# SUPPLEMENTAL INFORMATION

# Blockade of VEGF-C and VEGF-D modulates adipose tissue inflammation and improves metabolic parameters under high-fat diet

Sinem Karaman, Maija Hollmén, Marius R. Robciuc, Annamari Alitalo, Harri Nurmi, Bettina Morf, Dorina Buschle, H. Furkan Alkan, Alexandra M. Ochsenbein, Kari Alitalo, Christian Wolfrum and Michael Detmar

#### SUPPLEMENTAL EXPERIMENTAL METHODS

#### Whole-mount immunofluorescence stains

Approximately 10 mm long pieces of small intestine were dissected from mice. The intestines were cut longitudinally, rinsed with PBS and fixed in 4% PFA for 2 h at 4°C. Samples were washed with PBS several times for 30 min, followed by blocking with a mixture of 5% non-immune donkey serum, 0.1% Triton-X, 1% BSA and 0.05% NaN<sub>3</sub> in PBS for 2 h, and by incubation with the respective primary antibodies overnight. The samples were then incubated with Alexa 488- and 594- conjugated secondary antibodies (Invitrogen; 1:200 dilution) for 2 h, washed for 2 h with PBS and mounted on glass slides in VectaShield (Vector). Z-stack images were acquired with confocal imaging, which was performed on a Zeiss LSM 710-FCS confocal microscope equipped with a  $10 \times 0.3$ NA EC Plan-Neofluar objective and a  $20 \times 0.8$  NA Plan-Apochromat objective (Carl Zeiss). Images were acquired using the Zeiss ZEN 2009 software and processed using Imaris software (version 7.1.1, Bitplane) or ImageJ (NIH).

## Computed tomography analyses of adipose tissue

After 19 weeks of HFD, computed tomography (CT) scans were performed as described with slight modifications [1]. Briefly, mice were kept under isoflurane anesthesia and were imaged with a La Theta LCT-100 (Aloka) Micro-CT scanner. A pitch size of 1 mm was used and the tube voltage of the X-ray source was adjusted to 50 kV with 1 mA constant current. The volumes of SWAT and EWAT were corrected and analyzed with the LaTheta software (V 2.10, Aloka), and the corresponding weights were calculated using a density factor of 0.92 g/cm<sup>3</sup>.

#### Fasting insulin and glucose measurements

Mice that were on chow or HFD for 10, 20 or 38 weeks were fasted for 12 h during the light cycle (6 a.m. to 6 p.m.). Blood glucose was measured with a Contour glucometer (Bayer HealthCare). Serum insulin levels were measured with an ultra sensitive mouse Insulin ELISA kit (Crystal Chem Inc.). The homeostatic model assessment of insulin resistance (HOMA-IR) index was calculated using fasting glucose and insulin concentrations with the following formula: fasting blood glucose (mmol/L) × fasting insulin ( $\mu$ U/mL)/22.5.

### **Fasting - refeeding studies**

Mice that were on chow or HFD for 38 weeks were fasted for 12 h during the light cycle (6 a.m. to 6 p.m.). Mice were given access to food after collection of fasting blood. Three hours after re-feeding, blood samples were collected again. Free-fatty acid levels in the serum were measured with NEFA-HR kit (Wako) following the manufacturer's instructions.

## Cytokine analysis

For the conditioned media measurements,  $CD11b^+$  cells from the SVF were isolated by MACS (Miltenyi Biotech) from 24-week-old mice on chow diet (WT and sR3, *n*=4 each). Cells were cultured in DMEM (Gibco), supplemented with 1% mouse serum (obtained from the corresponding macrophage donor). After 24 h, conditioned media were collected, centrifuged and the supernatants were stored at -20 °C until used. The mouse Th1/Th2 10plex FlowCytomix kit (eBioscience) was used to measure the levels of cytokines in macrophage conditioned media, following the manufacturer's instructions.

## ELISA

The levels of sR3 protein were detected as described previously with slight modifications [2]. ELISA plates (Nunc) were coated with anti-human IgG (Sigma, 0.5  $\mu$ g/mL), washed 6 times and blocked with 1% non-fat milk powder, 1% gelatin (Hänseler AG) in PBS for 1 h. Lysates of SWAT, EWAT and liver were incubated for 1 h. After washing 4 times, biotinylated anti-human VEGFR-3 antibody (R&D Systems, 1:4,500) was added for 2 h. After washing 4 times, streptavidin conjugated horseradish peroxidase (GE Healthcare, 1:5,000) was added for 30 min, followed by addition of BM Blue POD substrate (Roche). The reaction was stopped with 2 M H<sub>2</sub>SO<sub>4</sub> and absorbance was determined at 450 nm using a Sunrise microplate reader (Tecan).

