

Functionally distinct groups of inherited PTEN mutations in autism and tumour syndromes

**Laura Spinelli, Fiona M. Black, Jonathan N. Berg, Britta J. Eickholt and
Nicholas R. Leslie**

Figure S1

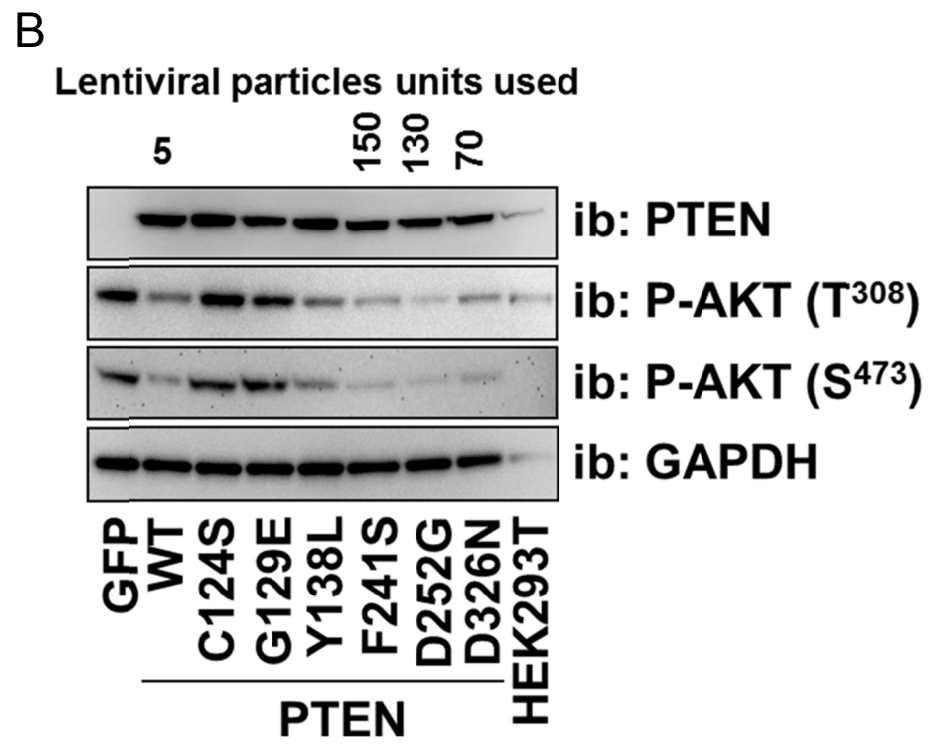
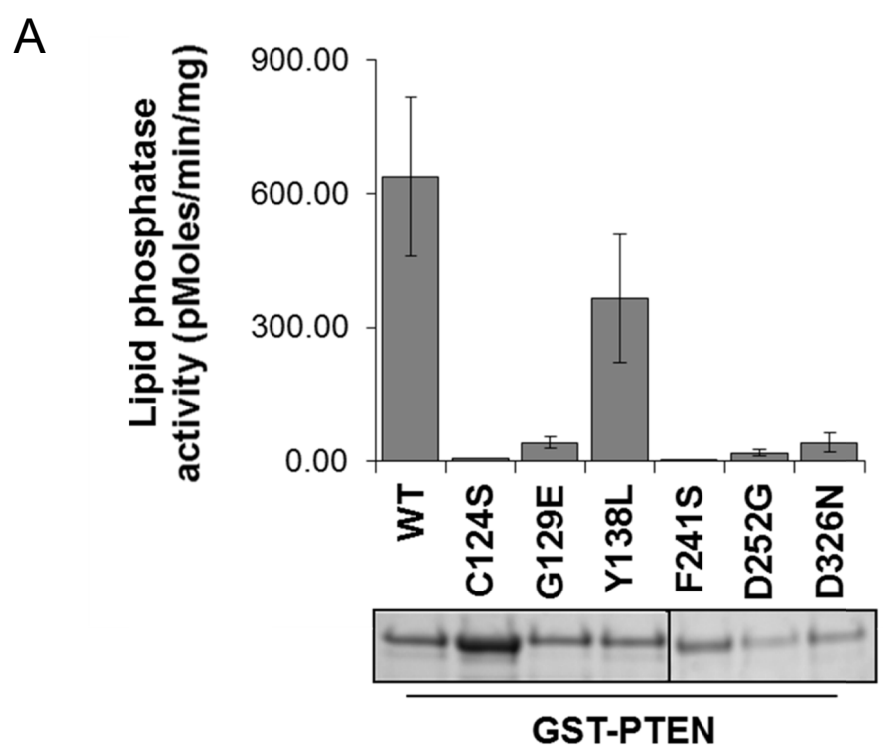


Figure S1. PTEN mutants characterisation in bacteria and U87MG cells

Autism-related mutations proteins drastically reduce PTEN phosphatase activity when expressed in bacteria but they are able to regulate AKT when expressed in U87MG cells. PTEN WT and PTEN mutants (C124S, G129E, Y138L, F241S, D252G and D326N) were expressed in bacteria and purified as GST-fusion proteins. (A) The proteins were separated by SDS-PAGE and stained with Coomassie Brilliant Blue R-250. Proteins were assayed against ^{33}P radiolabelled PIP_3 for 1 hour at 37°C . The activity is presented as the mean activity \pm s.e.m from three experiments performed in duplicates. (B) PTEN null U87MG cells were transduced for 48 hours with lentiviruses encoding GFP, PTEN WT or C124S, G129E, Y138L, F241S, D252G and D326N. PTEN expression and AKT phosphorylation were investigated by western blotting of total cell lysates using total and phospho-specific antibodies. Units of lentiviruses particles used to express PTEN are indicated (5 units are equal to $50\ \mu\text{l}$ of viral supernatant).

