

Gaik et al., <http://www.jcb.org/content/full/jcb.2014.11003/DC1>

Nup159-CTD

Multiple sequence alignment of Nup159-CTD from various species, including H2A, H2B, H3, and H4. The alignment shows conserved residues across different species and domains.

Linker

H1

Multiple sequence alignment of Linker and H1 domains. The alignment shows conserved residues across different species and domains.

H2

Multiple sequence alignment of H2 domain. The alignment shows conserved residues across different species and domains.

H3

H4

Multiple sequence alignment of H3 and H4 domains. The alignment shows conserved residues across different species and domains.

H4

Multiple sequence alignment of H4 domain. The alignment shows conserved residues across different species and domains.

tail

Multiple sequence alignment of tail domain. The alignment shows conserved residues across different species and domains.

Figure S1. α -Helically predicted domain of Nup159 are conserved among fungi. Multi-sequence alignment, using T-Coffee (<http://www.ebi.ac.uk/Tools/msa/tcoffee/>), and Jalview, of the α -helically predicted C domain of Nup159. The default color scheme of ClustalX/Jalview was used with e.g., hydrophobic residues in blue, acidic residues in violet, and basic residues in red.

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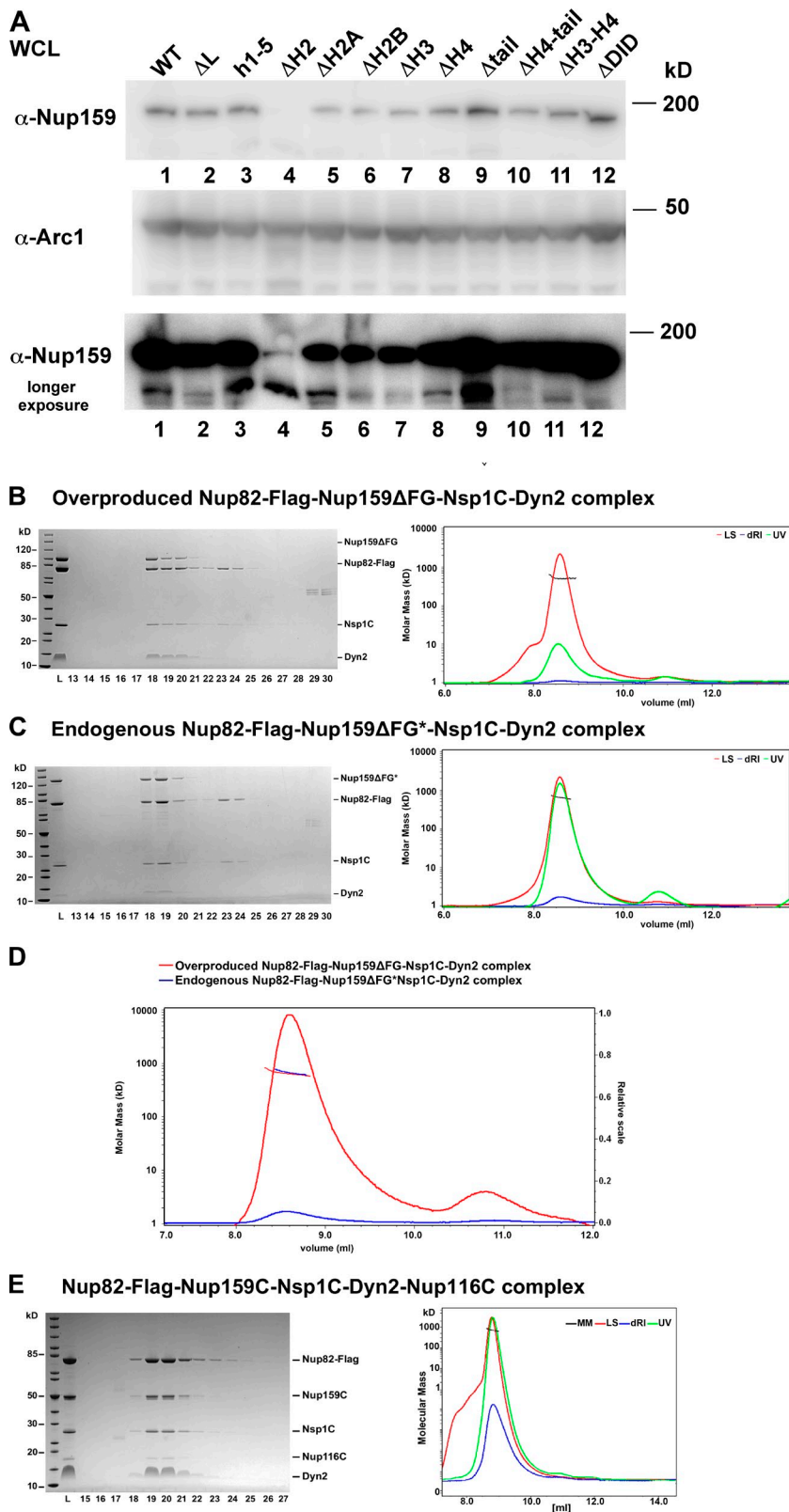


Figure S2. **Overproduced and endogenous Nup82-Nup159 Δ FG-Nsp1C-Dyn2 complexes have a similar gel filtration behavior and a related MALS value.** (A) Expression levels of the indicated Nup159 wild-type (WT) and mutant proteins used in this study, which were analyzed by SDS-PAGE of whole-cell lysates (WCL; cells were grown at 30°C) followed by Western blotting using a monoclonal anti-Nup159 antibody (mAB165C10), which allows detection of the Nup159 CTD constructs (Kraemer et al., 1995), and as loading control, we used anti-Arc1 antibodies. (B) SEC-MALS analysis of the overproduced Nup82-Flag-Nup159 Δ FG-Nsp1C-Dyn2 complex with a deduced molecular mass of 632 kD. (C) SEC-MALS analysis of the endogenous Nup82-Flag-Nup159 Δ FG*-Nsp1C-Dyn2 complex (indicated a molecular mass of 670 kD). The asterisk in Nup159 Δ FG* indicates that Nup159 carries additional amino acids between the β -propeller domain and the DID (see Tables S1 and S2). (D) Comparison of yield of overproduced versus endogenous Nup82-Flag-Nup159 Δ FG-Nsp1C-Dyn2 complex, as indicated by the differential refractive index (dRI). LS, light scattering; MM, molecular mass. (E) SEC-MALS analysis of the Nup82-Flag-Nup159C-Nsp1C-Dyn2-Nup116C complex (deduced molecular mass 693 kD). In A–D, data shown are from single representative experiments out of two repeats. L, load.