## Tamoxifen administration and X-gal staining

Flt4-Cre-ERT2;ROSA26-LSL-LacZ double knock-in mice on mixed (129/C57BL/6) background [3] were used to assess the Flt4 (VEGFR-3) expression in tissues. Tamoxifen injections (50  $\mu$ L of 1 mg/mL) were done on postnatal days P1, P2 and P3 intragastrically. The mice were sacrificed on day P11 and tissues were harvested for X-gal staining. The tissues were fixed in 2% formaldehyde, 0.2% glutaraldehyde and 0.02% NP-40 in PBS at 4 °C for 1 h, washed with PBS, and then stained overnight with 1 mg/mL (1:500) 5- bromo-4-chloro-indolyl- $\beta$ -D-galactopyranoside (X-gal) in the preheated (37 °C) X-gal staining buffer (5mM potassium ferrocyanide (K<sub>4</sub>[Fe(CN)<sub>6</sub>]), 5mM potassium ferricyanide (K<sub>3</sub>[Fe(CN)<sub>6</sub>]), 2mM MgCl<sub>2</sub>, 0.02% Np-40 in PBS) at 37 °C. The following day the tissues were post-fixed with 4% PFA in PBS for 1 h, washed with PBS for 1 h and imaged.

## SUPPLEMENTAL TABLES

**Table S1:** Serum parameters of 12-h fasted wildtype and sR3 mice on HFD or chow for 38 weeks. Data are expressed as mean  $\pm$  SEM. Parentheses indicate number of mice per group. \**P*<0.05, one-tailed Student's *t*-test, sR3 mice compared to WT under same diet conditions. † *P*<0.05, one-tailed Student's *t*-test, fasting level compared to post-prandial level.

	Chow		HFD	
	Wildtype	sR3	Wildtype	sR3
Fasting glucose (mmol/L)	7.1 ± 0.6 (5)	6.5 ± 0.4 (5)	8.0 ± 0.6 (5)	7.8 ± 0.4 (6)
Fasting insulin (ng/mL)	6.8 ± 2.2 (5)	1.6 ± 0.4 (6) *	12.7 ± 1.7 (5)	7.8 ± 1.5 (6) *
HOMA-IR	59.5 ± 23.2 (5)	13.0 ± 3.0 (6) *	119.5 ± 21.5 (5)	68.1 ± 11.2 (6) *
FFA (mmol/L) (fasting)	1.1 ± 0.1 (5)	$1.1 \pm 0.1$ (6)	$1.0 \pm 0.1$ (5)	$0.7 \pm 0.1$ (6)
FFA (mmol/L) (post-prandial)	0.8 ± 0.1 (5)	$0.6 \pm 0.2$ (6)†	1.1 ± 0.2 (5)	0.9 ± 0.1 (6)