Figure S2

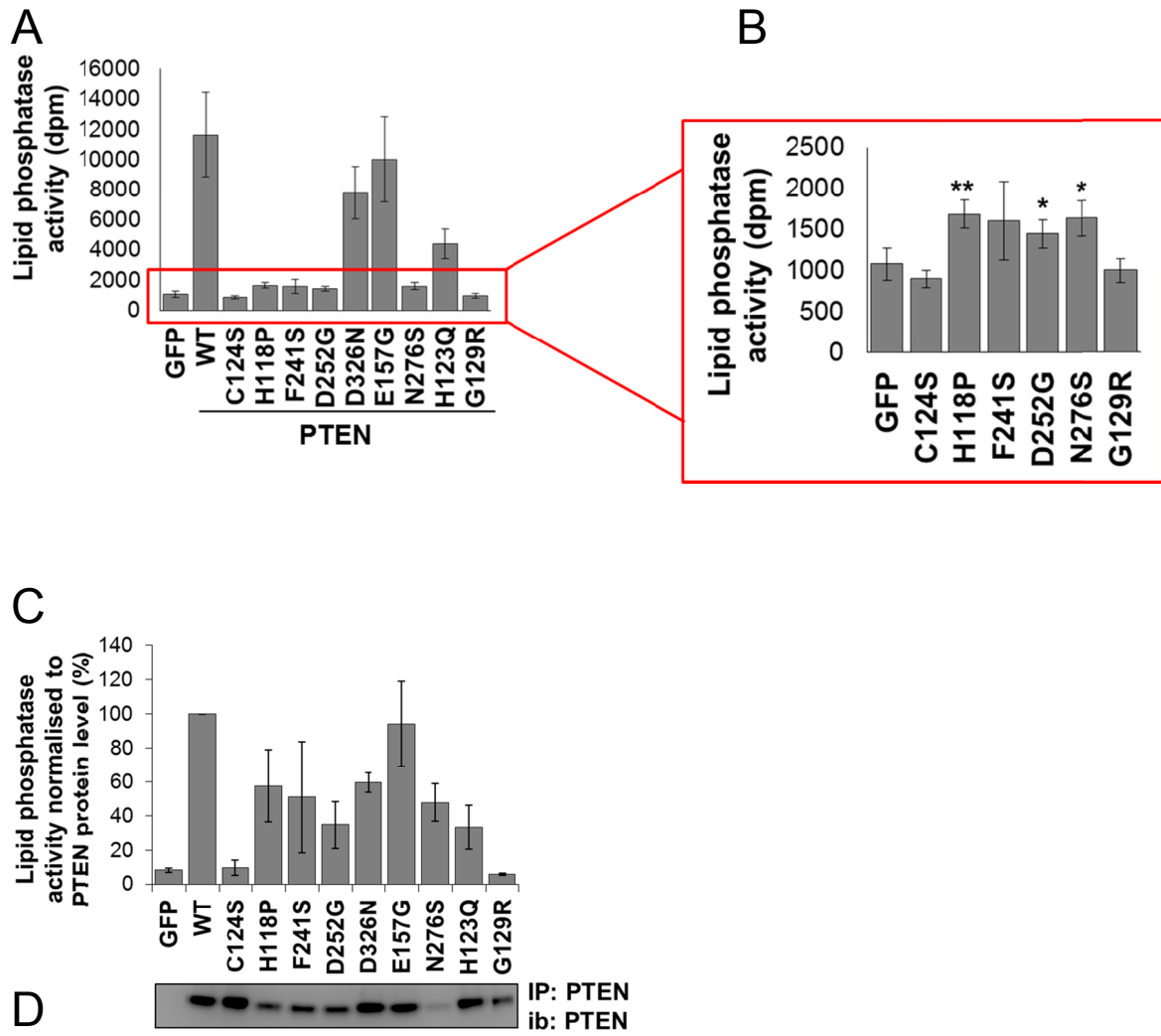


Figure S2. Raw data from the PTEN lipid phosphatase activity assay

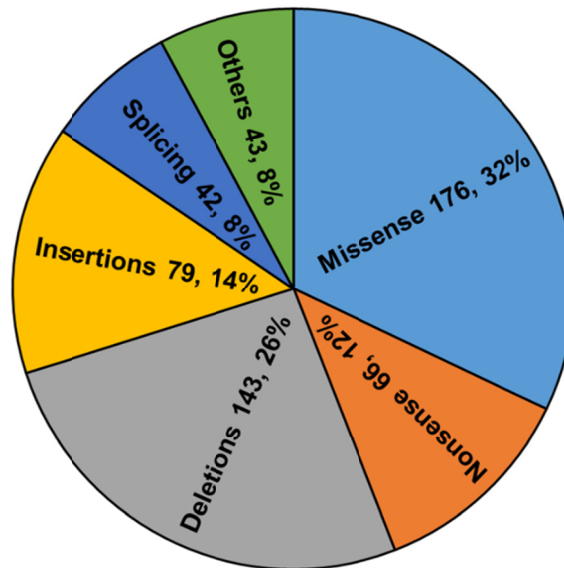
PTEN proteins or GFP were transiently over-expressed in U87MG cells and after cell lysis, immunoprecipitated using an antibody raised against the N-terminus of PTEN. Immune complexes were assayed against radiolabelled PIP₃ vesicles. (A) Mean raw phosphatase activity in dpm released phosphate is shown, derived from six experiments performed on different days, \pm s.e.m.

(B) This shows the same mean raw phosphatase activity data as (A) rescaled to show the differences between raw values with low recovered activity. Across the six experiments there was no overlap between the released phosphate values for any of the seven mutants and the data obtained using the inactive PTEN C124S control. Accordingly, for six of the seven mutants the measured released phosphate was significantly higher than detected with this inactive control (the exception being F241S). (** $p < 0.01$, * $p < 0.05$ compared to PTEN C124S; Student's *t*-test using Excel and GraphPad Prism software). (C) This shows the mean activity from six independent experiments \pm s.e.m as a % of wild-type PTEN activity after normalisation to the amount of protein immunoprecipitated. Immunoprecipitated protein was determined by densitometry (ImageQuant TL software) after immunoblotting and direct chemiluminescence detection using CCD camera. (D) A representative immunoblot of the immunoprecipitated proteins from one of the experiments.

Figure S3

A

All germline PTEN mutations



B

ASD-associated PTEN mutations

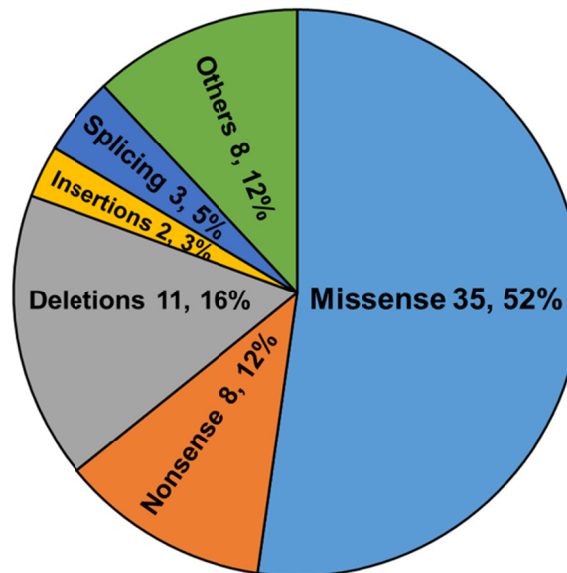


Figure S3. Classification of all inherited PTEN mutations (A) and selectively those associated with ASD (B). The germline PTEN mutations detailed in Tables S1, S3 and S4 were classified into the categories shown. (A) displays the number of apparently independent patients carrying PTEN mutations and (B) the classification of PTEN mutations in independent patients described with ASD, either in the presence or absence of other symptoms including PHTS. A significantly larger fraction of ASD associated mutations are missense mutations 35/67 and 176/549 ($p < 0.005$, Fishers Exact Test)

