

Supporting Information

Gehrke et al. 10.1073/pnas.1420208112

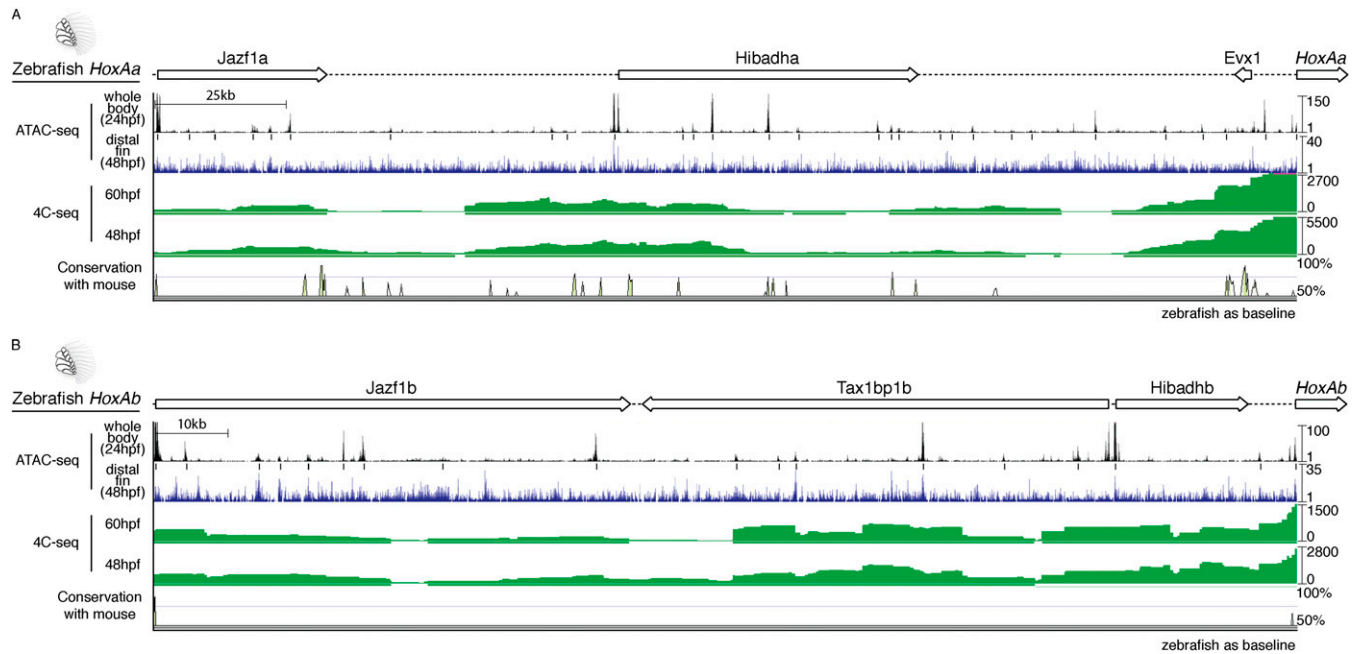


Fig. S1. Epigenetic profiling and sequence conservation of the zebrafish *hoxAa* (A) and *hoxAb* clusters (B). Schematic representations of the zebrafish *hoxA* clusters are shown at the top. ATAC-seq for both whole-body and distal fin are shown, combined with 4C-seq data to reveal areas of interaction with the *hoxa13a* and *hoxa13b* genes.

