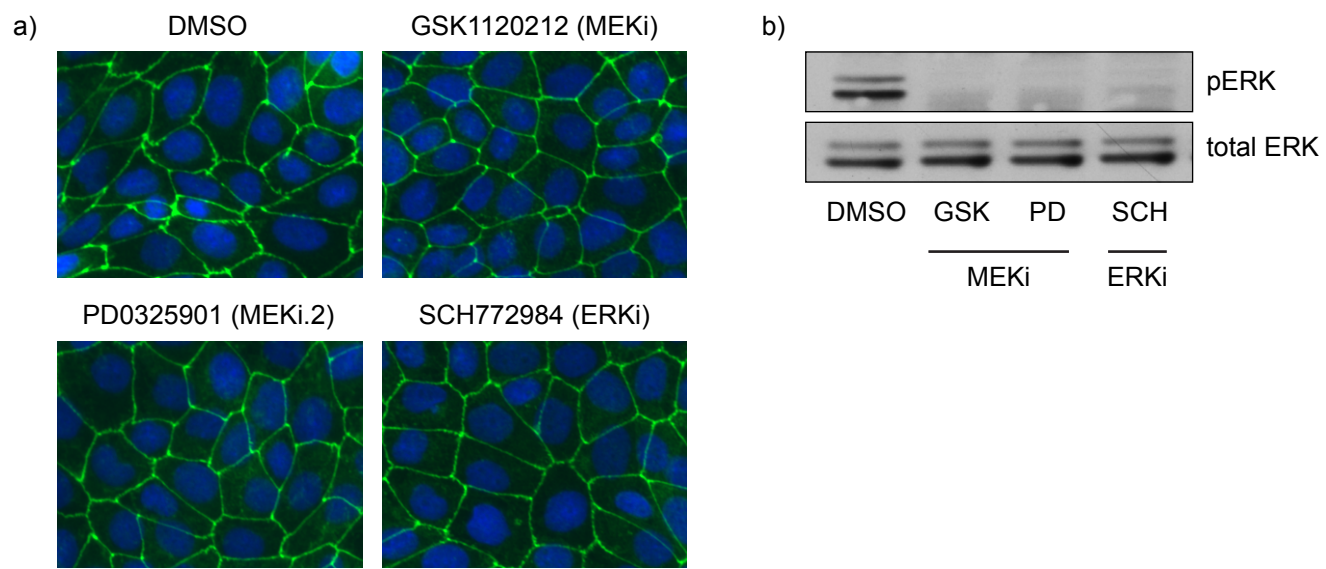
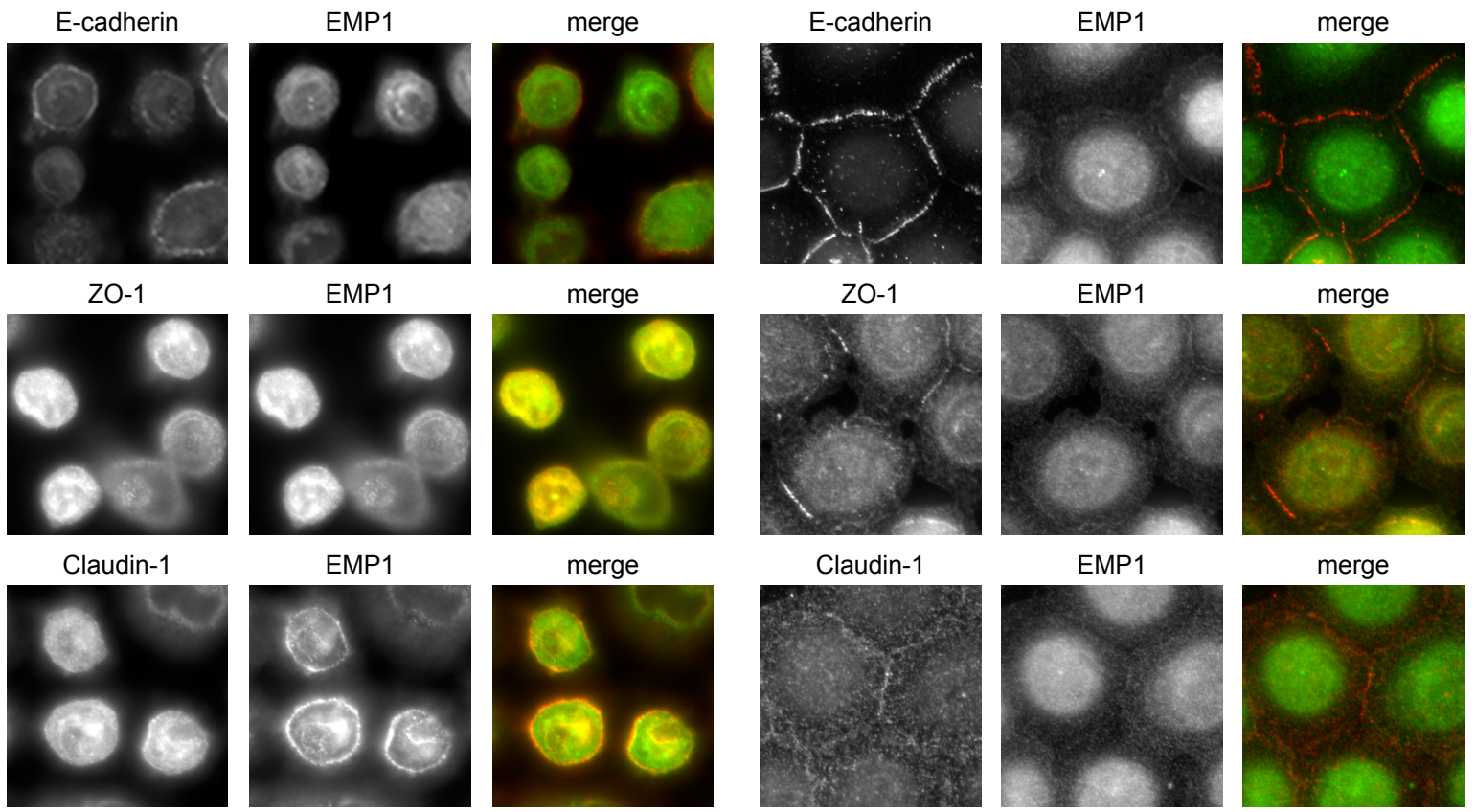