Figure S4

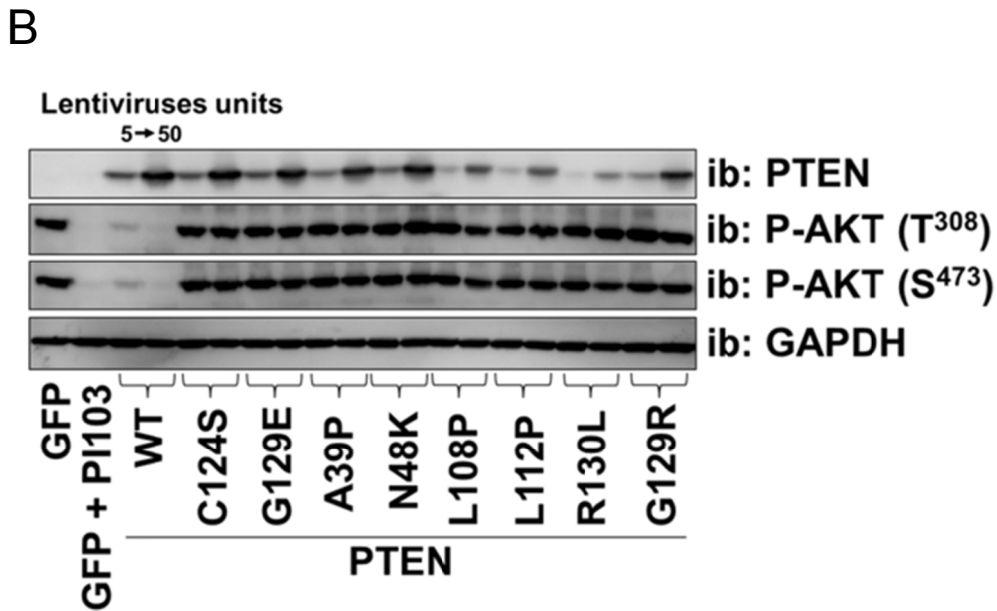
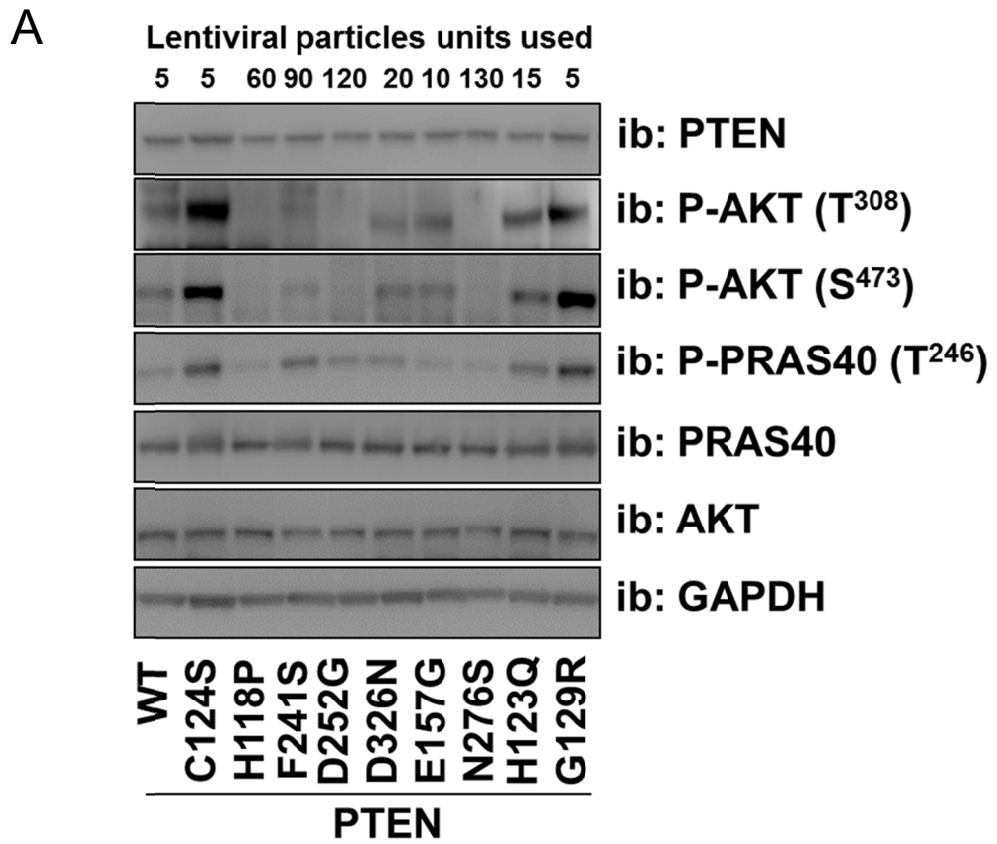


Figure S4. Regulation of AKT and downstream pathways by autism- and Cowden-related mutant proteins in U87MG cells

PTEN null U87MG cells were transduced for 48 hours with lentiviruses encoding PTEN WT or mutants using (A) different amount of lentiviral particles, as shown, for each mutant in order to obtain the same PTEN expression level or (B) two concentrations (low and high) of lentiviruses particles (5 units are equal to 50 μ l of viral supernatant). Cell expressing GFP and cells treated with the PI3K inhibitor PI103 (1 μ M for 30 minutes) were used as control. The evaluation of PTEN and phosphorylated and total AKT and PRAS40 in the lysates from transduced cells was carried out by western blotting using total and phospho-specific antibodies. The blotting panels shown are representative of three independent experiments.

Table S1. PTEN mutations associated with ASD

Missense	Nonsense	Deletions	Insertions	Splicing	Others	References
c.278A>G H93R						Butler et al., 2005
c.722T>C F241S						Butler et al., 2005
c.755A>G D252G						Butler et al., 2005
c.66C>G D22E						Buxbaum et al., 2007
					c._1088C>T	Buxbaum et al., 2007
					c._1084C>T	Buxbaum et al., 2007
					c. IVS3-9T>C	Buxbaum et al., 2007
c.976G>A D326N						Buxbaum et al., 2007
					c._1026C>A	Buxbaum et al., 2007
					c._903G>A	Buxbaum et al., 2007
c.353A>C H118P						Orrico et al., 2009
c.530T>G Y176C						Orrico et al., 2009
c.824A>G N276S						Orrico et al., 2009
			c.519-520insT			Varga et al., 2009
	c.388C>T R130*					Varga et al., 2009
	c.416T>A L139*					Varga et al., 2009
c.470A>G E157G						Varga et al., 2009
c.369C>G H123Q						McBride et al., 2010
c.518G>A R173H						McBride et al., 2010
c.232A>G T78A						Schaaf et al., 2011
c.75G>T L25F						Mester et al., 2012
	c.1003C>T R335*					Mester et al., 2012
		Whole gene deletion				Mester et al., 2012
			c.420-421insA			Mester et al., 2012
c.392C>T T131I						O'Roak et al., 2012
c.500C>A T167N						O'Roak et al., 2012
		c.158-159delTA				Bubien et al., 2013
		c.284delC				Bubien et al., 2013

