Table S1: Primers used for the construction of plasmids.

Plasmid	Primers for PCR1	Primers for PCR2
	Universal primer F1: 5'GATATACCATGGGCACCCTCAAAGAAGTGC	Universal primer R2: 5'GATATACCATGGGCACCCTCAAAGAAGTGC
pSNVN∆151-175	R1=5'AACATATAGATGTCTTTTGATTGTTTGACGGCCCCTAGTG	F2=5'CGTCAAACAATCAAAAGACATCTATATGTTTCTATGCCAACTGCT
pSNVN∆175-364	R1=5'CTGAGTACGCCTTAATGGCTTACGTATTCCATTAACTTCTTC	F2=5'GGAATACGTAAGCCATTAAGGCGTACTCAGTCAATGGGGATTC
pSNVN∆175-313	R1=5'TGCTGTAGGTGGACATGGCTTACGTATTCCATTAACTTCTTC	F2=5'GGAATACGTAAGCCATGTCCACCTACAGCATTATATGTGGCC
pSNVN∆175-262	R1=5'TGTTATAAAATAGGCTGGCTTACGTATTCCATTAACTTCTTC	F2=5'GGAATACGTAAGCCAGCCTATTTTATAACACGTCAATTACAG

Constructs: The deletion mutant pSNVN∆1-50, lacking the N-terminal fifty amino acids was constructed by generating a PCR product from pSNVN plasmid using a forward primer, 5'CATGCCATGGTGTCTGCATTGGAGACCAAACTCG3', and primer, **PCR** TGGTGGTG**CTCGAG**TTTAAGTGGTTCTTGGTTAGAGATTTCC3'. The product was gel purified and digested with Nco I and Xho I restriction enzymes, cloned between the same restriction sites in pTriEx1.1 backbone. The same strategy was used for the construction of all other N terminal deletion mutants. using the same reverse primer and a different forward primer depending upon the type of mutation. The forward primers used for the construction of pSNVN Δ 1-100. pSNVN∆1-150 and pSNVN∆1-175 were 5 CATGCCATGGTCCTTGATGTAAATTCCATTGACT3', 5' CATGCCATGGAAAATAAGGGAACAAGAATCCGATT3' CATGCCATGGGACATCTATATGTTTCTATGCCAAC3', respectively. The terminal deletion mutant pSNVN∆403-428, expressing N protein lacking twenty six amino acids at the C-terminus was constructed by generating a PCR product from NVN2q plasmid forward primer, using CATGCCATGGGCACCCTCAAAGAAGTGCAAG3' and reverse primer TATAATCTCGAGATCCATATCATCTCCAAGATGG 3'. The PCR product was gel purified and digested with Ncol and Xhol and cloned between the same restriction sites in pTriEx1.1 backbone. The same strategy was used for the construction of all the C-terminal deletion mutants, using the same forward primer and different reverse primers, depending upon the type of mutation. The reverse primers used for the construction of pSNVNΔ347-428, pSNVNΔ238-428, 5' pSNVN∆175-428 and pSNVN∆163-428 plasmids were TATAATCTCGAGAGATTTTGATGCCATTATGGTG3'. 5' 5' TATAAT**CTCGAG**ATCAATCCTTTCCATCCAATCT3'. 5 TATAAT**CTCGAG**TGGCTTACGTATTCCATTAACT3', TATAATCTCGAGATCATCCTTGAATCGGATTCTT3', respectively. A two-step PCR strategy was used for the construction of plasmids expressing N mutants containing internal deletion. For example, for the construction pSNVN∆151-175, a PCR product was generated from pSNVN plasmid using a forward universal primer F1: 5' GATATACCATGGGCACCCTCAAAGAAGTGC3' and reverse primer 5'AACATATAGATGTCTTTTGATTGTTTGACGGCCCCTAGTG3'. Similarly, second PCR product was generated from pSNVN plasmid using a forward primer F2: 5'CGTCAAACAATCAAAAGACATCTATATGTTTCTATGCCAACTGCT3', and primer reverse universal R2:

5'TGGTGGTGCTCGAGTTTAAGTGGTTCTTGGTTAGAGATTTCCCTGAC3'.

The two PCR products were gel purified and mixed together. The mixture was used as template and a third PCR product was generated using forward universal primer F1 and reverse universal primer R2. This final PCR product was again gel purified, digested with Ncol and Xhol restriction enzymes, cloned between the same restriction sites in pTriEX1.1 backbone. Using this cloning strategy, the mutations were incorporated through forward primer F2 and reverse primer R1. The same strategy was used for the construction of all other plasmids expressing N mutants containing internal deletions (Table S1). In all these constructs the forward universal primer F1 and reverse universal primer R2 were used. However, the sequences of reverse R1 and forward F2 primers were different depending upon the type of mutation (Table S1). The plasmid pFLuc was generated by the PCR amplification of the FLuc gene from PGL3 vector forward (Promega) using primer: GCCACCATGGAAGACGCCAAAAACATAAAGA and a reverse primer: AATAACTCGAGCACGGCGATCTTTCCGCCCTTCTTG, containing flanking NcoI and XhoI restriction sites. The PCR product was digested with NcoI and XhoI restriction enzymes and cloned between same sites in pTriEx1.1 backbone (Novagen).