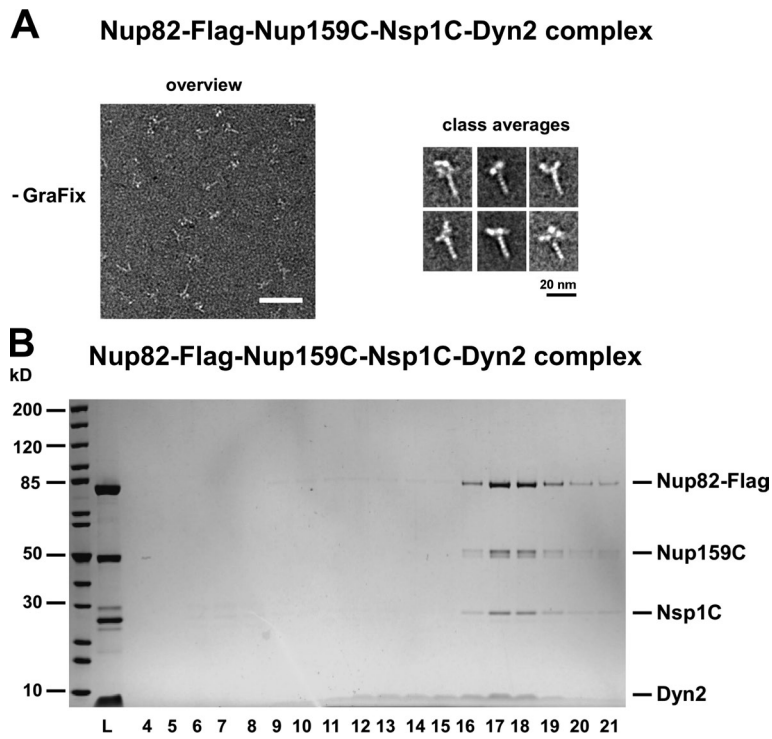


Figure S3. **Negative-staining EM analysis of the unfixed and GraFix-treated Nup82-Nup159C-Nsp1C-Dyn2 complex.** (A) Negative-staining EM of the unfixed Nup82-Nup159C-Nsp1C-Dyn2 complex showing an overview picture (left) and a gallery of class averages (right). Bar, 50 nm. (B) Glycerol gradient centrifugation of affinity-purified mNup82-Nup159C-Nsp1C-Dyn2 complex, analyzed by SDS-PAGE and Coomassie staining, to determine the relevant fractions (17-18) of glycerol-glutaraldehyde (GraFix) gradient used in the EM analysis (Fig. 4 A). L, load.

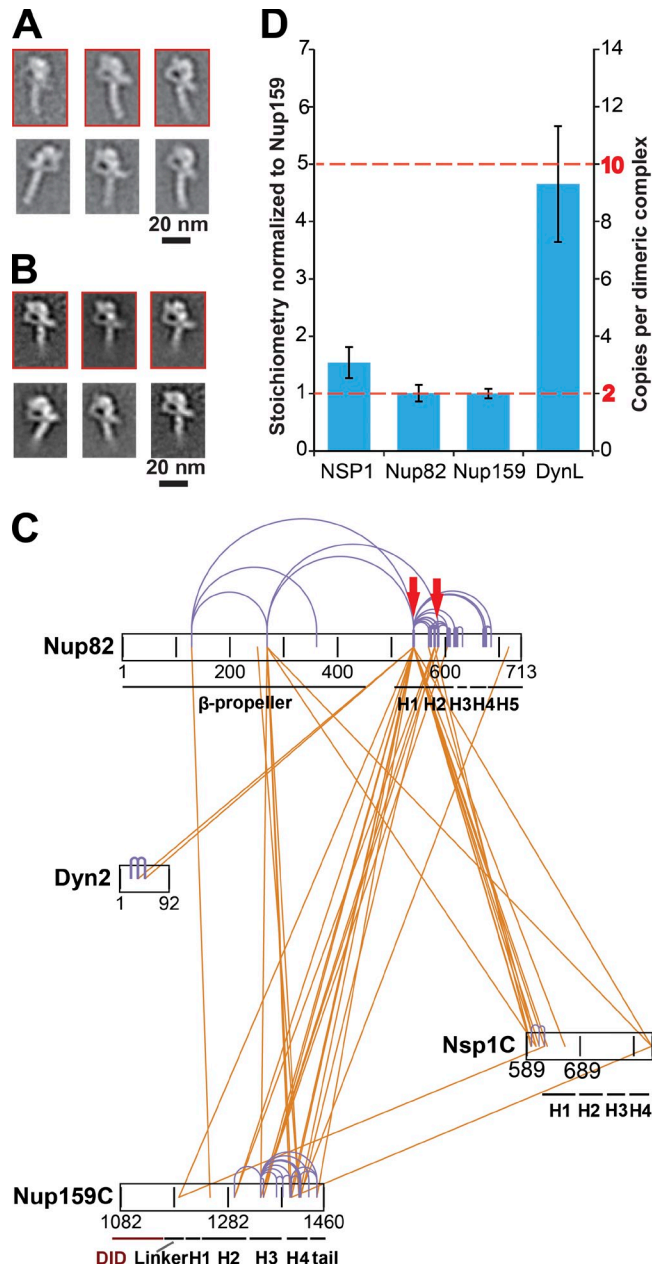


Figure S4. **EM structure, cross-linking, and stoichiometry of the purified Nup82 complex.** (A) Representative 2D class averages of affinity-purified Nup82–Nup159C–Nsp1C–Dyn2 complex based on subtomogram averaging. Parts of the DID_{Nup159}–Dyn2 stalk are averaged out in the first and the third class. Highlighted classes (red) were used for 3D reconstruction (B) Representative class averages of the affinity-purified Nup82–Nup159C–Nsp1C–Dyn2 complex based on subtomogram averaging focused by local masking to the head region such that it is resolved with higher detail. Highlighted classes (red) were used for 3D reconstruction. (C) XL-MS of the affinity-purified Nup82–Nup159C–Nsp1C–Dyn2 complex using DSG. The primary structure of the protein is shown, and specific regions are indicated. Interprotein cross-links are shown in orange; intraprotein cross-links are in purple. For the visualization of cross-links, the xiNET tool from the Rappsilber laboratory was used (<http://crosslinkviewer.org/index.php>). Because multiple copies of each protein are present, the latter might also occur across multiple instances of the same protein. Homodimeric cross-links that connect the two instances of the same lysine residue are indicated with red arrows. (D) Stoichiometric measurements of purified Nup82–Nup159C–Nsp1C–Dyn2 complex by quantitative, targeted proteomics within an early fraction of the gel filtration peak. Two heavy-labeled reference peptides per protein were used as intrinsic standards. The apparent values were normalized to the abundance of Nup159; error bars correspond to one standard deviation (Fig. 7).

