

Supporting Information

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SI Materials and Methods

Clonal Analysis. *wts* mutant clones were generated by crossing *hs-FLP; ubi-GFP, FRT82B/TM6B* with *wts^{XI} FRT82B/TM6B*. Flip-out ectopic expression clones were generated by crossing *UAS-transgenes with y w hs-FLP, act > y⁺>Gal4 UAS-GFP*. Clones were induced at the second instar: heat shock for 30 min at 37 °C 48–72 h after egg laying (AEL); dissection was performed 36 h or 72 h after clone induction.

Clone size measurements were done as follows: we randomly picked up four clones from each wing disk, excluding clones along the dorsal/ventral boundary (low Yki activity due to reduced tension), and four wing discs were examined for each genotype. In total, 16 clones of indicated genotype were picked in double-blind experiments, and the pixels of each clone were measured using Photoshop software (Adobe), and the graphs were generated using GraphPad Prism 5.

qRT-PCR. Total RNA was extracted from wing discs of indicated genotype or culture S2 cells using PureLink RNA Mini kit (Ambion); cDNA was synthesized using PrimeScript RT Master mix (TaKaRa); and quantitative PCR was performed using SYBR Premix Ex Taq II (TaKaRa) on a Stratagene MX3000P system (Stratagene). The following primer sequences were used for RT-PCR:

tublin-FW: GCTGTTCCACCCCGAGCAGCTGATC
tublin-RV: GGCGAACTCCAGCTTGGACTTCTTGC
Rho1-FW: GTGGATGGCAAACAGGTGGAGC
Rho1-RV: GCGAATCGGGTGAATCCACTGAG.

Luciferase Reporter Assay. The *luciferase* reporters were generated through cloning the dissected elements of Rho1 first intron (E1, E2, and E3 in Fig. 4I) into KpnI/XhoI sites of *pGL3-Basic* Vector containing the hsp70 basal promoter. For luciferase assays, S2 cells were transfected with the above luciferase reporters and *copia-renilla* reporter in 12-well plates together with yki-HA-expressing pAc5.1/V5-HisB expressing plasmids. Cells were incubated for 36 h after transfection. Luciferase assays were performed using the Dual-Luciferase Reporter Assay system (Promega). Dual-luciferase measurements were performed using FlexStation 3 (Molecular Devices).

Western Blotting and ChIP. For Western blot analysis, 20 third instar wing discs were lysed in 1× SDS/PAGE sample loading buffer (Beyotime) and proceeded by the standard Western blot protocol. Proteins were probed with the following primary antibodies: anti-Rho1 (1:1,000; DSHB) and β-tubulin (1:3,000; CMC Scientific). After incubation with secondary antibodies (1:3,000; CST), signals were visualized by enhanced chemiluminescence (Beyotime). Chromatin immunoprecipitation was carried out as described in the protocol of the Magna ChIP G Tissue kit (Millipore). Transient transfections were performed with equal amounts of DNA (2.5 μg per plasmids) using Effectene transfection reagent (Qiagen) for S2 cells in 100 mm × 20 mm. HA-Tag (CMCTAG), Sd (a kind gift from Kirsten A. Guss, Dickinson College, Carlisle, PA), and mouse IgG (CST) were used for immunoprecipitation of ChIP assay.