Primer	Sequence	Reference		
36b4 Forward	5'-AGATTCGGGATATGCTGTTGGC-3'	Primer bank ID: 6671569a1		
36b4 Reverse	5'-TCGGGTCCTAGACCAGTGTTC-3'			
Inos Forward	5'-CCTGGTACGGGCATTGCT-3'	F 4 1		
Inos Reverse	5'-GCTCATGCGGCCTCCTT-3'	[4]		
Mrc1 Forward	5'-GCTTCCGTCACCCTGTATGC-3'	Primer bank ID: 224967061c2		
Mrc1 Reverse	5'-TCATCCGTGGTTCCATAGACC-3'			
Tnf Forward	5'-CAGGCGGTGCCTATGTCTC-3'	[6]		
Tnf Reverse	5'-CGATCACCCCGAAGTTCAGTAG-3'	[5]		
CD11b Forward	5'-CCTTGTTCTCTTTGATGCAG-3'			
CD11b Reverse	5'-GTGATGACAACTAGGATCTT-3'	[6]		
CD11c Forward	5'-CCAAGACATCGTGTTCCTGATT -3'	Primer bank ID:		
CD11c Reverse	5'- ACAGCTTTAACAAAGTCCAGCA-3'	118130485c3		
CD68 Forward	5'- CCAATTCAGGGTGGAAGAAA -3'			
CD68 Reverse	5'- CTCGGGCTCTGATGTAGGTC-3'	[7]		
<i>Il6</i> Forward	5'-TCTATACCACTTCACAAGTCGGA-3'			
116 Reverse	5'- GAATTGCCATTGCACAACTCTTT-3'	[8]		
Arg1 Forward	5'-CTCCAAGCCAAAGTCCTTAGAG-3'	Primer bank ID:		
Argl Reverse	5'-AGGAGCTGTCATTAGGGACATC-3'	7106255a1		
Ym1 Forward	5'-GGGCATACCTTTATCCTGAG-3'	[9]		
Ym1 Reverse	5'-CCACTGAAGTCATCCATGTC-3'			
Vegfc Forward	5'-GGGGGCGAGGTCAAGGCTTTT-3'			
Vegfc Reverse	5'-CCTGGTATTGAGGGTGGGCTGC-3'	Self-designed		
Vegfd Forward	5'-AGCGAACATGGACCAGTGAAGGATT-3'			
Vegfd Reverse	5'-CCTCCAAACTAGAAGCTGCTCGGA-3'	Self-designed		
Vegfr2 Forward	5'-TTTGGCAAATACAACCCTTCAGA-3'	Primer bank ID:		
Vegfr2 Reverse	5'-GCAGAAGATACTGTCACCACC-3'	27777648a1		
Vegfr3 Forward	5'-CTGGCAAATGGTTACTCCATGA-3'	Primer bank ID:		
Vegfr3 Reverse	5'-ACAACCCGTGTGTCTTCACTG-3'	6679813a1		
Nrp1 Forward	5'-GACAAATGTGGCGGGACCATA-3'	Primer bank ID: 6679134a1		
Nrp1 Reverse	5'-TGGATTAGCCATTCACACTTCTC-3'			
Nrp2 Forward	5'-CCAGAACTGTGAGTGGATTGTC-3'	Self-designed		
Nrp2 Reverse	5'-CCATCCCGAATCTCAATGAAGTC-3'			

**Table S2:** Primers used for the quantitative real-time PCR analyses.

# SUPPLEMENTAL FIGURES and FIGURE LEGENDS

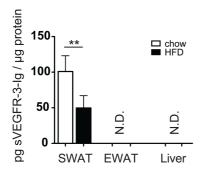


Figure S1: sR3 protein is detectable in SWAT, but not in EWAT or liver. ELISA analysis of adipose tissue and liver lysates showing detectable amounts of sR3 protein only in SWAT (n=3-4 each group). N.D., not-detected. \*\*P<0.01, two-tailed Student's *t*-test, WT under HFD compared to WT under chow diet. Data are mean  $\pm$  SD.

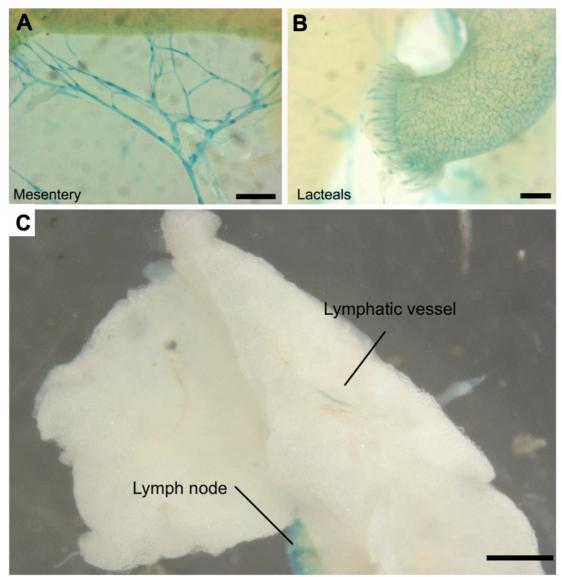


Figure S2: Whole mount X-gal staining in Flt4CreErt2 Rosa26LacZ double knock-in mice reveals lack of VEGFR-3 in adipose tissue. VEGFR-3 positive lymphatic vessels are visible in different tissues such as the mesentery (A) and the lacteals (B). (C) Lack of VEGFR-3 expressing cells in the adipose tissue except for the collecting lymphatic vessels and lymph nodes. Scale bars: A, B = 500  $\mu$ m; and C = 200  $\mu$ m.

