggcaggtgagg
Fragment 53	56,174,572	tgtaggctcctgggtattcattttgttctgtgg
Fragment 54	56,175,998	cctctgcaagtcataatgacttagttgcacagg
Fragment 55	56,178,759	gtctcaggatgccctccccatagtcc
Fragment 56	56,179,669	ggctcaagatgtgggaactggaactttaatgg
Fragment 57	56,181,895	cggtggctcatttgctgagtaaatatggagc
Fragment 58	56,183,388	agattttggagctgcagccaggttg
Fragment 59	56,188,350	ccatacatcacagctccttcatactgttgacc
Fragment 60	56,189,045	tactaagaatcagaattgtccaacacggaagg
Fragment 61	56,193,516	gtctatgaaaaaattcatccactggatgggagg
Fragment 62	56,211,583	tttctggattctgacatgaagactgtcacc

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**Table S2. Reporter gene construct cloning and overlapping mutagenesis primers**

Primer	Sequence
MAP3K1 promoter forward	<u>acgcgtccac</u> ctctctgcagtaacatag
MAP3K1 promoter reverse	<u>aagcttgcttc</u> ctctcggcaatctcg
rs74762363 forward	ccagttcaaccactg <u>a</u> caaacagtcaaaaatcag
rs74762363 reverse	ctgattttgactgtttgt <u>t</u> cagtgggtgaaactgg
PRE-A forward	<u>accggtgctca</u> actagccaaagcactcc
PRE-A reverse	<u>cctgcaggggg</u> caagtctaaagcagtgtgg
rs7709971 forward	cagttcagccactgtgg <u>a</u> aagcagtgagatctggag
rs7709971 reverse	ctccagatctcactgctt <u>t</u> ccacagtggctgaactg
PRE-B3 forward	<u>accggtgga</u> agatgggacaagccttgatgc
PRE-B3 reverse	<u>cctgcagggga</u> agaacaacctgtctcaatgatgg
rs17432750 forward	caacctggattctttcact <u>a</u> atcacacaagtcagg
rs17432750 reverse	cctgactgtgtgatt <u>a</u> gtgaaagaatccagggtg
rs11956804 forward	gctgagcagttatctttgtcatatt <u>a</u> gtaggatgaatg
rs11956804 reverse	cattcatcctact <u>t</u> aaatatgacaaagataaactgctcagc
PRE2c forward	<u>accggtaaa</u> agatagctttcaaagg
PRE2c reverse	<u>gcgtcgacc</u> atagttacttcaaagg
rs12659430 forward	<u>catgcatttg</u> atgtgtcctataaaaag
rs12659430 reverse	<u>ctttatagg</u> acacat <u>c</u> caaatgcatg
rs62355881 forward	<u>aaagtca</u> cggtgcttctggtagc
rs62355881 reverse	<u>gctaccaga</u> agcatcc <u>g</u> tgacttt
rs2113084 forward	<u>taaattt</u> gtggcat <u>g</u> caaatattaaac
rs2113084 forward	<u>gtttta</u> atatttgc <u>a</u> tgccacaaatta
PRE-C forward	<u>accggtcca</u> agtttcatgcatggctctgtgg
PRE-C reverse	<u>cctgcaggg</u> tggcctttccagtacagtgg
rs74345699 forward	ggctcacgcctgtaat <u>t</u> cagcactttgg
rs74345699 reverse	ccaaagtgtg <u>a</u> gattacaggcgtgagcc
rs62355900 forward	tgggggagcatc <u>c</u> gaggtggatgaag
rs62355900 reverse	cttcatccacctc <u>g</u> gatgctcccca
rs62355901 forward	ggagatcaagaccat <u>c</u> ccggctagcacg
rs62355901 reverse	cgtgctagccgg <u>g</u> atggtcttgatctcc
rs62355902 forward	tgggcaacagagcgagact <u>t</u> ccatctcaaaaaaca
rs62355902 reverse	tgtttttgagatgg <u>a</u> gtctcgctctgttgcca
PRE-D forward	<u>accggtggc</u> cattatagcagtgtctttgc
PRE-D reverse	<u>cctgcaggg</u> gctgatgcctagtagtcaattaagc
rs77371588 forward	ctgggcagtga <u>g</u> cccgtcttcagtg
rs77371588 reverse	cactggaagacggg <u>c</u> tactgcccag

Underlined sequences correspond to restriction enzyme recognition sites and bases highlighted in red denote SNP loci.

**Table S3. Primer pairs used in CHIP-RT-PCR and sequence confirmation**

<b>Primer</b>	<b>Sequence</b>	<b>Ref or comment</b>
rs17432750-for1	GGCCATCTGTTTTACCAACC	RT-PCR
rs17432750-rev1	ATTTGCACATGCCTTTCTCC	RT-PCR
rs17432750-for2	CAATGCAAATCTTCCTTGCTT	RT-PCR & sequencing
rs17432750-rev2	TGGGAAGGAGTCGTTGAGTT	RT-PCR & sequencing
ER- $\alpha$ -ENH-for	TGTAGGCTAGTTTTGTTTAACGATTTTT	Cancer Research 2007; 67:6477-6483
ER- $\alpha$ -ENH-rev	GGTGATGGGAGAATTGCTTAGAA	as above
<i>CCND1</i> -for	TGCCACACACCAGTGACTTT	Genes Dev. 2006; 20:2513- 2526.
<i>CCND1</i> -rev	ACAGCCAGAAGCTCCAAAAA	as above

**Table S6. Associations of the iCHAV representative SNPs with breast cancer risk in Asian and African American studies**

SNP	Position	Asian Studies						African American Studies					
		MAF	Imp $r^2$	P-trend	OR	LCI	UCI	MAF	Imp $r^2$	P-trend	OR	LCI	UCI
iCHAV1 rs62355902	56053723	0.38	0.94	3.30E-02	1.06	1.00	1.12	0.09	0.98	2.30E-01	1.14	0.92	1.36
iCHAV2a rs113317823	56087883	0.13	0.88	1.40E-05	1.19	1.11	1.27	0.06	0.78	7.80E-01	1.04	0.77	1.31
iCHAV2b rs62355899	56050465	0.26	1.00	7.46E-01	0.99	0.93	1.05	0.04	1.00	9.60E-01	0.99	0.72	1.36
iCHAV2c rs7721581	56087883	0.56	1.00	3.80E-01	0.98	0.93	1.03	0.06	0.78	7.78E-01	1.04	0.78	1.38
iCHAV3 rs11949391	56045081	0.05	1.00	2.60E-02	0.87	0.75	0.99	0.10	1.00	1.20E-01	0.84	0.62	1.06