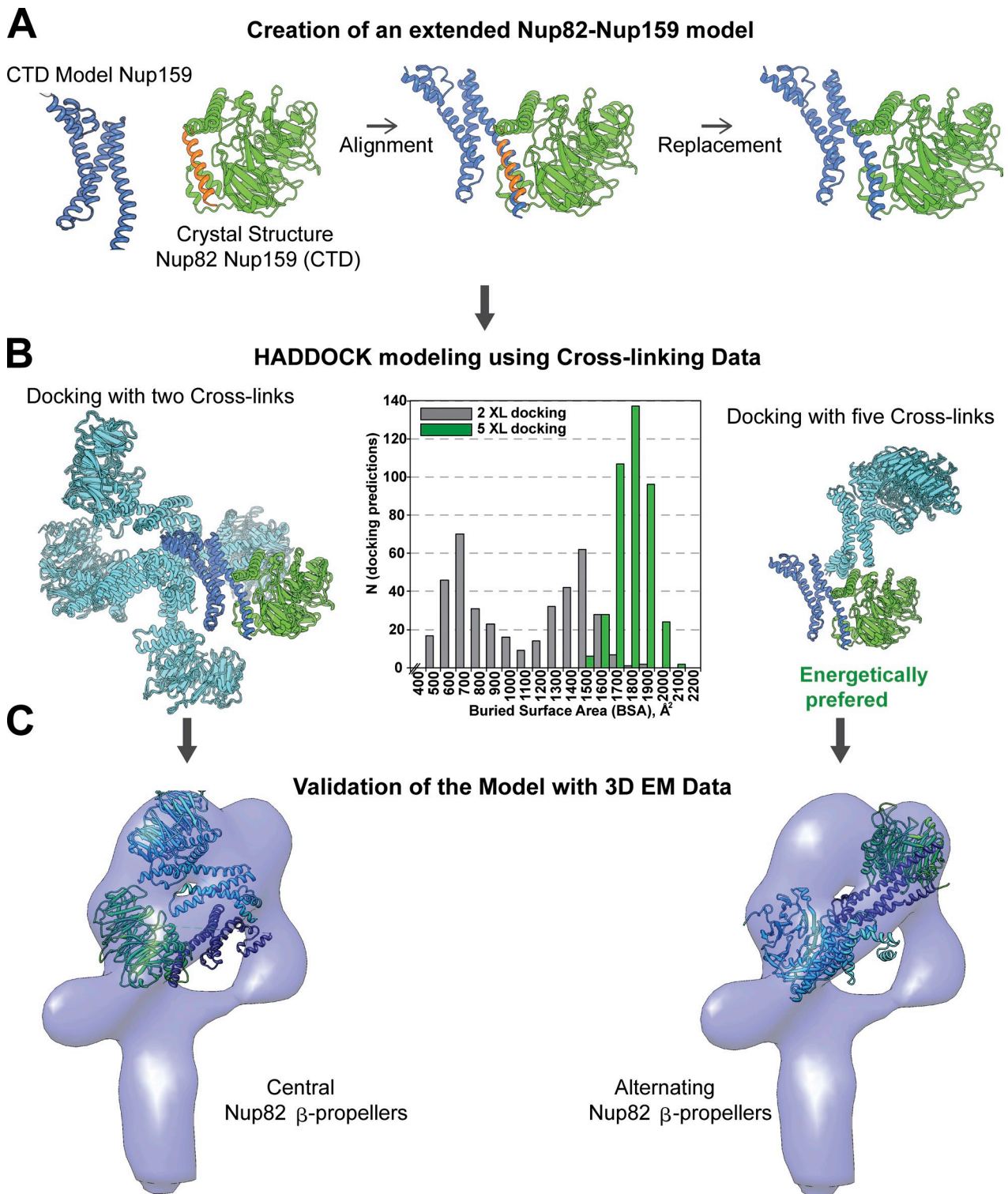


Figure S5. **Structural modeling of the interaction between Nup82 and Nup159.** (A) A structural model of Nup159C (amino acids 1,313–1,434) was generated using I-TASSER. An extended Nup82–Nup159 structural model was built after superposition on top of Nup82, present in the Nup159-Nup116 x-ray structure. The model was further refined using HADDOCK. (B) Interaction of four Nup82 propellers and two Nup159C domains (from amino acids 1,313 to 1,434) were modeled using docking by translating XL-MS information into spatial restraints as implemented in HADDOCK. When only homodimeric spatial restraints (Nup159 K1343 and Nup159 K1414) were considered (left), the approach did not converge into a single solution. When three additional distance restraints were considered (Nup82 K269 with Nup159 K1343, Nup82 K269 with Nup159 K1414, and Nup82 K274 with Nup159 K1343) under the assumption that they occur across two relevant instances of Nup82 and Nup159, an energetically favorable model was generated. (C) Structural models that satisfy the EM volume are shown. The right model is energetically preferred as highlighted in plot in B, showing that calculated buried surface area is much larger compared with the model on the left.

