BMJ Open

Effect of study design and setting on tuberculosis clustering estimates using Mycobacterial Interspersed Repetitive Units-Variable Number Tandem Repeats (MIRU-VNTR)

Journal:	BMJ Open
Manuscript ID:	bmjopen-2014-005636
Article Type:	Research
Date Submitted by the Author:	12-May-2014
Complete List of Authors:	Mears, Jessica; University College London, Department of Infection and Population Health Abubakar, Ibrahim; University College London, Department of Infection and Population Health; Public Health England, Centre for Infectious Disease Surveillance and Control Cohen, Ted; Harvard School of Public Health, Harvard University, Division of Global Health Equity, Brigham and Women's Hospital and Department of Epidemiology McHugh, Timothy; Centre for Clinical Microbiology, Research Department of Infection, Royal Free Campus, University College London Sonnenberg, Pamela; University College London, Department of Infection and Population Health
Primary Subject Heading :	Research methods
Secondary Subject Heading:	Infectious diseases, Public health
Keywords:	EPIDEMIOLOGY, Tuberculosis < INFECTIOUS DISEASES, MOLECULAR BIOLOGY

SCHOLARONE[™] Manuscripts

1		
2 3	1	Effect of study design and setting on tuberculosis clustering estimates using
4	2	Mycobacterial Interspersed Repetitive Units-Variable Number Tandem
5	-	Poposts (MIDII)/NTD)
0 7	5	
8 9	4	Jessica Mears ¹ , Ibrahim Abubakar ^{1,2} , Theodore Cohen ³ , Timothy D McHugh ⁴ & Pam Sonnenberg ^{1,*}
10 11	5	¹ Department of Infection and Population Health, University College London
12 13	6	² Centre for Infectious Disease Surveillance and Control, Public Health England
14 15 16	7	³ Division of Global Health Equity, Brigham and Women's Hospital and Department of Epidemiology,
17 18	8	Harvard School of Public Health, Harvard University
19 20	9	⁴ Centre for Clinical Microbiology, Department of Infection, University College London
21 22	10	*Corresponding author:
23 24 25	11	Dr Pam Sonnenberg
26 27	12	3° Floor Mortimer Market Centre
28 29	13	London WC1E 6IB
30 31 32	15	p.sonnenberg@ucl.ac.uk
33 34	16	
35 36 37	17	Word count: 1,733
38 39		
40 41		
42 43		
43 44		
45		

3	
4	
5	
6	
7	
2 Q	
0	
9	
10	
11	
12	
13	
14	
15	
16	
17	
18	
19	
20	
21	
22	
23	
24	
25	
26	
27	
28	
20	
20	
21	
27	
3Z	
აა ე⊿	
34	
35	
30	
37	
38	
39	
40	
41	
42	
43	
44	
45	
46	
47	
48	
49	
50	
51	
52	
53	
54	
55	
56	
57	
58	
59	
60	

18 Abstract

- 19 Objectives: To systematically review the evidence for the impact of study design and setting on the
- 20 interpretation of TB transmission using clustering derived from Mycobacterial Interspersed
- 21 Repetitive Units – Variable Number Tandem Repeats (MIRU-VNTR) strain typing.

22 Data sources: Medline, Embase, CINHAL, Web of Science and Scopus were searched for articles

- 23 published before November 2012
- 24 Review methods: Studies in humans that reported the proportion of clustering of TB isolates by
- 25 MIRU-VNTR were included in the analysis. Univariable meta-regression analyses were conducted to
- 26 assess the influence of study design and setting on the proportion of clustering.
- 27 Results: The search identified 14 eligible articles reporting clustering between 22.1% and 61.2%. The
- 28 proportion of culture positive isolates and the number of MIRU-VNTR loci typed explained 49% and
 - 34% of the between study variation, respectively, and had a significant association with the 29
- 30 proportion of clustering.
- 31 Conclusions: Although MIRU-VNTR typing is being adopted worldwide there is a paucity of data on
- 32 how study design and setting may influence estimates of clustering. We have highlighted study
- 33 design variables for consideration in the design and interpretation of future studies.
- 34

37

38

39

40

41

- 35 Strengths and Limitations of Study
- This is a timely evaluation of the impact of study design on estimates of TB clustering using 36 MIRU-VNTR strain typing because it has been incorporated into national typing services globally.
 - There were insufficient data available to fully explore the impact of study design and setting on estimates of clustering.

BMJ Open

2	
1	
4	
5	
6	
7	
8	
9	
10	
11	
12	
12	
13	
14	
15	
16	
17	
18	
19	
20	
21	
22	
22	
∠3 24	
24	
25	
26	
27	
28	
29	
30	
31	
32	
22	
22	
34	
35	
36	
37	
38	
39	
40	
41	
12	
42 12	
43	
44	
45	
46	
47	
48	
49	
50	
51	
52	
52	
55	
54	
55	
56	
57	
58	
59	
60	

43 Introduction

The introduction of molecular typing methods has improved our understanding of *Mycobacterium tuberculosis* (TB) transmission and has changed local and national control policies [1–5]. The proportion of cases that are clustered is often used to estimate the amount of ongoing transmission within the population, based on the assumption that cases with indistinguishable strain types are part of a chain of transmission. TB molecular typing methodology is changing rapidly and it is important that we better understand how to interpret the outputs and thus act.

TB molecular typing methods include Spoligotyping [6], insertion sequence *6110* (IS*6110*) restriction fragment length polymorphism (RFLP) analysis (the recent gold standard) [7], mycobacterial interspersed repetitive units-variable number tandem repeats (MIRU-VNTR) typing [8], and whole genome sequencing [9–11]. Published reviews have identified factors that might influence or bias clustering by IS*6110* RFLP [12,13]. No study has repeated this analysis using more up-to-date typing methods, which is important for understanding of the epidemiology of TB and to shape the application of molecular typing to improve TB control.

57 Published meta-analyses and modelling studies using IS6110 RFLP data show that the proportion of 58 clustering observed can be affected by 1) study design (affecting the proportion of eligible cases that 59 are included in the study); 2) features of the typing method (such as the ability to type isolates with 60 low copy numbers); and 3) study setting (such as characteristics of the study population). For 61 example, the proportion of clustering increases when the fraction of the total data sampled 62 increases [13–15] and when study duration increases [16].

63 MIRU-VNTR is currently the preferred method of molecular typing [17–21], and can be used 64 together with Spoligotyping [8]. Relative to IS6110 RFLP, MIRU-VNTR does not have to exclude 65 isolates with a low IS6110 copy number, has a faster turnaround time, is high throughput and the numeric strain types are more easily compared. MIRU-VNTR strain typing is increasingly being 66 67 adopted worldwide [1,22–27], yet unlike IS6110 RFLP, the evidence for the interpretation of the 68 findings such as the impact of study design and setting on clustering have not been reviewed. 69 Although the two typing methods have been shown to have a similar discriminatory value, the 70 markers evolve independently and at different rates, resulting in a difference in clustering between 71 the two methods [28]. This suggests that there could be differences in the way study design, typing 72 method and setting affects clustering by the two methods. We conducted a systematic review to 73 assess the evidence for the impact of study design and setting on the interpretation of TB

transmission using clustering derived from MIRU-VNTR strain typing – as has been shown using
IS*6110* RFLP typing.

76 Methods

Five electronic databases were searched (EMBASE, ISI Web of Science, CINHAL, Scopus and Medline
(Ovid)) up to 1 November 2012. The search strategy combined the following terms with Boolean
operators: Tuberculosis, strain typing, and transmission. The search was limited to studies using the
standard MIRU-VNTR method [8], in humans only, and in English.

All titles and abstracts from each of the searches were examined. The full text of each paper was obtained and reviewed if the study reported MIRU-VNTR strain typing of *M.tuberculosis* complex isolates with at least 15 of the standardised 24 loci [8,29,30].

Studies using fewer than 15 loci were not included because the level of discrimination is inadequate
for epidemiological use (n=97) [8]. Studies that used loci different to the standardised 15 and 24 set
were not included in the analysis in order to reduce the heterogeneity between studies (n=11). All
publication types were included in this first screen to ensure that no relevant data were missed.

Reviews, letters, editorials, outbreaks or case reports (n=99) were excluded in the second screen.
Studies that used incomplete sampling (e.g. random samples, studies using subsets of populations
such as MDR patients) (n=30) and studies that had a sample size of less than 50 (n=2) were also
excluded.

A reviewer extracted the following data items from all included studies using a form developed in Excel (Microsoft 2010): publication details (year, authors, study country), study details (study duration, loci typed, secondary typing method, study population), the proportion of total TB isolates clustered by MIRU-VNTR strain typing, and the covariates of interest: the number of clustered and unique isolates; the maximum size of clusters; the proportion of clusters containing two cases; the prevalence of culture-positivity among TB patients included in the study; the proportion of culture positive isolates typed; risk factors for clustering; and the Hunter Gaston Discriminatory Index (HGDI) [31]).

Authors were contacted if TB incidence rate was not reported. Where no response was receivedWHO country estimates of TB incidence for the study year were used [32].

Data were analysed in Stata 12. Where studies reported data from more than one set of loci, the
 method with the highest discriminatory value was included (i.e. MIRU-VNTR 24 would be chosen

BMJ Open

2	
3	
4	
5	
е С	
7	
1	
8	
9	
1	0
1	1
1	2
4	2 2
1	3
1	4
1	5
1	6
1	7
1	8
1	9
່າ	ñ
2	4
2	1
2	2
2	3
2	4
2	5
2	6
<u>っ</u>	7
2	<i>'</i>
2	8
2	9
3	0
3	1
3	2
3	3
2	о Л
ა ი	4 C
3	5
3	6
3	7
3	8
3	9
Δ	0
л Л	1
+ ^	י ר
4	2
4	3
4	4
4	5
4	6
4	7
т Л	Ω
+ /	0
4	9
5	U
5	1
5	2
5	3
5	4
5	5
ວ ເ	5
ວ	o T
5	1
5	8
5	9

60

104 over MIRU-VNTR 15, and MIRU-VNTR 15 plus Spoligotyping would be chosen over MIRU-VNTR 15 105 alone) (n=5). This review was not concerned with summary measures of clustering, but factors that 106 influenced clustering; therefore articles must have included at least one of the covariates. 107 Continuous variables were transformed where the distribution was skewed. The proportion 108 clustered was transformed using the Freeman Tukey transformation [33]. Univariable meta-109 regression analyses were carried out to determine the effect of the study design covariates on the 110 proportion of clustered isolates. All covariates in the analysis were hypothesised to influence the 111 proportion clustered *a priori*.

112 Results

The search identified 5607 references resulting in 12 journal articles and 2 conference abstracts
included after deduplication and title/abstract/full text screening (Figure 1). The main characteristics
of the included studies are shown in Table 1. Studies were published between 2007 and 2011 and
the clustering reported varied from 22.1% [34] to 61.2% [35].

117 The univariable meta-regression shows evidence for the proportion of clustering to decrease as the 118 prevalence of culture-positivity among TB patients included in the study increases (p=0.03; Table 2), 119 accounting for 49% of the between study variation. There was also evidence for the proportion of 120 clustering to decrease as the number of MIRU-VNTR loci typed increased from 15 to 24 (p=0.02), 121 explaining 34% of the between study variation. There was no evidence of the other study design or 122 study setting variables significantly influencing the proportion clustered. Though non-significant 123 (p>0.05), the size of the study and the maximum cluster size explained 15% and 27% of the between 124 study variation, respectively.

125 Discussion

126 This review identified 14 studies that met the inclusion criteria. We illustrate that the interpretation

127 of studies using MIRU-VNTR to estimate clustering is subject to bias relating to study design;

128 however, there were insufficient data available to fully explore the impact of study design and

129 setting on estimates of clustering.

130 As expected, we found that the proportion of clustering decreased with a greater number of MIRU-

131 VNTR loci typed. Our finding that the prevalence of culture-positivity among TB patients included in

the study influences the estimates of transmission within a population is counterintuitive and not

- 133 consistent with estimates of the influence of sampling on the proportion of clustering using *IS*6110
- 134 RFLP typing [36]. This may reflect the relationship between TB burden and resource poor/rich

settings and the consequent availability of culture diagnostic laboratory services; i.e. in resource
poor settings where there is a high burden of TB (and, therefore, high rates of clustering) the
prevalence of culture positive TB cases is low. The finding may also be due to chance, with only 8
studies included in the analysis of this variable.

The other study design variables included in this analysis, such as study duration, did not significantly influence the proportion of isolates that were clustered, contrary to previous findings [12]. This is likely to be because of a lack of good quality evidence: only 14 studies met the inclusion criteria for the review and of those only three reported all the variables of interest, reducing the power of the analysis and precluding multivariable meta-regression. In addition, the range of the variables may have been too limited to show any impact on clustering estimates. For example, the proportion of culture positive isolates typed had a narrow range from 81.9% to 100%. Furthermore, most of the studies were from low TB burden settings and therefore may be reflecting the rate at which imported cases have matching strain types by chance, rather than rates of recent transmission.

This study is a timely evaluation of the impact of study design on estimates of TB clustering using MIRU-VNTR strain typing because it has been incorporated into national typing services globally [23,37]. The findings are relevant where strain typing is used to evaluate TB control systems across different settings because the proportion of clustering is influenced by the prevalence of culture positive TB cases in the study setting. Given that strain typing methods are advancing beyond MIRU-VNTR typing and that the application of whole genome sequencing to TB control and public health strategies has been demonstrated [9–11,38], it is important that the biases in the analysis of such methods are explored and compared. Understanding how to design and compare research studies for public health will greatly improve the benefit gained from newer technologies.

157 This review has highlighted the need for better quality reporting in primary studies to enable future 158 reviews to be more robust. A lack of standards for the molecular epidemiology of infectious diseases 159 may explain the poor quality of reporting; this field would benefit from the introduction of such 160 standards (STROBE-ID, submitted).

The use of TB strain typing as a public health tool in TB control programmes is increasing globally.
We have identified a lack of good quality studies that can contribute to our understanding in
interpreting the molecular typing of TB. We have also shown that the proportion of clustering
derived from MIRU-VTNR typing is influenced by the number of loci typed and the prevalence of
culture-positivity among TB patients included in the study, highlighting these as important
considerations in the design and interpretation of future studies.

1														
2														
3	167	Conflict of interest												
5 6	168	Nothing to declare.												
7 8 9	169	Acknowledgements												
10 11	170	We would like to acknowledge Ross Harris from the Statistics Unit at Public Health England for his												
12 13	171	advice on meta-regression. Author contributions												
14 15	172													
16 17	173	All authors made substantial contributions to the conception and design of the review, and the												
18 10	174	analysis and interpretation of data. JM drafted the article and PS, IA, TM and TC revised it critically												
20 21	175	for important intellectual content. All authors approved the final version for publication.												
22 23	176	Funding												
24 25	177	JM is funded through a Public Health England and University College London Impact Studentship. IA												
26 27	178	is funded through a NIHR Senior Research Fellowship.												
28 29 30	179	Ethics												
31 32	180	Ethical approval was not required as this review analyses data that is in the public domain.												
33 34	181	Data sharing												
35 36 27	182	No additional data are available												
37 38 39	183	References												
40	184	1. Lambregts-van Weezenbeek CSB, Sebek MMGG, Van Gerven PJHJ, De Vries G, Verver S, et al.												
41	185	(2003) Tuberculosis contact investigation and DNA fingerprint surveillance in The Netherlands:												
4Z /3	186	6 years' experience with nation-wide cluster feedback and cluster monitoring. Int J Tuberc Lung												
44	187	Dis 7: S463–470.												
45	100	2 Borgdorff MW Van den Hof S. Kromer K. Verbagen L. Kalicupart N. et al. (2010) Progress												
46	180	towards tuberculosis elimination: secular trend immigration and transmission. Fur Respir 1.36												
47 48	100	339–347. doi:10.1183/09031936.00155409.												
40														
50	191	3. Kik SV, Verver S, Van Soolingen D, De Haas PEW, Cobelens FG, et al. (2008) Tuberculosis												
51	192	Outbreaks Predicted by Characteristics of First Patients in a DNA Fingerprint Cluster. Am J												
52 52	193	Respir Crit Care Med 178: 96–104. doi:10.1164/rccm.200708-1256OC.												
วง 54	10/	A Small PM McClenny NB Singh SP Schoolnik GK Tompkins (Set al. (1993) Molecular strain												
55	194	typing of Mycohacterium tuberculosis to confirm cross-contamination in the mycohacteriology												
56	155	Cyping of mycobacteriam tabercalosis to commit cross contamination in the mycobacteriology												
57														
58		7												
59 60														
00														

2			
3	196		laboratory and modification of procedures to minimize occurrence of false-positive cultures. J
4	197		Clin Microbiol 31: 1677–1682.
5		_	
6 7	198	5.	De Vries G, Van Hest RAH, Richardus JH (2007) Impact of mobile radiographic screening on
7	199		tuberculosis among drug users and homeless persons. Am J Respir Crit Care Med 176: 201–
0	200		207. doi:10.1164/rccm.200612-1877OC.
10		~	
11	201	6.	Kamerbeek J, Schouls L, Kolk A, Van Agterveld M, Van Soolingen D, et al. (1997) Simultaneous
12	202		detection and strain differentiation of Mycobacterium tuberculosis for diagnosis and
13	203		epidemiology. J Clin Microbiol 35: 907–914.
14	204	_	
15	204	7.	Van Embden JD, Cave MD, Crawford JT, Dale JW, Eisenach KD, et al. (1993) Strain identification
16	205		of Mycobacterium tuberculosis by DNA fingerprinting: recommendations for a standardized
17	206		methodology. J Clin Microbiol 31: 406–409.
18	207	~	Sumah, D. Allin C. Logicon, C. Condese, Oslamona, M. Düsch, Condes C. et al. (2006) Descended for
19	207	δ.	Supply P, Allix C, Lesjean S, Cardoso-Oelemann M, Rusch-Gerdes S, et al. (2006) Proposal for
20	208		standardization of optimized mycobacterial interspersed repetitive unit-variable-number
21	209		tandem repeat typing of Mycobacterium tuberculosis. J Clin Microbiol 44: 4498–4510.
22	210		doi:10.1128/JCM.01392-06.
23	244	~	
24	211	9.	Schurch AC, van Soolingen D (2011) DNA fingerprinting of Mycobacterium tuberculosis: From
20	212		phage typing to whole-genome sequencing. Infect Genet Evol. Available:
20	213		http://www.ncbi.nlm.nih.gov/pubmed/22067515. Accessed 13 March 2012.
28	214	10	Cardy II. Jakaster JC, U. S., CL, Carly VI, Chak L, et al. (2011) Whale some resource in a and
29	214	10.	Gardy JL, Jonnston JC, Ho Sul SJ, Cook VJ, Shan L, et al. (2011) Whole-genome sequencing and
30	215		social-network analysis of a tuberculosis outbreak. N Engl J Med 364: 730–739.
31	216		dol:10.1056/NEJM0a1003176.
32	217	11	Welling TM in CL. Herrell DL. Evens IT. Kenstei C. et al. (2012) Whele segure environments
33	217	11.	walker TM, Ip CL, Harrell RH, Evans JT, Kapatal G, et al. (2013) whole-genome sequencing to
34	218		delineate Mycobacterium tuberculosis outbreaks: a retrospective observational study. Lancet
35	219		Infect Dis 13: 137–146. doi:10.1016/S1473-3099(12)70277-3.
36	220	10	Hauban BMCL Churn IB (2000) A systematic region and mata analysis of molecular
37	220	12.	nouberi Rivigi, Giynn JR (2009) A systematic review and meta-analysis of molecular
38	221		Tranical Madicine & International Lealth 14, 802, 000, doi:10.1111/j.1205.2156.2000.02216.v.
39	222		Tropical Medicine & International Health 14: 892–909. doi:10.1111/j.1365-3156.2009.02316.X.
40	77 2	12	Fok A. Numata V. Schulzer M. EitzGerald MI (May) Bick factors for dustering of tubersulesis
41	225	15.	FOR A, Numata F, Schulzer M, FilzGerald MJ (May) Risk factors for clustering of tuberculosis
42 43	224		Article] The International Journal of Tuborculasic and Jung Disease 12: 480, 402
43	225		Articlej. The International Journal of Tuberculosis and Lung Disease 12, 480–492.
45	226	1/	Borgdorff MW/ Van Den Hof S. Kalisvaart N. Kremer K. Van Soolingen D. (2011) Influence of
46	220	14.	Sampling on Clustering and Accessitions With Pick Easters in the Melacular Enidemiology of
47	227		Sampling on clustering and Associations with Risk Factors in the Molecular Epidemiology of
48	220		Tuberculosis. All J Epidemiol. Available.
49	229		http://aje.oxfordjournals.org/content/early/2011/05/23/aje.kwr061. Accessed 29 March 2012.
50	220	15	Clupp IP Payor I Do Poor AS Porgdorff MW/ Fina DE at al (1000) Interpreting DNA fingerprint
51	250	15.	Givini JK, Bauer J, De Boer AS, Borguorri MW, Fille PE, et al. (1999) Interpreting DNA Ingerprint
52	251		Enidemiology and Control of Tuberculoris. Int L Tuberculor Dis 2: 1055 1060
53	232		Epidemiology and control of Tuberculosis. Int J Tuberc Lung Dis 3. 1055–1060.
54	222	16	Glypp IR Crampin AC Vates MD Tracre H. Mwaungulu ED, et al. (2005) The Importance of
55	233	10.	Recent Infection with Mycobacterium tuberculosis in an Area with High HIV Drevalance: A
56 57	234		Accent intection with mycobacterian tabercalosis in an Area with High hiv Frevalence. A
5/ 50			2
50			ŏ
60			

