

Supplementary Fig. 1. Exposure to saturating concentration of AHL does not affect stochastic AHL-induction at early stages of biofilm development of *P. putida* IsoF. Flow cells were initiated with IsoF carrying the GFP-based AHL sensor. (A) After microcolonies were formed, only a few cells start to produce AHLs. Furthermore, the appearance of induced-single-cells in the proximity of microcolonies suggests they left the colony once activated. (B) When the experiment was performed under uniform amount of autoinducer in the medium, 0.5μ M 3-oxo-C10-HSL, a similar pattern in terms of induction heterogeneity was observed, although induction occurred earlier and in a higher proportion of cells. Scale bar: 5 μ m.



Supplementary Fig. 2. Structure of putisolvin biosynthetic cluster and schematic outline of the construction of the conditional *psoABC* mutant PL2, in which natural *psoA* promoter has been replaced by the rhamnose-inducible promoter P_{rhaB} . In addition, the strain carries genes required for the activation of the P_{rhaB} promoter (*rhaS* and *rhaR*); aacC1, aminoglycoside acetyltranferase (gentamycin resistance); *oriT*, origin of transfer; SD, Shirne-Delgarno.



Supplementary Fig. 3. Swarming motility of mixed cultures. The wild type (IsoF) was combined with itself, the putisolvin-deficient PL11, or the flagella-deficient mutant GC25 strain on swarming plates. (*A*) In 1:1 mixes, the displacement of the swarming colony remains unaffected irrespective of the strain mixture. When mixing IsoF at different ratios with: (*B*) itself, IsoF::gfp; (*C*) with PL11::gfp; or (*D*) with GC25::gfp, the swarming performance was unaffected across most mixing ratios. Only when the proportion of IsoF was decreased to 10%, swarming displacement is slightly reduced in mixtures with PL11::gfp and GC25::gfp. These findings support our conclusions that PL11::gfp and GC25::gfp cannot swarm along with IsoF wild type. Mean values of three independent experiments are shown with s.e.m.



Supplementary Fig. 4. Putisolvins are not public goods in swarming colonies. In mixed colonies with IsoF, mutants deficient for (*A*) putisolvin (PL11:gfp) and (*B*) flagella (GC25:gfp) production remain located in the center of the plate, while IsoF swarms to the edges. This pattern remained consistent across a wide range of mixing ratios, confirming that putisolvin is a private good under swarming conditions. Mean values of three independent experiments are shown with s.e.m.

Supplementary Table 1. Bacterial strains and plasmids used in this study

Strain or plasmid	Relevant properties	Source or reference
E. coli		
MT102	araD139 (ara-leu)7679 ∆lac thi hsdR	This Laboratory
CC118(λpir)	Δ (ara-leu) araD Δ lacX74 galE galK phoA20 thi-1	1
	rpsE rpoB argE(Am) recA1 λpir lysogen	2
HB101	recA thi pro leu hsd M⁺; Sm ^ĸ	2
P. putida		3
	wild type; isolated from tomato roots; AHL	4
	ppul mutant of IsoF; Km ^R	This Laboratory
	ppuR mutant of IsoF; Km	This Laboratory
GC25	fliM mutant of IsoF: Gm ^R	This study
PI 2	conditional psoA mutant of IsoF' Press psoA' Gm ^R	This study
IsoF-mcherry	IsoF tagged with mcherry. Gm ^R	This laboratory
leer menery		ine laberatory
Plasmids		
pEX18	gene replacement vector; Gm ^R	5
pEX18psoA	pEX18 containing an internal fragment of <i>psoA</i> in the Smal site; Gm ^R	This study
pSHAFT2	pUTmini-Tn <i>5</i> Cm with deleted BgIII fragment;	This laboratory
pSHAFT2fliM	pSHAFT2 containing an internal fragment of <i>fliM</i> in the Stul site: Gm^R	This study
nSC200	oriR6K: rhampose-inducible promoter P _{are} : Tp ^R	6
pSC200 pSC200Gm	Gm ^R resistance cassette cloned into Xhal/Knnl sites	This study
p002000m	of pSC200; Gm ^R	The olday
pPO1	pSC200Gm carrying internal <i>psoA</i> fragment; Gm ^R	This study
pPLlas	pUT/mini with Km ^r :: <i>lasR</i> -P _{lac} -P _{lasB} -gfp(ASV)-T ₀ -T ₁ in the NotI site	7
pRPL4las	RP4 with Km ^R :: <i>las</i> R-P _{lac} -P _{lasB} -gfp(ASV)	7
pGA-G1	Promoter probe vector carrying a promoterless <i>gfp</i> - mut3 gene: Gm ^R	8
pLUM1	P_{nsoA} promoter cloned into pGA-G1: Gm ^R	This study
pLUM3	pBBR1MCS-3 carrying a P_{psoA} :: <i>cfp</i> fusion; Tc ^R	This study
pBAH8	P _{A1/04/03} -gfpmut3 fusion in pBBR1MCS-2; Km ^R	9
pBBR1MCS-3	Broad-host-range plasmid, <i>lac</i> Zα; Tc ^R	10
pBBR1MCS-5	Broad-host-range plasmid, <i>lac</i> Zα; Gm ^R	10
pRK600	ColE1 RK2-Mob ⁺ RK2-Tra ⁺ ; helper plasmid; Cm ^R	11

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