Supplementary Figure 1 SOS1, Ras, MEK and ERK are required for bronchial junction formation. 16HBE cells were stably infected with: (a) pSUPER control or shSOS1.1, (b) pQCXIP control or dominant negative myc-H Ras N17, or (c) Treated with DMSO or GSK1120212 (500nM) for 4 days. Cells were fixed and stained for occludin (tight junctions), E-cadherin (adherens junctions) and DNA (nuclei). Scale bars represent 20μm. d) 16HBE cells were fixed and stained for SOS1 and DNA. In a subset of cells, a junctional signal was detected as shown.



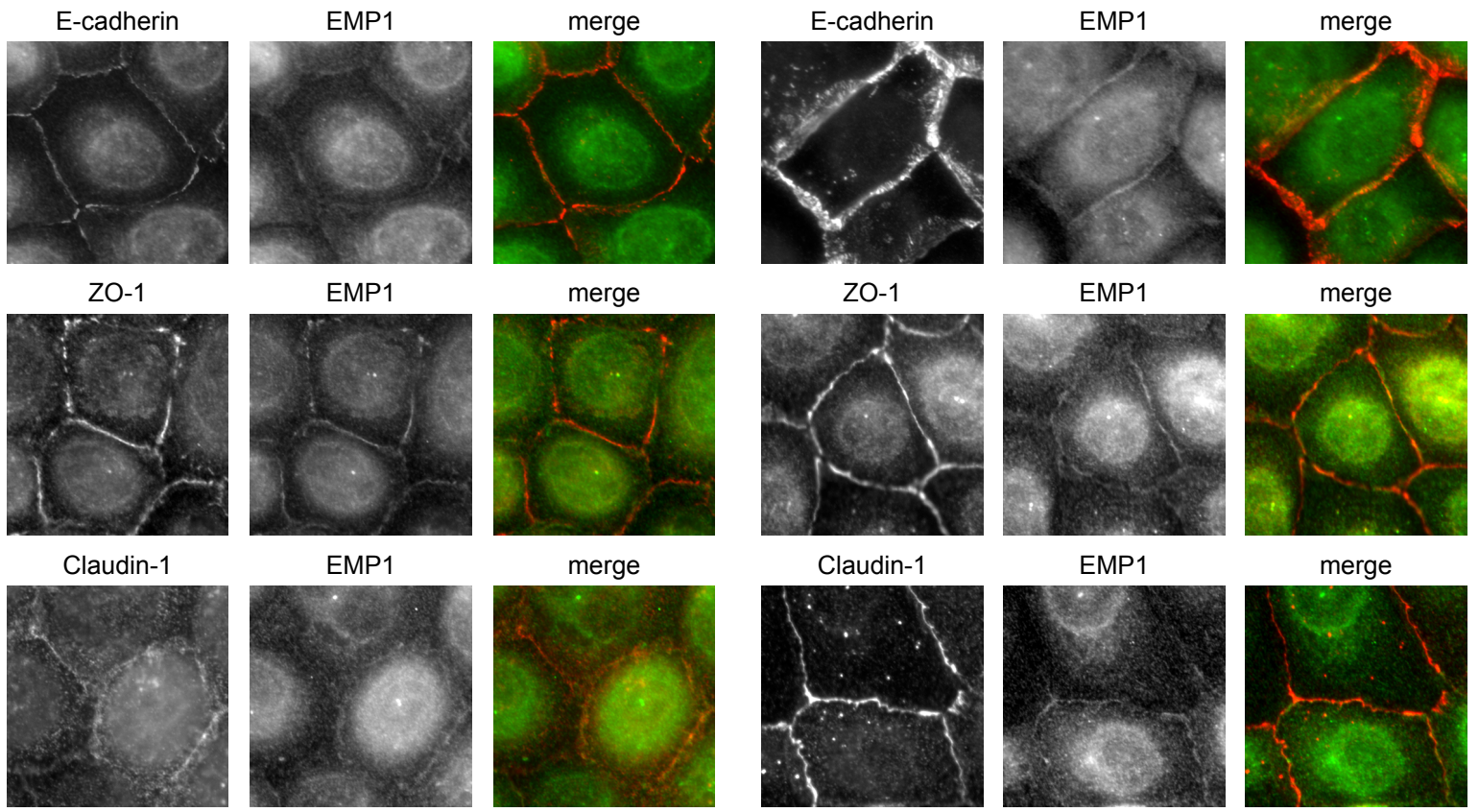
Supplementary Figure 2 MEK and ERK are not required for junctional maintenance in bronchial epithelia. 16HBE cells were seeded on glass coverslips and incubated for 4 days to allow apical junctions to form and mature. The cells were then treated with DMSO, GSK1120212 (500nM), PD0325901 (500nM), or SCH772984 (1 μ M), and incubated for a further 4 days. a) Cells were fixed and stained for ZO-1 (tight junctions) and DNA (nuclei). No obvious difference is detected upon MEK/ERK inhibition. b) Cell lysates were analysed by western blotting for p-ERK and total ERK to confirm efficient pathway inhibition under these conditions.