Missense	Nonsense	Deletions	Insertions	Splicing	Others	References
	c.388C>T R130*					Bubien et al., 2013
	c.388C>T R130*					Bubien et al., 2013
c.392C>T T131I						Bubien et al., 2013
c.403A>G I135V						Bubien et al., 2013
c.406T>C C136R						Bubien et al., 2013
c.518G>A R173H						Bubien et al., 2013
		c.586delC				Bubien et al., 2013
		c.586delC				Bubien et al., 2013
		c.586delC				Bubien et al., 2013
				c.635-1G>C		Bubien et al., 2013
				c.635-3C>G		Bubien et al., 2013
c.830C>G T277R						Bubien et al., 2013
		c.1004- 1005delGA				Bubien et al., 2013
c.202T>A Y68N						Klein et al., 2013
c.113C>A P38H						Klein et al., 2013
c.764T>C V255A						Klein et al., 2013
	c.959T>G L320*					Vanderver et al., 2014
c.17A>T L6I						Vanderver et al., 2014
c.194A>G Y65C						Vanderver et al., 2014
c.511C>G Q171E						Vanderver et al., 2014
	c.388C>T R130*					Vanderver et al., 2014
c.737C>T P246L						Vanderver et al., 2014
c.633C>G C211W						Vanderver et al., 2014
		c.43delA				Marchese et al., 2014
c.208C>G L70V						Hobert et al., 2014
					c.3G>T M1I	Hobert et al., 2014
					c.3G>T M1I	Hobert et al., 2014
	c.1003C>T R335*					Hobert et al., 2014
				c.209+5G>A		Hobert et al., 2014

Missense	Nonsense	Deletions	Insertions	Splicing	Others	References
c.40C>G R14G						Frazier et al., 2014
c.755A>T D252V						Frazier et al., 2014
c.202T>C Y68H						Frazier et al., 2014
c.37A>C N12T						Frazier et al., 2014
c.395G>A G132D						Frazier et al., 2014
c.277C>T H93Y						Frazier et al., 2014
		c.597delGTT				Frazier et al., 2014
					c._1177C>T	Frazier et al., 2014
		4-Mb deletion				Frazier et al., 2014
		10q deletion				Frazier et al., 2014

Tables S2 and S3. Severity index scoring system.

Based on the clinical description of the patients carrying PTEN germline mutations, the severity scores were created according to the number of sites affected as well as the presence of mental retardation/ development delay (MR/DD). The presence of macrocephaly was recorded but not included in the severity index score, as macrocephaly was seen in almost every subject, with the chance that it was not recorded in others. For each site affected by benign tumours or lesions it was assigned one point, two points for each site affected by malignant tumours, and one point for the presence of mental retardation. The severity index score is the sum of these numbers. Patients who scored one to three were classified as “mild”, four to five as “moderate” and six or above as “severe”.

The choice of mutations to analyse has been made based on the “index score average” resulting from the average of the scores of all patients carrying the same PTEN mutation. It should be noted that the characterised PTEN mutants associated with severe phenotypes (see figure 4) were selected at a mid-point of the progression of the project and that the data tabulated here contain very recently published information added since that time. Efforts have been made to minimise the chances of double counting but this cannot be excluded.

Table S2

Severity index scoring system			
Site affected	Benign lesion/tumour	Malignant tumour/cancer	MR/DD
CNS	1 point assigned for each site affected	2 points assigned for each site affected	1 point assigned for the presence of Mental Retardation/ Developmental Delay
Thyroid			
Breast			
Skin/Mucosa			
Gastrointestinal tract			
Others			
Total score	1-3 MILD 4-5 MODERATE 6 + SEVERE		

Table S3

Mutations	Index score average	Index score patients	Assigned Phenotype	References
M1I	1	1	ASD	[1] Hobert et al., 2014
		1	ASD	[1] Hobert et al., 2014
K6I	1	1	ASD	[2] Vanderver et al., 2014
K6E	1	1	ASD	[2] Vanderver et al., 2014
K13E	3	4	CS	[3] Bubien et al., 2013
		4	CS	[3] Bubien et al., 2013
		1	CS	[3] Bubien et al., 2013
R14G	0	0	ASD	[4] Frazier et al., 2014
R15S	3	3	CS	[5] Nagy et al., 2011
Y16H	4	4	CS	[6] Ngeow et al., 2014
D22E	0	0	ASD	[7] Buxbaum et al., 2007
D24Y	5	2	BRRS	[8] Celebi et al., 1999
		8	CS	[3] Bubien et al., 2013
D24G	4	4	CS	[6] Ngeow et al., 2014
D24H	6	6	CS	[6] Ngeow et al., 2014
D24V	5	5	CS	[6] Ngeow et al., 2014
L25F	0	0	ASD	[9] Mester et al., 2012
T26P	2	2	Unknown	[9] Mester et al., 2012
Y27N	4	4	CS	[6] Ngeow et al., 2014
Y27C	1	1	Unknown	[2] Vanderver et al., 2014
P30L	1	1	CS	[3] Bubien et al., 2013
A34D	2	2	BRRS	[10] Marsh et al., 1999
		2	BRRS	[10] Marsh et al., 1999
M35R	1	1	JPC	[11] Olschwang et al., 1998
M35T	4	4	PLS	[12] Zhou et al., 2001
G36R	5	5	CS	[13] Celebi et al., 2000
P38H	1	1	ASD	[14] Klein et al., 2013
A39P	7	7	CS	[15] Tate et al., 2008
G44D	2	2	Unknown	[16] Varga et al., 2009
N48K	8	8	CS	[17] Vega et al., 2003
I50T	1	1	Unknown	[2] Vanderver et al., 2014
H61D	0	0	VATER	[18] Reardon et al., 2001
H61R	4	4	CS	[6] Ngeow et al., 2014
Y65C	0	0	ASD	[2] Vanderver et al., 2014
		0	ASD	[2] Vanderver et al., 2014
I67R	7	7	CS	[3] Bubien et al., 2013
Y68N	2	2	ASD	[14] Klein et al., 2013
Y68D	0.5	1	PS	[19] Loffeld et al., 2006
		0	CS	[19] Loffeld et al., 2006
Y68H	1.33	1	BRRS	[20] Marsh et al., 1998
		3	CS	[20] Marsh et al., 1998
		0	ASD	[4] Frazier et al., 2014
Y68C	2	2	CS	[21] Lobo et al., 2009
L70P	2	2	CS	[20] Marsh et al., 1998
L70V	3	3	ASD	[1] Hobert et al., 2014
T78A	0	0	ASD	[22] Schaaf et al., 2011
A79T	3	3	BC	[23] Figer et al., 2002
N82T	3	3	BC	[23] Figer et al., 2002
Y88C	1	1	PHTS	[9] Mester et al., 2012
D92A	5	5	CS	[3] Bubien et al., 2013
H93Y	2.66	4	CS	[24] Kohno et al., 1998
		4	CS	[6] Ngeow et al., 2014
		0	ASD	[4] Frazier et al., 2014