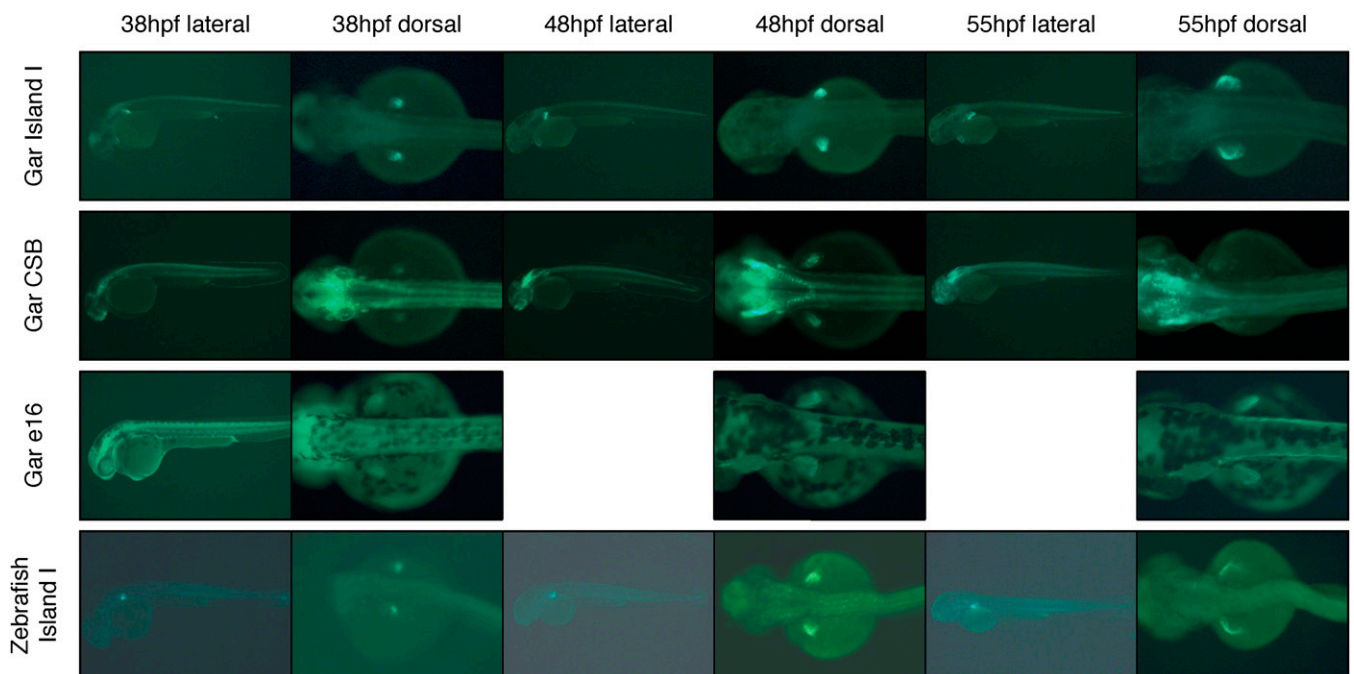


Fig. S2. Whole-body views of transgenic zebrafish. Lateral and dorsal views are shown for transgenic animals at 38, 48, and 55 hpf.

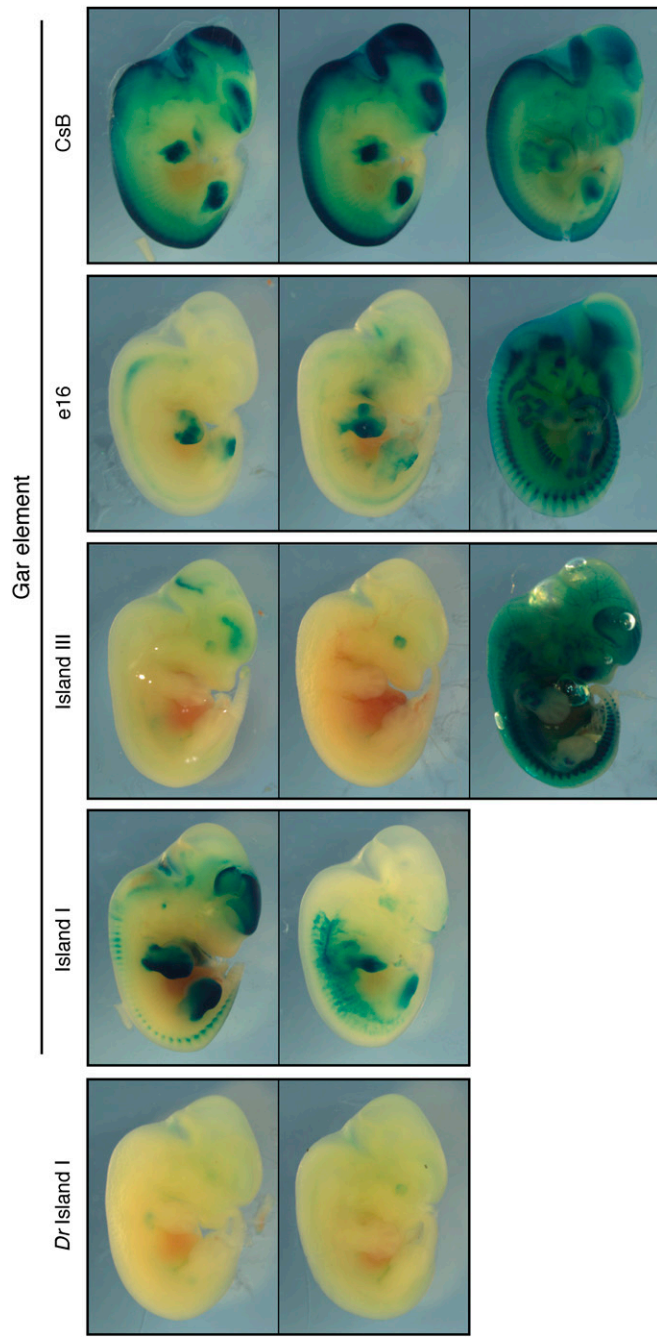


Fig. S3. Summary of mouse injections. All injected mouse embryos that were positive for LacZ staining are provided at stage e12.5.