2 3 4	235 236		Long-Term Molecular Epidemiological Study in Northern Malawi. J Infect Dis 192: 480–487. doi:10.1086/431517.
5 6 7 8 9	237 238 239	17.	De Beer JL, Kremer K, Ködmön C, Supply P, Van Soolingen D (2012) First Worldwide Proficiency Study on Variable-Number Tandem-Repeat Typing of Mycobacterium tuberculosis Complex Strains. Journal of Clinical Microbiology 50: 662–669. doi:10.1128/JCM.00607-11.
10 11 12 13	240 241 242	18.	Maes M, Kremer K, Van Soolingen D, Takiff H, De Waard JH (2008) 24-locus MIRU-VNTR genotyping is a useful tool to study the molecular epidemiology of tuberculosis among Warao Amerindians in Venezuela. Tuberculosis (Edinb) 88: 490–494. doi:10.1016/j.tube.2008.04.003.
14 15 16 17	243 244 245	19.	Sougakoff W (2011) Molecular epidemiology of multidrug-resistant strains of Mycobacterium tuberculosis. Clinical Microbiology and Infection 17: 800–805. doi:10.1111/j.1469-0691.2011.03577.x.
19 20 21 22	246 247 248	20.	Weniger T, Krawczyk J, Supply P, Niemann S, Harmsen D (2010) MIRU-VNTRplus: a web tool for polyphasic genotyping of Mycobacterium tuberculosis complex bacteria. Nucleic Acids Res 38: W326–331. doi:10.1093/nar/gkq351.
23 24 25	249 250	21.	Supply P (2010) MIRU-VNTR typing: the new international standard for TB molecular epidemiology Symposium of the Institut Pasteur de Tunisia.
26 27 28	251 252 253	22.	Van Soolingen D, Borgdorff MW, De Haas PE, Sebek MM, Veen J, et al. (1999) Molecular epidemiology of tuberculosis in the Netherlands: a nationwide study from 1993 through 1997. J Infect Dis 180: 726–736. doi:10.1086/314930.
29 30 31 32 33	254 255 256	23.	Cowan LS, Diem L, Monson T, Wand P, Temporado D, et al. (2005) Evaluation of a two-step approach for large-scale, prospective genotyping of Mycobacterium tuberculosis isolates in the United States. J Clin Microbiol 43: 688–695. doi:10.1128/JCM.43.2.688-695.2005.
34 35 36	257 258	24.	New CDC Program for Rapid Genotyping of Mycobacterium Tuberculosis Isolates (2005). JAMA 293: 2086–2086. doi:10.1001/jama.293.17.2086.
37 38 39 40	259 260 261	25.	Bauer J, Kok-Jensen A, Faurschou P, Thuesen J, Taudorf E, et al. (2000) A prospective evaluation of the clinical value of nation-wide DNA fingerprinting of tuberculosis isolates in Denmark. Int J Tuberc Lung Dis 4: 295–299.
41 42 43 44 45	262 263 264	26.	Bauer J, Yang Z, Poulsen S, Andersen AB (1998) Results from 5 years of nationwide DNA fingerprinting of Mycobacterium tuberculosis complex isolates in a country with a low incidence of M. tuberculosis infection. J Clin Microbiol 36: 305–308.
45 46 47 48	265 266	27.	Zolnir-Dovc M, Poljak M, Erzen D, Sorli J (2003) Molecular epidemiology of tuberculosis in Slovenia: results of a one-year (2001) nation-wide study. Scand J Infect Dis 35: 863–868.
49 50 51 52 53 54	267 268 269 270 271	28.	Hanekom M, Van der Spuy GD, Gey van Pittius NC, McEvoy CRE, Hoek KGP, et al. (2008) Discordance between mycobacterial interspersed repetitive-unit-variable-number tandem- repeat typing and IS6110 restriction fragment length polymorphism genotyping for analysis of Mycobacterium tuberculosis Beijing strains in a setting of high incidence of tuberculosis. J Clin Microbiol 46: 3338–3345. doi:10.1128/JCM.00770-08.
55 56 57 58 59 60	272 273	29.	Supply P, Lesjean S, Savine E, Kremer K, Van Soolingen D, et al. (2001) Automated high- throughput genotyping for study of global epidemiology of Mycobacterium tuberculosis based 9
			For near raviou only bits //bmianon bmi com/sita/about/auidalines ybtml

3	274		on mycobacterial interspersed repetitive units. J Clin Microbiol 39: 3563–3571.
4	275		doi:10.1128/JCM.39.10.3563-3571.2001.
5			
6	276	30.	Gopaul KK, Brown TJ, Gibson AL, Yates MD, Drobniewski FA (2006) Progression toward an
7	277		improved DNA amplification-based typing technique in the study of Mycobacterium
8 9	278		tuberculosis epidemiology. J Clin Microbiol 44: 2492–2498. doi:10.1128/JCM.01428-05.
10	279	31	Hunter PR Gaston MA (1988) Numerical index of the discriminatory ability of typing systems:
11	275	51.	an application of Simpson's index of diversity 1 Clin Microbiol 26: 2465–2466
12	200		
13	281	32	WHO I TB data (n d) WHO Available: http://www.who.int/tb/country/en/index.html
14	201	52.	Accessed 12 December 2012
15	202		Accessed 12 December 2012.
16	283	22	Freeman ME Tukey IW (1950) Transformations Related to the Angular and the Square Root
17	205	55.	Ann Math Statist 21: 607–611. doi:10.1214/20ms/1177720756
18	204		Ann Math Statist 21. 007–011. 001.10.1214/a0115/1177729750.
19	205	24	Oplomann MC Dial P. Vatin V. Haas W. Bücch Gordos S. at al. (2007) Assessment of an
20	205	54.	entimized mycebactorial interconcred repetitive, unit variable number tandem repeat tuning
21	200		optimized mycobacterial interspersed repetitive- unit-variable-humber tandem-repeat typing
22	287		system combined with spoligotyping for population-based molecular epidemiology studies of
23	288		tuberculosis. J Clin Microbiol 45: 691–697. doi:10.1128/JCM.01393-06.
24	200	25	E and (2010) And the four distant shorts in the second state in the United
20	289	35.	Evans J (2010) Analysis of prevalent Mycobacterium tuberculosis strains in the United
20	290		Kingdom: detection, distribution and expansion of MIRU-VNTR profiles containing high
21	291		numbers of isolates. European Society of Clinical Microbiology and Infectious Diseases. Vienna,
20	292		Austria.
29			
31	293	36.	Glynn JR, Vyonycky E, Fine PEM (1999) Influence of Sampling on Estimates of Clustering and
32	294		Recent Transmission of Mycobacterium tuberculosis Derived from DNA Fingerprinting
33	295		Techniques. American Journal of Epidemiology 149: 366–371.
34			
35	296	37.	TB Strain Typing Project Board HPA (2011) TB Strain Typing Cluster Investigation Handbook for
36	297		Health Protection Units. Available:
37	298		https://hpaintranet.hpa.org.uk/Content/ProgrammesProjects/HPAProgrammes/HPAKeyHealth
38	299		ProtectionProgrammes/Respiratory/TB/StrainTyping/. Accessed 30 November 2011.
39			
40	300	38.	Walker TM, Monk P, Grace Smith E, Peto TEA (2013) Contact investigations for outbreaks of
41	301		Mycobacterium tuberculosis: advances through whole genome sequencing. Clin Microbiol
42	302		Infect. doi:10.1111/1469-0691.12183.
43			
44	303	39.	Asgharzadeh M, Kafil HS, Roudsary AA, Hanifi GR (2011) Tuberculosis transmission in
45	304		Northwest of Iran: using MIRU-VNTR, ETR-VNTR and IS6110-RFLP methods. Infect Genet Evol
46	305		11: 124–131. doi:10.1016/j.meegid.2010.09.013.
47			
48	306	40.	Allix-Béguec C, Supply P, Wanlin M, Bifani P, Fauville-Dufaux M (2008) Standardised PCR-based
49	307		molecular epidemiology of tuberculosis. Eur Respir J 31: 1077–1084.
50	308		doi:10.1183/09031936.00053307.
51			
52	309	41.	Allix-Béguec C, Fauville-Dufaux M, Supply P (2008) Three-year population-based evaluation of
つづ F 4	310		standardized mycobacterial interspersed repetitive-unit-variable-number tandem-repeat
54 55	311		typing of Mycobacterium tuberculosis. J Clin Microbiol 46: 1398–1406.
00 56	312		doi:10.1128/JCM.02089-07.
50 57			-,
58			10
59			10
60			
~~			

60

2			
3	313	42.	Roetzer A, Schuback S, Diel R, Gasau F, Ubben T, et al. (2011) Evaluation of Mycobacterium
4	314		tuberculosis typing methods in a 4-year study in Schleswig-Holstein, Northern Germany. J Clin
5	315		Microbiol 49: 4173–4178. doi:10.1128/JCM.05293-11.
6			
7	316	43.	Oio OO. Sheehan S. Corcoran DG. Nikolayeysky V. Brown T. et al. (2010) Molecular
8	317		enidemiology of Mycohacterium tuberculosis clinical isolates in Southwest Ireland. Infect
9	318		Genet Evol 10: 1110–1116 doi:10.1016/i meegid 2010.07.008
10	510		Gener Evol 10. 1110 1110. doi.10.1010/j.mcc6id.2010.07.000.
11	210	11	Dymova MA Liashanko OO Poteiko PL Krutko VS Khranov FA et al. (2011) Genetic variation
12	220		of Mycobactorium tuborculoric circulating in Kbarkiy Oblact Ukraina, PMC Infact Dic 11: 77
13	320		of Mycobacterium tuberculosis circulating in Kharkiv Oblast, Okraine. Divic Imett Dis 11. 77.
14	321		00110.1186/14/1-2334-11-77.
15	222	4 5	Dide as Statistic Table Decision Canada D (2014) One consistent idea ad attaction of 24
16	322	45.	Bidovec-Stojković U, Zoinir-Dovć IVI, Supply P (2011) One year nationwide evaluation of 24-
17	323		locus MIRU-VNTR genotyping on Slovenian Mycobacterium tuberculosis isolates. Respir Med
18	324		105 Suppl 1: S67–73. doi:10.1016/S0954-6111(11)70014-2.
19			
20	325	46.	Alonso-Rodriguez N, Martínez-Lirola M, Sánchez ML, Herranz M, Peñafiel T, et al. (2009)
21	326		Prospective universal application of mycobacterial interspersed repetitive-unit-variable-
22	327		number tandem-repeat genotyping to characterize Mycobacterium tuberculosis isolates for
23	328		fast identification of clustered and orphan cases. J Clin Microbiol 47: 2026–2032.
24	329		doi:10.1128/JCM.02308-08.
25			
26	330	47.	Hamblion EL. Wynne-Edwards E. Anderson C. Anderson SR (2011) A summary of strain typing
27	331		and clustering of TB in London in 2010 and an analysis of the associated risk factors. Thorax 66:
28	332		$\Delta 88 = \Delta 89$ doi:10.1136/thoraxinl-2011-201054c 50
29	552		
30	222	18	Mandal S. Bradshaw I. Anderson J.F. Brown T. Evans IT. et al. (2011) Investigating Transmission
31	221	40.	of Mycobactorium Boyis in the LIK 2005 2008 1 Clin Microbiol Ayailable:
32	554 525		http://icm.acm.org/content/oorly/2011/02/22/JCM 02200.10. Accessed 24 April 2012
33	555		http://jcm.asm.org/content/eany/2011/03/23/JCM.02299-10. Accessed 24 April 2012.
34	220	40	Sails AD, Derrett A, Servinson S, Massa JC, Mayneyd D, et al. (2011) Malagular anidamialagy of
35	330	49.	Salis AD, Barrett A, Sarginson S, Magee JG, Maynard P, et al. (2011) Molecular epidemiology of
36	337		Nycobacterium tuberculosis in East Lancashire 2001-2009. Thorax 66: 709–713.
37	338		doi:10.1136/thx.2011.158881.
38			
39	339	50.	Nikolayevskyy VV, Brown TJ, Bazhora YI, Asmolov AA, Balabanova YM, et al. (2007) Molecular
40	340		epidemiology and prevalence of mutations conferring rifampicin and isoniazid resistance in
41	341		Mycobacterium tuberculosis strains from the southern Ukraine. Clin Microbiol Infect 13: 129–
42	342		138. doi:10.1111/j.1469-0691.2006.01583.x.
43			
44	343		
45			
46			
47			
48			
49			
50			
51			
ວ∠ 50			
00 54			
04 EE			
00 56			
00 57			
3/ E0			
50			11
59			

Tables

Table 1: Studies included in the analysis

Reference	Author	Country	Studysite ^a	Method ^b	Loci ^c	Study duration (months)	Clustered + unique isolates	TB incidence (per 100,000)	TB/HIV co-infection	Prevalence of culture positivity	% culture positive typed	No. clusters	Max cluster size	НСЛ	Proportion clustering	Recent transmission (%)
[39]	Asgharzadeh, M	Azerbaijan	r	15	ο	12	156	26.0		94.6	98.7	22	5	0.9966	32.7	18.6
[40]	Allix-Beguec, C	Belgium	r	24	n	24	530	35.2	5.1	86.1	87.9	53	23		29.6	19.6
[41]	Allix-Beguec, C	Belgium	r	24,S	n	39	802	35.2	5.1	81.8	84.7	82			28.8	19.6
[34]	Oelemann, M	Germany	ci	24,S	n	12	154	12.7			100	11			22.1	14.9
[42]	Roetzer, A	Germany	r	24,S	n	48	277	3.2	0.09		100	18	22		27.1	20.6
[43]	Ojo, OO	Ireland	r	24,S	n	36	171	15.3	3.3	79.5	96.1	15	12	0.9996	27.5	18.7
[44]	Dymova, MA	Russia	r	15	0	3	98	94.0	3.8		100	8		0.9900	31.6	23.5
[45]	Bidovec-Stojkovic, U	Slovenia	со	24,S	n	12	196	10.6	0.04		100	29	6	0.9965	36.2	21.4
[46]	Alonso-Rodriguez, N	Spain	r	15	n	27	281	26.0	6		81.9		8		43.1	24.4
[35]	Evans, J	UK	r	15	0	48	4207	15.0	8.2	58.3	100	439			61.2	50.8
[47]	Hamblion, E	UK	r	24	n	9	964	44.9	8.2		100				37.0	
[48]	Mandal, S	UK	со	15	0	48	102		8.2	90.7	87.2	8	12		30.4	22.6
[49]	Sails, A	UK	r	15	0	102	332	18.3	8.2	33.9	100	42	13		42.8	30.1
[50]	Nikolayevsky, V	Ukraine	r	15	0	4	225	80.4	3.9	39.2	97.4	31		0.9700	60.4	46.7

^a ci=city, r=region, co=country

^b 15=15 MIRU-VNTR loci, 24=24 MIRU-VNTR loci, S=with Spoligotyping

For peer review only - http://bmjopen.bmj.com/site/about/guidelines.xhtml

^c o= old 12 MIRU loci (MIRU 2, 4, 10, 16, 20, 23, 24, 26, 27, 30, 31, 39, 40), n=new 12 MIRU loci (MIRU 10, 16, 26, 31, 40 + Mtub 04, 21, 39 + ETR A C + QUB 11b, 26)

^d estimates from the literature of the prevalence of TB/HIV co-infection reported in the study area

2			
3			
4			
5			
6			
7			
8			
9			
10			
11			
12			
13			
14			
15			
16			
17			
18			
19			
20			
21			
22			
23			
24			
25			
26			
27			
28			
29			
30			
31			
32			
33			
34			
35			
36			
37			
38			
39			
40			
41			
42			
43			
44			
45			
46			
47			
48			
49			
50			
51			
52			

59 60

1

Table 2: Univariable metaregression showing the coefficients for change in the proportion of clustering and the percentage of between-study variation explained by variables describing the study design and setting.

	n	$Coefficient^{\pi}$	CI	р	Adj R ^{2 ¥}
Study design					
Study duration (months)	14	0.003	-0.063, 0.069	0.919	-8.47
Prevalence of culture positivity	8	-0.913	-1.732, -0.094	0.034	49.36
% culture positive typed	14	0.161	-0.731, 1.053	0.701	-6.99
Study size	14	-4.462	-10.000, 1.076	0.105	14.89
Number of loci (ref 15 loci)					
24 loci	14	-0.282	-0.519, -0.045	0.023	34.1
Study setting					
TB incidence	13	0.082	-0.097, 0.22	0.334	0.04
TB/HIV co-infection	12	0.088	-0.087, 0.263	0.288	3.28
Maximum cluster size	9	0.137	-0.035, 0.309	0.101	26.91
% clusters with 2 cases	7	0.004	-0.007, 0.016	0.396	-2.39

^{π} Coefficients for the change in the proportion of clustering for each covariate. E.g. for a one-month increase in study duration, the proportion of clustering increases by 0.003.

^{*} The proportion of between-study variation explained by the univariate metaregression.

Figure Caption

Figure 1: Results of systematic search, screening and data extraction.



Appendix: Medline/Embase search strategy

1. (tubercle adj3 (bacillus or bacilli)).mp. [mp=title, original title, abstract, name of substance word, subject heading word, unique identifier]

2. ((mycobacterium or mycobacteria) adj3 (bovis or africanum or microti or canetti)).mp.

3. exp tuberculosis/ or mycobacterium tuberculosis/ or tuberculosis.mp. or tb.mp. or Mtb.mp. or "M tuberculosis complex".mp.

4. or/1-3

5. Minisatellite Repeats/ or Genotype/ or Interspersed Repetitive Sequences/ or DNA Fingerprinting/ or Bacterial Typing Techniques/

6. "miru".mp. [mp=title, original title, abstract, name of substance word, subject heading word, unique identifier]

7. "vntr".mp. [mp=title, original title, abstract, name of substance word, subject heading word, unique identifier]

8. (miru adj3 vntr).mp. [mp=title, original title, abstract, name of substance word, subject heading word, unique identifier]

9. (mycobacterial adj3 interspersed adj3 repetitive adj3 units).mp. [mp=title, original title, abstract, name of substance word, subject heading word, unique identifier]

10. (dna adj3 fingerprinting).mp. [mp=title, original title, abstract, name of substance word, subject heading word, unique identifier]

11. ((strain adj3 type) or (strain adj3 typing) or (strain adj3 types)).mp. [mp=title, original title, abstract, name of substance word, subject heading word, unique identifier]

12. ((molecular adj3 typing) or (molecular adj3 strain adj3 typ*)).mp. [mp=title, original title, abstract, name of substance word, subject heading word, unique identifier]

13. (genotype or genotyping or genotypes).ti,ab.

14. (minisatellite adj3 repeat*).mp. [mp=title, original title, abstract, name of substance word, subject heading word, unique identifier]

15. molecular epidemiology/mt or (molecular adj3 epidemiology).mp. [mp=title, original title, abstract, name of substance word, subject heading word, unique identifier]

16. or/5-15

17. exp disease outbreaks/ or (outbreak adj3 analysis).mp. or (outbreak adj3 investigation).mp. or (outbreak adj3 management).mp. or (tuberculosis adj3 outbreak).mp. [mp=title, original title, abstract, name of substance word, subject heading word, unique identifier]

18. exp contact tracing/ or (contact adj3 tracing).mp. or (contact* adj3 traced).mp. or (contact adj3 screen*).mp. [mp=title, original title, abstract, name of substance word, subject heading word, unique identifier]

19. exp case management/ or (case adj3 management).mp. [mp=title, original title, abstract, name of substance word, subject heading word, unique identifier]

20. exp Risk Factors/

21. (risk adj3 factor*).mp. [mp=title, original title, abstract, name of substance word, subject heading word, unique identifier]

22. exp Epidemiologic Factors/

23. infectious disease transmission.mp. or exp Disease Transmission, Infectious/

24. exp case management/ or (case adj3 management).mp. [mp=title, original title, abstract, name of substance word, subject heading word, unique identifier]

25. program evaluation/ or evaluation studies as topic/ or (program adj3 evaluation).mp. or (programme adj3 evaluation).mp. [mp=title, original title, abstract, name of substance word, subject heading word, unique identifier]

26. public health practice/ or (public adj3 health).mp. [mp=title, original title, abstract, name of substance word, subject heading word, unique identifier]

27. ((tuberculosis adj3 control) or (tb adj3 control)).mp. [mp=title, original title, abstract, name of substance word, subject heading word, unique identifier]

28. (molecular adj3 surveillance).mp. [mp=title, original title, abstract, name of substance word, subject heading word, unique identifier]

29. exp cluster analysis/ or (cluster* adj3 rate*).mp. or (cluster* adj3 growth).mp. or (cluster* adj3 analysis).mp. or (cluster adj3 investigation).mp. or (proportion adj3 cluster*).mp. or (molecular adj3 cluster*).mp. [mp=title, original title, abstract, name of substance word, subject heading word, unique identifier]

30. ((recent adj3 transmission) or (transmission adj3 event*) or (transmission adj3 rate*) or (chain adj3 transmission) or (transmission adj3 setting*)).mp. [mp=title, original title, abstract, name of substance word, subject heading word, unique identifier]

31. or/17-30

Page 17 of 19

58 59 60

1	
2	
3	32. 4 and 16
4	33. 32 and 31
5	34. limit 33 to yr="1998-Current"
6	35. limit 34 to english language
7 8	
9	So. difficiency
10	37. humans/
11	38. 36 not 37
12	39. 35 not 38
13	
14	
15	
16	
18	
10	
20	
21	
22	
23	
24	
25	
26	
27	
20 20	
30	
31	
32	
33	
34	
35	
36	
3/	
30 30	
40	
41	
42	
43	
44	
45	
40 47	
48	
49	
50	
51	
52	
53	
54	
55 56	
00 57	
51	