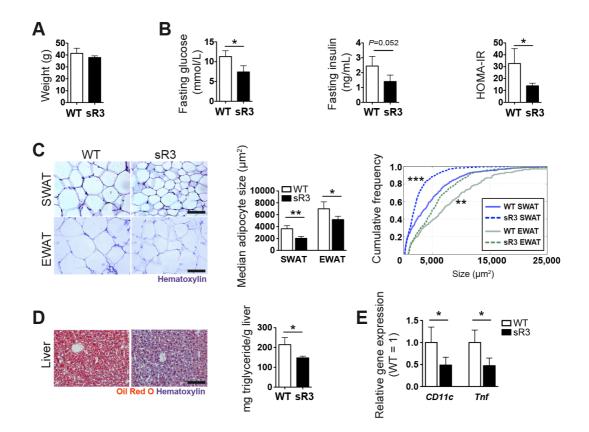


Figure S3: Increased insulin sensitivity of the adipose tissue of sR3 mice on the C57/BL6 background (sR3 B6) on HFD. (A) No difference in weight after 10 weeks on HFD (n=3-4). (B) Lower fasting glucose and insulin levels that result in reduced HOMA-IR indices (n=3-4). (C) sR3 B6 mice had smaller SWAT and EWAT adipocytes than WT mice, which was confirmed by the median adipocyte size comparison and cumulative frequency plots (n=3-4, \*\*\*P<0.001; two-sample Kolmogorov-Smirnov test). (D) sR3 B6 mice also had lower ectopic lipid accumulation in the liver as shown by Oil Red O staining and triglyceride measurements (n=3-4). (E) Macrophages isolated from SWAT of sR3 B6 mice showed reduced expression of the M1 markers *CD11c* and *Tnf* (n=3-4). Scale bars = 100 µm. \*P<0.05, \*\*P<0.01, two-tailed Student's *t*-test compared to WT. All data are mean±SD.

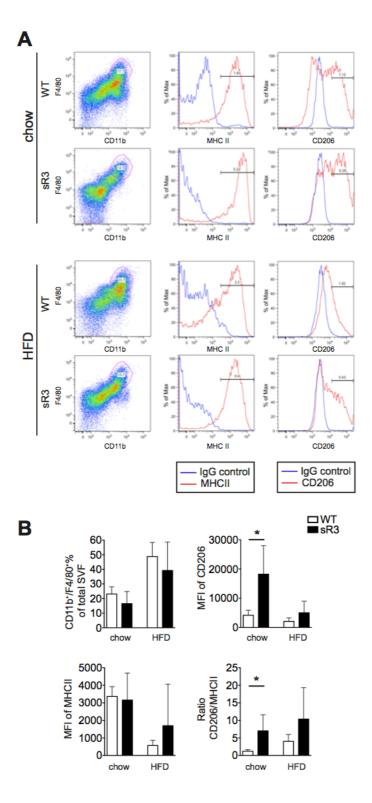


Figure S4: FACS analysis of EWAT macrophages. (A) Representative FACS plots. (B) FACS analyses showed a higher ratio of CD206/MHC II expression levels in EWAT macrophages of sR3 mice as compared to WT mice (n=3-7). \*P<0.05, two-tailed Student's *t*-test compared to WT under same diet. All data are mean±SD.

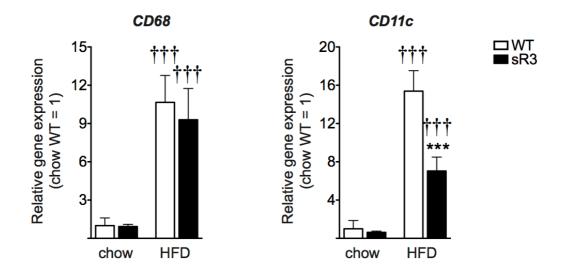


Figure S5: sR3 mice have reduced *CD11c* expression in SWAT under HFD. Comparable expression of *CD68* (common macrophage marker) between sR3 and WT mice under both chow and HFD. *CD68* expression levels were significantly upregulated after HFD. Expression of *CD11c* (an M1 marker) was significantly lower in sR3 mice than in WT mice after HFD (n=5-7 each group). \*\*\*P<0.001 two-tailed Student's *t*-test compared to WT of the same diet, †††P<0.001 two-tailed Student's *t*test compared to chow diet of the same genotype. All data are mean±SD.