5209 DHC2R4	0.0375594	0.155409	1.24937	1.24937 Control up vs	0.00629906	1.68388	1.68388 Control up vs	0.208589	1.21427	1.21427 Control up vs	4.9599	0.867314	0.503239	1
5540 DPK2L2	0.00980928	0.0598099	1.18723	1.18723 Control up vs	0.00015714	1.60055	1.60055 Control up vs	0.176588	1.110103	1.110103 Control up vs	16.5664	0.768251	0.124174	1
5709 EECM1	0.0311738	0.210445	1.19662	1.19662 Control up vs	0.00615862	1.6259	1.6259 Control up vs	0.407704	1.12209	1.12209 Control up vs	4.96124	0.807669	0.434122	1
5930 EMP3	0.00527459	0.0799013	1.3524	1.3524 Control up vs	0.00144279	1.78164	1.78164 Control up vs	0.655857	1.05786	1.05786 Control up vs	9.42923	1.30485	0.369024	1
5943 ENCL	0.00992632	0.0499746	1.39552	1.39552 Control up vs	0.0061562	1.65944	1.65944 Control up vs	0.0062945	0.888872	0.888872 Control up vs	7.88669	0.79505	0.102152	1
5996 EPHA2	1.69E-07	9.08E-06	1.43133	1.43133 Control up vs	2.65E-08	2.14801	2.14801 Control up vs	0.00074921	1.21039	1.21039 Control up vs	161.844	1.98485	0.0327038	1
6202 ETV5	6.36E-09	8.33E-09	1.58197	1.58197 Control up vs	1.10E-09	1.80829	1.80829 Control up vs	0.99E-08	1.5083	1.5083 Control up vs	37.1	1.22269	0.00878604	1
6246 FZRL1	0.00021392	0.00028264	1.55677	1.55677 Control up vs	4.45E-05	1.97295	1.97295 Control up vs	0.00402851	1.42423	1.42423 Control up vs	21.9489	1.49127	0.181181	1
6320 FAM107B	0.0059147	0.20391	1.38174	1.38174 Control up vs	0.0012832	1.79955	1.79955 Control up vs	0.433554	1.1052	1.1052 Control up vs	9.07829	1.25143	0.367597	1
6373 FAM129B	5.56E-06	6.92E-06	1.56438	1.56438 Control up vs	1.05E-06	1.77486	1.77486 Control up vs	0.0003855	1.28978	1.28978 Control up vs	65.879	1.16964	0.0473603	1
6919 FX1	0.0044169	0.0264734	1.29303	1.29303 Control up vs	0.00071289	1.65544	1.65544 Control up vs	0.00181165	1.46815	1.46815 Control up vs	9.47487	0.837361	0.23859	1
7439 FOXO1	5.75E-06	5.05E-06	1.57724	1.57724 Control up vs	1.07E-06	1.7222	1.7222 Control up vs	3.29E-05	1.41066	1.41066 Control up vs	65.2778	1.04599	0.0427299	1
7873 G8RA	0.00015238	0.255257	1.11772	1.11772 Control up vs	0.00012561	1.86982	1.86982 Control up vs	0.157524	0.867955	0.867955 Control up vs	27.1168	2.09465	0.205988	1
8087 GPI	0.00021334	0.0164118	1.39772	1.39772 Control up vs	0.0004702	1.61502	1.61502 Control up vs	0.00181165	1.46815	1.46815 Control up vs	12.2717	1.04599	0.176215	1
12732 IERS	0.00052863	0.0101194	1.36772	1.36772 Control up vs	7.33E-05	2.00577	2.00577 Control up vs	0.0350215	1.26754	1.26754 Control up vs	19.0775	1.56374	0.218581	1
12453 IGFBP4	0.002303	0.206653	1.13418	1.13418 Control up vs	0.00046346	1.68319	1.68319 Control up vs	0.00912491	1.36766	1.36766 Control up vs	12.2888	0.966255	0.299677	1
12455 IGFBP6	4.25E-05	0.38415	0.897781	0.897781 Control up vs	0.00204256	1.69093	1.69093 Control up vs	1.05E-05	0.320426	0.320426 Control up vs	70.6147	9.0726	0.342614	1
12408 IGF1L1	0.00018087	0.0521974	1.3983	1.3983 Control up vs	0.00140123	2.01874	2.01874 Control up vs	0.00612025	0.58093	0.58093 Control up vs	25.8553	1.72138	0.540768	1
12486 IGF2E	6.98E-05	0.00104981	1.36356	1.36356 Control up vs	1.53E-05	1.77278	1.77278 Control up vs	0.161293	1.10041	1.10041 Control up vs	33.6285	1.2104	0.0599821	1
12555 IL17RA	3.18E-05	0.00115757	1.50476	1.50476 Control up vs	4.07E-06	2.49724	2.49724 Control up vs	0.00386961	1.39535	1.39535 Control up vs	41.5489	0.269656	0.171983	1
12580 L18	5.87E-05	0.014088	1.43209	1.43209 Control up vs	9.26E-06	3.11214	3.11214 Control up vs	0.0232994	1.37892	1.37892 Control up vs	35.2132	4.35158	0.325942	1