PRISMA 2009 Checklist

Section/topic	#	Checklist item	Reported on page #
TITLE			
Title	1	Identify the report as a systematic review, meta-analysis, or both.	1
ABSTRACT			
Structured summary	2	Provide a structured summary including, as applicable: background; objectives; data sources; study eligibility criteria, participants, and interventions; study appraisal and synthesis methods; results; limitations; conclusions and implications of key findings; systematic review registration number.	2
INTRODUCTION			
Rationale	3	Describe the rationale for the review in the context of what is already known.	3
Objectives	4	Provide an explicit statement of questions being addressed with reference to participants, interventions, comparisons, outcomes, and study design (PICOS).	3
METHODS			
Protocol and registration	5	Indicate if a review protocol exists, if and where it can be accessed (e.g., Web address), and, if available, provide registration information including registration number.	n/a
Eligibility criteria	6	Specify study characteristics (e.g., PICOS, length of follow-up) and report characteristics (e.g., years considered, language, publication status) used as criteria for eligibility, giving rationale.	4
Information sources	7	Describe all information sources (e.g., databases with dates of coverage, contact with study authors to identify additional studies) in the search and date last searched.	4
Search	8	Present full electronic search strategy for at least one database, including any limits used, such that it could be repeated.	14
Study selection	9	State the process for selecting studies (i.e., screening, eligibility, included in systematic review, and, if applicable, included in the meta-analysis).	4
Data collection process	10	Describe method of data extraction from reports (e.g., piloted forms, independently, in duplicate) and any processes for obtaining and confirming data from investigators.	4
Data items	11	List and define all variables for which data were sought (e.g., PICOS, funding sources) and any assumptions and simplifications made.	5
Risk of bias in individual studies	12	Describe methods used for assessing risk of bias of individual studies (including specification of whether this was done at the study or outcome level), and how this information is to be used in any data synthesis.	4
Summary measures	13	State the principal summary measures (e.g., risk ratio, difference in means).	5
Synthesis of results	14	Describe the methods of handling data and combining results of studies, if done, including measures of consistency (e.g., I^2) for each meta-analysis.	5
3		For peer review only - http://bmjopen.bmj.com/site/about/guidelines.xhtml Page 1 of 2	



PRISMA 2009 Checklist

Section/topic	#	Checklist item	Reported on page #
Risk of bias across studies	15	Specify any assessment of risk of bias that may affect the cumulative evidence (e.g., publication bias, selective reporting within studies).	4
Additional analyses	16	Describe methods of additional analyses (e.g., sensitivity or subgroup analyses, meta-regression), if done, indicating which were pre-specified.	5
2 RESULTS	•		
³ Study selection	17	Give numbers of studies screened, assessed for eligibility, and included in the review, with reasons for exclusions at each stage, ideally with a flow diagram.	5
6 Study characteristics	18	For each study, present characteristics for which data were extracted (e.g., study size, PICOS, follow-up period) and provide the citations.	12
⁸ Risk of bias within studies	19	Present data on risk of bias of each study and, if available, any outcome level assessment (see item 12).	12
0 Results of individual studies	20	For all outcomes considered (benefits or harms), present, for each study: (a) simple summary data for each intervention group (b) effect estimates and confidence intervals, ideally with a forest plot.	12
2 Synthesis of results	21	Present results of each meta-analysis done, including confidence intervals and measures of consistency.	14
4 Risk of bias across studies	22	Present results of any assessment of risk of bias across studies (see Item 15).	12
6 Additional analysis	23	Give results of additional analyses, if done (e.g., sensitivity or subgroup analyses, meta-regression [see Item 16]).	14
9 9 Summary of evidence 9	24	Summarize the main findings including the strength of evidence for each main outcome; consider their relevance to key groups (e.g., healthcare providers, users, and policy makers).	5
Limitations	25	Discuss limitations at study and outcome level (e.g., risk of bias), and at review-level (e.g., incomplete retrieval of identified research, reporting bias).	6
4 Conclusions	26	Provide a general interpretation of the results in the context of other evidence, and implications for future research.	6
Funding	27	Describe sources of funding for the systematic review and other support (e.g., supply of data); role of funders for the systematic review.	7
IO IO I1 <i>From:</i> Moher D, Liberati A, Tetzlaf I2 doi:10.1371/journal.pmed1000097	f J, Altm	an DG, The PRISMA Group (2009). Preferred Reporting Items for Systematic Reviews and Meta-Analyses: The PRISMA Statement. PLoS Med	6(6): e1000097
13		For more information, visit: <u>www.prisma-statement.org</u> .	
14		Page 2 of 2	

BMJ Open

Page 2 of 2

For peer review only - http://bmjopen.bmj.com/site/about/guidelines.xhtml

BMJ Open

Effect of study design and setting on tuberculosis clustering estimates using Mycobacterial Interspersed Repetitive Units-Variable Number Tandem Repeats (MIRU-VNTR): A systematic review

Journal:	BMJ Open
Manuscript ID:	bmjopen-2014-005636.R1
Article Type:	Research
Date Submitted by the Author:	25-Nov-2014
Complete List of Authors:	Mears, Jessica; University College London, Department of Infection and Population Health Abubakar, Ibrahim; University College London, Department of Infection and Population Health; Public Health England, Centre for Infectious Disease Surveillance and Control Cohen, Ted; Harvard School of Public Health, Harvard University, Division of Global Health Equity, Brigham and Women's Hospital and Department of Epidemiology McHugh, Timothy; Centre for Clinical Microbiology, Research Department of Infection, Royal Free Campus, University College London Sonnenberg, Pamela; University College London, Department of Infection and Population Health
Primary Subject Heading :	Research methods
Secondary Subject Heading:	Infectious diseases, Public health
Keywords:	EPIDEMIOLOGY, Tuberculosis < INFECTIOUS DISEASES, MOLECULAR BIOLOGY

SCHOLARONE[™] Manuscripts



BMJ Open

2		
3	1	Effect of study design and setting on tuberculosis clustering estimates using
4	2	Mycobacterial Interspersed Repetitive Units-Variable Number Tandem
5	2	
6	3	Repeats (MIRU-VNIR): A systematic review
7 8 0	4	Jessica Mears ¹ , Ibrahim Abubakar ^{1,2,3} , Theodore Cohen ⁴ , Timothy D McHugh ⁵ & Pam Sonnenberg ^{1,*}
9 10 11	5	¹ Department of Infection and Population Health, University College London
12 13	6	² Centre for Infectious Disease Surveillance and Control, Public Health England
14 15	7	³ Clinical Trials Unit, Medical Research Council, London
16 17	8	⁴ Division of Global Health Equity, Brigham and Women's Hospital and Department of Epidemiology,
18 19	9	Harvard School of Public Health, Harvard University
20 21 22	10	⁵ Centre for Clinical Microbiology, Department of Infection, University College London
22 23 24	11	*Corresponding author:
25 26	12	Dr Pam Sonnenberg
27 28	13	3 rd Floor Mortimer Market Centre
29 30	14	University College London
31 32 33	15	London WC1E 6JB
34 35	16	p.sonnenberg@ucl.ac.uk
36 37	17	
38 39	18	Word count: 2439
40		
41		
42 13		
43 11		
44 45		
46		
47		
48		
49		
50		
51		
52		
53		
54		
55		
56		
57		
58		1
59		
60		

19 Abstract

- 20 Objectives: To systematically review the evidence for the impact of study design and setting on the
- 21 interpretation of TB transmission using clustering derived from Mycobacterial Interspersed
- 22 Repetitive Units Variable Number Tandem Repeats (MIRU-VNTR) strain typing.

23 Data sources: Medline, Embase, CINHAL, Web of Science and Scopus were searched for articles

- 24 published before 21st October 2014.
- 25 Review methods: Studies in humans that reported the proportion of clustering of TB isolates by
- 26 MIRU-VNTR were included in the analysis. Univariable meta-regression analyses were conducted to
- 27 assess the influence of study design and setting on the proportion of clustering.
- 28 Results: The search identified 27 eligible articles reporting clustering between 0% and 63%. The
- 29 number of MIRU-VNTR loci typed, requiring consent to type patient isolates (as a proxy for sampling
 - 30 fraction), the TB incidence and the maximum cluster size explained 14%, 14%, 27% and 48%,
 - 31 respectively, and had a significant association with the proportion of clustering .
 - 32 Conclusions: Although MIRU-VNTR typing is being adopted worldwide there is a paucity of data on
 - 33 how study design and setting may influence estimates of clustering. We have highlighted study
 - 34 design variables for consideration in the design and interpretation of future studies.

36 Strengths and Limitations of Study

- This is a timely evaluation of the impact of study design on estimates of TB clustering using MIRU-VNTR strain typing because it has been incorporated into national typing services globally.
- The strength of this meta-analysis was limited by the lack of detail reported by the included studies, highlighting the need for better quality reporting in primary studies.

BMJ Open

44 Introduction

The introduction of molecular typing methods has improved our understanding of *Mycobacterium tuberculosis* (TB) transmission and has changed local and national control policies [1–5]. The proportion of cases that are clustered is often used to estimate the amount of ongoing transmission within the population, based on the assumption that cases with indistinguishable strain types are part of a chain of transmission. TB molecular typing methodology is changing rapidly and it is important that we better understand how to interpret the outputs and thus act.

TB molecular typing methods include Spoligotyping [6], insertion sequence *6110* (IS*6110*) restriction fragment length polymorphism (RFLP) analysis (the recent gold standard) [7], mycobacterial interspersed repetitive units-variable number tandem repeats (MIRU-VNTR) typing [8], and whole genome sequencing [9–11]. Published reviews have identified factors that might influence or bias clustering by IS*6110* RFLP [12,13]. No study has repeated this analysis using more up-to-date typing methods, which is important for understanding of the epidemiology of TB and to shape the application of molecular typing to improve TB control.

Published meta-analyses and modelling studies using IS*6110* RFLP data show that the proportion of clustering observed can be affected by 1) study design (affecting the proportion of eligible cases that are included in the study); 2) features of the typing method (such as the ability to type isolates with low copy numbers); and 3) study setting (such as characteristics of the study population). For example, the proportion of clustering increases when the fraction of the total data sampled increases [13–15] and when study duration increases [16].

MIRU-VNTR is currently the preferred method of molecular typing [17-21], and can be used together with Spoligotyping [8]. Relative to IS6110 RFLP, MIRU-VNTR does not have to exclude isolates with a low IS6110 copy number, has a faster turnaround time, is high throughput and the numeric strain types are more easily compared. MIRU-VNTR strain typing is increasingly being adopted worldwide [1,22–27], yet unlike IS6110 RFLP, the evidence for the interpretation of the findings such as the impact of study design and setting on clustering have not been reviewed. Although the two typing methods have been shown to have a similar discriminatory value, the markers evolve independently and at different rates, resulting in a difference in clustering between the two methods [28]. This suggests that there could be differences in the way study design, typing method and setting affects clustering by the two methods. We conducted a systematic review to assess the evidence for the impact of study design and setting on the interpretation of TB

transmission using clustering derived from MIRU-VNTR strain typing – as has been shown using
IS*6110* RFLP typing.

77 Methods

Five electronic databases were searched (EMBASE, ISI Web of Science, CINHAL, Scopus and Medline (Ovid)) up to 20th October 2014. The search strategy combined the following terms with Boolean operators: Tuberculosis, strain typing, and transmission. The search was limited to studies using the standard MIRU-VNTR method [8], in humans only, and in English.

All titles and abstracts from each of the searches were examined. The full text of each paper was obtained and reviewed if the study reported MIRU-VNTR strain typing of *M.tuberculosis* complex isolates with at least 15 of the standardised 24 loci (ETR A, B, C, D, E; MIRU 2, 10, 16, 20, 23, 24, 26, 27, 39, 40; VNTR 424, 1955, 2163b, 2347, 2401, 3171, 3690, 4052, 4156) [8,29,30].

Studies using fewer than 15 loci were not included because the level of discrimination is inadequate for epidemiological use (n=121) [8]. Studies that used loci different to the standardised 15 and 24 set were not included in the analysis in order to reduce the heterogeneity between studies (n=19). All publication types were included in this first screen to ensure that no relevant data were missed.

Reviews, letters, editorials, outbreaks or case reports (n=103) were excluded in the second screen.
Studies that used incomplete sampling (e.g. random samples, studies using subsets of populations
such as multidrug-resistant patients) (n=47) and studies that had a sample size of less than 50 (n=4)
were also excluded.

A reviewer (JM) extracted the following data items from all included studies using a form developed in Excel (Microsoft 2010): publication details (year, authors, study country), study details (study duration, loci typed, secondary typing method, study population, whether participant consent was required (a characteristic of the study design that was used as proxy for sampling fraction, assuming that where consent was required the sampling fraction was low)), the number of clustered and unique isolates, and the covariates of interest: the maximum size of clusters; the proportion of clusters containing two cases; the proportion of the population that was culture positive; the proportion of culture positive isolates typed; risk factors for clustering; and the Hunter Gaston Discriminatory Index (HGDI) [31]). IA extracted data from 10% of the papers for external validity, disagreements were discussed and a consensus agreed upon.

BMJ Open

The main outcome measure – the proportion of TB isolates clustered by MIRU-VNTR strain typing –
 was calculated as the number of clustered isolates/number of clustered+unique isolates. Where
 there were uncertainties JM consulted with IA

Authors were contacted if TB incidence rate was not reported. Where no response was received WHO country estimates of TB incidence for the study year were used [32]. As so few studies reported the proportion coinfected with TB/HIV, these estimates for the study country were taken from an EU-wide survey and WHO country profiles.[33,34] Due to poor recording of the sampling fraction (the number of isolates typed/ the total number of culture positive TB cases diagnosed during the study period (n=19)), whether the study required the consent of participants (yes/no) was included as a proxy for (high/low) sampling fraction. The risk of bias within each study was assessed using the STROME-ID checklist. [35]

Data were analysed in Stata 12. Where studies reported data from more than one set of loci, the method with the highest discriminatory value was included (i.e. MIRU-VNTR 24 would be chosen over MIRU-VNTR 15, and MIRU-VNTR 15 plus Spoligotyping would be chosen over MIRU-VNTR 15 alone) (n=8). This review was not concerned with summary measures of clustering, but factors that influenced clustering; therefore articles must have included at least one of the covariates. Continuous variables were transformed where the distribution was skewed. The proportion clustered was transformed using the Freeman Tukey transformation [36]. Study heterogeneity was assessed using a forest plot and the chi² test of heterogeneity. Univariable meta-regression analyses were carried out to determine the effect of the study design covariates on the proportion of clustered isolates. All covariates in the analysis were hypothesised to influence the proportion clustered *a priori*.

Sensitivity analyses were conducted to see the effect of removing studies reporting 0% clustering, with only extra-pulmonary TB cases, only *M.bovis* cases, studies using the 'old 12' MIRU loci as part of their 15 loci, and studies assessed as having a high likelihood of bias (STROME-ID score less than 20).

130 Results

The search identified 7274 references resulting in 27 studies (25 journal articles and 2 conference abstracts) included after deduplication and title/abstract/full text screening (Figure 1). The main characteristics of the included studies are shown in Table 1. Studies were published between 2007 and 2014 and the clustering reported varied from 0% [37] to 62.8% [38]. In all studies, clustered

isolates were defined as having identical strain types based on the MIRU-VNTR loci typed, with or without Spoligotyping. 17 studies included isolates from newly diagnosed TB cases, three studies reported including isolates from new and chronic cases of TB, and seven did not report this information. In addition, ten studies did not include repeat isolates from the same patient, one study included a repeat isolate from one patient, and the remaining 17 did not report whether repeat isolates were included or not. Furthermore, four studies included isolates with missing loci in the cluster analysis, whereas four excluded isolates with missing loci, and the remaining 20 did not report how they dealt with missing loci. The number of studies reporting each variable of interest is shown in Table 2.

A forest plot shows the spread of clustering reported by number of loci and additional typing
method (Figure 2). Significant heterogeneity was identified between the studies (p<0.001),
suggesting that a meta-regression would be an appropriate analysis.

The univariable meta-regression shows evidence for the proportion of clustering to decrease as the number of MIRU-VNTR loci typed increased from 15 to 24 (p=0.04; Table 3), accounting for 14% of the between study variation, and to increase when the study participants consented to being included in the study (p=0.03), accounting for 14% of the between study variation. The proportion of clustering increased as the TB incidence in the population increased (p=0.007, Adj R² = 26.7). There was also evidence for the proportion of clustering to increase as the maximum cluster size increased (p=0.001), accounting for 48% of between study variation. There was no evidence of the other study design or study setting variables significantly influencing the proportion clustered. Though non-significant (p>0.05), the TB/HIV coinfection rate in the population explained 2% of the between study variation. Too few studies included information on the proportion of clusters containing two cases, proportion of the study sample with previous TB or with pulmonary TB, and the proportion of the population with culture positive TB, so these could not be included in the analysis (Table 2).

Sensitivity analyses to examine the effect of excluding studies reporting 0% clustering,[37] only M.bovis cases, [39] studies using the 'old 12' MIRU loci, [39–44] and studies assessed as having a high risk of bias, [37,45–48] did not generally change the results. The proportion of culture positive TB in the population remained insignificant but explained 2.6% of the between study variation when excluding 0% clustering (p=0.278 and Adj R²=2.62). Similarly, the proportion of culture positive TB in the population remained insignificant but explained 2.6% of the between study variation when excluding studies with the highest risk of bias (p=0.278 and Adj $R^2=2.62$). The number of loci typed became non-significant, but explained 9.6% and 10.5% of the between study variation when

BMJ Open

167 excluding studies using the 'old 12' loci and the highest risk of bias, respectively (p=0.106, Adj
168 R²=9.63; p=0.111, Adj R²=10.51, respectively).

169 Discussion

This review identified 27 studies that met the inclusion criteria. We illustrate that the interpretation
of studies using MIRU-VNTR to estimate clustering is subject to bias relating to study design and
setting; however, there were insufficient data available to fully explore this impact.

As expected, we found that the proportion of clustering decreased with a greater number of MIRU-VNTR loci typed, with increasing TB incidence and with increasing maximum cluster size. We found that requiring consent to type patient isolates reduced the proportion of clustering, which is expected, given that the sampling fraction would be lower in these studies.

The other study design variables included in this analysis, such as study duration, did not significantly influence the proportion of isolates that were clustered, contrary to previous findings [12]. This is likely to be because of a lack of good quality evidence: of the 27 studies that met the inclusion criteria for the review, none reported all the variables of interest, reducing the power of the analysis and precluding multivariable meta-regression (Table 2). Importantly, key details of cluster analyses were not reported consistently across the studies, such as whether repeat isolates from the same patients were included, or typing profiles with missing loci were included, introducing new, unmeasured biases. In addition, the range of the variables may have been too limited to show any impact on clustering estimates. For example, the proportion of culture positive isolates typed ranged from 34.5% to 100%, with 17 of the 19 studies reporting this variable from 81.9% to 100%. Furthermore, most of the studies (17/27=63%) were from low TB burden settings and therefore may be reflecting the rate at which imported cases have matching strain types by chance, rather than rates of recent transmission.

The sensitivity analysis suggested that, when excluding the studies with the greatest risk of bias, the culture-positivity in the population might explain a small amount of the between study variation. This is counterintuitive and not consistent with estimates of the influence of sampling on the proportion of clustering using IS6110 RFLP typing [49]. This may reflect the relationship between TB burden and resource poor/rich settings and the consequent availability of culture diagnostic laboratory services; i.e. in resource poor settings where there is a high burden of TB (and, therefore, high rates of clustering) the prevalence of culture positive TB cases is low. The finding may also be due to chance, with only 14 studies included in the analysis of this variable. In the sensitivity analysis

excluding studies that used the 'old 12' loci, the effect of the number of loci typed becomes nonsignificant. This is likely because studies using the 'old 12' accounted for six out of ten studies
reporting 15 loci, reducing the number of studies and the power of the model.

This study is a timely evaluation of the impact of study design on estimates of TB clustering using MIRU-VNTR strain typing because it has been incorporated into national typing services globally [23,50]. The findings are relevant where strain typing is used to evaluate TB control systems across different settings because the proportion of clustering is influenced by the number of loci typed, the TB incidence and the maximum cluster size. Given that strain typing methods are advancing beyond MIRU-VNTR typing and that the application of whole genome sequencing to TB control and public health strategies has been demonstrated [9–11,51], it is important that the biases in the analysis of such methods are explored and compared. Understanding how to design and compare research studies for public health will greatly improve the benefit gained from newer technologies.

The strength of this meta-analysis was limited by (a lack of) detail reported by the included studies. This review has highlighted the need for better quality reporting in primary studies to enable future reviews to be more robust. Recently published standards for reporting of molecular epidemiology for infectious diseases should improve the quality of reporting.[35] This review is further limited by our inability to access 58 of the title/abstract screened articles for full text screening.

The use of TB strain typing as a public health tool in TB control programmes is increasing globally. We have identified a lack of good quality studies that can contribute to our understanding in interpreting the molecular typing of TB. We have also shown that the proportion of clustering derived from MIRU-VTNR typing is influenced by the number of loci typed, whether consent is required to type isolates, TB incidence in the study setting, and the maximum cluster size, highlighting these as important considerations in the design and interpretation of future studies.

221 Conflict of interest

- 222 Nothing to declare.
- 223 Acknowledgements

We would like to acknowledge Ross Harris from the Statistics Unit at Public Health England for hisadvice on meta-regression.

226 Author contributions

228

229

230

231

232

233

234

235

236

237

238

239

240

241

242

243

244

245

246 247

248

249

250

251

252

253

254

255

256

257

258

259

260

Funding

Ethics

Data sharing

References

1.

2.

3.

4.

5.

6.

7.

No additional data are available

Dis 7: S463-470.

1 2

BMJ Open

All authors made substantial contributions to the conception and design of the review, and the

analysis and interpretation of data. JM drafted the article and PS, IA, TM and TC revised it critically

JM is funded through a Public Health England and University College London Impact Studentship. IA

Lambregts-van Weezenbeek CSB, Sebek MMGG, van Gerven PJHJ, de Vries G, Verver S, et al.