Mutations	Index score average	Index score patients	Assigned Phenotype	References
H93R	1	1	ASD	[25] Butler et al., 2005
P96Q	2.5	3	CS	[26] Bussaglia et al., 2002
		2	CS	[26] Bussaglia et al., 2002
P96R	3.5	3	CS	[9] Mester et al., 2012
		4	CS	[27] Busch et al., 2013
C105Y	2	2	BRRS	[10] Marsh et al., 1999
D107G	1	1	Unknown	[2] Vanderver et al., 2014
L108P	6.7	6	PHTS	[28] Tan et al., 2007
		7	PHTS	[3] Bubien et al., 2013
		7	CS	[29] Ngeow et al., 2013
L112P	11	11	CS	[30] Tsou et al., 1998
H118P	1	1	ASD	[31] Orrico et al., 2009
V119I	4	4	Multiple cancers	[32] De Vivo et al., 2000
		4	Multiple cancers	[32] De Vivo et al., 2000
H123Q	2.5	1	ASD	[33] McBride et al., 2010
		4	CS	[34] Kerseboom et al., 2012
H123R	4	4	CS	[35] Nelen et al., 1997
H123D	3	3	CS	[26] Bussaglia et al., 2002
C124R	4	2	CS	[35] Nelen et al., 1997
		4	CS	[35] Nelen et al., 1997
		6	CS	[35] Nelen et al., 1997
A126P	3	3	CS	[9] Mester et al., 2012
K128E	4	4	CS	[27] Busch et al., 2013
K128N	3	3	CS	[29] Ngeow et al., 2013
G129R	3.5	5	CS	[36] Elia et al., 2012
		2	Unknown	unpublished patient
G129E	4	4	CS	[37] Liaw et al., 1997
		5	CS	[37] Liaw et al., 1997
		7	CS	[37] Liaw et al., 1997
		2	CS	[37] Liaw et al., 1997
		2	CS	[29] Ngeow et al., 2013
R130Q	3.9	5	CS	[38] Kurose et al., 1999
		4	CS/BRRS	[38] Kurose et al., 1999
		3	CS/BRRS	[38] Kurose et al., 1999
		3	CS/BRRS	[38] Kurose et al., 1999
		5	CS	[39] Heindl et al., 2012
		4	CS	[39] Heindl et al., 2012
		5	CS	[29] Ngeow et al., 2013
		4	CS	[6] Ngeow et al., 2014
		2	CS	[6] Ngeow et al., 2014
R130G	1.8	2	CS	[21] Lobo et al., 2009
		4	Unknown	[9] Mester et al., 2012
		1	CS	[3] Bubien et al., 2013
		1	CS	[3] Bubien et al., 2013
		1	Unknown	[3] Bubien et al., 2013
R130L	5	2	CS	[3] Bubien et al., 2013
		3	CS	[3] Bubien et al., 2013
		3	CS	[3] Bubien et al., 2013
		10	CS	[20] Marsh et al., 1998
T131I	1.7	4	CS	[6] Ngeow et al., 2014
		1	CS/ASD	[3] Bubien et al., 2013
		3	Unknown	[3] Bubien et al., 2013
G132A	1	1	ASD	[40] O'Roak et al., 2012
		1	PHTS	[28] Tan et al., 2007

Mutations	Index score average	Index score patients	Assigned Phenotype	References
G132D	3.16	2	CS	[41] Derrey et al., 2004
		6	CS	[3] Bubien et al., 2013
		4	CS	[3] Bubien et al., 2013
		3	CS	[3] Bubien et al., 2013
		4	CS	[39] Heindl et al., 2012
		0	ASD	[4] Frazier et al., 2014
G132V	3	3	PHTS	[42] Tekin et al., 2006
M134R	0.75	0	BRRS	[23] Figer et al., 2002
		0	BRRS	[23] Figer et al., 2002
		0	BRRS	[23] Figer et al., 2002
		3	Unknown	[9] Mester et al., 2012
M134I	0	0	Unknown	[43] Busa et al., 2013
		0	Unknown	[43] Busa et al., 2013
		0	Unknown	[43] Busa et al., 2013
		0	Unknown	[43] Busa et al., 2013
M134T	4.66	1	MR	[33] McBride et al., 2010
		5	CS	[29] Ngeow et al., 2013
		8	Unknown	[27] Busch et al., 2013
I135R	3	3	BRRS	[44] Boccone et al., 2006
I135V	2.7	1	BRRS	[10] Marsh et al., 1999
		3	Unknown	[3] Bubien et al., 2013
		2	Unknown	[3] Bubien et al., 2013
		5	Unknown	[3] Bubien et al., 2013
		3	Unknown	[3] Bubien et al., 2013
		1	Unknown	[3] Bubien et al., 2013
		4	CS	[3] Bubien et al., 2013
C136R	5.6	7	CS	[45] Kubo et al., 2000
		7	CS	[3] Bubien et al., 2013
		2	CS	[3] Bubien et al., 2013
		6	CS	[3] Bubien et al., 2013
		6	CS	[6] Ngeow et al., 2014
R142P	3	3	CS	[3] Bubien et al., 2013
A151B	3	3	CS	[3] Bubien et al., 2013
Y155N	4	4	CS	[3] Bubien et al., 2013
Y155C	3.66	2	CS	[46] Gicquel et al., 2003
		7	CS	[3] Bubien et al., 2013
		2	CS	[29] Ngeow et al., 2013
E157G	1	1	ASD	[16] Varga et al., 2009
V158I	3	4	Unknown	[32] De Vivo et al., 2000
		2	Unknown	[32] De Vivo et al., 2000
D162E	1	1	CS	[9] Mester et al., 2012
G165E	5	5	CS	[47] Nelen et al., 1999
G165V	4	4	CS	[20] Marsh et al., 1998
G165R	2	2	CS	[48] Banneau et al., 2010
T167N	1	1	ASD	[40] O'Roak et al., 2012
S170R	4	2	BRRS	[49] Marsh et al., 1997
		7	CS	[3] Bubien et al., 2013
		4	Unknown	[3] Bubien et al., 2013
		3	CS	[3] Bubien et al., 2013
S170I	4	4	CS	[6] Ngeow et al., 2014
Q171E	0	0	ASD	[2] Vanderver et al., 2014
R173C	2.5	1	BRRS	[50] Lachlan et al., 2007
		4	CS	[6] Ngeow et al., 2014
R173P	3.5	3	LDD	[51] Kirches et al., 2010
		4	Unknown	[51] Kirches et al., 2010