Table S1. **Plasmids used in this study**

Plasmid	Markers and construction details	Reference
pRS414-NUP159	CEN, TRP1, AmpR, P _{NUP159} NUP159, T _{ADH1}	Stelter et al., 2007
pRS414-nup159ΔDID	See above, P _{NUP159} Nup159 ΔΔ1,086–1,185 aa	Stelter et al., 2007
pRS414-nup159ΔLinker	See above, P _{NUP159} Nup159 ΔΔ1,179–1,210 aa	This study
pRS414-nup159ΔH1	See above, P _{NUP159} Nup159 ΔΔ1,210–1,244 aa	This study
pRS414-nup159ΔH2	See above, P _{NUP159} Nup159 ΔΔ1,245–1,330 aa	This study
pRS414-nup159ΔH3	See above, P _{NUP159} Nup159 ΔΔ1,330–1,381 aa	This study
pRS414-nup159ΔH4	See above, P _{NUP159} Nup159 ΔΔ1,381–1,425 aa	This study
pRS414-nup159Δtail	See above, P _{NUP159} Nup159 ΔΔ1,425–1,460 aa	This study
pRS414-nup159ΔH3-H4	See above, P _{NUP159} Nup159 ΔΔ1,330–1,425 aa	This study
pRS414-nup159ΔH4-tail	See above, P _{NUP159} Nup159 ΔΔ1,381–1,460 aa	This study
pRS414-nup159 h1-5	See above, P _{NUP159} Nup159 Δ I1232>D,M1235>E	This study
pRS414-GFP-NUP159	CEN, TRP1, AmpR, P _{NOP1} NUP159, T _{ADH1}	This study
pRS414-GFP-nup159 h1-5	CEN, TRP1, AmpR, P _{NOP1} nup159h1-5, T _{ADH1}	This study
pRS414-NUP159-Flag-TEV-ProtA	CEN, TRP1, AmpR, P _{NOP1} NUP159-FLAG-TEV-pA, T _{ADH1}	This study
pRS414-Nup159-h1-5-Flag-TEV-ProtA	CEN, TRP1, AmpR, P _{NOP1} nup159h1-5-FLAG-TEV-pA, T _{ADH1}	This study
YEplac181-NUP159	2μ, LEU2, AmpR, P _{GAL1-10} NUP159 T _{ADH1}	This study
YEplac181-Nup159ΔFG	2μ, LEU2, AmpR, P _{GAL1-10} NUP159 ΔΔ459–1,083 aa T _{ADH1}	This study
YEplac181-Nup159C	2μ, LEU2, AmpR, P _{GAL1-10} NUP159 ΔΔ1–1,082 aa, T _{ADH1}	This study
Yep351-GAL1-Dyn2	2μ, LEU2, AmpR, P _{GAL1-10} DYN2, T _{ADH1}	This study
YEplac195-P1GAL-Dyn2-P2-Nup159C	YEplac195-P1 _{GAL1-10} -DYN2-P2-NUP159 ΔΔ1–1,082 aa, URA3	This study
YEplac181-P1GAL-Nsp1C-P2-Nup82-Flag-TEV-ProtA	YEplac181-P1 _{GAL1-10} -NSP1 ΔΔ1–572 aa-P2-NUP82-FLAG-TEV-pA, LEU2	This study
YEplac195-P1GAL-Dyn2-P2-Nup159Δtail	YEplac195-P1 _{GAL1-10} -DYN2-P2-NUP159 ΔΔ1–1,082 aa and 1,425–1,460 aa, URA3	This study
YEplac195-P1GAL-Dyn2-P2-Nup159ΔFG	YEplac195-P1 _{GAL1-10} -DYN2-P2-NUP159 ΔΔ459–1,083 aa, URA3	This study
YEplac181-P1GAL-HIS-Nup116C	YEplac181-P1 _{GAL1-10} -NUP116 ΔΔ1–967 aa, LEU2	This study
YEplac181-P1GAL-Nsp1C-Flag-TEV-ProtA-P2-Nup82	YEplac181-P1 _{GAL1-10} -NSP1 ΔΔ1–572 aa-FLAG-TEV-pA-P2-NUP82, LEU2	This study
YEplac195-P1GAL-Dyn2-P2-Nup159tail-Flag	YEplac195-P1 _{GAL1-10} -DYN2-P2-NUP159 ΔΔ1–1,425 aa-FLAG, URA3	This study
YEplac112-P1GAL-Nsp1C-P2-Nup82-Flag-TEV-ProtA	YEplac112-P1 _{GAL1-10} -NSP1 ΔΔ1–572 aa-P2-NUP82-FLAG-TEV-pA, TRP1	This study
YEplac112-P1GAL-Nsp1C-P2-Nup82-TEV-ProtA	YEplac112-P1 _{GAL1-10} -NSP1 ΔΔ1–572 aa-P2-NUP82-TEV-pA, TRP1	This study
pET-15b-HIS-TEV-Nup159-QT ₄₋₅ -Linker-H1	KanR, P _{T7} HIS ₆ -TEV-nup159-1,153–1,241 aa	This study
pET-15b-HIS-TEV-Nup159-QT ₄₋₅ -Linker-h1-5	KanR, P _{T7} HIS ₆ -TEV-nup159-1,153–1,241 aa I1232>D,M1235>E	This study
pRS414-ADHI-NSP1C	CEN, TRP1, AmpR, P _{ADH1} NSP1-ΔΔ 1-605 aa, T _{ADH1}	Nehrbass et al., 1990
pAM1 (Nup159ΔFG*)	CEN4, LEU2, AmpR, YCplac111 (Δ[HindIII-SmaI]) containing the RAT7-ΔRp/NUP159 gene [pLG7]	Del Priore et al., 1997

Table S2. **Yeast strains used in this study**

Strain	Genotype	Reference
Ds1-2b	his3-Δ200, leu2-Δ1, trp1-Δ63, ura3-52, MAT α	Grandi et al., 1995
Nup159Δ shuffle	nup159::natNT2, his3-Δ200, ura3-52, leu2-Δ1, trp1-Δ63, MAT α, pLG-URA3-NUP159	Gorsch et al., 1995
Nup159Δ shuffle, Nup82-Flag-TEV-ProtA	nup159::natNT2, his3-Δ200, ura3-52, leu2-Δ1, trp1-Δ63, MAT α, Nup82-Flag-TEV-ProtA::HIS3, pLG-URA3-NUP159	This study
Nup82-Flag-TEV-ProtA, Nup159Δ, Nsp1Δ shuffle	nup159::HIS3, nsp1::HIS3, ura3-52, leu2-Δ1, trp1-Δ63, MAT α, Nup82-Flag-TEV-ProtA::natNT2, pLG-URA3-NUP159, pRS316-URA3-NSP1	This study