12739 ISG15L1	0.00070631	0.00059017	1.46461	1.46461 Control up vs	0.0014239	1.60258	1.60258 Control up vs	0.00227229	1.35818	1.35818 Control up vs	17.2565	0.784802	0.121276	1
13309 KLIF4	6.98E-05	0.0891431	1.19184	1.19184 Control up vs	0.00055105	1.65229	1.65229 Control up vs	0.00260073	0.676683	0.676683 Control up vs	33.9593	2.59061	0.205632	1
13460 KRT6A	0.00699632	0.788113	0.892028	0.892028 Control up vs	0.0206236	3.26257	3.26257 Control up vs	0.0491127	0.415113	0.415113 Control up vs	8.58144	13.5838	4.22114	1
13473 KRT8	0.00518239	0.0103945	1.39239	1.39239 Control up vs	0.00073279	1.68918	1.68918 Control up vs	0.0233089	1.31987	1.31987 Control up vs	9.48361	0.878057	0.246898	1
13675 LCN2	0.0077825	0.82999	1.09228	1.09228 Control up vs	0.00567197	4.40035	4.40035 Control up vs	0.411078	0.709502	0.709502 Control up vs	8.27848	1.2431	3.91154	1
15193 LOC10013224	0.00013875	0.0248614	1.22783	1.22783 Control up vs	1.98E-05	1.94385	1.94385 Control up vs	0.00496169	1.33998	1.33998 Control up vs	27.8296	1.44639	0.138614	1
16246 LOC149501	0.124706	0.234823	1.2913	1.2913 Control up vs	0.0305576	1.69028	1.69028 Control up vs	0.0706909	1.51766	1.51766 Control up vs	2.93731	0.974947	1.00098	1
19501 LOC455553	0.152919	0.777403	1.06654	1.06654 Control up vs	0.0428594	1.7396	1.7396 Control up vs	0.486754	1.18277	1.18277 Control up vs	2.31001	1.1469	1.32398	1
23953 LTR	0.00078642	0.0105502	1.32865	1.32865 Control up vs	0.0007161	1.65296	1.65296 Control up vs	0.0172712	1.29186	1.29186 Control up vs	11.5782	0.793229	0.182695	1
23926 UIN	0.0333378	0.645283	1.08655	1.08655 Control up vs	0.0201502	1.65176	1.65176 Control up vs	0.454344	0.898397	0.898397 Control up vs	47.1362	1.33101	0.125299	1
24032 MAL2	0.00088426	0.449206	1.08732	1.08732 Control up vs	0.0030172	1.89246	1.89246 Control up vs	0.802519	1.02758	1.02758 Control up vs	16.5279	7.17337	0.276507	1
24091 MAP3K8	4.17E-05	0.0029179	1.14048	1.14048 Control up vs	6.25E-06	1.61226	1.61226 Control up vs	0.00277481	1.21548	1.21548 Control up vs	38.6329	0.760534	0.0249664	1
24192 MBRAT7	0.00091642	0.540285	1.05947	1.05947 Control up vs	0.00091642	1.65989	1.65989 Control up vs	0.0891764	1.19428	1.19428 Control up vs	15.0749	0.15033	0.069258	1
25290 MP2L2	1.44E-05	9.37E-05	1.45994	1.45994 Control up vs	1.73E-06	1.91561	1.91561 Control up vs	0.00020209	1.39699	1.39699 Control up vs	51.2639	1.33141	0.069258	1
25631 MYO1B	0.00693229	0.264037	1.14031	1.14031 Control up vs	0.0012297	1.70411	1.70411 Control up vs	0.0898974	1.2347	1.2347 Control up vs	8.61188	0.96374	0.298422	1
25856 NDRG1	0.00182191	0.0143807	1.32591	1.32591 Control up vs	0.00022558	1.72146	1.72146 Control up vs	0.00174841	1.51716	1.51716 Control up vs	13.2102	1.01606	0.205106	1
25942 NET1	0.00149208	0.0111192	1.36344	1.36344 Control up vs	0.00024084	1.79461	1.79461 Control up vs	0.150328	1.16202	1.16202 Control up vs	14.0391	1.17169	0.222556	1
26455 NEU1	0.00003624	0.00003624	1.35988	1.35988 Control up vs	1.15E-05	1.65042	1.65042 Control up vs	0.001207	1.16876	1.16876 Control up vs	26.9977	0.797464	0.680559	1
26977 NFKB1	0.00203067	0.00243613	1.50565	1.50565 Control up vs	0.00036069	1.75016	1.75016 Control up vs	0.0186191	1.31877	1.31877 Control up vs	12.777	1.05775	0.207062	1
27134 PARR4	0.00033389	0.0109331	1.47492	1.47492 Control up vs	0.00099028	1.8209	1.8209 Control up vs	0.0188412	1.41362	1.41362 Control up vs	8.87336	1.11557	0.347316	1
27615 PKCICD	0.00688628	0.00688628	1.24228	1.24228 Control up vs	1.54E-05	1.60792	1.60792 Control up vs	0.0410017	1.16876	1.16876 Control up vs	19.0784	0.733888	0.102592	1