Mutations	Index score average	Index score patients	Assigned Phenotype	References
R173H	0.75	0	Unknown	[50] Lachlan et al., 2007
		2	Unknown	[50] Lachlan et al., 2007
		1	CS/ASD	[3] Bubien et al., 2013
		0	ASD	[33] McBride et al., 2010
R173G	2	2	CS	[6] Ngeow et al., 2014
Y176C	1	1	ASD	[31] Orrico et al., 2009
L181P	1	1	CS	[52] Thiffault et al., 2004
T202I	2	2	Unknown	[16] Varga et al., 2009
C211W	1	1	ASD	[2] Vanderver et al., 2014
V217D	2	2	CS	[53] Kim et al., 2005
R234Q	2	2	Multiple cancers	[54] Staal et al., 2002
F241S	0	0	ASD	[25] Butler et al., 2005
P246L	1	2	BRRS	[10] Marsh et al., 1999
		1	BRRS	[9] Mester et al., 2012
		0	ASD	[2] Vanderver et al., 2014
D252G	1	1	ASD	[25] Butler et al., 2005
		1	CS	[3] Bubien et al., 2013
D252V	0	0	ASD	[4] Frazier et al., 2014
V255A	0	0	ASD	[14] Klein et al., 2013
W274L	1	1	DD	[33] McBride et al., 2010
N276S	1	1	ASD	[31] Orrico et al., 2009
T277R	1.66	1	CS	[3] Bubien et al., 2013
		2	CS	[3] Bubien et al., 2013
		2	CS	[3] Bubien et al., 2013
		3	CS	[3] Bubien et al., 2013
		0	CS/ASD	[3] Bubien et al., 2013
		2	CS	[48] Banneau et al., 2010
K289E	1	1	CS	[55] Chi et al., 1998
D326N	1	1	AUTISM	[7] Buxbaum et al., 2007
R335L	4	4	CS	[56] Sawada et al., 2000
F337S	2	2	CS	[50] Lachlan et al., 2007
K342N	2	3	HTS	[3] Bubien et al., 2013
		1	CS	[3] Bubien et al., 2013
V343E	3.5	4	CS	[57] Lynch et al., 1997
		4	CS	[57] Lynch et al., 1997
		5	CS	[57] Lynch et al., 1997
		3	CS	[57] Lynch et al., 1997
		2	CS	[57] Lynch et al., 1997
		3	CS	[57] Lynch et al., 1997
L345V	4.5	4	CS	[6] Ngeow et al., 2014
		5	CS	[27] Busch et al., 2013

ASD, Autism spectrum disorder; BC, Breast cancer; BRRS, Bannayan-Riley-Ruvalcaba Syndrome; CS, Cowden Syndrome; DD, Developmental Delay; HTS, Hamartoma Tumour Syndrome; JPS, Juvenile Polyposis Coli; LDD, Lhermitte-Duclos disease; MR, Mental Retardation; PHTS, PTEN Hamartoma-Tumour Syndrome; PLS, Proteus-Like Syndrome; PS; Proteus Syndrome.

MILD	MODERATE	SEVERE
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Table S4 Clinical features of the 204 patients carrying germline-derived PTEN missense mutations.

Mutation	M11	M11	K6I	K6E	N12T	K13E	K13E	K13E	K13E	R14G	R15S
Diagnosis	ASD	ASD	ASD		ASD	CS	CS	CS	CS	ASD	CS
Sex	M	M	M	F	M					F	
Age	11y	12y	3y	11m	5y2m					3y3m	
CNS											
Macrocephaly	X	X	X	X	X	X	X	X	X	X	X
LDD											
Benign Tumour											
Malignant Tumour											
MR/DD			X	X							
THYROID											
Benign (adenoma, goitre)						X	X	X	X		X
Malignant											
BREASTS											
Benign											
Malignant Tumour											
SKIN/MUCOSA											
Trichilemmona											
MCL, FP, AK, F	X	X				X	X	X	X		
OP	X	X				X	X	X	X		
SP											
Lipomas											
Malignant Tumour											
GASTROINTESTINAL TRACT											
GI polyps											
Benign (hamartoma)											
Malignant Tumour											X
OTHER BENIGN											
OTHER CANCERS											
SI SCORE	1	1	1	1	1	0	4	4	4	1	0
REFERENCES	Hobert et al., 2014	Hobert et al., 2014	Vanderver et al., 2014	Vanderver et al., 2014	Frazier et al., 2014	Bubien et al., 2013	Bubien et al., 2013	Bubien et al., 2013	Bubien et al., 2013	Frazier et al., 2014	Nagy et al., 2011

Mutation	Y16H	D22E	D24Y	D24Y	D24Y	D24G	D24H	D24V	L25F	T26P	Y27N
Diagnosis	CS	ASD	BRRS	CS	CS	CS	CS	CS	ASD		CS
Sex	F	M	M		F	F	F	F	M	F	F
Age	65y		4y		49y	46y	28y	7y	19y		46y
CNS											
Macrocephaly		X	X				X	X	X	X	
LDD										X	
Benign Tumour											
Malignant Tumour			X								
MR/DD											
THYROID											
Benign (adenoma, goitre)				X			X			X	
Malignant											
BREASTS											
Benign				X							
Malignant Tumour	X			X	X	X	X	X			X
SKIN/MUCOSA											
Trichilemmona											
MCL, FP, AK, PPK, F				X			X				
OP				X							
SP											
Lipomas			X								
			X								
Malignant Tumour				X							
GASTROINTESTINAL TRACT											
GI polyps											
Benign (hamartoma)											
Malignant Tumour											
OTHER BENIGN											
OTHER CANCERS											
SI SCORE	4	0	2	8	4	6	5	0	2	4	
	ENCA	GU les	ENCA	ENCA-RC	UTLM	RC					

Ngeow et al., 2007 al., 2009 al., 2013 al., 2014 al., 2014 al., 2012 al., 2012 al., 2014 al., 2014

REFERENCES

Mutation	Y27C	P30L	A34D	A34D	A34D	M35R	M35T	G36R	P38H	A39P	G44D
Diagnosis	CS	CS	BRRS	BRRS	BRRS	JPC	PLS	CS	ASD	CS	
Sex	F		M	M	M	M	M	F	M	F	F
Age	3y10m		10y	10y	10y	7y	7y	47y	15y	50-59y	27m
CNS											
Macrocephaly	X	X	X	X	X		X	X	X	X	X
LDD	X										
Benign Tumour											
Malignant Tumour	X		X	X	X		X				X
MR/DD											
THYROID											
Benign (adenoma, goitre)										X	
Malignant											
BREASTS											
Benign											
Malignant Tumour								X		FBD	
SKIN/MUCOSA											
Trichilemmona								X			
MCL, FP, AK, PPK, F		X						X		X	
OP								X		X	
SP		X	X	X	X		X		X		
Lipomas			X	X	X				X		
Malignant Tumour											
GASTROINTESTINAL TRACT											
GI polyps								X		X	
Benign (hamartoma)						JPC					
Malignant Tumour											
OTHER BENIGN											
OTHER CANCERS											
SI SCORE	1	1	2	2	2	1	1	4	5	1	7

VanderVer Bubien et al., 2013
 Marsh et al., 1999
 Marsh et al., 1999
 Marsh et al., 1999
 Olschwang Zhou et al., 2000
 Celebi et al., 2000
 Klein et al., 2013
 Klein et al., 2013
 Tate et al., 2008
 Varga et al., 2009

Mutation	Y68H	Y68H	Y68C	L70P	L70V	T78A	A79T	N82T	Y88C	D92A
Diagnosis	CS	ASD	CS	CS	ASD	ASD	BC	BC	PHTS	CS
Sex	F	M	F	M	M	F	F	F	M	
Age	75y	2y	46y	31y	13y		36y	35y	1y	
CNS										
Macrocephaly		X			X	X			X	X
LDD										
Benign Tumour										
Malignant Tumour										
MR/DD					X			X		
THYROID										
Benign (adenoma, goitre)	X				X					X
Malignant			X	X						
BREASTS										
Benign	X						X	X		X
Malignant Tumour			X				X	X		
SKIN/MUCOSA										
Trichilemmona	X									
MCL, FP, AK, PPK, F	X									X
OP	X				X					X
SP										
Lipomas					X					X
Malignant Tumour										