Borgdorff MW, van den Hof S, Kremer K, Verhagen L, Kalisvaart N, et al. (2010) Progress

Kik SV, Verver S, Van Soolingen D, De Haas PEW, Cobelens FG, et al. (2008) Tuberculosis Outbreaks Predicted by Characteristics of First Patients in a DNA Fingerprint Cluster. Am J

Small PM, McClenny NB, Singh SP, Schoolnik GK, Tompkins LS, et al. (1993) Molecular strain

De Vries G, van Hest RAH, Richardus JH (2007) Impact of mobile radiographic screening on

tuberculosis among drug users and homeless persons. Am J Respir Crit Care Med 176: 201-

Kamerbeek J, Schouls L, Kolk A, van Agterveld M, van Soolingen D, et al. (1997) Simultaneous

Van Embden JD, Cave MD, Crawford JT, Dale JW, Eisenach KD, et al. (1993) Strain identification

of Mycobacterium tuberculosis by DNA fingerprinting: recommendations for a standardized

detection and strain differentiation of Mycobacterium tuberculosis for diagnosis and

typing of Mycobacterium tuberculosis to confirm cross-contamination in the mycobacteriology

laboratory and modification of procedures to minimize occurrence of false-positive cultures. J

Respir Crit Care Med 178: 96–104. doi:10.1164/rccm.200708-1256OC.

(2003) Tuberculosis contact investigation and DNA fingerprint surveillance in The Netherlands:

6 years' experience with nation-wide cluster feedback and cluster monitoring. Int J Tuberc Lung

towards tuberculosis elimination: secular trend, immigration and transmission. Eur Respir J 36:

for important intellectual content. All authors approved the final version for publication.

Ethical approval was not required as this review analyses data that is in the public domain.

is funded through a NIHR Senior Research Fellowship.

339-347. doi:10.1183/09031936.00155409.

Clin Microbiol 31: 1677–1682.

207. doi:10.1164/rccm.200612-1877OC.

epidemiology. J Clin Microbiol 35: 907–914.

methodology. J Clin Microbiol 31: 406–409.

2	
3	
4	
5	
6	
7	
<i>'</i>	
8	
9	
10	
11	
10	
12	
13	
14	
15	
16	
17	
10	
18	
19	
20	
21	
21	
22	
23	
24	
25	
26	
20	
21	
28	
29	
30	
31	
22	
32	
33	
34	
35	
36	
27	
31	
38	
39	
40	
41	
12	
42	
43	
44	
45	
46	
10	
47	
48	
49	
50	
51	
52	
52	
53	
54	
55	
56	
57	
57	
58	
59	
60	

1			
2	261	Q	Supply P. Alliy C. Leciean S. Cardoso-Oelemann M. Rüsch-Gerdes S. et al. (2006) Proposal for
4	262	0.	standardization of ontimized mycohacterial interspersed renetitive unit-variable-number
5	263		tandem repeat typing of Mycobacterium tuberculosis. J Clin Microbiol 44: 4498–4510.
6	263		doi:10.1128/ICM 01392-06
7	201		
8	265	9.	Schürch AC, van Soolingen D (2011) DNA fingerprinting of Mycobacterium tuberculosis: From
9	266		phage typing to whole-genome sequencing. Infect Genet Evol. Available:
10	267		http://www.ncbi.nlm.nih.gov/pubmed/22067515. Accessed 13 March 2012.
11			
12	268	10.	Gardy JL, Johnston JC, Ho Sui SJ, Cook VJ, Shah L, et al. (2011) Whole-genome sequencing and
13	269		social-network analysis of a tuberculosis outbreak. N Engl J Med 364: 730–739.
15	270		doi:10.1056/NEJMoa1003176.
16			
17	271	11.	Walker TM, Ip CL, Harrell RH, Evans JT, Kapatai G, et al. (2013) Whole-genome sequencing to
18	272		delineate Mycobacterium tuberculosis outbreaks: a retrospective observational study. Lancet
19	273		Infect Dis 13: 137–146. doi:10.1016/S1473-3099(12)70277-3.
20			
21	274	12.	Houben RMGJ, Glynn JR (2009) A systematic review and meta-analysis of molecular
22	275		epidemiological studies of tuberculosis: development of a new tool to aid interpretation.
23	276		Tropical Medicine & International Health 14: 892–909. doi:10.1111/j.1365-3156.2009.02316.x.
24			
25	277	13.	Fok A, Numata Y, Schulzer M, FitzGerald MJ (May) Risk factors for clustering of tuberculosis
26	278		cases: a systematic review of population-based molecular epidemiology studies [Review
27	279		Article]. The International Journal of Tuberculosis and Lung Disease 12: 480–492.
28			
29	280	14.	Borgdorff MW, Van Den Hof S, Kalisvaart N, Kremer K, Van Soolingen D (2011) Influence of
30	281		Sampling on Clustering and Associations With Risk Factors in the Molecular Epidemiology of
32	282		Tuberculosis. Am J Epidemiol. Available:
33	283		http://aje.oxfordjournals.org/content/early/2011/05/23/aje.kwr061. Accessed 29 March 2012.
34			
35	284	15.	Glynn JR, Bauer J, de Boer AS, Borgdorff MW, Fine PE, et al. (1999) Interpreting DNA fingerprint
36	285		clusters of Mycobacterium tuberculosis. European Concerted Action on Molecular
37	286		Epidemiology and Control of Tuberculosis. Int J Tuberc Lung Dis 3: 1055–1060.
38			
39	287	16.	Glynn JR, Crampin AC, Yates MD, Traore H, Mwaungulu FD, et al. (2005) The Importance of
40	288		Recent Infection with Mycobacterium tuberculosis in an Area with High HIV Prevalence: A
41	289		Long-Term Molecular Epidemiological Study in Northern Malawi. J Infect Dis 192: 480–487.
42	290		doi:10.1086/431517.
43			
44	291	17.	De Beer JL, Kremer K, Ködmön C, Supply P, van Soolingen D (2012) First Worldwide Proficiency
45	292		Study on Variable-Number Tandem-Repeat Typing of Mycobacterium tuberculosis Complex
40	293		Strains. Journal of Clinical Microbiology 50: 662–669. doi:10.1128/JCM.00607-11.
47 48			
40 49	294	18.	Maes M, Kremer K, van Soolingen D, Takiff H, de Waard JH (2008) 24-locus MIRU-VNTR
50	295		genotyping is a useful tool to study the molecular epidemiology of tuberculosis among Warao
51	296		Amerindians in Venezuela. Tuberculosis (Edinb) 88: 490–494. doi:10.1016/j.tube.2008.04.003.
52			
53	297	19.	Sougakott W (2011) Molecular epidemiology of multidrug-resistant strains of Mycobacterium
54	298		tuberculosis. Clinical Microbiology and Infection 17: 800–805. doi:10.1111/j.1469-
55	299		0691.2011.03577.x.
56			
57			
58			10
59			
60			

2			
3	300	20.	Weniger T, Krawczyk J, Supply P, Niemann S, Harmsen D (2010) MIRU-VNTRplus: a web tool for
4	301		polyphasic genotyping of Mycobacterium tuberculosis complex bacteria. Nucleic Acids Res 38:
5	302		W326–331. doi:10.1093/nar/gkq351.
6			
/ 0	303	21.	Supply P (2010) MIRU-VNTR typing: the new international standard for TB molecular
0 0	304		epidemiology Symposium of the Institut Pasteur de Tunisia.
10	205	22	Marcharles D. Davida (CAMA) de Harr DE Cabal AANA Marcharles (4000) AA davida
11	305	22.	van Soolingen D, Borgdorff MW, de Haas PE, Sebek MM, veen J, et al. (1999) Molecular
12	300		Unfact Dic 180: 726, 726, doi:10.1096/214020
13	307		J IIIIect DIS 180. 720–730. 001.10.1080/314930.
14	308	23	Cowan LS Diem L Monson T Wand P Temporado D et al. (2005) Evaluation of a two-sten
15	309	23.	approach for large-scale, prospective genotyping of Mycobacterium tuberculosis isolates in the
16	310		United States Clin Microbiol 43: 688–695, doi:10.1128/ICM 43.2.688-695.2005
1/	010		
19	311	24.	Centers for Disease Control and Prevention (2005) New CDC Program for Rapid Genotyping of
20	312		Mycobacterium Tuberculosis Isolates. JAMA 293: 2086–2086. doi:10.1001/jama.293.17.2086.
21			
22	313	25.	Bauer J, Kok-Jensen A, Faurschou P, Thuesen J, Taudorf E, et al. (2000) A prospective evaluation
23	314		of the clinical value of nation-wide DNA fingerprinting of tuberculosis isolates in Denmark. Int J
24	315		Tuberc Lung Dis 4: 295–299.
25			
26	316	26.	Bauer J, Yang Z, Poulsen S, Andersen AB (1998) Results from 5 years of nationwide DNA
21	317		fingerprinting of Mycobacterium tuberculosis complex isolates in a country with a low
20 20	318		incidence of M. tuberculosis infection. J Clin Microbiol 36: 305–308.
30	210	27	Zalnia Dava M. Daliah M. Erran D. Carli I (2002) Malagular anidamialamu of tubarnulasis in
31	319	27.	Zoinir-Dovc M, Poijak M, Erzen D, Sorii J (2003) Molecular epidemiology of tuberculosis in
32	320		Slovenia: results of a one-year (2001) nation-wide study. Scand J Infect Dis 35: 863–868.
33	321	28	Hanekom M. van der Snuv GD. Gev van Pittius NC. McEvov CRE. Hoek KGP. et al. (2008)
34	322	20.	Discordance between mycobacterial interspersed repetitive-unit-variable-number tandem-
35	323		repeat typing and IS6110 restriction fragment length polymorphism genotyping for analysis of
36	324		Mycobacterium tuberculosis Beijing strains in a setting of high incidence of tuberculosis. J Clin
37 38	325		Microbiol 46: 3338–3345. doi:10.1128/JCM.00770-08.
39			
40	326	29.	Supply P, Lesjean S, Savine E, Kremer K, van Soolingen D, et al. (2001) Automated high-
41	327		throughput genotyping for study of global epidemiology of Mycobacterium tuberculosis based
42	328		on mycobacterial interspersed repetitive units. J Clin Microbiol 39: 3563–3571.
43	329		doi:10.1128/JCM.39.10.3563-3571.2001.
44			
45	330	30.	Gopaul KK, Brown TJ, Gibson AL, Yates MD, Drobniewski FA (2006) Progression toward an
40 47	331		improved DNA amplification-based typing technique in the study of Mycobacterium
48	332		tuberculosis epidemiology. J Clin Microbiol 44: 2492–2498. doi:10.1128/JCM.01428-05.
49	222	21	Uniter DD. Coston MA (1088) Numerical index of the discriminatory chility of tuning systems
50	333	31.	Hunter PR, Gaston MA (1988) Numerical index of the discriminatory ability of typing systems:
51	334		an application of simpson's muck of diversity. J Clin Microbiol 26: 2465–2466.
52	335	32	WHO TB data (n d) WHO Available: http://www.who.int/tb/country/en/index.html
53	336	52.	Accessed 12 December 2012
54	550		
55 56			
57			
58			11
59			
60			

		BMJ Open
337	33.	Kruijshaar ME, Pimpin L, Abubakar I, Rice B, Delpech V, et al. (2011) The burden of TB-HIV in
338 339		the EU: how much do we know? A survey of surveillance practices and results. Eur Respir J 38: 1374–1381. doi:10.1183/09031936.00198310.
340 341	34.	World Health Organization (n.d.) WHO Tuberculosis Country Profiles. Available: http://www.who.int/tb/country/data/profiles/en/.
342	35.	Field N, Cohen T, Struelens MJ, Palm D, Cookson B, et al. (2014) Strengthening the Reporting of Molecular Enidemiology for Infortious Diseases (STROME ID); an extension of the STROPE
343 344		statement. The Lancet Infectious Diseases 14: 341–352. doi:10.1016/S1473-3099(13)70324-4.
345 346	36.	Freeman MF, Tukey JW (1950) Transformations Related to the Angular and the Square Root. Ann Math Statist 21: 607–611. doi:10.1214/aoms/1177729756.
347	37.	Guang-ming D, Zhi-guo Z, Peng-ju D, Qian Z, Li W, et al. (2013) Differences in the population of
348		genetics of Mycobacterium tuberculosis between urban migrants and local residents in Beijing,
349		China. Chinese Medical Journal 126: 4066–4071. doi:10.3760/cma.j.issn.0366-6999.20130216.
350	38.	Zmak L, Obrovac M, Katalinic Jankovic V (2014) First insights into the molecular epidemiology
351		of tuberculosis in Croatia during a three-year period, 2009 to 2011. Scandinavian Journal of
352		Infectious Diseases 46: 123–129.
353	39.	Mandal S, Bradshaw L, Anderson LF, Brown T, Evans JT, et al. (2011) Investigating transmission
354		of Mycobacterium bovis in the United Kingdom in 2005 to 2008. J Clin Microbiol 49: 1943–
355		1950. doi:10.1128/JCM.02299-10.
356	40.	Asgharzadeh M, Kafil HS, Roudsary AA, Hanifi GR (2011) Tuberculosis transmission in
357		Northwest of Iran: using MIRU-VNTR, ETR-VNTR and IS6110-RFLP methods. Infect Genet Evol
358		11: 124–131. doi:10.1016/j.meegid.2010.09.013.
359	41.	Dymova MA, Liashenko OO, Poteiko PI, Krutko VS, Khrapov EA, et al. (2011) Genetic variation
360 361		of Mycobacterium tuberculosis circulating in Kharkiv Oblast, Ukraine. BMC Infect Dis 11: 77. doi:10.1186/1471-2334-11-77.
262	40	Sails AD Derrott A Corginson C Magoe IC Maynord D at al (2011) Malagular anidamialagy of
302	42.	Salis AD, Barrett A, Sarginson S, Magee JG, Maynard P, et al. (2011) Molecular epidemiology of Mycobactorium tuborculosis in East Lancachiro 2001, 2000, Thorax 66, 700, 712
264		doi:10.1126/thy 2011.158881
504		u01.10.1130/U1X.2011.138881.
365	43.	Evans J (2010) Analysis of prevalent Mycobacterium tuberculosis strains in the United
366		Kingdom: detection, distribution and expansion of MIRU-VNTR profiles containing high
367		numbers of isolates. European Society of Clinical Microbiology and Infectious Diseases. Vienna,
368		Austria.
369	44.	Nikolayevskyy VV, Brown TJ, Bazhora YI, Asmolov AA, Balabanova YM, et al. (2007) Molecular
370		epidemiology and prevalence of mutations conferring rifampicin and isoniazid resistance in
371		Mycobacterium tuberculosis strains from the southern Ukraine. Clin Microbiol Infect 13: 129-
372		138. doi:10.1111/j.1469-0691.2006.01583.x.
373	45.	L. Z, M. O, V. KJ (2014) First insights into the molecular epidemiology of tuberculosis in Croatia
374		during a three-year period, 2009 to 2011. Scandinavian Journal of Infectious Diseases.
		12
		12
	337 338 339 340 341 342 343 344 345 346 347 348 349 350 351 352 353 354 355 356 357 358 359 360 361 362 363 364 365 366 367 368 369 370 371 372 373	337 33. 338 34. 340 34. 341 35. 342 35. 343 36. 344 37. 345 36. 347 37. 348 37. 349 37. 350 38. 351 39. 355 40. 355 40. 355 40. 355 41. 360 361 361 42. 363 43. 364 43. 365 43. 366 367 368 43. 369 44. 370 371 373 45.

3	375	46.	Roetzer A, Schuback S, Diel R, Gasau F, Ubben T, et al. (2011) Evaluation of Mycobacterium
4	376		tuberculosis typing methods in a 4-year study in Schleswig-Holstein, Northern Germany. J Clin
5	377		Microbiol 49: 4173–4178. doi:10.1128/JCM.05293-11.
6			
7	378	47.	Dymova MA, Kinsht VN, Cherednichenko AG, Khrapov EA, Svistelnik AV, et al. (2011) Highest
8	379		prevalence of the Mycobacterium tuberculosis Beijing genotype isolates in patients newly
9	380		diagnosed with tuberculosis in the Novosibirsk oblast Russian Federation Med Microbiol 60:
10	201		
11	201		1005-1009. 001.10.1039/JIIIII.0.027995-0.
12	202	40	Alexan Deditions, N. Martíne, Linda M. Cínster, M. Hanner, M. D. S. Colt, et al. (2000)
13	382	48.	Alonso-Rodriguez N, Martinez-Lirola M, Sanchez ML, Herranz M, Penafiel T, et al. (2009)
14	383		Prospective universal application of mycobacterial interspersed repetitive-unit-variable-
15	384		number tandem-repeat genotyping to characterize Mycobacterium tuberculosis isolates for
16	385		fast identification of clustered and orphan cases. J Clin Microbiol 47: 2026–2032.
17	386		doi:10.1128/JCM.02308-08
18			
10	387	10	Glypp JR Vyopycky E Fine PEM (1999) Influence of Sampling on Estimates of Clustering and
19	207	45.	Becont Transmission of Muschasterium tubersulasis Derived from DNA Eingerprinting
20	200		
21	389		Techniques. American Journal of Epidemiology 149: 366 –371.
22			
23	390	50.	TB Strain Typing Project Board HPA (2011) TB Strain Typing Cluster Investigation Handbook for
24	391		Health Protection Units 1st Edition. Available:
25	392		https://hpaintranet.hpa.org.uk/Content/ProgrammesProjects/HPAProgrammes/HPAKeyHealth
26	393		ProtectionProgrammes/Respiratory/TB/StrainTyping/. Accessed 30 November 2011.
27			
28	394	51	Walker TM Monk P. Grace Smith F. Peto TFA (2013) Contact investigations for outbreaks of
29	205	51.	Mycohacterium tuberculosis: advances through whole genome sequencing. Clin Microhiol
30	206		Infact doi:10.1111/1460.0601.12192
31	590		IIIect. doi.10.1111/1409-0091.12165.
32	207	52	C. do H. H. K. K. D. M. Coll, or N. Marste D. Circular (17) (2014) To see which as wise of
33	397	52.	Gurjav U, Jeits P, Miccallum N, Marais BJ, Sintchenko V (2014) Temporal dynamics of
34	398		Mycobacterium tuberculosis genotypes in New South Wales, Australia. BMC infectious
35	399		diseases 14: 455–455. doi:10.1186/1471-2334-14-455.
36			
37	400	53.	Allix-Béguec C, Fauville-Dufaux M, Supply P (2008) Three-year population-based evaluation of
38	401		standardized mycobacterial interspersed repetitive-unit-variable-number tandem-repeat
39	402		typing of Mycobacterium tuberculosis. J Clin Microbiol 46: 1398–1406.
40	403		doi:10.1128/JCM.02089-07.
41			
42	404	54.	Allix-Béguec C. Supply P. Wanlin M. Bifani P. Fauville-Dufaux M (2008) Standardised PCR-based
43	405	• · ·	molecular enidemiology of tuberculosis Fur Respir I 31: 1077–1084
44	405		doi:10 1192/00021026 00052207
45	400		uui.10.1185/05051550.00055507.
46	407		Tuite AD Cuthrie II. Alexander DC Whelen MC Lee D et al. (2012) Enidemiological evaluation
47	407	55.	Tuite AR, Guthne JL, Alexander DC, Whelan WS, Lee B, et al. (2013) Epidemiological evaluation
48	408		of spatiotemporal and genotypic clustering of mycobacterium tuberculosis in Ontario, Canada.
49	409		International Journal of Tuberculosis and Lung Disease 17: 1322–1327.
5 0			
50	410	56.	Tessema B, Beer J, Merker M, Emmrich F, Sack U, et al. (2013) Molecular epidemiology and
52	411		transmission dynamics of Mycobacterium tuberculosis in Northwest Ethiopia: new
52	412		phylogenetic lineages found in Northwest Ethiopia. Bmc Infectious Diseases 13: 131.
55	413		doi:10 1186/1471-2334-13-131
04 55	.10		
55			
56			
5/			
58			13
59			
60			