Table S3. **Cross-links identified within the Nup82 complex using DSS**

Peptide	Protein 1	Protein 2	AA 1	AA 2	Id score	Comment
KINSWDQVLV-KMLEIDSK	Nsp1	Nup82	661	685	37.2	
IVKAQTLGVSIHNR-TEESSTGKSTADV	Nup82	Nsp1	541	607	35.5	
IVKAQTLGVSIHNR-K ⁺ TEESSTGKSTADV	Nup82	Nsp1	541	607	36.0	Missed-cleavage derivative to previous peptide
QINSIKK-MQKTL	Nsp1	Nup159	822	1,343	31.1	
SLDDNSTSLEKQINSIK-MQKTLR	Nsp1	Nup159	816	1,343	31.5	
SLDDNSTSLEKQINSIK ⁺ -K-MQKTLR	Nsp1	Nup159	816	1,343	32.3	Missed-cleavage derivative to previous peptide
FGKVDIQK-MQKTLR	Nup82	Nup159	269	1,343	45.3	Restraint used for modeling
FGKVDIQK ⁺ EYR-MQKTLR	Nup82	Nup159	269	1,343	39.3	Missed-cleavage derivative to previous peptide
FGKVDIQKEYR-MQKTLR	Nup82	Nup159	274	1,343	32.8	Restraint used for modeling
FGKVDIQK-LQLEEKGGK	Nup82	Nup159	269	1,414	37.2	Restraint used for modeling
FGKVDIQK-LQLEEKGGK ⁺	Nup82	Nup159	269	1,414	37.5	Missed-cleavage derivative to previous peptide
IEAETIKVDKK-LFTVKNK	Nup82	Nup159	674	1,372	24.7	
IEAETIKVDKK-MQKTLR	Nup82	Nup159	671	1,343	36.7	
IISKDDDLRR-MQKTLR	Nup82	Nup159	573	1,343	29.7	
QLQSTCK ⁺ IISKDDDLR-MQKTLR	Nup82	Nup159	573	1,343	25.3	Missed-cleavage derivative to previous peptide
IVKAQTLGVSIHNR-DALDKYQLER	Nup82	Dyn2	541	34	40.4	
IVKAQTLGVSIHNR-DGLLKEIK	Nup82	Nup159	541	1,397	34.8	
IVKAQTLGVSIHNR-DIAGTVKK	Nup82	Dyn2	541	46	31.2	
IVKAQTLGVSIHNR-DNEKTEESSTGK	Nup82	Nsp1	541	598	36.3	
IVKAQTLGVSIHNR-EIKLLR	Nup82	Nup159	541	1,400	35.3	
IVKAQTLGVSIHNR-EKVTDYVR	Nup82	Nup159	541	1,294	32.9	
IVKAQTLGVSIHNR-KINSWDQVLV	Nup82	Nsp1	541	661	36.9	
IVKAQTLGVSIHNR-KTEESSTGK	Nup82	Nsp1	541	599	38.8	
IVKAQTLGVSIHNR-LQLEEKGGK	Nup82	Nup159	541	1,414	37.6	
IVKAQTLGVSIHNR-LQLEEKGGK ⁺	Nup82	Nup159	541	1,414	26.0	Missed-cleavage derivative to previous peptide
IVKAQTLGVSIHNR-MQKTLR	Nup82	Nup159	541	1,343	42.9	
IVKAQTLGVSIHNR-QINSIKK	Nup82	Nsp1	541	822	32.0	
IVKAQTLGVSIHNR-QLKEYYTSK	Nup82	Nup159	541	1,191	34.9	
IVKAQTLGVSIHNR-STADV ⁺ KSSDSLK	Nup82	Nsp1	541	613	39.3	
KFEAQNK-MQKTLR	Nup82	Nup159	580	1,343	36.1	
KFEAQNK ⁺ -K-MQKTLR	Nup82	Nup159	580	1,343	33.1	Missed-cleavage derivative to previous peptide
KFEAQNK-MQKTLR	Nup82	Nup159	586	1,343	34.3	
KMLEIDSK-MQKTLR	Nup82	Nup159	685	1,343	39.5	
KWDAQLSR-LQLEEKGGK	Nup82	Nup159	587	1,414	27.4	
KWDAQLSR-MQKTLR	Nup82	Nup159	587	1,343	42.5	
MLEIDSKIIK-EIKLLR	Nup82	Nup159	692	1,400	37.7	
QLQSTCKIISK-MQKTLR	Nup82	Nup159	569	1,343	31.4	
QLQSTCKIISK ⁺ DDDLR-MQKTLR	Nup82	Nup159	569	1,343	26.7	Missed-cleavage derivative to previous peptide
TQGSYDKDHGDYKDDDDK-MQKTLR	Nup82	Nup159	718	1,343	27.1	
EKVTDYVRK-KFEAQNK	Nup159	Nup82	1,294	586	29.4	
EYYTSK ⁺ SNIPFVSQNSTLR-STADV ⁺ KSSDSLK	Nup159	Nsp1	1,198	613	24.8	
NMDTFFTDQSSIPV ⁺ KR-IVKAQTLGVSIHNR	Nup159	Nup82	1,249	541	34.4	
NMDTFFTDQSSIPV ⁺ KR-KFEAQNK	Nup159	Nup82	1,249	580	25.3	
NMDTFFTDQSSIPV ⁺ KR-KFEAQNK ⁺	Nup159	Nup82	1,249	580	26.3	Missed-cleavage derivative to previous peptide
DALDKYQLER-DIAGTVKK	Dyn2		34	46	29.7	
DALDKYQLER-DIAGTVKK ⁺ QLDVK	Dyn2		34	46	33.6	Missed-cleavage derivative to previous peptide
DALDKYQLER-KQLDVK	Dyn2		34	47	41.2	
DALDKYQLER ⁺ -DIAGTVK-KQLDVK	Dyn2		34	47	38.3	Missed-cleavage derivative to previous peptide
DALDKYQLER-LKEDILTISK	Dyn2		34	21	40.7	

Table S3. **Cross-links identified within the Nup82 complex using DSS** (Continued)

Peptide	Protein 1	Protein 2	AA 1	AA 2	ld score	Comment
DNEK K TEESSTGK-KTEESSTGK	Nsp1		598	599	25.2	
DNEK K TEESSTGK-STADV K SSDSLK	Nsp1		598	613	25.0	
STADV K SSDSLK-KTEESSTGK	Nsp1		613	599	38.0	
STADV K SSDSLK-DNEK K TEESSTGK	Nsp1		613	599	33.5	Missed-cleavage derivative to previous peptide
KTEESSTG K STADV K -KTEESSTGK	Nsp1		607	599	27.7	
KTEESSTG K STADV K -MGAQ K DNEK	Nsp1		607	594	28.5	
LNS K PVELKPVSLDNK-STADV K SSDSLK	Nsp1		623	613	33.1	
NMDTFFTDQSSIP V KR-MQ K TLR	Nup159		1,249	1,343	24.6	
E K VTDYVR-MQ K TLR	Nup159		1,294	1,343	33.7	
MQ K TLR-MQ K TLR	Nup159		1,343	1,343	37.5	Homodimeric peptide topology; restraint used for modeling
Q K LFDVSAK-MQ K TLR	Nup159		1,348	1,343	41.2	
LDDNPLV A KLAK-DGL L KEIK	Nup159		1,384	1,397	42.2	
LDDNPLV A KLAK-EI K LLR	Nup159		1,384	1,400	33.8	
LDDNPLV A KLAK-LQLEEK G K	Nup159		1,384	1,414	31.5	
LDDNPLV A KLAK-MQ K TLR	Nup159		1,384	1,343	32.7	
DGL L KEIK-EI K LLR	Nup159		1,397	1,400	35.0	
DGL L KEIK-LQLEEK G K	Nup159		1,397	1,414	38.8	
DGL L KEIK-MQ K TLR	Nup159		1,397	1,343	36.8	
EI K LLR-MQ K TLR	Nup159		1,400	1,343	29.6	
LQLEEK G K-DM K G F K	Nup159		1,414	1,432	38.5	
LQLEEK G K-EI K LLR	Nup159		1,414	1,400	36.0	
LQLEEK G K K -EI K LLR	Nup159		1,414	1,400	31.5	Missed-cleavage derivative to previous peptide
LQLEEK G K-LQLEEK G K	Nup159		1,414	1,414	35.8	Homodimeric peptide topology; restraint used for modeling
LQLEEK G K K -LQLEEK G K	Nup159		1,414	1,414	38.2	Missed-cleavage derivative to previous peptide
LQLEEK G K K -LQLEEK G K K	Nup159		1,414	1,414	35.7	Missed-cleavage derivative to previous peptide
LQLEEK G K-MQ K TLR	Nup159		1,414	1,343	44.4	
LQLEEK G K K -MQ K TLR	Nup159		1,414	1,343	36.5	Missed-cleavage derivative to previous peptide
LQLEEK G K K -Q K LFDVSAK	Nup159		1,414	1,348	24.8	
LQLEEK G K-LFT V KNK	Nup159		1,414	1,372	30.4	
LQLEEK G K K -MQ K TLR	Nup159		1,416	1,343	31.5	
K ASSFDASSSIT K -LQLEEK G K	Nup159		1,417	1,414	46.4	
K ASSFDASSSIT K -LQLEEK G K K	Nup159		1,417	1,416	29.3	
K ASSFDASSSIT K -MQ K TLR	Nup159		1,417	1,343	35.7	
ASSFDASSSIT K DM K -LQLEEK G K	Nup159		1,429	1,414	27.7	
DM K G F K-MQ K TLR	Nup159		1,432	1,343	28.0	
K QIGDFF K -LQLEEK G K	Nup159		1,447	1,414	32.9	
FG K V D I Q K- K F E A Q N K	Nup82		269	580	26.1	
FG K V D I Q K- K W D A Q L S R	Nup82		269	587	28.0	
IV K A Q T L G V SI H N R -FG K V D I Q K	Nup82		541	269	35.5	
IV K A Q T L G V SI H N R -FS K L S K	Nup82		541	604	39.7	
IV K A Q T L G V SI H N R -IE A ET I K V D K K	Nup82		541	674	35.1	
IV K A Q T L G V SI H N R -I S K D D D L R R	Nup82		541	573	35.9	
IV K A Q T L G V SI H N R -IV K A Q T L G V SI H N R	Nup82		541	541	39.7	Homodimeric peptide topology
IV K A Q T L G V SI H N R - K F E A Q N K	Nup82		541	580	43.1	
IV K A Q T L G V SI H N R - K F E A Q N K K	Nup82		541	586	35.5	
IV K A Q T L G V SI H N R - K M L E I D S K	Nup82		541	685	43.9	
IV K A Q T L G V SI H N R - K S Q N E W D E L R	Nup82		541	675	35.2	
IV K A Q T L G V SI H N R - K S Q N E W D E L R K	Nup82		541	675	27.4	Missed-cleavage derivative to previous peptide
IV K A Q T L G V SI H N R - K W D A Q L S R	Nup82		541	587	48.4	
IV K A Q T L G V SI H N R -QL Q ST C K I IS K	Nup82		541	569	36.7	
IV K A Q T L G V SI H N R -QL Q ST C K I IS K [^] DD D L R	Nup82		541	569	38.6	Missed-cleavage derivative to previous peptide
QL Q ST C K I IS K - K F E A Q N K	Nup82		569	580	38.7	