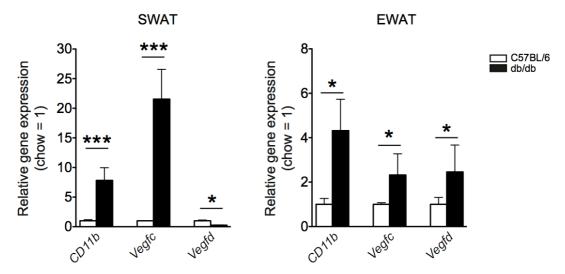


Figure S6: VEGF-C and -D are upregulated in the EWAT of db/db mice. Quantitative real-time PCR analyses of *Vegfc* and *Vegfd* expression together with *CD11b* in total SWAT and EWAT of lean (C57BL/6) and obese (db/db) mice showed that *Vegfc* and *Vegfd* mRNA levels were also upregulated in EWAT in obesity (n=3-5 each group). \*P<0.05, \*\*\*P<0.001 one-tailed Student's *t*-test compared to lean (C57BL/6) mice. All data are mean±SD.

## SUPPLEMENTAL REFERENCES

- [1] Hillebrand, J.J.G., Langhans, W., and Geary, N., 2010. Validation of computed tomographic estimates of intra-abdominal and subcutaneous adipose tissue in rats and mice. Obesity (Silver Spring) 18:848–853.
- [2] Mäkinen, T., Jussila, L., Veikkola, T., Karpanen, T., Kettunen, M., Pulkkanen, K., et al., 2001. Inhibition of lymphangiogenesis with resulting lymphedema in transgenic mice expressing soluble VEGF receptor-3. Nature Medicine 7:199–205.
- [3] Aschen, S.Z., Farias-Eisner, G., Cuzzone, D.A., Albano, N.J., Ghanta, S., Weitman, E.S., et al., 2014. Lymph node transplantation results in spontaneous lymphatic reconnection and restoration of lymphatic flow. Plastic and Reconstructive Surgery 133:301–310.
- [4] Hamalainen, M., Nieminen, R., Vuorela, P., Heinonen, M., and Moilanen, E., 2007. Anti-inflammatory effects of flavonoids: genistein, kaempferol, quercetin, and daidzein inhibit STAT-1 and NF-kappaB activations, whereas flavone, isorhamnetin, naringenin, and pelargonidin inhibit only NF-kappaB activation along with their inhibitory effect on iNOS expression and NO production in activated macrophages. Mediators of Inflammation 2007:45673.
- [5] Roth Flach, R.J., Matevossian, A., Akie, T.E., Negrin, K.A., Paul, M.T., and Czech, M.P., 2012. Beta3-adrenergic receptor stimulation induces E-selectinmediated adipose tissue inflammation. Journal of Biological Chemistry 288:2882–2892.
- [6] Downer, E.J., Clifford, E., Gran, B., Nel, H.J., Fallon, P.G., and Moynagh, P.N., 2011. Identification of the synthetic cannabinoid R(+)WIN55,212-2 as a novel regulator of IFN regulatory factor 3 activation and IFN-beta expression: relevance to therapeutic effects in models of multiple sclerosis. Journal of Biological Chemistry 286:10316–10328.
- [7] Mansuy-Aubert, V., Zhou, Q.L., Xie, X., Gong, Z., Huang, J.Y., Khan, A.R., et al., 2013. Imbalance between neutrophil elastase and its inhibitor alpha1-antitrypsin in obesity alters insulin sensitivity, inflammation, and energy expenditure. Cell Metabolism 17:534–548.
- [8] Die, L., Yan, P., Jun Jiang, Z., Min Hua, T., Cai, W., and Xing, L., 2012. Glycogen synthase kinase-3 beta inhibitor suppresses Porphyromonas gingivalis lipopolysaccharide-induced CD40 expression by inhibiting nuclear factor-kappa B activation in mouse osteoblasts. Molecular Immunology 52:38–49.
- [9] Raes, G., De Baetselier, P., Noel, W., Beschin, A., Brombacher, F., and Hassanzadeh Gh, G., 2002. Differential expression of FIZZ1 and Ym1 in alternatively versus classically activated macrophages. Journal of Leukocyte Biology 71:597–602.