			BMJ Open
1			
2 3	414	57.	Smit PW, Haanpera M, Rantala P, Couvin D, Lyytikainen O, et al. (2013) Molecular
4	415		Epidemiology of Tuberculosis in Finland, 2008-2011. Plos One 8: e85027.
5	416		doi:10.1371/journal.pone.0085027.
6			
7	417	58.	Oelemann MC, Diel R, Vatin V, Haas W, Rüsch-Gerdes S, et al. (2007) Assessment of an
8	418		optimized mycobacterial interspersed repetitive- unit-variable-number tandem-repeat typing
9	419		system combined with spoligotyping for population-based molecular epidemiology studies of
10	420		tuberculosis. J Clin Microbiol 45: 691–697. doi:10.1128/JCM.01393-06.
12		-0	
13	421	59.	Ojo OO, Sheehan S, Corcoran DG, Nikolayevsky V, Brown T, et al. (2010) Molecular
14	422		epidemiology of Mycobacterium tuberculosis clinical isolates in Southwest Ireland. Infect
15	423		Genet Evol 10: 1110–1116. doi:10.1016/j.meegid.2010.07.008.
16	121	60	Aleksic F. Merker M. Cox H. Reiher B. Sekawi Z. et al. (2013) First Molecular Enidemiology Study
1/	424	00.	of Mycobacterium tuberculosis in Kiribati. PLoS ONE 8. Available:
10	425		http://www.scopus.com/inward/record.url?eid=2-s2.0-
20	420		84873163328& nartnerID=40&md5=3994b8e5638129b621abc4d7d6d5e3b8
20	727		040751055284partnenD=404ma5=5554b0c5050125002100c4070005c500.
22	428	61.	De Beer JL, van Ingen J, de Vries G. Erkens C. Sebek M, et al. (2013) Comparative study of
23	429		IS6110 restriction fragment length polymorphism and variable-number tandem-repeat typing
24	430		of Mycobacterium tuberculosis isolates in the Netherlands, based on a 5-year nationwide
25	431		survey. J Clin Microbiol 51: 1193–1198. doi:10.1128/JCM.03061-12.
26			
27	432	62.	Varghese B, Supply P, Shoukri M, Allix-Beguec C, Memish Z, et al. (2013) Tuberculosis
28	433		Transmission among Immigrants and Autochthonous Populations of the Eastern Province of
29	434		Saudi Arabia. PLoS ONE 8. Available: http://www.scopus.com/inward/record.url?eid=2-s2.0-
31	435		84885784886&partnerID=40&md5=4fdbf4015a999a9fcd1a1c31207a75a2.
32			
33	436	63.	Lim LK-Y, Sng LH, Win W, Chee CB-E, Hsu LY, et al. (2013) Molecular Epidemiology of
34	437		Mycobacterium tuberculosis Complex in Singapore, 2006-2012. Plos One 8: e84487.
35	438		doi:10.1371/journal.pone.0084487.
36	420	64	Bidevec Steikevic II. Zelniz Deve M. Supply P. (2011) One year nationwide evaluation of 24
37	439	64.	Bidovec-Stojković U, Zoinir-Dovć W, Supply P (2011) One year hationwide evaluation of 24-
38	440		10Cus MiRO-VNTR genotyping on Slovenian Mycobacterium tuberculosis isolates. Respir Med
39 40	441		105 Suppl 1. 307–75. 001.10.1010/30954-0111(11)/0014-2.
41	442	65.	Jonsson J., Hoffner S., Berggren J., Bruchfeld J., Ghebremichael S., et al. (2014) Comparison
42	443		between RFLP and MIRU-VNTR genotyping of mycobacterium tuberculosis strains isolated in
43	444		stockholm 2009 to 2011. PLoS ONE. Available:
44	445		http://www.plosone.org/article/fetchObiect.action?uri=info%3Adoi%2F10.1371%2Fiournal.po
45	446		ne.0095159&representation=PDF.
46	-		
47	447	66.	Muwonge A, Malama S, Johansen TB, Kankya C, Biffa D, et al. (2013) Molecular Epidemiology,
48	448		Drug Susceptibility and Economic Aspects of Tuberculosis in Mubende District, Uganda. PLoS
49 50	449		ONE 8. Available: http://www.scopus.com/inward/record.url?eid=2-s2.0-
51	450		84878608813&partnerID=40&md5=babbd6d006ca64e327fb19e01b6bc697.
52			
53	451	67.	Hamblion EL, Wynne-Edwards E, Anderson C, Anderson SR (2011) A summary of strain typing
54	452		and clustering of TB in London in 2010 and an analysis of the associated risk factors. Thorax 66:
55	453		A88–A89. doi:10.1136/thoraxjnl-2011-201054c.50.
56			
57			
58			14
60 60			
00			

2			
3	454	68.	Hang NTL, Maeda S, Lien LT, Thuong PH, Hung NV, et al. (2013) Primary Drug-Resistant
4	455		Tuberculosis in Hanoi, Viet Nam: Present Status and Risk Factors, Plos One 8: UNSP e71867.
5	156		doi:10.1371/journal.none.0071867
6	450		
7			
7	457		
8			
9			
10			
11			
12			
13			
14			
15			
10			
10			
17			
18			
19			
20			
21			
22			
23			
24			
27 25			
25			
26			
27			
28			
29			
30			
31			
32			
33			
34			
35			
36			
30			
37			
38			
39			
40			
41			
42			
43			
44			
45			
46			
40			
47			
4ð			
49			
50			
51			
52			
53			
54			
55			
56			
50			
57			
58			15
59			
60			
Tables

Table 1: The study setting and design characteristics of the included articles

Ref		Study se	tting							Stu	udy design				Risk of bias ^d	Clustering (%) ^e
	Study area and country	TB incidence (per 100,000)	TB/HIV (per 100,000) ^a	Previous TB treatment (%)	Pulmonary TB (%)	Maximum cluster size	Clusters of size 2 (%)	Study duration (months)	Study size (clustered + unique isolates)	Culture positive in study population (%)	Culture positive isolates typed (%)	Typing method b	Loci typed $^{\rm c}$	Consent required		
[52]	New South Wales, Australia	6.7	0.2	0.0	63.7		b .	36	1128			m24	Ν	no	low	20.1
[40]	Tabriz and Orumieh, Azarbaijan	26.0		5.2	87.0	5	81.8	12	156		94.5	m15	0	no	low	32.7
[53]	Brussels-Capital Region, Belgium	35.2	5.1	10.8		23	64.2	24	530	86.1	87.9	m24	Ν	no	low	29.6
[54]	Brussels-Capital Region, Belgium	35.2	5.1		100			39	802	81.8	84.7	m24s	Ν	no	low	28.8
[55]	Ontario, Canada	4.8	0.4			18	58.8	65	2016			m24s	Ν	no	low	23.1
[37]	Changping District, Beijing, China	•	0.3		100	0		30	318	31.5	94.6	m24	Ν	no	high	0.0
[38]	Croatia	19.0	0.1			45	48.3	36	1587			m15	Ν	no	high	62.8
[56]	Amhara region, Northwest Ethiopia		24.0	17.6	100	13		5	244			m24	N	yes	low	45.1
[57]	Finland	5.0	0.0			20		48	1048	75.4	99.4	m15s		no	low	33.9
[58]	Hamburg, Germany	12.7					45.5	12	154	78.2	91.1	m24s	N	no	low	22.1
[46]	Schleswig-Holstein, Germany	3.2	0.1			22	44.4	48	277			m24s	N	no	high	27.1
[59]	South West Ireland	15.3	3.3		82.7	12		36	171	79.5	96.1	m24s	Ν	no	low	27.5
[60]	South Tawara, Kiribati	370.0		4.1	100	25	55.6	24	73	45.4	98.6	m24s	Ν	yes	low	75.3
[61]	Netherlands	6.5	0.2				57.2	60	3978		100.1	m24	Ν	no	low	46.7
[41]	Kharkiv, Russia	94.0	3.8	63.3	100	10	50.0	3	98		100	m15	0	yes	high	31.6
[62]	Eastern province, Saudi Arabia	4.0			73.1	24	19.0	24	522			m24s	Ν	no	low	40.2
							16									

For peer review only - http://bmjopen.bmj.com/site/about/guidelines.xhtml

ngapore	40.5	1.2			21	48.0	24	1128	82.0	34.5	m24s	N	no	low	30.8
ovenia	10.6	0.0			6		12	196	94.4	97.5	m24s	Ν	no	low	36.2
meria, Spain	26.0	6.0			8		27	281		81.9	m15	Ν	no	high	43.1
veden	4.8	0.1			10		36	406			m24s	Ν	no	low	21.2
ubende, Uganda		86.0	31.1	87.8	11	70.0	6	67	21.5	90.5	m15s	Ν	yes	low	35.8
ist Lancashire, UK	18.3	8.2			13	58.3	102	332	48.5	69.9	m15	0	no	low	42.8
K		8.2		42.3	12	50.0	48	102	90.7	87.2	m15	0	no	low	30.4
ndon, UK	44.9	8.2					9	964	36.0	100	m24	Ν	no		37.0
idlands, UK	15.0	8.2					48	4207	58.3	100	m15	0	no		61.2
dessa and Nikolaev, Ukraine	80.4	3.9	34.2	100			4	225			m15	0	yes ^f	low	60.4
anoi, Vietnam	146.0	10.0	0.0	100			20	465	92.7	91.9	m15s	Ν	yes	low	55.3
nı or u u s s c r i u u s	gapore venia neria, Spain eden ibende, Uganda it Lancashire, UK ndon, UK dlands, UK essa and Nikolaev, Ukraine noi, Vietnam	gapore 40.5 venia 10.6 neria, Spain 26.0 eden 4.8 ibende, Uganda . it Lancashire, UK 18.3 ndon, UK 44.9 dlands, UK 15.0 essa and Nikolaev, Ukraine 80.4 noi, Vietnam 146.0	gapore 40.5 1.2 venia 10.6 0.0 neria, Spain 26.0 6.0 eden 4.8 0.1 ibende, Uganda . 86.0 it Lancashire, UK 18.3 8.2 ndon, UK 44.9 8.2 dlands, UK 15.0 8.2 essa and Nikolaev, Ukraine 80.4 3.9 noi, Vietnam 146.0 10.0	gapore 40.5 1.2 . venia 10.6 0.0 . neria, Spain 26.0 6.0 . eden 4.8 0.1 . ibende, Uganda . 86.0 31.1 it Lancashire, UK 18.3 8.2 . ndon, UK 44.9 8.2 . dlands, UK 15.0 8.2 . essa and Nikolaev, Ukraine 80.4 3.9 34.2 noi, Vietnam 146.0 10.0 0.0	gapore 40.5 1.2 . venia 10.6 0.0 . neria, Spain 26.0 6.0 . eden 4.8 0.1 . ibende, Uganda . 86.0 31.1 it Lancashire, UK 18.3 8.2 . . 8.2 . . . 8.2 . . . 8.2 . . . 8.2 . . . 8.2 . . . 8.2 . . . 15.0 8.2 . . . sesa and Nikolaev, Ukraine 80.4 3.9 34.2 100 noi, Vietnam 146.0 10.0 0.0 100	gapore 40.5 1.2 . . 21 venia 10.6 0.0 . . 6 neria, Spain 26.0 6.0 . . 8 eden 4.8 0.1 . . 10 ibende, Uganda . 86.0 31.1 87.8 11 it Lancashire, UK 18.3 8.2 . . 13 ndon, UK 44.9 8.2 . . . dlands, UK 15.0 8.2 . . . essa and Nikolaev, Ukraine 80.4 3.9 34.2 100 . noi, Vietnam 146.0 10.0 0.0 100 .	gapore40.51.22148.0venia10.60.06.neria, Spain26.06.08.eden4.80.110.ibende, Uganda.86.031.187.81170.0it Lancashire, UK18.38.21358.38.21358.38.28.215.08.2934.2100146.010.00.0100	gapore40.51.22148.024venia10.60.06.21neria, Spain26.06.08.27eden4.80.110.36ibende, Uganda.86.031.187.81170.06it Lancashire, UK18.38.21358.3102ndon, UK44.98.242.31250.048essa and Nikolaev, Ukraine80.43.934.21004noi, Vietnam146.010.00.010020	gapore40.51.22148.0241128venia10.60.06.12196neria, Spain26.06.08.27281eden4.80.110.36406ibende, Uganda.86.031.187.81170.0667it Lancashire, UK18.38.21358.3102332ndon, UK44.98.2.42.31250.048102essa and Nikolaev, Ukraine80.43.934.21004225noi, Vietnam146.010.00.010020465	gapore40.51.22148.024112882.0venia10.60.06.1219694.4neria, Spain26.06.08.27281.eden4.80.110.36406.ibende, Uganda.86.031.187.81170.066721.5it Lancashire, UK18.38.21358.310233248.5ndon, UK44.98.21358.310230.7dlands, UK15.08.248420758.3essa and Nikolaev, Ukraine80.43.934.21004225.noi, Vietnam146.010.00.01002046592.7	gapore40.51.22148.024112882.034.5venia10.60.06.1219694.497.5neria, Spain26.06.08.27281.81.9eden4.80.110.36406ibende, Uganda.86.031.187.81170.066721.590.5it Lancashire, UK18.38.21358.310233248.569.9ndon, UK44.98.21358.310290.787.2ndon, UK15.08.248420758.3100essa and Nikolaev, Ukraine80.43.934.210044225noi, Vietnam146.010.00.01002046592.791.9	gapore40.51.22148.024112882.034.5m24svenia10.60.06.1219694.497.5m24sneria, Spain26.06.088.27281.81.9m15eden4.80.110.36406m24sibende, Uganda.86.031.187.81170.066721.590.5m15sit Lancashire, UK18.38.21358.310233248.569.9m15ndon, UK44.98.2996436.0100m24dlands, UK15.08.248420758.3100m15essa and Nikolaev, Ukraine80.43.934.21004225m15noi, Vietnam146.010.00.01002046592.791.9m15	gapore40.51.22148.024112882.034.5m24sNvenia10.60.061219694.497.5m24sNneria, Spain26.06.062728181.9m15Neden4.80.11036406m24sNibende, Uganda86.031.187.81170.066721.590.5m15sNit Lancashire, UK18.38.21358.310233248.569.9m15Ondon, UK44.98.2996436.0100m24Ndlands, UK15.08.248420758.3100m15Oessa and Nikolaev, Ukraine80.43.934.210044225m15Onoi, Vietnam146.010.00.010010010010046592.791.9m15sN	gapore40.51.22148.024112882.034.5m24sNnovenia10.60.06.1219694.497.5m24sNnoneria, Spain26.06.08.27281.81.9m15Nnoeden4.80.110.36406m24sNnoibende, Uganda.86.031.187.81170.066721.590.5m15sNyesit Lancashire, UK18.38.21358.310233248.569.9m15Onondon, UK44.98.2996436.0100m24Nnodlands, UK15.08.248420758.3100m15Onoessa and Nikolaev, Ukraine80.43.934.210044225m15Oyesresno, Vietnam146.010.00.01002046592.791.9m15sNyes	gapore 40.5 1.2 21 48.0 24 1128 82.0 34.5 m24s N no low venia 10.6 0.0 6 12 196 94.4 97.5 m24s N no low neria, Spain 26.0 6.0 8 27 281 81.9 m15 N no high eden 4.8 0.1 10 36 406 m15. N no high ibende, Uganda 86.0 31.1 87.8 11 70.0 6 67 21.5 90.5 m15s N yes low ibende, Uganda 86.2 13 58.3 102 332 48.5 69.9 m15 0 no low idon, UK 48.3 8.2 12 50.0 48 102 90.7 87.2 m15 0

^a Estimates from of the prevalence of TB/HIV co-infection in the study country [33,34]

^b 15=15 MIRU-VNTR loci (made up of the 'old 12' or 'new 12' defined in the footnote below), 24=24 MIRU-VNTR loci (ETR A, B, C, D, E; MIRU 2, 10, 16, 20, 23, 24, 26, 27, 39, 40; VNTR 424, 1955, 2163b, 2347, 2401, 3171, 3690, 4052, 4156), S=with Spoligotyping

^c O= old 12 MIRU loci (MIRU 2, 4, 10, 16, 20, 23, 24, 26, 27,30, 31, 39, 40), N=new 12 MIRU loci (MIRU 10, 16, 26, 31, 40 + Mtub 04, 21, 39 + ETR A C + QUB 11b, 26)

^d Risk of bias was assessed using the STROME-ID checklist. Studies scoring <20 were categorised as have a high risk of bias

^e The proportion of clustering was calculated as the number of clustered isolates/number of clustered + unique isolates

^f 11.3% did not consent to being part of the study. The other studies that required consent for isolates to be typed did not report the refusal rate

For peer review only - http://bmjopen.bmj.com/site/about/guidelines.xhtml

Table 2: The number of studies that reported the variables of interest

	Reported	Missing
Study setting		
TB incidence	8	15
TB/HIV co-infection	5	22
Previous TB treatment	9	18
Proportion pulmonary TB	14	13
Maximum cluster size	19	8
% clusters with 2 cases	14	13
Study design		
Study duration	27	0
Study size	27	0
% population that is culture	15	12
% culture positive typed	19	8
24 loci (compared to 15)	27	0
Repeat isolates	12	15
Missing loci	8	19
Double alleles	1	26
Consent required	6 ^a	21
Epidemiological information	6	21
^a Only one study reported the consent rat	e	

BMJ Open

2
3
4
5
6
7
1
8
9
10
11
12
12
13
14
15
16
17
10
10
19
20
21
22
23
21
24
25
26
27
28
29
20
30
31
32
33
34
35
36
07
31
38
39
40
41
42
12
40
44
45
46
47
48
10
49 50
50
51
52
53
54
55
55
56
57
58

59 60 Table 3: Univariable metaregression showing the coefficients for change in the proportion of clustering and the percentage of between-study variation explained by variables describing the study design and setting.

	n	Coefficient ^a	CI	р	Adj R ^{2 b}
Study setting					
TB incidence	23	0.14	0.04-0.24	0.007	26.74
TB/HIV co-infection	23	0.04	-0.03-0.11	0.246	2.00
Maximum cluster size	19	0.20	0.09-0.30	0.001	48.20
Study design					
Study duration	27	-0.02	-0.09-0.06	0.677	-3.37
% population that is culture positive	15	0.34	-1.23-1.96	0.661	-5.92
% culture positive typed	19	0.22	-1.08-1.52	0.725	-5.41
Study size	27	0.03	-0.11-0.16	0.702	-3.31
24 loci (compared to 15)	27	-0.30	-0.590.01	0.04	13.58
Consent required	27	0.38	0.04-0.72	0.029	14.41

^a Coefficients for the change in the proportion of clustering for each covariate. E.g. for a one-month increase in study duration, the proportion of clustering increases by 0.003.

^b The proportion of between-study variation explained by the univariate meta-regression.

Figure Caption

Figure 1: Results of systematic search, screening and data extraction.

Figure 2: Forest plot showing the proportion of clustering reported in each study by the number of MIRU-VNTR loci typed

The number of loci typed is categorised into 15 loci (m15), 15 loci with Spoligotyping (m15s), 24 loci (m24) and Spoligotyping 24 loci with Spoligotyping (m24s). The study reference is shown in the right hand column.



190x254mm (300 x 300 DPI)

2	
3	
4	
5	
6	
0	
1	
8	
9	
1(0
1	1
4	ו ר
1	2
1:	3
1.	4
1	5
1(6
1	7
4	/ 0
10	B
1	9
2	0
2	1
2	2
2	2
2.	2
2	4
2	5
2	6
2	7
2	Q
2	
2	9
3	U
3	1
3	2
3	3
2	1
3	+
3	D C
3	6
3	7
3	8
3	9
1	n
4	4
4	1
4	2
4	3
4	4
4	5
4	6
· ד-	7
4	1 0
4	B
4	9
5	0
5	1
5	2
5	2
່ວ. ກ	0
5	4
5	5
5	6
5	7
5	R
5	0
0	J

1



190x254mm (300 x 300 DPI)

BMJ Open

Appendix 1: Medline/Embase search strategy

1. (tubercle adj3 (bacillus or bacilli)).mp. [mp=title, original title, abstract, name of substance word, subject heading word, unique identifier]

2. ((mycobacterium or mycobacteria) adj3 (bovis or africanum or microti or canetti)).mp.

3. exp tuberculosis/ or mycobacterium tuberculosis/ or tuberculosis.mp. or tb.mp. or Mtb.mp. or "M tuberculosis complex".mp.

4. or/1-3

5. Minisatellite Repeats/ or Genotype/ or Interspersed Repetitive Sequences/ or DNA Fingerprinting/ or Bacterial Typing Techniques/

6. "miru".mp. [mp=title, original title, abstract, name of substance word, subject heading word, unique identifier]

7. "vntr".mp. [mp=title, original title, abstract, name of substance word, subject heading word, unique identifier]

8. (miru adj3 vntr).mp. [mp=title, original title, abstract, name of substance word, subject heading word, unique identifier]

9. (mycobacterial adj3 interspersed adj3 repetitive adj3 units).mp. [mp=title, original title, abstract, name of substance word, subject heading word, unique identifier]

10. (dna adj3 fingerprinting).mp. [mp=title, original title, abstract, name of substance word, subject heading word, unique identifier]

11. ((strain adj3 type) or (strain adj3 typing) or (strain adj3 types)).mp. [mp=title, original title, abstract, name of substance word, subject heading word, unique identifier]

12. ((molecular adj3 typing) or (molecular adj3 strain adj3 typ*)).mp. [mp=title, original title, abstract, name of substance word, subject heading word, unique identifier]

13. (genotype or genotyping or genotypes).ti,ab.