Table S3. **Cross-links identified within the Nup82 complex using DSS** (Continued)

Peptide	Protein 1	Protein 2	AA 1	AA 2	ld score	Comment
QLQSTCK IISK -KFEAQNKK	Nup82		569	586	26.2	
IISKDDDLR-R KFEAQN K	Nup82		573	580	25.2	
IISKDDDLR [^] R-KFEAQN K	Nup82		573	580	41.9	Missed-cleavage derivative to previous peptide
IISKDDDLR [^] R-KFEAQN K [^] K	Nup82		573	580	28.9	Missed-cleavage derivative to previous peptide
QLQSTCK [^] IISKDDDLR-R KFEAQN K	Nup82		573	580	31.6	Missed-cleavage derivative to previous peptide
IISKDDDLRR-KFEAQN KK	Nup82		573	586	29.4	
IISKDDDLRR-K WDAQLSR	Nup82		573	587	38.3	
KFEAQN KK - KMLEIDSK	Nup82		586	685	25.2	
KWDAQLSR -FEAQN KK	Nup82		587	586	36.6	
FEAQN K [^] KWDAQLSR -KFEAQN K [^] K	Nup82		587	586	26.5	Missed-cleavage derivative to previous peptide
KWDAQLSR -FS KLSK	Nup82		587	604	40.0	
KWDAQLSR - KFEAQN K	Nup82		587	580	42.4	
KWDAQLSR -R [^] KFEAQN K	Nup82		587	580	35.1	Missed-cleavage derivative to previous peptide
KLSQIAESNK -FS KLSK	Nup82		608	604	40.6	
KLSQIAESNK [^] FK-FS KLSK	Nup82		608	604	41.9	Missed-cleavage derivative to previous peptide
KISHGEMK -WF KEIR	Nup82		622	632	30.3	
IEAETIKVD KK -FG KVDIGK	Nup82		671	269	37.3	
IEAETIKVD KK -FS KLSK	Nup82		671	604	28.7	
IEAETIKVD KK - KFEAQN K	Nup82		671	580	28.5	
IEAETIKVD KK - KMLEIDSK	Nup82		671	685	35.4	
IEAETIKVD KK - KWDAQLSR	Nup82		671	587	37.2	
IEAETIKVD K - KSQNEWDEL R	Nup82		671	675	38.3	
SELTR [^] IEAETIKVD K - KSQNEWDEL R K	Nup82		671	675	35.0	Missed-cleavage derivative to previous peptide
IEAETIKVD K - KSQNEWDEL R [^] K	Nup82		671	675	42.0	Missed-cleavage derivative to previous peptide
IEAETIKVD KK - KFEAQN K	Nup82		674	580	34.4	
IEAETIKVD KK - KMLEIDSK	Nup82		674	685	31.7	
IEAETIKVD KK - KWDAQLSR	Nup82		674	587	28.3	
KSQNEWDEL R- KMLEIDSK	Nup82		675	685	38.2	
KSQNEWDEL R K - MLEIDSK I K	Nup82		675	692	29.5	
TQGS DYKD H DGDYK DDDD K - IV K A QTLG VS IHN R	Nup82		718	541	34.2	
TQGS DYKD H DGDYK DDDD K -FEAQN KK	Nup82		725	586	26.7	

Cross-linked peptides are separated by hyphens; cross-linked lysine residues of the peptides are in bold. [^] indicates a missed-cleavage derivative of previous peptide, which comprises an independent experimental evidence. ld, linear discriminant.