27628 PIM1	0.00031189	0.00112669	1.39561	1.39561 Control up vs	4.17E-05	1.72055	1.72055 Control up vs	0.00040321	1.30794	1.30794 Control up vs	22.0644	0.937931	0.113449	1
27765 PLEK2	0.00445605	0.0944397	1.28617	1.28617 Control up vs	0.00119494	1.91564	1.91564 Control up vs	0.732936	1.048	1.048 Control up vs	9.96441	1.64306	0.439714	1
27777 PLEKH1	0.00061683	0.0372951	1.24225	1.24225 Control up vs	0.00010958	1.84277	1.84277 Control up vs	0.159764	1.14432	1.14432 Control up vs	18.2433	1.29275	0.188965	1
27811 PLEKHG3	1.51E-05	2.23E-05	1.58234	1.58234 Control up vs	2.54E-06	1.84731	1.84731 Control up vs	0.0006486	1.32615	1.32615 Control up vs	50.681	1.29872	0.0683343	1
27900 PDKXL	0.00014094	0.00116186	1.55021	1.55021 Control up vs	9.92E-05	2.12414	2.12414 Control up vs	0.117027	1.16393	1.16393 Control up vs	27.7091	2.05928	0.198181	1
28318 PRDM2	0.00080746	0.0161224	1.29505	1.29505 Control up vs	0.00012416	1.81517	1.81517 Control up vs	0.00402957	1.40389	1.40389 Control up vs	16.6529	1.13001	0.180951	1
28715 RAB38	0.00019736	0.0053184	1.18638	1.18638 Control up vs	2.90E-05	1.77444	1.77444 Control up vs	0.0101769	1.25394	1.25394 Control up vs	25.2334	1.08256	0.114405	1
28765 RAC2	0.00059668	0.00112939	1.55992	1.55992 Control up vs	0.00010412	1.89015	1.89015 Control up vs	0.0440047	1.23955	1.23955 Control up vs	18.8933	1.43127	0.202014	1
29127 RASD2	0.0007895	0.0012572	1.42594	1.42594 Control up vs	0.00014157	1.63998	1.63998 Control up vs	0.028307	1.21539	1.21539 Control up vs	16.9736	0.847754	0.133188	1
29288 RNF145	2.23E-05	0.467969	1.04338	1.04338 Control up vs	6.16E-06	1.78687	1.78687 Control up vs	0.0270566	1.16205	1.16205 Control up vs	45.6229	1.32294	0.0772582	1
29328 RNF24	0.00091642	0.0294565	1.21708	1.21708 Control up vs	0.00017605	1.62801	1.62801 Control up vs	0.200939	1.10903	1.10903 Control up vs	15.942	0.823371	0.137728	1
29688 S100A8	0.00749563	0.427002	1.20094	1.20094 Control up vs	0.0257546	1.81824	1.81824 Control up vs	0.964623	0.99037	0.99037 Control up vs	13.7618	1.51406	1.19588	1
29687 S100A9	0.00080503	0.0950221	1.47672	1.47672 Control up vs	0.00013128	4.09738	4.09738 Control up vs	0.0523112	1.59835	1.59835 Control up vs	16.8754	6.70445	1.05944	1
29690 S100P	0.202949	0.565142	1.20226	1.20226 Control up vs	0.0493808	2.03497	2.03497 Control up vs	0.295904	1.40582	1.40582 Control up vs	9.12228	1.70515	2.25458	1
30034 SERPINA1	0.00034845	0.00044857	1.59022	1.59022 Control up vs	0.00011696	1.76033	1.76033 Control up vs	0.123267	1.15017	1.15017 Control up vs	21.4983	1.37023	0.164772	1
30051 SERPINE2	0.00127282	0.0628355	1.49018	1.49018 Control up vs	0.0178389	1.82133	1.82133 Control up vs	0.533885	0.877351	0.877351 Control up vs	5.71798	2.12007	1.01202	1
30692 SLC20A1	0.00025393	0.04489884	1.23975	1.23975 Control up vs	0.0008113	1.60677	1.60677 Control up vs	0.383	1.01393	1.01393 Control up vs	11.8844	0.924221	0.260976	1
30695 SLC6A1	0.00135638	0.197837	1.11934	1.11934 Control up vs	0.00024112	1.6539	1.6539 Control up vs	0.0485432	1.2052	1.2052 Control up vs	14.4497	0.872189	0.160961	1
30780 SMOX	0.00056846	0.00227331	1.34572	1.34572 Control up vs	0.00011384	1.60191	1.60191 Control up vs	0.112352	1.12776	1.12776 Control up vs	18.8808	0.795212	0.113516	1
31370 SREBF2	7.98E-05	0.00028282	1.36988	1.36988 Control up vs	1.02E-05	1.65123	1.65123 Control up vs	0.00007602	1.31783	1.31783 Control up vs	32.392	0.802397	0.0660572	1
31464 SFA2	3.15E-05													