14. (minisatellite adj3 repeat*).mp. [mp=title, original title, abstract, name of substance word, subject heading word, unique identifier]

15. molecular epidemiology/mt or (molecular adj3 epidemiology).mp. [mp=title, original title, abstract, name of substance word, subject heading word, unique identifier]

16. or/5-15

17. exp disease outbreaks/ or (outbreak adj3 analysis).mp. or (outbreak adj3 investigation).mp. or (outbreak adj3 management).mp. or (tuberculosis adj3 outbreak).mp. [mp=title, original title, abstract, name of substance word, subject heading word, unique identifier]

18. exp contact tracing/ or (contact adj3 tracing).mp. or (contact* adj3 traced).mp. or (contact adj3 screen*).mp. [mp=title, original title, abstract, name of substance word, subject heading word, unique identifier]

19. exp case management/ or (case adj3 management).mp. [mp=title, original title, abstract, name of substance word, subject heading word, unique identifier]

20. exp Risk Factors/

21. (risk adj3 factor*).mp. [mp=title, original title, abstract, name of substance word, subject heading word, unique identifier]

22. exp Epidemiologic Factors/

23. infectious disease transmission.mp. or exp Disease Transmission, Infectious/

24. exp case management/ or (case adj3 management).mp. [mp=title, original title, abstract, name of substance word, subject heading word, unique identifier]

25. program evaluation/ or evaluation studies as topic/ or (program adj3 evaluation).mp. or (programme adj3 evaluation).mp. [mp=title, original title, abstract, name of substance word, subject heading word, unique identifier]

26. public health practice/ or (public adj3 health).mp. [mp=title, original title, abstract, name of substance word, subject heading word, unique identifier]

27. ((tuberculosis adj3 control) or (tb adj3 control)).mp. [mp=title, original title, abstract, name of substance word, subject heading word, unique identifier]

28. (molecular adj3 surveillance).mp. [mp=title, original title, abstract, name of substance word, subject heading word, unique identifier]

29. exp cluster analysis/ or (cluster* adj3 rate*).mp. or (cluster* adj3 growth).mp. or (cluster* adj3 analysis).mp. or (cluster adj3 investigation).mp. or (proportion adj3 cluster*).mp. or (molecular adj3 cluster*).mp. [mp=title, original title, abstract, name of substance word, subject heading word, unique identifier]

30. ((recent adj3 transmission) or (transmission adj3 event*) or (transmission adj3 rate*) or (chain adj3 transmission) or (transmission adj3 setting*)).mp. [mp=title, original title, abstract, name of substance word, subject heading word, unique identifier]

31. or/17-30

1
ว
2
3
4
5
6
7
, 0
0
9
10
11
12
12
10
14
15
16
17
18
10
19
20
21
22
23
24
24
25
26
27
28
20
20
30
31
32
33
34
25
30
36
37
38
39
10
40
41
42
43
44
45
46
40
47
48
49
50
51
51
52
53
54
54 55
54 55 56
54 55 56 57
54 55 56 57
54 55 56 57 58
54 55 56 57 58 59

32. 4 and 16
33. 32 and 31
34. limit 33 to yr="1998-Current"
35. limit 34 to english language
36. animals/
37. humans/
38. 36 not 37
39. 35 not 38

Appendix 2: STROME-ID scores for the included studies

Author	STROME-ID score ^a
Aleksic, E	24
Alliex-Beguec, C	32
Allix-Beguec, C	25
Alonso-Rodriguez, N	18
Asgharzadeh, M	28
Bidovec-Stojkovic, U	31
De Beer, JL	30
Dymova, MA	19
Evans, J	b
Grujav, U	32
Guang-ming, DAI	19
Hamblion, E	b
Hang, NTHL	31
Jonsson, J	22
Lim, LKY	30
Mandal, S	32
Muwonge, A	25
Nikolayevsky, V	23
Oelemann, M	34
Ojo, OO	36
Roetzer, A	16
Sails, A	23
Smit, PW	29
Tessema, B	26
Tuite, AR	31
Varghese, B	23
Zmak, L	19

^aIndividual studies score 1 for each element of checklist they had address

^bConference abstracts

PRISMA 2009 Checklist

Section/topic	#	Checklist item	Reported on page #
TITLE			
Title	1	Identify the report as a systematic review, meta-analysis, or both.	1
ABSTRACT			
2 Structured summary 3 4	2	Provide a structured summary including, as applicable: background; objectives; data sources; study eligibility criteria, participants, and interventions; study appraisal and synthesis methods; results; limitations; conclusions and implications of key findings; systematic review registration number.	2
Rationale	3	Describe the rationale for the review in the context of what is already known.	3
) Objectives	4	Provide an explicit statement of questions being addressed with reference to participants, interventions, comparisons, outcomes, and study design (PICOS).	3
METHODS			
Protocol and registration	5	Indicate if a review protocol exists, if and where it can be accessed (e.g., Web address), and, if available, provide registration information including registration number.	n/a
Eligibility criteria	6	Specify study characteristics (e.g., PICOS, length of follow-up) and report characteristics (e.g., years considered, language, publication status) used as criteria for eligibility, giving rationale.	4
Information sources	7	Describe all information sources (e.g., databases with dates of coverage, contact with study authors to identify additional studies) in the search and date last searched.	4
) Search	8	Present full electronic search strategy for at least one database, including any limits used, such that it could be repeated.	appendix
Study selection	9	State the process for selecting studies (i.e., screening, eligibility, included in systematic review, and, if applicable, included in the meta-analysis).	4
Data collection process	10	Describe method of data extraction from reports (e.g., piloted forms, independently, in duplicate) and any processes for obtaining and confirming data from investigators.	4
) Data items	11	List and define all variables for which data were sought (e.g., PICOS, funding sources) and any assumptions and simplifications made.	5
Risk of bias in individual studies	12	Describe methods used for assessing risk of bias of individual studies (including specification of whether this was done at the study or outcome level), and how this information is to be used in any data synthesis.	5
Summary measures	13	State the principal summary measures (e.g., risk ratio, difference in means).	5
Synthesis of results	14	Describe the methods of handling data and combining results of studies, if done, including measures of consistency (e.g., 1 ² for each meta-analysis, bttp://bmiopen.bmi.com/site/about/guidelines.xbtml	5

BMJ Open



PRISMA 2009 Checklist

_		-	
Page	1	of	2
1 446		01	-

4			Page 1 of 2	
5 6 7	Section/topic	#	Checklist item	Reported on page #
8 9	Risk of bias across studies	15	Specify any assessment of risk of bias that may affect the cumulative evidence (e.g., publication bias, selective reporting within studies).	5
10 1 12	Additional analyses	16	Describe methods of additional analyses (e.g., sensitivity or subgroup analyses, meta-regression), if done, indicating which were pre-specified.	5
1:	RESULTS			
14 14 10	Study selection	17	Give numbers of studies screened, assessed for eligibility, and included in the review, with reasons for exclusions at each stage, ideally with a flow diagram.	Figure 1
1	Study characteristics	18	For each study, present characteristics for which data were extracted (e.g., study size, PICOS, follow-up period) and provide the citations.	15
2	Risk of bias within studies	19	Present data on risk of bias of each study and, if available, any outcome level assessment (see item 12).	15
2	Results of individual studies	20	For all outcomes considered (benefits or harms), present, for each study: (a) simple summary data for each intervention group (b) effect estimates and confidence intervals, ideally with a forest plot.	Figure 2
2	Synthesis of results	21	Present results of each meta-analysis done, including confidence intervals and measures of consistency.	18
2	Risk of bias across studies	22	Present results of any assessment of risk of bias across studies (see Item 15).	15
2	Additional analysis	23	Give results of additional analyses, if done (e.g., sensitivity or subgroup analyses, meta-regression [see Item 16]).	18
2	DISCUSSION			
3	Summary of evidence	24	Summarize the main findings including the strength of evidence for each main outcome; consider their relevance to key groups (e.g., healthcare providers, users, and policy makers).	7
3	3 Limitations	25	Discuss limitations at study and outcome level (e.g., risk of bias), and at review-level (e.g., incomplete retrieval of identified research, reporting bias).	7-8
3: 3(Conclusions	26	Provide a general interpretation of the results in the context of other evidence, and implications for future research.	8
3	FUNDING			
39	Funding	27	Describe sources of funding for the systematic review and other support (e.g., supply of data); role of funders for the systematic review.	8
4	1			

From: Moher D, Liberati A, Tetzlaff J, Altman DG, The PRISMA Group (2009). Preferred Reporting Items for Systematic Reviews and Meta-Analyses: The PRISMA Statement. PLoS Med 6(6): e1000097. 43 doi:10.1371/journal.pmed1000097

For more information, visit: www.prisma-statement.org.

Page 2 of 2 For peer review only - http://bmjopen.bmj.com/site/about/guidelines.xhtml

BMJ Open

BMJ Open

Effect of study design and setting on tuberculosis clustering estimates using Mycobacterial Interspersed Repetitive Units-Variable Number Tandem Repeats (MIRU-VNTR): A systematic review

Journal:	BMJ Open
Manuscript ID:	bmjopen-2014-005636.R2
Article Type:	Research
Date Submitted by the Author:	23-Dec-2014
Complete List of Authors:	Mears, Jessica; University College London, Department of Infection and Population Health Abubakar, Ibrahim; University College London, Department of Infection and Population Health; Public Health England, Centre for Infectious Disease Surveillance and Control Cohen, Ted; Harvard School of Public Health, Harvard University, Division of Global Health Equity, Brigham and Women's Hospital and Department of Epidemiology McHugh, Timothy; Centre for Clinical Microbiology, Research Department of Infection, Royal Free Campus, University College London Sonnenberg, Pamela; University College London, Department of Infection and Population Health
Primary Subject Heading :	Research methods
Secondary Subject Heading:	Infectious diseases, Public health
Keywords:	EPIDEMIOLOGY, Tuberculosis < INFECTIOUS DISEASES, MOLECULAR BIOLOGY

SCHOLARONE[™] Manuscripts



BMJ Open

2		
3	1	Effect of study design and setting on tuberculosis clustering estimates using
4	-	Mycobactorial Intersported Ponetitive Units Variable Number Tandom
5	Z	
6	3	Repeats (MIRU-VNTR): A systematic review
7		
8	4	Jessica Mears ¹ , Ibrahim Abubakar ^{1,2,3} , Theodore Cohen ⁴ , Timothy D McHugh ⁵ & Pam Sonnenberg ^{1,*}
9		
10	5	¹ Department of Infection and Population Health, University College London
11		
12	6	² Centre for Infectious Disease Surveillance and Control, Public Health England
13		
14	7	³ Clinical Trials Unit, Medical Research Council, London
10		
10	8	⁴ Division of Global Health Equity, Brigham and Women's Hospital and Department of Epidemiology.
18		
10	9	Harvard School of Public Health, Harvard University
20		
20	10	⁵ Centre for Clinical Microbiology Department of Infection, University College London
22	10	centre for ennear wherobology, Department of infection, oniversity conege condon
23	11	*Corresponding author:
24	11	
25	10	
26	12	Dr Pam Sonnenberg
27	4.2	
28	13	3 Floor Mortimer Market Centre
29		
30	14	University College London
31	. –	
32	15	London WC1E 6JB
33		
34	16	p.sonnenberg@ucl.ac.uk
35		
30	17	
31		
აი 20	18	Word count: 2392
39 40		
40		
42		
43		
44		
45		
46		
47		
48		
49		
50		
51		
52		
53 54		
04 55		
50 56		
50		
58		1
59		1
60		

19 Abstract

- 20 Objectives: To systematically review the evidence for the impact of study design and setting on the
- 21 interpretation of TB transmission using clustering derived from Mycobacterial Interspersed
- 22 Repetitive Units Variable Number Tandem Repeats (MIRU-VNTR) strain typing.

23 Data sources: Medline, Embase, CINHAL, Web of Science and Scopus were searched for articles

- 24 published before 21st October 2014.
- 25 Review methods: Studies in humans that reported the proportion of clustering of TB isolates by
- 26 MIRU-VNTR were included in the analysis. Univariable meta-regression analyses were conducted to
- 27 assess the influence of study design and setting on the proportion of clustering.
- 28 Results: The search identified 27 eligible articles reporting clustering between 0% and 63%. The
- 29 number of MIRU-VNTR loci typed, requiring consent to type patient isolates (as a proxy for sampling
 - 30 fraction), the TB incidence and the maximum cluster size explained 14%, 14%, 27% and 48% of
 - 31 between-study variation, respectively, and had a significant association with the proportion of
 - 32 clustering.
 - 33 Conclusions: Although MIRU-VNTR typing is being adopted worldwide there is a paucity of data on
 - 34 how study design and setting may influence estimates of clustering. We have highlighted study
 - 35 design variables for consideration in the design and interpretation of future studies.

37 Strengths and Limitations of Study

- This is a timely evaluation of the impact of study design on estimates of TB clustering using MIRU-VNTR strain typing because it has been incorporated into national typing services globally.
- The strength of this meta-analysis was limited by the lack of detail reported by the included studies, highlighting the need for better quality reporting in primary studies.

BMJ Open

45 Introduction

The introduction of molecular typing methods has improved our understanding of *Mycobacterium tuberculosis* (TB) transmission and has changed local and national control policies [1–5]. The proportion of cases that are clustered is often used to estimate the amount of ongoing transmission within the population, based on the assumption that cases with indistinguishable strain types are part of a chain of transmission. TB molecular typing methodology is changing rapidly and it is important that we better understand how to interpret the outputs and thus act.

TB molecular typing methods include Spoligotyping [6], insertion sequence *6110* (IS*6110*) restriction fragment length polymorphism (RFLP) analysis (the recent gold standard) [7], mycobacterial interspersed repetitive units-variable number tandem repeats (MIRU-VNTR) typing [8], and whole genome sequencing [9–11]. Published reviews have identified factors that might influence or bias clustering by IS*6110* RFLP [12,13]. No study has repeated this analysis using more up-to-date typing methods, which is important for understanding of the epidemiology of TB and to shape the application of molecular typing to improve TB control.

Published meta-analyses and modelling studies using IS*6110* RFLP data show that the proportion of clustering observed can be affected by 1) study design (affecting the proportion of eligible cases that are included in the study); 2) features of the typing method (such as the ability to type isolates with low copy numbers); and 3) study setting (such as characteristics of the study population). For example, the proportion of clustering increases when the fraction of the total data sampled increases [13–15] and when study duration increases [16].

MIRU-VNTR is currently the preferred method of molecular typing [17-21], and can be used together with Spoligotyping [8]. Relative to IS6110 RFLP, MIRU-VNTR does not have to exclude isolates with a low IS6110 copy number, has a faster turnaround time, is high throughput and the numeric strain types are more easily compared. MIRU-VNTR strain typing is increasingly being adopted worldwide [1,22–27], yet unlike IS6110 RFLP, the evidence for the interpretation of the findings such as the impact of study design and setting on clustering have not been reviewed. Although the two typing methods have been shown to have a similar discriminatory value, the markers evolve independently and at different rates, resulting in a difference in clustering between the two methods [28]. This suggests that there could be differences in the way study design, typing method and setting affects clustering by the two methods. We conducted a systematic review to assess the evidence for the impact of study design and setting on the interpretation of TB

transmission using clustering derived from MIRU-VNTR strain typing – as has been shown using
 IS6110 RFLP typing.

78 Methods

Five electronic databases were searched (EMBASE, ISI Web of Science, CINHAL, Scopus and Medline (Ovid)) up to 20th October 2014. The search strategy combined the following terms with Boolean operators: Tuberculosis, strain typing, and transmission (Appendix 1). The search was limited to studies using the standard MIRU-VNTR method [8], in humans only, and in English.

All titles and abstracts from each of the searches were examined. The full text of each paper was obtained and reviewed if the study reported MIRU-VNTR strain typing of *M.tuberculosis* complex isolates with at least 15 of the standardised 24 loci (ETR A, B, C, D, E; MIRU 2, 10, 16, 20, 23, 24, 26, 27, 39, 40; VNTR 424, 1955, 2163b, 2347, 2401, 3171, 3690, 4052, 4156) [8,29,30].

Studies using fewer than 15 loci were not included because the level of discrimination is inadequate for epidemiological use (n=121) [8]. Studies that used loci different to the standardised 15 and 24 set were not included in the analysis in order to reduce the heterogeneity between studies (n=19). All publication types were included in this first screen to ensure that no relevant data were missed.

Reviews, letters, editorials, outbreaks or case reports (n=103) were excluded in the second screen.
Studies that used incomplete sampling (e.g. random samples, studies using subsets of populations
such as multidrug-resistant patients) (n=47) and studies that had a sample size of less than 50 (n=4)
were also excluded.

A reviewer (JM) extracted the following data items from all included studies using a form developed in Excel (Microsoft 2010): publication details (year, authors, study country), study details (study duration, loci typed, secondary typing method, study population, whether participant consent was required (a characteristic of the study design that was used as proxy for sampling fraction, assuming that where consent was required the sampling fraction was low)), the number of clustered and unique isolates, and the covariates of interest: the maximum size of clusters; the proportion of clusters containing two cases; the proportion of the population that was culture positive; the proportion of culture positive isolates typed; risk factors for clustering; and the Hunter Gaston Discriminatory Index (HGDI) [31]). IA extracted data from 10% of the papers for external validity, disagreements were discussed and a consensus agreed upon.

BMJ Open

The main outcome measure – the proportion of TB isolates clustered by MIRU-VNTR strain typing –
 was calculated as the number of clustered isolates/number of clustered+unique isolates. Where
 there were uncertainties JM consulted with IA.

Authors were contacted if TB incidence rate was not reported. Where no response was received WHO country estimates of TB incidence for the study year were used.[32] As so few studies reported the proportion coinfected with TB/HIV, these estimates for the study country were taken from an EU-wide survey and WHO country profiles.[33,34] Due to poor recording of the sampling fraction (the number of isolates typed/the total number of culture positive TB cases diagnosed during the study period (n=19)), whether the study required the consent of participants (yes/no) was included as a proxy for (low/high) sampling fraction. The risk of bias within each study was assessed using the STROME-ID checklist.[35]

Data were analysed in Stata 12. Where studies reported data from more than one set of loci, the method with the highest discriminatory value was included (i.e. MIRU-VNTR 24 would be chosen over MIRU-VNTR 15, and MIRU-VNTR 15 plus Spoligotyping would be chosen over MIRU-VNTR 15 alone) (n=8). This review was not concerned with summary measures of clustering, but factors that influenced clustering; therefore articles must have included at least one of the covariates. Continuous variables were transformed where the distribution was skewed. The proportion clustered was transformed using the Freeman Tukey transformation [36]. Study heterogeneity was assessed using a forest plot and the chi² test of heterogeneity. Univariable meta-regression analyses were carried out to determine the effect of the study design covariates on the proportion of clustered isolates. All covariates in the analysis were hypothesised to influence the proportion clustered *a priori*.

Sensitivity analyses were conducted to see the effect of removing studies reporting 0% clustering, with only extra-pulmonary TB cases, only *M.bovis* cases, studies using the 'old 12' MIRU loci as part of their 15 loci, and studies assessed as having a high likelihood of bias (STROME-ID score less than 20).

131 Results

The search identified 7274 references resulting in 27 studies (25 journal articles and 2 conference abstracts) included after deduplication and title/abstract/full text screening (Figure 1). The main characteristics of the included studies are shown in Table 1. Studies were published between 2007 and 2014 and the clustering reported varied from 0% [37] to 62.8% [38]. In all studies, clustered

BMJ Open

isolates were defined as having identical strain types based on the MIRU-VNTR loci typed, with or without Spoligotyping. 17 studies included isolates from newly diagnosed TB cases, three studies reported including isolates from new and chronic cases of TB, and seven did not report this information. In addition, ten studies did not include repeat isolates from the same patient, one study included a repeat isolate from one patient, and the remaining 17 did not report whether repeat isolates were included or not. Furthermore, four studies included isolates with missing loci in the cluster analysis, whereas four excluded isolates with missing loci, and the remaining 20 did not report how they dealt with missing loci. The number of studies reporting each variable of interest is shown in Table 2. STROME-ID scores can be found in Appendix 2.

A forest plot shows the spread of clustering reported by number of loci and additional typing
method (Figure 2). Significant heterogeneity was identified between the studies (p<0.001),
suggesting that a meta-regression would be an appropriate analysis.

The univariable meta-regression shows evidence for the proportion of clustering to decrease as the number of MIRU-VNTR loci typed increased from 15 to 24 (p=0.04; Table 3), accounting for 14% of the between study variation, and to increase when the study participants consented to being included in the study (p=0.03), accounting for 14% of the between study variation. The proportion of clustering increased as the TB incidence in the population increased (p=0.007, Adj R² = 26.7). There was also evidence for the proportion of clustering to increase as the maximum cluster size increased (p=0.001), accounting for 48% of between study variation. There was no evidence of the other study design or study setting variables significantly influencing the proportion clustered. Though non-significant (p>0.05), the TB/HIV coinfection rate in the population explained 2% of the between study variation. Too few studies included information on the proportion of clusters containing two cases, proportion of the study sample with previous TB or with pulmonary TB, so these could not be included in the analysis (Table 2).

Sensitivity analyses to examine the effect of excluding studies reporting 0% clustering,[37] only M.bovis cases, [39] studies using the 'old 12' MIRU loci, [39–44] and studies assessed as having a high risk of bias, [37,45–48] did not generally change the results. The proportion of culture positive TB in the population remained insignificant but explained 2.6% of the between study variation when excluding 0% clustering (p=0.278 and Adj R²=2.62). Similarly, the proportion of culture positive TB in the population remained insignificant but explained 2.6% of the between study variation when excluding studies with the highest risk of bias (p=0.278 and Adj $R^2=2.62$). The number of loci typed became non-significant, but explained 9.6% and 10.5% of the between study variation when

BMJ Open

168 excluding studies using the 'old 12' loci and the highest risk of bias, respectively (p=0.106, Adj
 169 R²=9.63; p=0.111, Adj R²=10.51, respectively).

170 Discussion

This review identified 27 studies that met the inclusion criteria. We illustrate that the interpretation
of studies using MIRU-VNTR to estimate clustering is subject to bias relating to study design and
setting; however, there were insufficient data available to fully explore this impact.

As expected, we found that the proportion of clustering decreased with a greater number of MIRU-VNTR loci typed, with increasing TB incidence and with increasing maximum cluster size. We found that requiring consent to type patient isolates increased the proportion of clustering, which is not expected, given that the sampling fraction would be lower in these studies.

The other study design variables included in this analysis, such as study duration, did not significantly influence the proportion of isolates that were clustered, contrary to previous findings [12]. This is likely to be because of a lack of good quality evidence: of the 27 studies that met the inclusion criteria for the review, none reported all the variables of interest, reducing the power of the analysis and precluding multivariable meta-regression (Table 2). Importantly, key details of cluster analyses were not reported consistently across the studies, such as whether repeat isolates from the same patients were included, or typing profiles with missing loci were included, introducing new, unmeasured biases. In addition, the range of the variables may have been too limited to show any impact on clustering estimates. For example, the proportion of culture positive isolates typed ranged from 34.5% to 100%, with 17 of the 19 studies reporting this variable from 81.9% to 100%. Furthermore, most of the studies (17/27=63%) were from low TB burden settings and therefore may be reflecting the rate at which imported cases have matching strain types by chance, rather than rates of recent transmission.

The sensitivity analysis suggested that, when excluding the studies with the greatest risk of bias, the culture-positivity in the population might explain a small amount of the between study variation. This is consistent with estimates of the influence of sampling on the proportion of clustering using *IS*6110 RFLP typing [49]. In the sensitivity analysis excluding studies that used the 'old 12' loci, the effect of the number of loci typed becomes non-significant. This is likely because studies using the 'old 12' accounted for six out of ten studies reporting 15 loci, reducing the number of studies and the power of the model.