GASTROINTESTINAL TRACT

GI polyps

Benign (hamartoma)

Malignant Tumour

OTHER BENIGN

OTHER CANCERS

SI SCORE



Marsh et al., 1998
 Frazier et al., 2014
 Ngeow et al., 2014
 Marsh et al., 1998
 Hobert et al., 2014
 Schaaf et al., 2011
 Figer et al., 2002
 2002
 al., 2012
 al., 2013

REFERENCES

Mutation	L108P	L108P	L108P	L112P	H118P	V119I	V119I	H123Q	H123Q	H123Q	H123R
Diagnosis	PHTS	PHTS	CS	CS	ASD	Mcancer	Mcancer	ASD	CS	CS	CS
Sex			F	F		F	F	F	F	F	M
Age		50y	26y					2y6m	28y		
CNS											
Macrocephaly	X		X	X	X			X	X	X	X
LDD			X	X							
Benign Tumour											
Malignant Tumour			X	X							
MR/DD	X		X	X	X			X			
THYROID											
Benign (adenoma, goitre)		X	X	X						X	X
Malignant											X
BREASTS											
Benign	FBD	X	X	X					X		
Malignant Tumour		X	X			X	X				
SKIN/MUCOSA											
Trichilemmona			X	X							
MCL, FP,AK, PPK, F			X	X							X
OP			X	X							
SP											
Lipomas	X	X									
Malignant Tumour			X								
GASTROINTESTINAL TRACT											
GI polyps		X									
Benign (hamartoma)											
Malignant Tumour										X	
OTHER BENIGN											
OTHER CANCERS	VASC les	GU les									
SI SCORE	OVCA	ENCA	ENCA	ENCA	ENCA	LUNG	ENCA	ENCA	ENCA	ENCA	ENCA
	6	7	7	7	11	1	4	4	1	4	4
REFERENCES	Tan et al., 2007	Bubien et al., 2013	Ngeow et al., 2013	Tsou et al., 1998	Orrico et al., 2009	deVivo et al., 2000	deVivo et al., 2000	McBride et al., 2010	Kerseboom Nelen et al., 1997		

Mutation	H123D	C124R	C124R	C124R	C124R	A126P	K128E	K128N	G129R	G129R	G129E
Diagnosis	CS	CS	CS	CS	CS	CS	CS	CS	CS	CS	CS
Sex	F	F				M	M	M	F	M	
Age	53y					35y	23y	35y	14y7m		
CNS											
Macrocephaly	X	X				X	X		X	X	
LDD											
Benign Tumour							X		X		
Malignant Tumour									X		
MR/DD									X		
THYROID											
Benign (adenoma, goitre)	X	X	X	X	X	X	X		X	X	X
Malignant											
BREASTS											
Benign			X	X	X						X
Malignant Tumour											
SKIN/MUCOSA											
Trichilemmona	X										
MCL, FP, AK, PPK, F	X	X	X	X	X						X
OP	X						X				
SP						X	X				
Lipomas						X					
Malignant Tumour											
GASTROINTESTINAL TRACT											
GI polyps									X		
Benign (hamartoma)			X	X	X	X		X			X
Malignant Tumour				X	X			X			
OTHER BENIGN											
OTHER CANCERS	OC						VASC les			VASC les	
SI SCORE	3	2	4	4	6	3	3	4	3	5	2
REFERENCES	Bussaglia et al., 2002	Nelen et al., 1997	Nelen et al., 1997	Nelen et al., 1997	Mester et al., 2012	Mester et al., 2012	Buschet et al., 2013	Ngeow et al., 2013	Elia et al., 2012	UNPUB	Liaw et al., 1997

Mutation	R130Q	R130Q	R130Q	R130Q	R130G	R130G	R130G	R130G	R130G	R130G	R130L
Diagnosis	CS	CS	CS	CS	CS	CS	CS	CS	CS	CS	CS
Sex	F	F	F	F	F	F					
Age	42y	29y	48y	40y							
CNS											
Macrocephaly	X			X	X	X	X	X	X	X	X
LDD											
Benign Tumour											
Malignant Tumour											
MR/DD											
THYROID											
Benign (adenoma, goitre)	X			X							X
Malignant											
BREASTS											
Benign	X										
Malignant Tumour	X	X	X	X							
SKIN/MUCOSA											
Trichilemmona											
MCL, FP, AK, PPK, F							X	X	X	X	X
OP							X				
SP											
Lipomas	X							X	X	X	
Malignant Tumour											
GASTROINTESTINAL TRACT											
GI polyps											
Benign (hamartoma)											
Malignant Tumour											
OTHER BENIGN											
OTHER CANCERS											
SI SCORE											
		OVCA		UF							
	5	4	4	2	2	4	4	1	1	1	2
	3										

Ngeowet al., 2013
Ngeowet al., 2014
Ngeowet al., 2014
Ngeowet al., 2014
Mester et al., 2012
Bubien et al., 2013
Bubien et al., 2013
Bubien et al., 2013
Bubien et al., 2013
Bubien et al., 2013
Bubien et al., 2013
Bubien et al., 2013

REFERENCES

Mutation	R130L	R130L	R130L	R130L	T131I	T131I	T131I	T131I	G132A	G132D	G132D	G132D	G132D
Diagnosis	CS	CS	CS	CS	CS/ASD	ASD	PHTS	CS	CS	CS	CS	CS	CS
Sex	F	F	F	F	F	F	M	M	M	M	M	M	M
Age	21y	21y	21y	21y	4y1m	4y1m	27y	27y	27y	53y	53y	53y	53y
CNS													
Macrocephaly	X			X	X	X	X		X		X		X
LDD													
Benign Tumour													
Malignant Tumour													
MR/DD				X		X		DGCC					
THYROID													
Benign (adenoma, goitre)	X				X					X			X
Malignant	X												X
BREASTS													
Benign	X												X
Malignant Tumour	X												X
SKIN/MUCOSA													
Trichilemmona													
MCL, FP, AK, PPK, F	X												X
OP	X												X
SP	X												X
Lipomas													
Malignant Tumour													
GASTROINTESTINAL TRACT													
GI polyps	X												
Benign (hamartoma)					X								
Malignant Tumour													
OTHER BENIGN													
OTHER CANCERS													
SI SCORE	3	10	4	1	3	1	1	1	2	4	6		
REFERENCES													
	Bubien et al., 2013	Marsh et al., 1998	Ngeow et al., 2014	Bubien et al., 2013	Bubien et al., 2013	O'Roak et al., 2012	Tan et al., 2007	Derrey et al., 2004	Heindl et al., 2012	Bubien et al., 2013	Heindl et al., 2012	Bubien et al., 2013	Bubien et al., 2013