no calcium

30 mins recovery



60 mins recovery

120 mins recovery

Supplementary Figure 4 Recruitment during junction formation. 16HBE cells were incubated for 4 days to form mature monolayers, then subjected to a calcium switch to initiate *de novo* junction formation. During the recovery period, cells were fixed at the indicated timepoints, then co-stained for EMP1 with E-cadherin, ZO-1 or claudin-1.

Supplementary Materials and Methods:

Cloning

Dominant negative HRas N17 was subcloned into pQCXIP using Bam HI/Eco RI and fully sequenced. This mutant acts as a dominant negative with respect to all 3 mammalian Ras isoforms [30].

RNAi

An shRNA library was constructed in pSUPERpuro, comprising 3 hairpins per gene, for 86 predicted human Rho GEFs, and a Cdc42 control. EMP1 shRNAs were obtained from the TRC collection in pLKO.1 (MSKCC RNAi core facility). The specific hairpins reported in this study are as follows: shARHGEF18 (gaagctgttagtcattaca), shCdc42 (gatgaccctctactattg), shSOS1.1 (acagttgagtgatataa), shSOS1.2 (ttacatagtagcagtctta), shSOS1.3 (ggcagaaattcgacaatat), shEMP1.1 (ccacatcgtactgttattat), shEMP1.2 (gtgtccatctacactagtcacat). The following siRNA reagents were purchased from Dharmacon: siControl (siLamin A/C; D-001620-02) and siSOS1.1 (5'-acaguugaguggcauuaauu-3')

Cell culture and treatment

16HBE cells were a kind gift from Dr Dieter Gruenert (California Pacific Medical Center, San Francisco) [28]. Cells were cultured as described previously except that FBS was purchased from Omega Scientific (FB-11, Lot 169905) [22]. BCi-NS1.1 cells were cultured in Bronchial Epithelial Growth Media (BEGM, Lonza, CA). For siRNA transfection, 2.5×10^4 16HBE cells were seeded/24 well and treated with 50nM siRNA + 1.25 μ l Lipofectamine LTX (Invitrogen) in antibiotic free media, for 16hrs. The medium was then replaced and the cells incubated for 4 days. For calcium-switch experiments, confluent 16HBE cells were washed 3 times in PBS and incubated in calcium free medium for 4 hrs (DMEM without CaCl₂, Invitrogen 21068, plus 10% FBS pretreated with Chelex 100 Resin, Biorad). Normal growth media, containing calcium, was then replaced for a further 4 hrs. Inhibitor treatments were performed using 500nM GSK1120212 (MEKi), 500nM PD0325901 or 1 μ M Sch772984 (ERKi); DMSO (1:20,000) was used as a carrier control. GSK1120212 and PD0325901 were purchased from Selleck; SCH772984 was kindly provided by Neal Rosen.