Table S1. Comparisons between mouse and zebrafish ATAC-seq and previously published reports

| Animal | Overlap with ATAC-seq |
|--------------------------------|-----------------------|
| Mouse | |
| Peaks | |
| H3K4Me1 (LICR ENCODE) | 26,210/52,705 (49.7) |
| H3K27ac (1) | 18,082/25,861 (69.9) |
| Vista Enhancer Database (limb) | 54/67 (80.6) |
| Vista Enhancer Database (all) | 138/250 (55.2) |
| Zebrafish | |
| Peaks | |
| H3K27ac (2) | 20,387/21,839 (93.3) |

Values are number (percentage).

1. Cotney J, et al. (2012) Chromatin state signatures associated with tissue-specific gene expression and enhancer activity in the embryonic limb. *Genome Res* 22(6):1069–1080.
2. Bogdanovic O, et al. (2012) Dynamics of enhancer chromatin signatures mark the transition from pluripotency to cell specification during embryogenesis. *Genome Res* 22(10): 2043–2053.

Table S2. Summary of zebrafish injections

| Genomic region | No. injected | Stage screened (hpf) | No. fin positive | No. fin negative | % w/fin signal | Raise for line? |
|----------------|--------------|----------------------|------------------|------------------|----------------|-----------------|
| Lo CsB | 172 | 48 | 66 | 106 | 62.2 | Yes |
| Lo Island I | 152 | 48 | 21 | 131 | 13.8 | Yes |
| Lo e16 | 183 | 48 | 26 | 157 | 14.2 | Yes |
| Lo Island II | 160 | 48 | 4 | 156 | 2.5 | No |
| Lo Island III | 186 | 36, 48 | 12 | 174 | 6.4 | No |
| Lo Island IV.1 | 171 | 48 | 6 | 165 | 3.4 | No |
| Lo Island IV.2 | 168 | 48 | 4 | 164 | 2.4 | No |
| Dr Island I | 136 | 48 | 3 | 133 | 2.2 | Yes |

Table S3. List of primers used

| Name | Locus | Sequence |
|------------------|------------------------------------------|------------------------|
| Zebrafish | | |
| Dr_Island_I_F | Zebrafish Island I | AGCAACGCATGTCTTTCAACA |
| Dr_Island_I_R | Zebrafish Island I | ATAACGTTGTGTGCCTGCTG |
| Gar | | |
| Lo_Island_I_F | Gar Island I | TGGCCTACAACACCAGTGAA |
| Lo_Island_I_R | Gar Island I | CAGATTTTGTGCGTTTCTCCT |
| Lo_CsB_F | Gar CsB | GGAGTCTCCCACAAGGTGAA |
| Lo_CsB_R | Gar CsB | CGAAGGCTCTGCACTACTCA |
| Lo_Island_II_F2 | Genomic “area” of mouse Island II | GAGGTTGTGGGCTGTCCAAA |
| Lo_Island_II_R2 | Genomic “area” of mouse Island II | CCACATTTGTGAAAATTCTCTG |
| Lo_Island_IV_F2 | Genomic “area” of mouse Island IV part 1 | GCTTGAAAGCAACTGCATC |
| I4_Split_R | Genomic “area” of mouse Island IV part 1 | GAGATGGCAACGCCATTATGT |
| I4_Split_F | Genomic “area” of mouse Island IV part 2 | CCGTGTGATCCAAAGCAATA |
| Lo_Island_IV_R | Genomic “area” of mouse Island IV part 2 | TTGGGCTGACCTGCTTTTAT |
| Lo_e16_R | HoxA enhancer e16 | GTGATTTTCTCGGCATTTGG |
| Lo_e16_F | HoxA enhancer e16 | CACCGACTTTGCTGTGTCAT |
| Gar Island III | Lo_Island_III_Long_F | GTGAGCCATGAGATGTACCG |
| Gar Island III | Lo_Island_III_Long_F | GTA AACACTCCGGCACCTT |
| Mouse | | |
| Tg1_F | Mouse Island I | TTCAGACTAGGCCCTCCAGA |
| Tg1_R | Mouse Island I | GACAGTGGGGACAACCCCTAA |
| Tg2_F | Mouse Island II | GTGTGTGGCTAGGGATTCT |
| Tg2_R | Mouse Island II | AGAGAGGGCTCGTCACTCAA |
| Tg4_F | Mouse Island IV | GAGTTAGCCACTCAGCCATGT |
| Tg4_R | Mouse Island IV | TGGTGTCTTCTCCATTCTG |