Table S4. **Cross-links identified within the Nup82 complex using DSG**

Peptide	Protein 1	Protein 2	AA 1	AA 2	ld score	Comment
KTEESSTGKSTADV K-KQLDVK	Nsp1	Dyn2	607	47	24.9	
KTEESSTGKSTADV K-FGKVDIQK	Nsp1	Nup82	607	269	27.6	
KTEESSTGK-IVKAQTLGVSIHNR	Nsp1	Nup82	541	599	36.6	
KTEESSTGK^STADV K-IVKAQTLGVSIHNR	Nsp1	Nup82	599	541	28.5	Missed-cleavage derivative to previous peptide
LNSKPVELKPVSLDNK-IVKAQTLGVSIHNR	Nsp1	Nup82	623	541	29.7	
LNSKPVELKPVSLDNK-IVKAQTLGVSIHNR	Nsp1	Nup82	628	541	28.2	
KTEESSTGKSTADV K-QLGSTCKIISK	Nsp1	Nup82	607	569	25.3	
DNEKKTEESSTGK-KFEAQNK	Nsp1	Nup82	598	580	28.2	
LNSKPVELKPVSLDNK-QLKEYYTSAK	Nsp1	Nup159	623	1,191	27.6	
QINSIKK-EIKLLR	Nsp1	Nup159	822	1,400	28.0	
NMDTFFTDQSSIVLVR-KR-KVLFHPK	Nup159	Nup82	1,249	129	25.3	
DGLLKEIK-FGKVDIQK	Nup159	Nup82	1,397	269	25.4	
LQLEEKGK-FGKVDIQK	Nup159	Nup82	1,414	269	25.8	
LQLEEKGK^K-FGKVDIQK	Nup159	Nup82	1,414	269	28.5	Missed-cleavage derivative to previous peptide
EKVTDYVRK-KFEAQNK	Nup159	Nup82	1,294	580	29.4	
KASSFDASSITK-KFEAQNK	Nup159	Nup82	1,417	580	28.5	
QLKEYYTSAK-KTEESSTGK	Nup159	Nsp1	1,191	599	24.5	
IVKAQTLGVSIHNR-DALDKYQLER	Nup82	Dyn2	541	34	26.5	
IVKAQTLGVSIHNR-KQLDVK	Nup82	Dyn2	541	47	29.8	
IVKAQTLGVSIHNR-DNEKKTEESSTGK	Nup82	Nsp1	541	598	31.0	
IVKAQTLGVSIHNR-DNEK^KTEESSTGK	Nup82	Nsp1	541	599	32.2	Missed-cleavage derivative to previous peptide
IVKAQTLGVSIHNR-TEESSTGKSTADV K	Nup82	Nsp1	541	607	25.6	
IVKAQTLGVSIHNR-K^TEESSTGKSTADV K	Nup82	Nsp1	541	607	37.5	Missed-cleavage derivative to previous peptide
IVKAQTLGVSIHNR-STADV KSSDSLK	Nup82	Nsp1	541	613	33.9	
IVKAQTLGVSIHNR-KINSWDQVVLK	Nup82	Nsp1	541	661	27.9	
FGKVDIQK-QINSIKK	Nup82	Nsp1	269	822	26.3	
IVKAQTLGVSIHNR-QINSIKK	Nup82	Nsp1	541	822	28.3	
IVKAQTLGVSIHNR-QLKEYYTSAK	Nup82	Nup159	541	1,191	31.1	
IVKAQTLGVSIHNR-EKVTDYVR	Nup82	Nup159	541	1,294	32.1	
IVKAQTLGVSIHNR-EKVTDYVR^K	Nup82	Nup159	541	1,294	36.9	Missed-cleavage derivative to previous peptide
FGKVDIQK-MQKTLR	Nup82	Nup159	269	1,343	37.1	
FGKVDIQK^EYR-MQKTLR	Nup82	Nup159	269	1,343	31.5	Missed-cleavage derivative to previous peptide
IVKAQTLGVSIHNR-MQKTLR	Nup82	Nup159	541	1,343	37.2	
IISKDDLR-MQKTLR	Nup82	Nup159	573	1,343	24.8	
KFEAQNK-MQKTLR	Nup82	Nup159	580	1,343	27.2	
KFEAQNK^K-MQKTLR	Nup82	Nup159	580	1,343	26.3	Missed-cleavage derivative to previous peptide
KWDAQLSR-MQKTLR	Nup82	Nup159	587	1,343	26.7	
FSKLSK-MQKTLR	Nup82	Nup159	604	1,343	25.9	
IVKAQTLGVSIHNR-QKLFVSAK	Nup82	Nup159	541	1,348	32.7	
IVKAQTLGVSIHNR-DGLLKEIK	Nup82	Nup159	541	1,397	30.2	
FGKVDIQK-EIKLLR	Nup82	Nup159	269	1,400	29.2	
IVKAQTLGVSIHNR-EIKLLR	Nup82	Nup159	541	1,400	31.	
NVIKQLQFVSK-LQLEEKGK	Nup82	Nup159	251	1,414	26.4	
IVKAQTLGVSIHNR-LQLEEKGK	Nup82	Nup159	541	1,414	27.8	
TQGS DYKDHDGDYKDDDDK-LQLEEKGKK	Nup82	Nup159	718	1,414	27.3	
IVKAQTLGVSIHNR-KQIGDFFK	Nup82	Nup159	541	1,447	32.5	
DALDKYQLER-KQLDVK	Dyn2		34	47	31.9	
DALDKYQLER-LKEDILTISK	Dyn2		34	21	27.5	
STADV KSSDSLK-KTEESSTGK	Nsp1		613	599	26.5	
LNSKPVELKPVSLDNK-STADV KSSDSLK	Nsp1		623	613	26.4	
EKVTDYVR-MQKTLR	Nup159		1,294	1,343	26.7	

Table S4. **Cross-links identified within the Nup82 complex using DSG** (Continued)

Peptide	Protein 1	Protein 2	AA 1	AA 2	ld score	Comment
EKVTDYVR [^] K-MQKTLR	Nup159		1,294	1,343	28.4	Missed-cleavage derivative to previous peptide
QKLFVDSAK-MQKTLR	Nup159		1,348	1,343	31.7	
LFTVKNK-MQKTLR	Nup159		1,372	1,343	25.5	
RLDDNPLVAKLAK-LFTVKNK	Nup159		1,384	1,372	29.2	
LDDNPLVAKLAK-EIKLLR	Nup159		1,384	1,400	27.7	
LDDNPLVAKLAK-MQKTLR	Nup159		1,384	1,343	25.8	
LDDNPLVAKLAK-DGLLKEIK	Nup159		1,384	1,397	24.8	
DGLLKEIK-MQKTLR	Nup159		1,397	1,343	25.5	
DGLLKEIK-EIKLLR	Nup159		1,397	1,400	25.1	
LQLEEKGK-EIKLLR	Nup159		1,414	1,400	32.0	
EIKLLR [^] EQVSR-LQLEEKGK	Nup159		1,400	1,414	26.9	Missed-cleavage derivative to previous peptide
EIKLLREQVSR-MQKTLR	Nup159		1,400	1,343	24.6	
LQLEEKGK-MQKTLR	Nup159		1,414	1,343	28.5	
LQLEEKGKK-MQKTLR	Nup159		1,416	1,343	25.9	
KASSFDASSSITK-LQLEEKGK	Nup159		1,417	1,414	32.9	
KASSFDASSSITK-MQKTLR	Nup159		1,417	1,343	26.1	
DMKGFK-MQKTLR	Nup159		1,432	1,343	27.0	
KQIGDFFK-LQLEEKGK	Nup159		1,447	1,414	28.6	
KQIGDFFK-EIKLLR	Nup159		1,447	1,400	27.3	
KQIGDFFK-MQKTLR	Nup159		1,447	1,343	25.5	
FGKVDIQK-KVLFHPK	Nup82		269	129	34.4	
FGKVDIQK-KWDAQLSR	Nup82		269	587	25.9	
EIKSLITLPEQLGK-KVLFHPK	Nup82		361	129	26.1	
IVKAQTLGVSIHNR-KWDAQLSR	Nup82		541	587	39.5	
IVKAQTLGVSIHNR-FSKLSK	Nup82		541	604	38.5	
IVKAQTLGVSIHNR-FGKVDIQK	Nup82		541	269	37.8	
IVKAQTLGVSIHNR-IEAETIKVDKK	Nup82		541	674	33.1	
IVKAQTLGVSIHNR-KFEAQNK	Nup82		541	580	32.2	
IVKAQTLGVSIHNR-R [^] KFEAQNK	Nup82		541	580	29.2	Missed-cleavage derivative to previous peptide
IVKAQTLGVSIHNR-KVLFHPK	Nup82		541	129	31.8	
IVKAQTLGVSIHNR-KMLEIDSK	Nup82		541	685	31.3	
IVKAQTLGVSIHNR-IISKDDDLR	Nup82		541	573	28.0	
IVKAQTLGVSIHNR-IISKDDDLR [^] R-	Nup82		541	573	30.0	Missed-cleavage derivative to previous peptide
IVKAQTLGVSIHNR-IVKAQTLGVSIHNR	Nup82		541	541	28.0	Homodimeric peptide topology
IVKAQTLGVSIHNR-QLQSTCKIISK	Nup82		541	569	28.0	
IVKAQTLGVSIHNR-LSQIAESNKFK	Nup82		541	617	27.7	
IVKAQTLGVSIHNR-IEAETIKVDKK	Nup82		541	671	26.6	
IVKAQTLGVSIHNR-KFEAQNKK	Nup82		541	586	26.1	
QLQSTCKIISK-KWDAQLSR	Nup82		569	587	29.1	
QLQSTCKIISKDDDLR-IVKAQTLGVSIHNR	Nup82		569	541	26.3	
QLQSTCKIISK-KFEAQNK	Nup82		569	580	25.8	
IISKDDDLR-KFEAQNK	Nup82		573	580	29.6	
IISKDDDLR-FSKLSK	Nup82		573	604	27.0	
IISKDDDLR-KWDAQLSR	Nup82		573	587	25.7	
IISKDDDLR [^] R-KWDAQLSR	Nup82		573	587	25.5	Missed-cleavage derivative to previous peptide
KFEAQNK-FSKLSK	Nup82		580	604	32.4	
KFEAQNK-KFEAQNK	Nup82		580	580	24.9	Homodimeric peptide topology
KFEAQNK-KFSKLSK	Nup82		586	604	25.9	
KWDAQLSR-R [^] KFEAQNK	Nup82		587	580	26.3	Missed-cleavage derivative to previous peptide
KWDAQLSR-KFEAQNK	Nup82		587	580	33.0	
KWDAQLSR-FSKLSK	Nup82		587	604	29.4	
FSKLSK-KFEAQNK	Nup82		604	580	28.8	
KLQIAESNK-FSKLSK	Nup82		608	604	35.2	