Virus production and preparation of stable cells

Retroviral or lentiviral particles were prepared to deliver cDNA or shRNA for stable expression, as described previously [52]. 16HBE infections were performed by centrifugation. 10^5 cells were seeded in a 6-well dish, incubated with 1.5ml of viral suspension + 8 μ g/ml polybrene and spun at 2250rpm for 30mins. Two days post-infection, stable pools were selected using 1.5 μ g/ml puromycin for 4-5 days. To avoid clonal effects, experiments were performed with freshly prepared, short term cultures of selected stable pools, and separate pools were used for each independent experimental repeat. Stable cells were typically reseeded on glass coverslips or plastic dishes for subsequent experiments. However, expression of HRas N17 prevented reattachment after trypsinisation, so these cells were reseeded on coverslips/plastic 1 day post-infection, and selected directly. For BCi-NS1.1, cells were infected overnight in media + 2 μ g/ml polybrene with purified, high titer lentivirus, harbouring control or EMP1 specific shRNA, at an equal multiplicity of infection.

QPCR

10⁵ cells were seeded per 6-well. The media was replaced after 24hrs, +/- inhibitors, and the cells were incubated for a further 3 days. RNA was prepared using the RNeasy kit, with an on-column DNase I digestion, according to the manufacturer's instructions (Qiagen). cDNA was synthesized using Superscript III First-Strand Synthesis Supermix, with random hexamers (Invitrogen, 18080-400). QPCR was performed using Taqman Universal PCR mastermix (Applied Biosystems, 4304437) and the following probes (Applied Biosystems): EMP1 (Hs00608055_m1) and GAPDH (Hs02758991). Samples were loaded onto a 96-well plate (Applied Biosystems, 4346906) and analyzed using a Biorad iQ5 Multicolor RT-PCR Detection System. The data represent 3 independent experiments, each performed in triplicate, with EMP1 expression normalized against the GAPDH control.

Western blotting

Total cell extracts were prepared and western blotting was performed as described previously [53]. Primary antibodies were used as follows: mouse anti-beta actin (AC-74, Sigma), 1:50,000; mouse anti-pERK (M8159, Sigma), 1:1000; rabbit anti-total ERK (M5670), 1:2000; rat anti-Ras (sc-35, Santa Cruz), 1:1000; rabbit anti-p90RSK (S380, Cell Signaling, 9335P); rabbit anti-RSK1 (D6D5, Cell Signaling, 8408S); rabbit anti-SOS1 (sc-256, Santa Cruz), 1:1000. HRP-conjugated secondary antibodies (Dako) were used at 1:5000. Blots were scanned using a Biorad GS-800 Calibrated Densitometer and molecular weight markers are indicated in figures. All western blots are representative of data obtained from three independent experiments.

Immunofluorescence & epifluorescent imaging

16HBE cells were grown on glass coverslips and incubated as described in figure legends. All subsequent steps were performed at room temperature in PBS, unless otherwise indicated. For ZO-1, occludin and E-cadherin staining, cells were fixed using 3.7% formaldehyde (10mins). For MEK, pERK or EMP1 staining, cells were fixed with MeOH at -20°C (10mins). Cells were permeabilized using 0.5% Triton X-100 (5mins), rinsed and then stained with primary antibody overnight at 4°C. Primary antibodies were used as follows: rat anti-E-cadherin (13-1900, Invitrogen), 1:100; mouse anti-EMP1 (Abnova, H00002012-A01), 1:100; rabbit anti-EMP1 (Abgent, AP17400a), 1:100; rabbit anti-pERK (4370, Cell Signaling); mouse anti-MEK (4694, Cell Signaling), 1:100; rabbit anti-ZO-1 (61-7300, Invitrogen), 1:25-100 (according to lot#). Cells were washed, incubated with Alexa Fluor 488/568 goat anti-rabbit (H+L) secondary (1:500; Invitrogen) and Hoechst 33342 (1µg/ml) for 45mins, washed again with PBS and then with water. Coverslips were mounted using Aqua Polymount (Polysciences, Inc) and visualized using an upright AX10 Imager.A1 epifluorescent microscope (Zeiss), equipped with an EC-Plan-NEOFLUAR 20x/0,5 objective (420350-9900) or a Plan-APOCHROMAT 63x/1,4 oil objective (420780-9900) and a Hamamatsu Orca-ER 1394 C4742-80 camera, controlled by Axiovision software (Zeiss). Scale bars were added using Metamorph software (Molecular Devices). All images are representative of data obtained from three independent experiments.