BMJ Open

This study is a timely evaluation of the impact of study design on estimates of TB clustering using MIRU-VNTR strain typing because it has been incorporated into national typing services globally [23,50]. The findings are relevant where strain typing is used to evaluate TB control systems across different settings because the proportion of clustering is influenced by the number of loci typed, the TB incidence and the maximum cluster size. Given that strain typing methods are advancing beyond MIRU-VNTR typing and that the application of whole genome sequencing to TB control and public health strategies has been demonstrated [9–11,51], it is important that the biases in the analysis of such methods are explored and compared. Understanding how to design and compare research studies for public health will greatly improve the benefit gained from newer technologies.

The strength of this meta-analysis was limited by (a lack of) detail reported by the included studies. This review has highlighted the need for better quality reporting in primary studies to enable future reviews to be more robust. Recently published standards for reporting of molecular epidemiology for infectious diseases should improve the quality of reporting.[35] This review is further limited by our inability to access 58 of the title/abstract screened articles for full text screening.

The use of TB strain typing as a public health tool in TB control programmes is increasing globally. We have identified a lack of good quality studies that can contribute to our understanding in interpreting the molecular typing of TB. We have also shown that the proportion of clustering derived from MIRU-VTNR typing is influenced by the number of loci typed, whether consent is required to type isolates, TB incidence in the study setting, and the maximum cluster size, highlighting these as important considerations in the design and interpretation of future studies.

Conflict of interest

 Nothing to declare.

Acknowledgements

- We would like to acknowledge Ross Harris from the Statistics Unit at Public Health England for his advice on meta-regression.
- Author contributions

All authors made substantial contributions to the conception and design of the review, and the analysis and interpretation of data. JM drafted the article and PS, IA, TM and TC revised it critically for important intellectual content. All authors approved the final version for publication.

2	227	Funding
4	227	runung
5 6	228	JM is funded through a Public Health England and University College London Impact Studentship. IA
7 8	229	is funded through a NIHR Senior Research Fellowship.
9 10	230	Ethics
11 12 13	231	Ethical approval was not required as this review analyses data that is in the public domain.
14 15	232	Data sharing
16		
17 18	233	No additional data are available
19 20	234	References
21	7 25	1 Jambragts van Waazanbaak CSB, Sabak MMGG, van Garvan DIHL de Vries G, Verver S, et al
22	235	1. Lambregts-vall weezembeek CSB, Sebek Minide, vall Gerven PJHJ, de Viles G, Verver S, et al.
23	230	(2003) Tuberculosis contact investigation and DNA fingerprint surveillance in The Netherlands:
24	237	6 years' experience with nation-wide cluster feedback and cluster monitoring. Int J Tuberc Lung
25 26	238	Dis /: \$463-470.
20	239	2. Borgdorff MW, van den Hof S, Kremer K, Verhagen L, Kalisvaart N, et al. (2010) Progress
28	240	towards tuberculosis elimination: secular trend, immigration and transmission. Eur Respir J 36:
29 30	241	339–347. doi:10.1183/09031936.00155409.
31	242	3. Kik SV, Verver S, Van Soolingen D, De Haas PEW, Cobelens FG, et al. (2008) Tuberculosis
32	243	Outbreaks Predicted by Characteristics of First Patients in a DNA Fingerprint Cluster. Am J
33 34	244	Respir Crit Care Med 178: 96–104. doi:10.1164/rccm.200708-1256OC.
35	245	4 Small PM McClenny NB Singh SP Schoolnik GK Tompkins IS et al. (1993) Molecular strain
36	246	typing of Mycobacterium tuberculosis to confirm cross-contamination in the mycobacteriology
37	240	laboratory and modification of procedures to minimize occurrence of false-positive cultures.
38 39	248	Clin Microbiol 31: 1677–1682.
40	249	5 De Vries G van Hest RAH, Richardus IH (2007) Impact of mobile radiographic screening on
41	250	tuberculosis among drug users and homeless persons. Am L Respir Crit Care Med 176: 201–
42 43	251	207. doi:10.1164/rccm.200612-1877OC.
44	252	6. Kamerbeek J. Schouls L. Kolk A. van Agterveld M. van Soolingen D. et al. (1997) Simultaneous
45	252	detection and strain differentiation of Mycohacterium tuberculosis for diagnosis and
46 47	254	epidemiology. J Clin Microbiol 35: 907–914.
48	255	
49	255	7. Van Embden JD, Cave MD, Crawford JT, Dale JW, Elsenach KD, et al. (1993) Strain identification
50	256	of Mycobacterium tuberculosis by DNA fingerprinting: recommendations for a standardized
51 52	257	methodology. J Clin Microbiol 31: 406–409.
53	258	8. Supply P, Allix C, Lesjean S, Cardoso-Oelemann M, Rüsch-Gerdes S, et al. (2006) Proposal for
54	259	standardization of optimized mycobacterial interspersed repetitive unit-variable-number
55	260	tandem repeat typing of Mycobacterium tuberculosis. J Clin Microbiol 44: 4498–4510.
56	261	doi:10.1128/JCM.01392-06.
57		
58		9
59		
60		

BMJ Open

3	262	9.	Schürch AC, van Soolingen D (2011) DNA fingerprinting of Mycobacterium tuberculosis: From
4	263		phage typing to whole-genome sequencing. Infect Genet Evol. Available:
5	264		http://www.ncbi.nlm.nih.gov/pubmed/22067515. Accessed 13 March 2012.
0 7		4.0	
8	265	10.	Gardy JL, Johnston JC, Ho Sui SJ, Cook VJ, Shah L, et al. (2011) Whole-genome sequencing and
9	266		social-network analysis of a tuberculosis outbreak. N Engl J Med 364: 730–739.
10	267		doi:10.1056/NEJMoa1003176.
11	268	11.	Walker TM, Ip CL, Harrell RH, Evans JT, Kapatai G, et al. (2013) Whole-genome sequencing to
12	269		delineate Mycobacterium tuberculosis outbreaks: a retrospective observational study. Lancet
13	270		Infect Dis 13: 137–146. doi:10.1016/S1473-3099(12)70277-3.
14			
15	271	12.	Houben RMGJ, Glynn JR (2009) A systematic review and meta-analysis of molecular
16	272		enidemiological studies of tuberculosis: development of a new tool to aid interpretation
17	272		Tropical Medicine & International Health 1/: 802–909, doi:10.1111/i.1365-3156.2009.02316 v
18	275		Topical wedicine & international reach 14. 052 (505. doi.10.1111/j.1505 (5150.2005.02510.x.
19	27/	12	Fok A Numata V Schulzer M EitzGerald MI (May) Risk factors for clustering of tuberculosis
20	275	15.	cases: a systematic review of nonulation-based molecular enidemiology studies [Review
21	275		Article] The International Journal of Tuberculasis and Lung Disease 12: 480, 402
22	270		Articlej. The international journal of ruberculosis and Lung Disease 12. 480–492.
23	277	11	Pergdorff MW/ Van Den Hof S. Kalisyaart N. Kromer K. Van Seelingen D. (2011) Influence of
2 4 25	277	14.	Sampling on Clustering and Associations With Disk Easters in the Melacular Enidemiology of
25	278		Sampling on clustering and Associations with Risk Factors in the Molecular Epidemiology of
27	279		
28	280		http://aje.oxfordjournals.org/content/early/2011/05/23/aje.kwr061. Accessed 29 March 2012.
29	201	4 -	Church ID, Deven L, de Deven AC, Devendeuff MAN, Since DE, et al. (1000) Internetine DNA fingenerint
30	281	15.	Glynn JR, Bauer J, de Boer AS, Borgdorff MW, Fine PE, et al. (1999) Interpreting DNA fingerprint
31	282		clusters of Mycobacterium tuberculosis. European Concerted Action on Molecular
32	283		Epidemiology and Control of Tuberculosis. Int J Tuberc Lung Dis 3: 1055–1060.
33			
34	284	16.	Glynn JR, Crampin AC, Yates MD, Traore H, Mwaungulu FD, et al. (2005) The Importance of
35	285		Recent Infection with Mycobacterium tuberculosis in an Area with High HIV Prevalence: A
36	286		Long-Term Molecular Epidemiological Study in Northern Malawi. J Infect Dis 192: 480–487.
37	287		doi:10.1086/431517.
38			
39	288	17.	De Beer JL, Kremer K, Ködmön C, Supply P, van Soolingen D (2012) First Worldwide Proficiency
40	289		Study on Variable-Number Tandem-Repeat Typing of Mycobacterium tuberculosis Complex
41	290		Strains. Journal of Clinical Microbiology 50: 662–669. doi:10.1128/JCM.00607-11.
42			
43	291	18.	Maes M, Kremer K, van Soolingen D, Takiff H, de Waard JH (2008) 24-locus MIRU-VNTR
44	292		genotyping is a useful tool to study the molecular epidemiology of tuberculosis among Warao
45	293		Amerindians in Venezuela. Tuberculosis (Edinb) 88: 490–494. doi:10.1016/j.tube.2008.04.003.
46			
47	294	19.	Sougakoff W (2011) Molecular epidemiology of multidrug-resistant strains of Mycobacterium
48	295		tuberculosis. Clinical Microbiology and Infection 17: 800–805. doi:10.1111/j.1469-
49	296		0691.2011.03577.x.
50			
51	297	20.	Weniger T, Krawczyk J, Supply P, Niemann S, Harmsen D (2010) MIRU-VNTRplus: a web tool for
じZ 52	298		polyphasic genotyping of Mycobacterium tuberculosis complex bacteria. Nucleic Acids Res 38:
00 E4	299		W326–331. doi:10.1093/nar/gkq351.
54	-		, , , , , ,
56	300	21.	Supply P (2010) MIRU-VNTR typing: the new international standard for TB molecular
57	301	-	epidemiology Symposium of the Institut Pasteur de Tunisia.
58			10
59			10
60			

BMJ Open

2			
3	302	22.	Van Soolingen D, Borgdorff MW, de Haas PE, Sebek MM, Veen J, et al. (1999) Molecular
4	303		epidemiology of tuberculosis in the Netherlands: a nationwide study from 1993 through 1997.
5	304		Linfect Dis 180: 726–736. doi:10.1086/314930
6	504		5 meet bis 100.720 750. doi:10.1000/514550.
7	205	22	Cowan IS Diam L Monson T Wand P Tomporado D at al (2005) Evaluation of a two stop
8	202	25.	cowait LS, Dieni L, Monson T, Wanu P, Temporado D, et al. (2005) Evaluation of a two-step
9	300		approach for large-scale, prospective genotyping of Mycobacterium tuberculosis isolates in the
10	307		United States. J Clin Microbiol 43: 688–695. doi:10.1128/JCM.43.2.688-695.2005.
11		~ .	
12	308	24.	Centers for Disease Control and Prevention (2005) New CDC Program for Rapid Genotyping of
13	309		Mycobacterium Tuberculosis Isolates. JAMA 293: 2086–2086. doi:10.1001/jama.293.17.2086.
14			
15	310	25.	Bauer J, Kok-Jensen A, Faurschou P, Thuesen J, Taudorf E, et al. (2000) A prospective evaluation
16	311		of the clinical value of nation-wide DNA fingerprinting of tuberculosis isolates in Denmark. Int J
17	312		Tuberc Lung Dis 4: 295–299.
18			Ĵ
10	313	26.	Bauer J. Yang Z. Poulsen S. Andersen AB (1998) Results from 5 years of nationwide DNA
20	314		fingerprinting of Mycobacterium tuberculosis complex isolates in a country with a low
20	215		incidence of M tuberculosis infection. I Clin Microbiol 36: 305–308
21	515		
22	216	77	Zalpir Dave M. Daliak M. Erzan D. Sarli I (2002) Malagular anidamialogy of tubargulasis in
23	217	27.	201111-DOVC IVI, POIJak IVI, El 2010, Solil J (2003) Molecular epiderniology of tuberculosis in
24	317		Slovenia: results of a one-year (2001) hation-wide study. Scand J miect Dis 35: 863–868.
20	24.0	20	
20	318	28.	Hanekom M, van der Spuy GD, Gey van Pittius NC, Micevoy CRE, Hoek KGP, et al. (2008)
27	319		Discordance between mycobacterial interspersed repetitive-unit-variable-number tandem-
28	320		repeat typing and IS6110 restriction fragment length polymorphism genotyping for analysis of
29	321		Mycobacterium tuberculosis Beijing strains in a setting of high incidence of tuberculosis. J Clin
30	322		Microbiol 46: 3338–3345. doi:10.1128/JCM.00770-08.
31			
32	323	29.	Supply P, Lesjean S, Savine E, Kremer K, van Soolingen D, et al. (2001) Automated high-
33	324		throughput genotyping for study of global epidemiology of Mycobacterium tuberculosis based
34	325		on mycobacterial interspersed repetitive units I Clin Microbiol 39: 3563–3571
35	326		doi:10.1128/JCM 39.10.3563-3571.2001
36	520		doi.10.1120/3civi.55.10.5505 5571.2001.
37	277	30	Gonaul KK, Brown TL, Gibson AL, Vates MD, Drobniewski EA (2006) Progression toward an
38	220	50.	improved DNA amplification based typing technique in the study of Mysebasterium
39	320		tubersularia anidamialamu L Clin Minrahial 44, 2402, 2400, daiuto 1120/JCM 01420.05
40	329		tuberculosis epidemiology. J Clin Microbiol 44: 2492–2498. doi:10.1128/JCM.01428-05.
41		24	
42	330	31.	Hunter PR, Gaston MA (1988) Numerical index of the discriminatory ability of typing systems:
43	331		an application of Simpson's index of diversity. J Clin Microbiol 26: 2465–2466.
44			
45	332	32.	WHO TB data (n.d.). WHO. Available: http://www.who.int/tb/country/en/index.html.
46	333		Accessed 12 December 2012.
47			
48	334	33.	Kruijshaar ME, Pimpin L, Abubakar I, Rice B, Delpech V, et al. (2011) The burden of TB-HIV in
49	335		the EU: how much do we know? A survey of surveillance practices and results. Eur Respir J 38:
50	336		1374–1381. doi:10.1183/09031936.00198310.
51			
52	337	34.	World Health Organization (n.d.) WHO Tuberculosis Country Profiles. Available:
53	338		http://www.who.int/tb/country/data/profiles/en/.
54 55			
22 50			
50			
57 E0			11
50			11
59			
00			

			BMJ Open
1			
2 3 4 5	339 340 341	35.	Field N, Cohen T, Struelens MJ, Palm D, Cookson B, et al. (2014) Strengthening the Reporting of Molecular Epidemiology for Infectious Diseases (STROME-ID): an extension of the STROBE statement. The Lancet Infectious Diseases 14: 341–352. doi:10.1016/S1473-3099(13)70324-4
6 7	342	36.	Freeman ME, Tukey IW (1950) Transformations Related to the Angular and the Square Root.
8 9	343	50.	Ann Math Statist 21: 607–611. doi:10.1214/aoms/1177729756.
10 11	344 345	37.	Guang-ming D, Zhi-guo Z, Peng-ju D, Qian Z, Li W, et al. (2013) Differences in the population of genetics of Mycobacterium tuberculosis between urban migrants and local residents in Beijing,
12 13	346		China. Chinese Medical Journal 126: 4066–4071. doi:10.3760/cma.j.issn.0366-6999.20130216.
14 15 16	347 348 349	38.	Zmak L, Obrovac M, Katalinic Jankovic V (2014) First insights into the molecular epidemiology of tuberculosis in Croatia during a three-year period, 2009 to 2011. Scandinavian Journal of
17 18 10	350	39	Mandal S. Bradshaw I. Anderson J.F. Brown T. Evans JT. et al. (2011) Investigating transmission
20 21 22	351 352	55.	of Mycobacterium bovis in the United Kingdom in 2005 to 2008. J Clin Microbiol 49: 1943– 1950. doi:10.1128/JCM.02299-10.
23 24	353 354	40.	Asgharzadeh M, Kafil HS, Roudsary AA, Hanifi GR (2011) Tuberculosis transmission in Northwest of Iran: using MIRU-VNTR, ETR-VNTR and IS6110-RFLP methods. Infect Genet Evol
25 26	355		11: 124–131. doi:10.1016/j.meegid.2010.09.013.
27 28 29 30	356 357 358	41.	Dymova MA, Liashenko OO, Poteiko PI, Krutko VS, Khrapov EA, et al. (2011) Genetic variation of Mycobacterium tuberculosis circulating in Kharkiv Oblast, Ukraine. BMC Infect Dis 11: 77. doi:10.1186/1471-2334-11-77.
31 32 33	359 360 361	42.	Sails AD, Barrett A, Sarginson S, Magee JG, Maynard P, et al. (2011) Molecular epidemiology of Mycobacterium tuberculosis in East Lancashire 2001-2009. Thorax 66: 709–713. doi:10.1136/thx.2011.158881.
34 35 36	362	43.	Evans J (2010) Analysis of prevalent Mycobacterium tuberculosis strains in the United
37 38	363 364		Kingdom: detection, distribution and expansion of MIRU-VNTR profiles containing high numbers of isolates. European Society of Clinical Microbiology and Infectious Diseases. Vienna,
39 40	305		Austria.
41 42 42	367	44.	epidemiology and prevalence of mutations conferring rifampicin and isoniazid resistance in
43 44 45	368 369		138. doi:10.1111/j.1469-0691.2006.01583.x.
46 47	370	45.	L. Z, M. O, V. KJ (2014) First insights into the molecular epidemiology of tuberculosis in Croatia
48 49	371		during a three-year period, 2009 to 2011. Scandinavian Journal of Infectious Diseases.
50 51 52	372 373 374	46.	Roetzer A, Schuback S, Diel R, Gasau F, Ubben T, et al. (2011) Evaluation of Mycobacterium tuberculosis typing methods in a 4-year study in Schleswig-Holstein, Northern Germany. J Clin Microbiol 49: 4173–4178. doi:10.1128/JCM.05293-11.
53 54	375 376	47.	Dymova MA, Kinsht VN, Cherednichenko AG, Khrapov EA, Svistelnik AV, et al. (2011) Highest prevalence of the Mycobacterium tuberculosis Beijing genotype isolates in patients newly
55 56 57	377 378		diagnosed with tuberculosis in the Novosibirsk oblast, Russian Federation. J Med Microbiol 60: 1003–1009. doi:10.1099/jmm.0.027995-0.
58 59 60			12

BMJ Open

2			
3	379	48.	Alonso-Rodriguez N, Martínez-Lirola M, Sánchez ML, Herranz M, Peñafiel T, et al. (2009)
4	380		Prospective universal application of mycobacterial interspersed repetitive-unit-variable-
5	381		number tandem-repeat genotyping to characterize Mycobacterium tuberculosis isolates for
6	201		fact identification of clustered and emban cases. J Clin Microbiol 47: 2026, 2022
7	202		last ruentinication of clustered and orphan cases. J Clin Microbiol 47. 2020–2052.
8	383		doi:10.1128/JCM.02308-08.
9	281	10	Glypp IR Myonychy F. Fine DEM (1999) Influence of Sampling on Estimates of Clustering and
10	205	49.	Depend Trepresies of Muschesterium tubersulesis Devived from DNA Fingersuisting
11	385		Recent Transmission of Mycobacterium tuberculosis Derived from DNA Fingerprinting
12	386		lechniques. American Journal of Epidemiology 149: 366 –371.
13			
14	387	50.	TB Strain Typing Project Board HPA (2011) TB Strain Typing Cluster Investigation Handbook for
15	388		Health Protection Units 1st Edition. Available:
16	389		https://hpaintranet.hpa.org.uk/Content/ProgrammesProjects/HPAProgrammes/HPAKeyHealth
17	390		ProtectionProgrammes/Respiratory/TB/StrainTyping/, Accessed 30 November 2011
18	550		
10	391	51	Walker TM Monk P. Grace Smith F. Peto TEA (2013) Contact investigations for outbreaks of
19	202	51.	Muchastarium tubarculacis, advances through whole genome seguencing. Clin Microbiol
20	392		Nycobacterium tuberculosis, advances through whole genome sequencing. Clin Microbiol
21	393		Infect. doi:10.1111/1469-0691.12183.
22			
23	394	52.	Gurjav U, Jelfs P, McCallum N, Marais BJ, Sintchenko V (2014) Temporal dynamics of
24	395		Mycobacterium tuberculosis genotypes in New South Wales, Australia. BMC infectious
25	396		diseases 14: 455–455. doi:10.1186/1471-2334-14-455.
26			
27	397	53	Allix-Béguec C. Fauville-Dufaux M. Supply P (2008) Three-year population-based evaluation of
28	308	55.	standardized mycobacterial interspersed repetitive-unit-variable-number tandem-repeat
29	200		standardized mycobacterial interspersed repetitive-dilit-variable-number tandem-repeat
30	399		typing of Mycobacterium tuberculosis. J Clin Microbiol 46: 1398–1406.
31	400		doi:10.1128/JCM.02089-07.
32			
33	401	54.	Allix-Béguec C, Supply P, Wanlin M, Bifani P, Fauville-Dufaux M (2008) Standardised PCR-based
34	402		molecular epidemiology of tuberculosis. Eur Respir J 31: 1077–1084.
35	403		doi:10.1183/09031936.00053307.
36			
37	404	55.	Tuite AR, Guthrie JL, Alexander DC, Whelan MS, Lee B, et al. (2013) Epidemiological evaluation
38	405		of spatiotemporal and genotypic clustering of mycobacterium tuberculosis in Optario. Canada
30	406		International Journal of Tuborculosis and Lung Disease 17: 1222–1227
39	400		International Journal of Tuberculosis and Lung Disease 17, 1522–1527.
40	407	БС	Tessame D. Dear J. Marker M. Emmrich F. Cock J. et al. (2012) Malagular enidemiology and
41	407	50.	ressentia B, Beer J, Merker M, Enfinition F, Sack O, et al. (2013) Molecular epidemiology and
42	408		transmission dynamics of Mycobacterium tuberculosis in Northwest Ethiopia: new
43	409		phylogenetic lineages found in Northwest Ethiopia. Bmc Infectious Diseases 13: 131.
44	410		doi:10.1186/1471-2334-13-131.
45			
46	411	57.	Smit PW, Haanpera M, Rantala P, Couvin D, Lyytikainen O, et al. (2013) Molecular
47	412		Epidemiology of Tuberculosis in Finland, 2008-2011, Plos One 8: e85027.
48	413		doi:10.1371/journal.none.0085027
49	110		
50	<i>411</i>	58	Oelemann MC Diel R Vatin V Haas W Rüsch-Gerdes S et al. (2007) Assessment of an
51	714 /15	50.	ontimized muchaetarial interparted reportitive unit veriable number tanders report turing
52	415		optimized mycobacterial interspersed repetitive- unit-variable-number tandem-repeat typing
53	416		system combined with spoligotyping for population-based molecular epidemiology studies of
54	417		tuberculosis. J Clin Microbiol 45: 691–697. doi:10.1128/JCM.01393-06.
55			
56			
57			
57 58			13
57 58 59			13

