



Supple. Fig. 2









CD115



EL4





D

WT

B16F10

miR-155^{-/-}







۸

30**-**

20-

10-

С

% splenic Gr1⁺CD11b⁺





В







А







Supplemental Figure 1. Tumor growth in miR-155-deficient mice. WT or miR-155^{-/-} mice (n=5) were inoculated s.c. with 10⁶ B16-SIY cells (**A**). Tumor volume was measured every 3 days. (**B**) The suppressive activity of MDSCs sorted from B16-SIY-bearing WT or miR-155^{-/-} mice. Sorted WT or miR-155^{-/-} tumor-infiltrating Gr1⁺CD11b⁺ MDSCs were added at different ratios to OT-I splenocytes stimulated with OVA-I peptides for 3d, and 3[H] thymidine uptake was measured. *, *p*<0.05; **, *p*<0.01.

Supplemental Figure 2. Host miR-155 deficiency enhanced antigen-specific antitumor T cell immunity. (A) Percent CD4⁺, CD8⁺, Gr1⁺CD11b⁺, CD45⁺CD19⁺ and CD49b⁺NK1.1⁺ cells in tumor infiltrates of WT or miR-155^{-/-} mice collected 14-21 days after inoculation with LLC1-OVA tumor cells (n=10-19). (B) CD8⁺IFN- γ^+ T cell frequency in spleen, DLN and tumor from LLC1-OVAbearing WT or miR-155-/- mice 14-21 days after tumor inoculation (n=11-15). (C) Representative flow analysis of tumor antigen-specific CD8⁺ T cell tumor infiltrates from LLC1-OVA-bearing WT or miR-155^{-/-} mice. Frequency of tetramer⁺ cells specific for the OVA epitope SIINFEKL in CD8⁺ infiltrates from mice in B, was summarized (n = 5). *, *p*<0.05; **, *p*<0.01; ***, *p*<0.001.

Supplemental Figure 3. miR-155 is required for optimal function of DCs and T cells from tumor-bearing mice. (A) Flow analysis for cell surface expression of IA/IE, CD80, CD40 and CD86 on tumor-infiltrating myeloid DC (CD11b⁺CD11c⁺) from LLC1-OVA tumor-bearing mice. The results are

summarized from three repeated experiments. (**B**) Sorted tumor-infiltrating DCs from WT or miR-155^{-/-} tumor-bearing mice were added at different ratios to stimulate CFSE-labeled OT-I CD8⁺ T cells for 3d. The dilution of CFSE was detected by flow cytometry and the results were summarized. (**C**) Sorted tumor-infiltrating CD8⁺ T cells from WT or miR-155^{-/-} tumor-bearing mice were labeled with eFluor450 and added at 1:2 to WT DCs stimulated with OVA-I peptides for 3d. The dilution of eFluor450 was detected by flow cytometry and the results were summarized. if were summarized. *, *p*<0.05; **, *p*<0.01.

Supplemental Figure 4. miR-155 is required for splenic MDSC accumulation in different tumor models. (A) Percentages of splenic $Gr1^+CD11b^+$ MDSCs were determined by flow cytometry from EL4-, B16F10- and (B) LLC1 tumorbearing mice (n=5). *, *p*<0.05. (C) Percentages of splenic CD11b⁺Ly6G⁺Ly6C^{low} (granulocytic) and CD11b⁺Ly6G⁻Ly6C^{high} (monocytic) MDSCs from WT or miR-155^{-/-} LLC1 tumor-bearing mice were determined by flow cytometry. (D) Expression of CD115 and CD124 on both MDSC subsets above was determined by flow cytometry. The MFI (mean fluorescence intensity) of CD115 or CD124 expression was summarized (n = 5).

Supplemental Figure 5. miR-155 regulates BM-derived MDSC. (**A**) BM cells were cultured with GM-CSF or GM-CSF+IL-6 for 3 d and miR-155 expression was assayed by quantitative real-time PCR (n=3). (**B**) The suppressive activity of the GM-CSF and IL-6-conditioned BM-derived MDSCs from miR-155^{-/-} mice

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versus WT mice. (**C**) The suppressive activity of the GM-CSF and IL-6conditioned BM-derived MDSCs by the transfection with pre-miR-155/BIC (P-MDSC), miR-155 inhibitor miRNA (I-MDSC) or control oligonucleotides (C-MDSC) by AMAXA. *, p<0.05.

Supplemental Figure 6. Decreased accumulation of Tregs in miR-155deficient tumor-bearing mice. (A) Representative dot plots of Foxp3 expression in EG7 tumor-infiltrating CD4⁺ cells. (B) Percent Foxp3⁺ cells among CD4⁺ cells from spleen and tumor tissues of EG7-bearing WT or miR-155^{-/-} mice were summarized (n=5). *, p<0.05; **, p<0.01.

Supplemental Figure 7. Suppressive activity and tumor-promoting role of miR-155-deficient Tregs. (A) WT or miR-155^{-/-} CD4⁺CD25⁺ Tregs were sorted and added at different ratios to CFSE-labeled CD4⁺CD25⁻ Tres cells stimulated with anti-CD3 and WT DCs for 3d. The dilution of CFSE was detected by flow cytometry and the results were summarized. (B) Percentages of CD39, CD73, GITR, CD44, CD62L and CTLA4 expression among splenic CD4⁺CD25⁺Foxp3⁺ Tregs from WT or miR-155^{-/-} LLC1-bearing mice were measured by flow cytometry. (C) Sorted WT or miR-155^{-/-} splenic CD4⁺CD25⁺ Tregs were injected i.v. into LLC1-bearing mice on d7, d14 and d20. Tumor volumes were measured every 3 days.

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