hs-Flp; act-Gal4 UAS-GFP; UAS-Yki^{S168A}, puc-LacZ

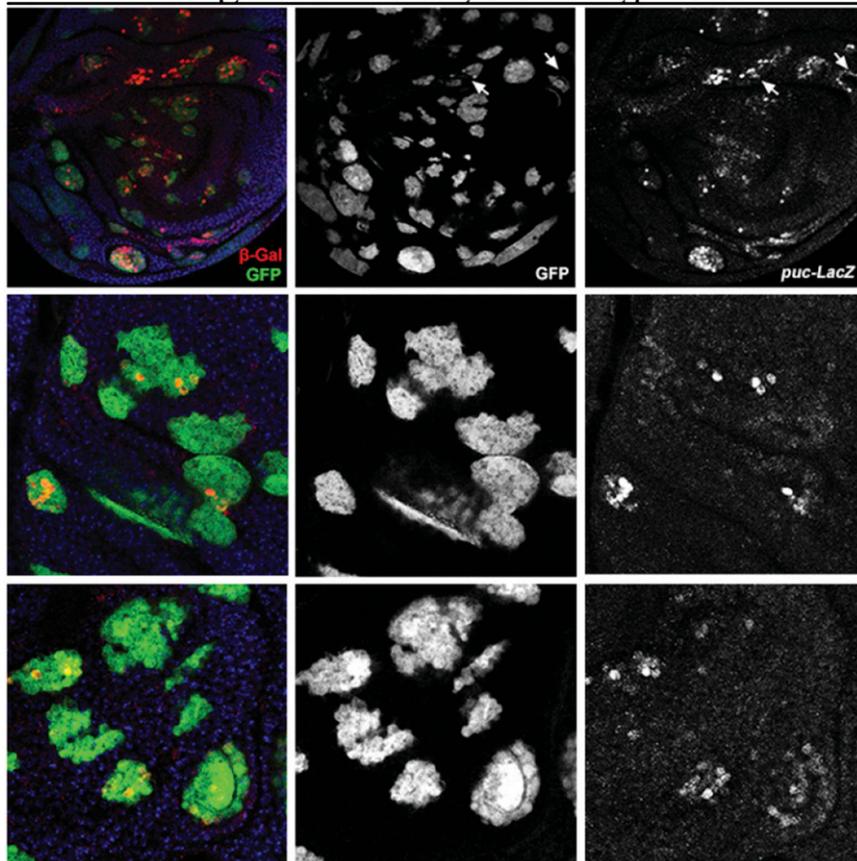


Fig. S1. Yki-expressing clones induces JNK activation throughout the wing disk. Yki^{S168A}-expressing Flp-out clones up-regulate *puc* transcription throughout the wing disk in an autonomous manner, although occasionally, nonautonomous JNK activation is observed in surrounding cells (first row, indicated by white arrow).

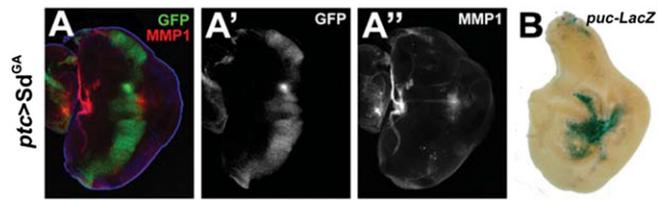


Fig. 58. Sd^{GA} overexpression induces JNK activation. Third instar wing discs stained with MMP1 (A) or X-Gal (B). Expression of an activated form of Sd (Sd^{GA}) up-regulates MMP1 (A) and *puc*-LacZ expression (B).

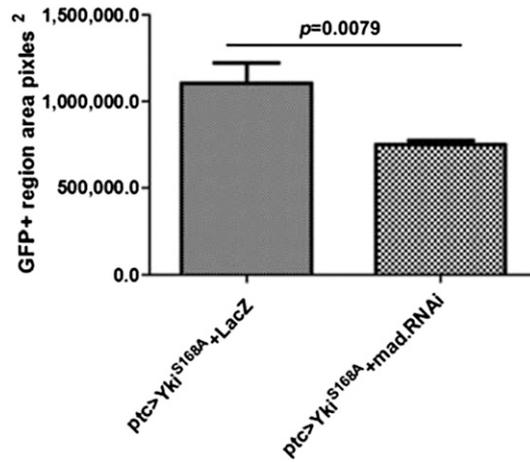


Fig. 59. *mad* is required for Yki-induced overgrowth. Statistical analysis of GFP⁺ regions shown in Fig. 4 A and C. Error bars are +SD; two-tailed, unpaired Student's *t* test.