Table S4. **Cross-links identified within the Nup82 complex using DSG** (Continued)

Peptide	Protein 1	Protein 2	AA 1	AA 2	ld score	Comment
KLSQIAESNK [^] FK-FSKLSK	Nup82		608	604	31.3	Missed-cleavage derivative to previous peptide
LSQIAESNKFK-EKKISHGEMK	Nup82		617	621	25.7	
LSQIAESNKFK-KISHGEMK	Nup82		617	622	25.4	
KISHGEMK-WFKEIR	Nup82		622	632	28.2	
IEAETIKVDK-KSQNEWDELK	Nup82		671	675	27.5	
SELTRIEAETIKVDK-KSQNEWDELK [^]	Nup82		671	675	29.5	Missed-cleavage derivative to previous peptide
IEAETIKVDK-KMLEIDSK	Nup82		671	685	26.5	
KSQNEWDELK-KMLEIDSK	Nup82		675	685	35.8	

Cross-linked peptides are separated by hyphens; cross-linked lysine residues of the peptides are in bold. [^] indicates missed-cleavage derivative of previous peptide, which comprises an independent experimental evidence. ld, linear discriminant.

Table S5. **Protein stoichiometry of the Nup82 complex using SRM-MS**

Protein	Peptide	Early gel filtration fraction		Apex gel filtration fraction	
		Intensity per peptide ^a	Abundance per protein ^b	Intensity per peptide ^a	Abundance per protein ^b
Nsp1	IDQSLQYIER	0.24	3.08	7.66	1.78
Nsp1	ILNSHFDALR	0.35	–	8.51	–
Nup82	AQTLGVSIIHNR	0.22	2.02	13.19	2.39
Nup82	SLQQDLSYK	0.17	–	8.54	–
Nup159	DLSTHQFR	0.21	2.00	9.16	2.00
Nup159	SINNLYTWR	0.18	–	8.98	–
Dyn2	NFGSYVTHEK	0.70	9.31	35.27	10.56
Dyn2	YGNTWHVIVGK	1.09	–	60.49	–

^aSummed intensity of the five most intense transitions per peptide.

^bPeptide intensities corresponding to the same protein were averaged and normalized to the Nup159 signal, resulting in the final stoichiometric read out per protein.

References

- Del Priore, V., C. Heath, C. Snay, A. MacMillan, L. Gorsch, S. Dagher, and C. Cole. 1997. A structure/function analysis of Rat7p/Nup159p, an essential nucleoporin of *Saccharomyces cerevisiae*. *J. Cell Sci.* 110:2987–2999.
- Gorsch, L.C., T.C. Dockendorff, and C.N. Cole. 1995. A conditional allele of the novel repeat-containing yeast nucleoporin RAT7/NUP159 causes both rapid cessation of mRNA export and reversible clustering of nuclear pore complexes. *J. Cell Biol.* 129:939–955. <http://dx.doi.org/10.1083/jcb.129.4.939>
- Grandi, P., S. Emig, C. Weise, F. Hucho, T. Pohl, and E.C. Hurt. 1995. A novel nuclear pore protein Nup82p which specifically binds to a fraction of Nsp1p. *J. Cell Biol.* 130:1263–1273. <http://dx.doi.org/10.1083/jcb.130.6.1263>
- Kraemer, D.M., C. Strambio-de-Castillia, G. Blobel, and M.P. Rout. 1995. The essential yeast nucleoporin NUP159 is located on the cytoplasmic side of the nuclear pore complex and serves in karyopherin-mediated binding of transport substrate. *J. Biol. Chem.* 270:19017–19021. <http://dx.doi.org/10.1074/jbc.270.32.19017>
- Nehrbass, U., H. Kern, A. Mutvei, H. Horstmann, B. Marshallsay, and E.C. Hurt. 1990. NSP1: a yeast nuclear envelope protein localized at the nuclear pores exerts its essential function by its carboxy-terminal domain. *Cell.* 61:979–989. [http://dx.doi.org/10.1016/0092-8674\(90\)90063-K](http://dx.doi.org/10.1016/0092-8674(90)90063-K)
- Stelter, P., R. Kunze, D. Flemming, D. Höpfner, M. Diepholz, P. Philippson, B. Böttcher, and E. Hurt. 2007. Molecular basis for the functional interaction of dynein light chain with the nuclear-pore complex. *Nat. Cell Biol.* 9:788–796. <http://dx.doi.org/10.1038/ncb1604>