			BMJ Open
1			
2	418	59.	Ojo OO, Sheehan S, Corcoran DG, Nikolayevsky V, Brown T, et al. (2010) Molecular
4	419		epidemiology of Mycobacterium tuberculosis clinical isolates in Southwest Ireland. Infect
5	420		Genet Evol 10: 1110–1116. doi:10.1016/j.meegid.2010.07.008.
6			
7	421	60.	Aleksic E, Merker M, Cox H, Reiher B, Sekawi Z, et al. (2013) First Molecular Epidemiology Study
8	422		of Mycobacterium tuberculosis in Kiribati. PLoS ONE 8. Available:
9	423		http://www.scopus.com/inward/record.url?eid=2-s2.0-
10	424		84873163328&partnerID=40&md5=3994b8e5638129b621abc4d7d6d5e3b8.
12			
13	425	61.	De Beer JL, van Ingen J, de Vries G, Erkens C, Sebek M, et al. (2013) Comparative study of
14	426		IS6110 restriction fragment length polymorphism and variable-number tandem-repeat typing
15	427		of Mycobacterium tuberculosis isolates in the Netherlands, based on a 5-year nationwide
16	428		survey. J Clin Microbiol 51: 1193–1198. doi:10.1128/JCM.03061-12.
1/	429	62	Varghese B. Supply P. Shoukri M. Allix-Begues C. Memish 7, et al. (2013) Tuberculosis
10	430	02.	Transmission among Immigrants and Autochthonous Populations of the Fastern Province of
20	431		Saudi Arabia. PLoS ONE 8. Available: http://www.scopus.com/inward/record.url?eid=2-s2.0-
21	432		84885784886&partnerID=40&md5=4fdbf4015a999a9fcd1a1c31207a75a2.
22			
23	433	63.	Lim LK-Y, Sng LH, Win W, Chee CB-E, Hsu LY, et al. (2013) Molecular Epidemiology of
24	434		Mycobacterium tuberculosis Complex in Singapore, 2006-2012. Plos One 8: e84487.
25	435		doi:10.1371/journal.pone.0084487.
26			
28	436	64.	Bidovec-Stojkovic U, Zolnir-Dovc M, Supply P (2011) One year nationwide evaluation of 24-
29	437		locus MIRU-VNTR genotyping on Slovenian Mycobacterium tuberculosis isolates. Respir Med
30	438		105 Suppl 1: S67–73. doi:10.1016/S0954-6111(11)70014-2.
31	420	CE	Jansson J. Hoffmar C. Deversion J. Druchfold J. Chabramishael C. et al. (2014) Comparison
32	439	05.	between PELP and MIPLE VNTP genetyping of mycebactorium tuberculosis strains isolated in
33	440		stockholm 2000 to 2011. BLoS ONE, Available:
34	441		http://www.plosone.org/article/fetchOhject.action?uri=info%3Adoi%2E10.1371%2Ejournal.no
35	442		ne 0095159&representation=PDF
30			
38	444	66.	Muwonge A, Malama S, Johansen TB, Kankya C, Biffa D, et al. (2013) Molecular Epidemiology,
39	445		Drug Susceptibility and Economic Aspects of Tuberculosis in Mubende District, Uganda. PLoS
40	446		ONE 8. Available: http://www.scopus.com/inward/record.url?eid=2-s2.0-
41	447		84878608813&partnerID=40&md5=babbd6d006ca64e327fb19e01b6bc697.
42			
43	448	67.	Hamblion EL, Wynne-Edwards E, Anderson C, Anderson SR (2011) A summary of strain typing
44 45	449		and clustering of TB in London in 2010 and an analysis of the associated risk factors. Thorax 66:
46	450		A88–A89. doi:10.1136/thoraxjnl-2011-201054c.50.
47	451	60	Hang NTL Maada S. Lion LT. Thuang DH. Hung NV, et al. (2012) Drimary Drug Desistant
48	451	00.	Tuborculoris in Hanoi Viot Nam: Prosent Status and Pick Factors, Plos One 8: UNSD e71967
49	452		doi:10.1371/journal.none.0071867
50	433		doi.10.13/1/journal.pone.00/100/.
51	454		
52 52			
54			
55			
56			
57			
58			14
59			
00			
			For poor rovious only http://bmionon.hmi.com/cita/about/auidalines.yhtml

Tables

Table 1: The study setting and design characteristics of the included articles

Ref		Study se	tting							Stu	udy design				Risk of bias ^d	Clustering (%) ^e
	Study area and country	TB incidence (per 100,000)	TB/HIV (per 100,000) ^a	Previous TB treatment (%)	Pulmonary TB (%)	Maximum cluster size	Clusters of size 2 (%)	Study duration (months)	Study size (clustered + unique isolates)	Culture positive in study population (%)	Culture positive isolates typed (%)	Typing method ^b	Loci typed $^{\rm c}$	Consent required		
[52]	New South Wales, Australia	6.7	0.2	0.0	63.7		b .	36	1128			m24	Ν	no	low	20.1
[40]	Tabriz and Orumieh, Azarbaijan	26.0		5.2	87.0	5	81.8	12	156		94.5	m15	0	no	low	32.7
[53]	Brussels-Capital Region, Belgium	35.2	5.1	10.8		23	64.2	24	530	86.1	87.9	m24	Ν	no	low	29.6
[54]	Brussels-Capital Region, Belgium	35.2	5.1		100			39	802	81.8	84.7	m24s	Ν	no	low	28.8
[55]	Ontario, Canada	4.8	0.4			18	58.8	65	2016	•		m24s	Ν	no	low	23.1
[37]	Changping District, Beijing, China		0.3		100	0		30	318	31.5	94.6	m24	Ν	no	high	0.0
[38]	Croatia	19.0	0.1	•	•	45	48.3	36	1587			m15	Ν	no	high	62.8
[56]	Amhara region, Northwest Ethiopia		24.0	17.6	100	13		5	244			m24	N	ves	low	45 1
[57]	Finland	5.0	0.0			20		48	1048	75.4	99.4	m15s		no	low	33.9
[58]	Hamburg, Germany	12.7					45.5	12	154	78.2	91.1	m24s	N	no	low	22.1
[46]	Schleswig-Holstein, Germany	3.2	0.1			22	44.4	48	277			m24s	N	no	high	27.1
[59]	South West Ireland	15.3	3.3		82.7	12		36	171	79.5	96.1	m24s	N	no	low	27.5
[60]	South Tawara, Kiribati	370.0		4.1	100	25	55.6	24	73	45.4	98.6	m24s	N	yes	low	75.3
[61]	Netherlands	6.5	0.2				57.2	60	3978		100.1	m24	N	no	low	46.7
[41]	Kharkiv, Russia	94.0	3.8	63.3	100	10	50.0	3	98		100	m15	о	yes	high	31.6
[62]	Eastern province, Saudi Arabia	4.0			73.1	24	19.0	24	522			m24s	Ν	no	low	40.2
							15									

	i l						i									
[63]	Singapore	40.5	1.2			21	48.0	24	1128	82.0	34.5	m24s	Ν	no	low	30.8
[64]	Slovenia	10.6	0.0			6		12	196	94.4	97.5	m24s	Ν	no	low	36.2
[48]	Almeria, Spain	26.0	6.0			8		27	281		81.9	m15	Ν	no	high	43.1
[65]	Sweden	4.8	0.1			10		36	406			m24s	Ν	no	low	21.2
[66]	Mubende, Uganda		86.0	31.1	87.8	11	70.0	6	67	21.5	90.5	m15s	Ν	yes	low	35.8
[42]	East Lancashire, UK	18.3	8.2			13	58.3	102	332	48.5	69.9	m15	0	no	low	42.8
[39]	UK		8.2		42.3	12	50.0	48	102	90.7	87.2	m15	0	no	low	30.4
[67]	London, UK	44.9	8.2					9	964	36.0	100	m24	Ν	no		37.0
[43]	Midlands, UK	15.0	8.2	•				48	4207	58.3	100	m15	0	no		61.2
[44]	Odessa and Nikolaev, Ukraine	80.4	3.9	34.2	100			4	225			m15	0	yes ^f	low	60.4
[68]	Hanoi, Vietnam	146.0	10.0	0.0	100			20	465	92.7	91.9	m15s	Ν	yes	low	55.3

^a Estimates from of the prevalence of TB/HIV co-infection in the study country [33,34]

 ^b 15=15 MIRU-VNTR loci (made up of the 'old 12' or 'new 12' defined in the footnote below), 24=24 MIRU-VNTR loci (ETR A, B, C, D, E; MIRU 2, 10, 16, 20, 23, 24, 26, 27, 39, 40; VNTR 424, 1955, 2163b, 2347, 2401, 3171, 3690, 4052, 4156), S=with Spoligotyping

^c O= old 12 MIRU loci (MIRU 2, 4, 10, 16, 20, 23, 24, 26, 27,30, 31, 39, 40), N=new 12 MIRU loci (MIRU 10, 16, 26, 31, 40 + Mtub 04, 21, 39 + ETR A C + QUB 11b, 26)

^d Risk of bias was assessed using the STROME-ID checklist. Studies scoring <20 were categorised as have a high risk of bias. See Appendix 2 for STROME-ID scores

^e The proportion of clustering was calculated as the number of clustered isolates/number of clustered + unique isolates

^f 11.3% did not consent to being part of the study. The other studies that required consent for isolates to be typed did not report the refusal rate

For peer review only - http://bmjopen.bmj.com/site/about/guidelines.xhtml

Page 17 of 25

	Reported	Missing
Study setting		
TB incidence	8	15
TB/HIV co-infection	5	22
Previous TB treatment	9	18
Proportion pulmonary TB	14	13
Maximum cluster size	19	8
% clusters with 2 cases	14	13
Study design	27	0
Study duration	27	0
Study size	27	0
positive	15	12
% culture positive typed	19	8
24 loci (compared to 15)	27	0
Repeat isolates	12	15
Missing loci	8	19
Double alleles	1	26
Consent required	6ª	21
Epidemiological information	6	21
^a Only one study reported the consent rate		

	n	Coefficient ^a	CI	р	Adj R ^{2 b}
Study setting					
TB incidence	23	0.14	0.04-0.24	0.007	26.74
TB/HIV co-infection	23	0.04	-0.03-0.11	0.246	2.00
Maximum cluster size	19	0.20	0.09-0.30	0.001	48.20
Study design					
Study duration	27	-0.02	-0.09-0.06	0.677	-3.37
% population that is culture positive	15	0.34	-1.23-1.96	0.661	-5.92
% culture positive typed	19	0.22	-1.08-1.52	0.725	-5.41
Study size	27	0.03	-0.11-0.16	0.702	-3.31
24 loci (compared to 15)	27	-0.30	-0.590.01	0.04	13.58
Consent required	27	0.38	0.04-0.72	0.029	14.41

Table 3: Univariable metaregression showing the coefficients for change in the proportion of clustering and the percentage of between-study variation explained by variables describing the study design and setting.

^a Coefficients for the change in the proportion of clustering for each covariate. E.g. for a one unit increase in maximum cluster size, the proportion of clustering increases by 0.2.

^b The proportion of between-study variation explained by the univariate meta-regression.

Figure Caption

Figure 1: Results of systematic search, screening and data extraction.

Figure 2: Forest plot showing the proportion of clustering reported in each study by the number of MIRU-VNTR loci typed

The number of loci typed is categorised into 15 loci (m15), 15 loci with Spoligotyping (m15s), 24 loci (m24) and poligotyping 24 loci with Spoligotyping (m24s). The study reference is shown in the right hand column.



190x254mm (300 x 300 DPI)



190x254mm (300 x 300 DPI)

BMJ Open

Appendix 1: Medline/Embase search strategy

1. (tubercle adj3 (bacillus or bacilli)).mp. [mp=title, original title, abstract, name of substance word, subject heading word, unique identifier]

2. ((mycobacterium or mycobacteria) adj3 (bovis or africanum or microti or canetti)).mp.

3. exp tuberculosis/ or mycobacterium tuberculosis/ or tuberculosis.mp. or tb.mp. or Mtb.mp. or "M tuberculosis complex".mp.

4. or/1-3

5. Minisatellite Repeats/ or Genotype/ or Interspersed Repetitive Sequences/ or DNA Fingerprinting/ or Bacterial Typing Techniques/

6. "miru".mp. [mp=title, original title, abstract, name of substance word, subject heading word, unique identifier]

7. "vntr".mp. [mp=title, original title, abstract, name of substance word, subject heading word, unique identifier]

8. (miru adj3 vntr).mp. [mp=title, original title, abstract, name of substance word, subject heading word, unique identifier]

9. (mycobacterial adj3 interspersed adj3 repetitive adj3 units).mp. [mp=title, original title, abstract, name of substance word, subject heading word, unique identifier]

10. (dna adj3 fingerprinting).mp. [mp=title, original title, abstract, name of substance word, subject heading word, unique identifier]

11. ((strain adj3 type) or (strain adj3 typing) or (strain adj3 types)).mp. [mp=title, original title, abstract, name of substance word, subject heading word, unique identifier]

12. ((molecular adj3 typing) or (molecular adj3 strain adj3 typ*)).mp. [mp=title, original title, abstract, name of substance word, subject heading word, unique identifier]

13. (genotype or genotyping or genotypes).ti,ab.

14. (minisatellite adj3 repeat*).mp. [mp=title, original title, abstract, name of substance word, subject heading word, unique identifier]

15. molecular epidemiology/mt or (molecular adj3 epidemiology).mp. [mp=title, original title, abstract, name of substance word, subject heading word, unique identifier]

16. or/5-15

17. exp disease outbreaks/ or (outbreak adj3 analysis).mp. or (outbreak adj3 investigation).mp. or (outbreak adj3 management).mp. or (tuberculosis adj3 outbreak).mp. [mp=title, original title, abstract, name of substance word, subject heading word, unique identifier]

18. exp contact tracing/ or (contact adj3 tracing).mp. or (contact* adj3 traced).mp. or (contact adj3 screen*).mp. [mp=title, original title, abstract, name of substance word, subject heading word, unique identifier]

19. exp case management/ or (case adj3 management).mp. [mp=title, original title, abstract, name of substance word, subject heading word, unique identifier]

20. exp Risk Factors/

21. (risk adj3 factor*).mp. [mp=title, original title, abstract, name of substance word, subject heading word, unique identifier]

22. exp Epidemiologic Factors/

23. infectious disease transmission.mp. or exp Disease Transmission, Infectious/

24. exp case management/ or (case adj3 management).mp. [mp=title, original title, abstract, name of substance word, subject heading word, unique identifier]

25. program evaluation/ or evaluation studies as topic/ or (program adj3 evaluation).mp. or (programme adj3 evaluation).mp. [mp=title, original title, abstract, name of substance word, subject heading word, unique identifier]

26. public health practice/ or (public adj3 health).mp. [mp=title, original title, abstract, name of substance word, subject heading word, unique identifier]

27. ((tuberculosis adj3 control) or (tb adj3 control)).mp. [mp=title, original title, abstract, name of substance word, subject heading word, unique identifier]

28. (molecular adj3 surveillance).mp. [mp=title, original title, abstract, name of substance word, subject heading word, unique identifier]

29. exp cluster analysis/ or (cluster* adj3 rate*).mp. or (cluster* adj3 growth).mp. or (cluster* adj3 analysis).mp. or (cluster adj3 investigation).mp. or (proportion adj3 cluster*).mp. or (molecular adj3 cluster*).mp. [mp=title, original title, abstract, name of substance word, subject heading word, unique identifier]

30. ((recent adj3 transmission) or (transmission adj3 event*) or (transmission adj3 rate*) or (chain adj3 transmission) or (transmission adj3 setting*)).mp. [mp=title, original title, abstract, name of substance word, subject heading word, unique identifier]

31. or/17-30

32. 4 and 16	
33. 32 and 31	
34. limit 33 to yr="1998-Current"	
35. limit 34 to english language	
36. animals/	
37. humans/	
38. 36 not 37	

39. 35 not 38

Appendix 2: STROME-ID scores for the included studies

STROME-ID score ^a	
24	
32	
25	
18	
28	
31	
30	
19	
b	
32	
19	
b	
31	
22	
30	
32	
25	
23	
34	
36	
16	
23	
29	
26	
31	
23	
19	
	STROME-ID score ^a 24 32 25 18 28 31 30 19 b 32 19 b 31 22 30 32 25 23 34 36 16 23 29 26 31 23 19

^aIndividual studies score 1 for each element of checklist they had address

^bConference abstracts

PRISMA 2009 Checklist

Section/topic	#	Checklist item	Reported on page #
TITLE			
Title	1	Identify the report as a systematic review, meta-analysis, or both.	1
ABSTRACT	·		
2 Structured summary	2	Provide a structured summary including, as applicable: background; objectives; data sources; study eligibility criteria, participants, and interventions; study appraisal and synthesis methods; results; limitations; conclusions and implications of key findings; systematic review registration number.	
INTRODUCTION			
Rationale	3	Describe the rationale for the review in the context of what is already known.	3
) Objectives	4	Provide an explicit statement of questions being addressed with reference to participants, interventions, comparisons, outcomes, and study design (PICOS).	3
METHODS			
Protocol and registration	5	Indicate if a review protocol exists, if and where it can be accessed (e.g., Web address), and, if available, provide registration information including registration number.	
Eligibility criteria	6	Specify study characteristics (e.g., PICOS, length of follow-up) and report characteristics (e.g., years considered, language, publication status) used as criteria for eligibility, giving rationale.	4
Information sources	7	Describe all information sources (e.g., databases with dates of coverage, contact with study authors to identify additional studies) in the search and date last searched.	4
) Search	8	Present full electronic search strategy for at least one database, including any limits used, such that it could be repeated.	appendix
Study selection	9	State the process for selecting studies (i.e., screening, eligibility, included in systematic review, and, if applicable, included in the meta-analysis).	4
Data collection process	10	Describe method of data extraction from reports (e.g., piloted forms, independently, in duplicate) and any processes for obtaining and confirming data from investigators.	4
3 Data items	11	List and define all variables for which data were sought (e.g., PICOS, funding sources) and any assumptions and simplifications made.	5
Risk of bias in individual studies	12	Describe methods used for assessing risk of bias of individual studies (including specification of whether this was done at the study or outcome level), and how this information is to be used in any data synthesis.	5
Summary measures	13	State the principal summary measures (e.g., risk ratio, difference in means).	5
Synthesis of results	14	Describe the methods of handling data and combining results of studies, if done, including measures of consistency (e.g., I ² for each meta-analysis.	5
Page 25 of 25

Page 1 of 2



PRISMA 2009 Checklist

5 6 7	Section/topic	#	Checklist item	Reported on page #
8 9	Risk of bias across studies	15	Specify any assessment of risk of bias that may affect the cumulative evidence (e.g., publication bias, selective reporting within studies).	5
1(1 ⁻ 12	Additional analyses	16	Describe methods of additional analyses (e.g., sensitivity or subgroup analyses, meta-regression), if done, indicating which were pre-specified.	5
13	RESULTS			
14 15 16	Study selection	17	Give numbers of studies screened, assessed for eligibility, and included in the review, with reasons for exclusions at each stage, ideally with a flow diagram.	Figure 1
17	Study characteristics	18	For each study, present characteristics for which data were extracted (e.g., study size, PICOS, follow-up period) and provide the citations.	15
20	Risk of bias within studies	19	Present data on risk of bias of each study and, if available, any outcome level assessment (see item 12).	15
22	Results of individual studies	20	For all outcomes considered (benefits or harms), present, for each study: (a) simple summary data for each intervention group (b) effect estimates and confidence intervals, ideally with a forest plot.	Figure 2
24	Synthesis of results	21	Present results of each meta-analysis done, including confidence intervals and measures of consistency.	18
25	Risk of bias across studies	22	Present results of any assessment of risk of bias across studies (see Item 15).	15
27	Additional analysis	23	Give results of additional analyses, if done (e.g., sensitivity or subgroup analyses, meta-regression [see Item 16]).	18
28				
30	Summary of evidence	24	Summarize the main findings including the strength of evidence for each main outcome; consider their relevance to key groups (e.g., healthcare providers, users, and policy makers).	7
33 34	Limitations	25	Discuss limitations at study and outcome level (e.g., risk of bias), and at review-level (e.g., incomplete retrieval of identified research, reporting bias).	7-8
35 36	Conclusions	26	Provide a general interpretation of the results in the context of other evidence, and implications for future research.	8
37	FUNDING			
39 40	Funding	27	Describe sources of funding for the systematic review and other support (e.g., supply of data); role of funders for the systematic review.	8
41 42 <i>From:</i> Moher D, Liberati A, Tetzlaff J, Altman DG, The PRISMA Group (2009). Preferred Reporting Items for Systematic Reviews and Meta-Analyses: The PRISMA Statement. PLoS M 43 doi:10.1371/journal.pmed1000097				6(6): e1000097.

For more information, visit: www.prisma-statement.org.

Page 2 of 2 For peer review only - http://bmjopen.bmj.com/site/about/guidelines.xhtml