16HBE tight junction formation assay

16HBE cells were seeded at 2.5x10⁴ cells/coverslip in a 24-well plate, treated +/- inhibitors, and incubated for 3-4 days. Cells were fixed and stained for ZO-1/DNA.

For each sample, 5 random, non-overlapping images were acquired at 20x as described above; >500 cells were counted/condition/experiment, across three independent experiments. Apical junction formation was quantified using Metamorph software. Cells with a continuous ring of ZO-1 at cell-cell contacts were counted as having intact apical junctions, cells with punctate, discontinuous or absent ZO-1 at cell-cell contacts were defined as not having apical junctions.

BCi-NS1.1 tight junction formation assay

BCi-NS1.1 cells were seeded at 4.5×10^5 cells/cm² on transwell inserts (0.4 μ m size pore; Corning) coated with human type IV collagen (Sigma) in media consisting of a 1:1 mixture of DMEM (Cellgro, Manassas, VA) and Ham's F-12 Nutrient Mix (Invitrogen) supplemented with 100 U/ml penicillin, 5% fetal bovine serum 100 μ g/ml streptomycin, 0.1% gentamycin and 0.5% amphotericin B. Following overnight incubation, the media was replaced with 1:1 DMEM/Ham's F12 (including antibiotics described above) supplemented with 2% of the serum substitute Ultrosor G (BioSerpa S.A., Cergy-Saint-Christophe, France). Two days post seeding, the cells were fixed and stained for ZO-1/DNA. For each sample, z-stacks were acquired from 6 non-overlapping fields of view; this was repeated across 3 independent experiments. Images were acquired using a Nikon Ti-E inverted microscope attached to a CoolSNAP CCD camera (Photometrics) with a ELWD 20x Plan Fluor objective and NIS Elements software (Nikon). Nuclei were counted to confirm that cell number was consistent across different stable cell lines. Apical junction formation was quantified by counting continuous rings of ZO-1 staining/field of view.

Transepithelial resistance assay

5×10^4 16HBE cells were seeded on collagen coated, 0.4 μ m PTFE membrane filters in 6.5mm inserts (Costar, 3495), on 24-well plates (Corning). 24 hours later, the cells were confluent and the media was replaced +/- inhibitors for a further 3 days. Transepithelial resistance was measured using an EVOM volttohmmeter and STX2 electrode (World Precision Instruments), according to the manufacturer's instructions. The data were acquired through three independent experiments. For BCi-NS1.1 cells, air-liquid interface (ALI) cultures were analyzed on day 21 post-seeding, as described previously [42].

FM4-64 dye assay and confocal imaging

10^5 16HBE cells were seeded per glass bottomed, 35mm dish (MatTek). 24 hours later, the media was replaced +/- inhibitors, and the cells were incubated for a further 3 days. The lipophilic probe, FM4-64 (5 μ g/ml; Invitrogen) was added to the media for 10mins at 37°C. Confocal z-stacks were acquired from 10 fields of view using an Ultraview Vox spinning disc confocal system (Perkin Elmer), equipped with a Yokogawa CSU-X1 spinning disc head, and EMCCD camera (Hamamatsu C9100-13), coupled with a Nikon Ti-E microscope. For quantification, 3 x/z slices per image were analyzed using ImageJ software. To generate fluorescence ratios, line profiles were drawn over apical and basolateral surfaces to calculate average fluorescent intensities. The data were acquired through three independent experiments.

Microarray analysis

Differential gene expression analysis was performed using 4 parallel samples: control (pQCXIP cells, DMSO), dominant negative HRas (pQCXIP-HRas N17 cells), MEK inhibitor (pQCXIP cells, GSK1120212/MEKi), ERK inhibitor (pQCXIP cells,

SCH772984/ERKi). The cells were seeded at $10^5/6$ -well. 24 hours later, the medium was replaced +/- inhibitors, and the cells were incubated for a further 3 days. Three independent samples were prepared for each condition. RNA was isolated as described above. Quality control and microarray analysis were performed by the MSKCC Genomics Core Laboratory, using an Illumina gene expression array (Human HT-12, 47 000 transcripts). The data were analyzed using Partek software. Briefly, gene level data were subjected to quantile normalization and each group was compared to the control to detect differentially expressed genes using 1-way ANOVA. We selected genes that were downregulated by >1.6-fold, relative to the control, with an unadjusted p-value <0.05.

Statistics

Unless otherwise indicated, statistical significance was evaluated using Prism software.

Unpaired t-tests were performed, with 2-tailed P-values and 95% confidence intervals.