UBC

RC

Mutation	I135V	I135V	I135V	I135V	C136R	C136R	C136R	C136R	C136R	C136R	R142P	A151D
Diagnosis		ASD	CS	CS	CS	CS	D	CS	CS	CS	CS	CS
Sex				F					F			
Age				30y					33y			
CNS												
Macrocephaly	X	X	X			X					X	X
LDD								X				
Benign Tumour												
Malignant Tumour												
MR/DD						X						
THYROID												
Benign (adenoma, goitre)	X		X	X				X			X	X
Malignant						X		X				
BREASTS												
Benign			X	X								
Malignant Tumour					X							
SKIN/MUCOSA										X		
Trichilemmona												
MCL, FP, AK, PPK, F	X		X	X		X		X			X	X
OP	X		X	X				X			X	X
SP								X				
Lipomas		X	X	X			X					X
Malignant Tumour												
GASTROINTESTINAL TRACT												
GI polyps	X		X	X		X		X			X	X
Benign (hamartoma)												
Malignant Tumour				X								
OTHER BENIGN				GU les								
OTHER CANCERS												
SI SCORE	3	1	4	7	7	2	6	6	3	3	3	3
REFERENCES	ENCA											
	Bubienet al., 2013	Bubienet al., 2013	Bubienet al., 2013	Kubo et al., 2000	Bubienet al., 2013	Bubienet al., 2013	Bubienet al., 2013	Bubienet al., 2013	Bubienet al., 2013	Ngeowet al., 2014	Bubienet al., 2013	Bubienet al., 2013

Mutation	Y155N	Y155C	Y155C	Y155C	E157G	V158I	V158I	D162E	G165R	G165E
Diagnosis	CS	CS	CS	CS	ASD	F	F	CS	CS	CS
Sex	F	F	F	F	3y8m			M		F
Age	50y	50y	66y	66y				3y	59y	
CNS										
Macrocephaly		X	X	X	X			X		X
LDD										
Benign Tumour										
Malignant Tumour										
MR/DD					X			X		
THYROID										
Benign (adenoma, goitre)	X	X								X
Malignant										X
BREASTS										
Benign		X	X	X						X
Malignant Tumour		X	X			X			X	
SKIN/MUCOSA										
Trichilemmona										
MCL, FP, AK, PPK, F	X	X								X
OP	X	X								
SP										
Lipomas										
Malignant Tumour	X			X						
GASTROINTESTINAL TRACT										
GI polyps										
Benign (hamartoma)										
Malignant Tumour										
OTHER BENIGN										
OTHER CANCERS										
SI SCORE										
	4	2	7	2	2	1	4	2	1	2
	MT						LGTC			L
REFERENCES	Bubien et al., 2013	Gicquel et al., 2003	Bubien et al., 2013	Ngeow et al., 2013	Varga et al., 2009	Varga et al., 2000	de Vivo et al., 2000	Mester et al., 2012	Bannean et al., 2010	Nelen et al., 1999

Mutation	R173G	R173P	R173P	R173H	R173H	R173H	R173H	R173H	R173H	R173H	Y176C	L181P	T202I
Diagnosis	CS	LD	F	CS	M	M	M	ASD	ASD	ASD	ASD	CS	
Sex	F	F	F		M	M	M	M	M	M			M
Age	44y		42y	9y	43y	4y5m							1y8m
CNS													
Macrocephaly		X	X	X	X	X	X	X	X	X	X	X	
LDD		X	X										
Benign Tumour													
Malignant Tumour													
MR/DD			X	X						X			X
THYROID													
Benign (adenoma, goitre)		X	X										
Malignant													
BREASTS													
Benign													
Malignant Tumour													
SKIN/MUCOSA		X											
Trichilemmona													
MCL, FP, AK, PPK, F		X	X										X
OP									X				
SP									X				
Lipomas													
Malignant Tumour													
GASTROINTESTINAL TRACT													
GI polyps													
Benign (hamartoma)													
Malignant Tumour									X				
OTHER BENIGN													
OTHER CANCERS													
SI SCORE	2	3	4	4	1	1	0	2	0	1	1	1	2

VASC les

Ngeow et Kirches et Bubien et Lachlan et Lachlan et McBride et Orrico et Thiffault et Varga et al., 2014 al., 2010 al., 2010 al., 2013 al., 2007 al., 2007 al., 2010 al., 2009 al., 2004 al., 2009

Mutation	C211W	V217D	R234Q	F241S	P246L	P246L	P246L	P246L	D252V	D252G	D252G
Diagnosis	ASD	CS	Mcancer	ASD	BRRS	BRRS	BRRS	ASD	ASD	ASD	CS
Sex	M		M	M	M	M	M	M	F	M	
Age	3y10m		38y	2y5m	2y	2y	5y	5y	15y5m	3y5m	
CNS											
Macrocephaly	X				X		X	X	X	X	X
LDD											
Benign Tumour											
Malignant Tumour			OD, MnG								
MR/DD	X				X					X	
THYROID											
Benign (adenoma, goitre)											
Malignant											
BREASTS											
Benign											
Malignant Tumour											
SKIN/MUCOSA											
Trichilemmona											
MCL, FP, AK, PPK, F											X
OP		X									X
SP											
Lipomas											
Malignant Tumour											
GASTROINTESTINAL TRACT											
GI polyps											
Benign (hamartoma)											
Malignant Tumour											
OTHER BENIGN											
OTHER CANCERS											
SI SCORE	1	2	2	0	2	2	1	0	0	1	1

VASC les

Vanderver Kim et al., 2014
 et al., 2014 2005
 Staal et al., 2002
 Butler et al., 2005
 Butler et al., 2005
 Marsh et al., 1999
 Mester et al., 2012
 Vanderver et al., 2014
 Frazier et al., 2014
 Butler et al., 2005
 Bubien et al., 2013

Mutation	V255A	W274L	N276S	T277R	T277R	T277R	T277R	T277R	T277R	T277R	T277R	T277R	K289E
Diagnosis	ASD	DD	ASD	CS	CS	CS	CS	CS	CS	CS/ASD	CS	CS	CS
Sex	M	F								F			
Age	4y4m	8m											27y
CNS													
Macrocephaly	X	X	X	X	X	X	X	X	X	X	X	X	X
LDD													
Benign Tumour													
Malignant Tumour													
MR/DD		X	X										
THYROID													
Benign (adenoma, goitre)					X	X	X	X	X				
Malignant													
BREASTS													
Benign									X				
Malignant Tumour													X
SKIN/MUCOSA													
Trichilemmona													
MCL, FP, AK, PPK, F				X	X	X	X	X	X	X	X	X	X
OP				X	X	X	X	X	X	X	X	X	X
SP													
Lipomas													
Malignant Tumour				X									
GASTROINTESTINAL TRACT													
GI polyps													X
Benign (hamartoma)													X
Malignant Tumour													
OTHER BENIGN													
OTHER CANCERS													
SI SCORE	0	1	1	1	1	1	2	2	2	3	0	2	1

Klein et al., McBride et Orrico et Bubien et Bubien et Bubien et Bubien et Bubien et Bubien et Bubien et Banneau Chi et al., 2013 al., 2010 al., 2009 al., 2013 al., 2013 al., 2013 al., 2013 al., 2013 al., 2010 1998

REFERENCES

Mutation	D326N	R335L	F337S	K342N	K342N	V343E	V343E	V343E	V343E	V343E	V343E	V343E	L345V	L345V	
Diagnosis	ASD	CS	CS	CS	HTS	CS	CS	CS	CS	CS	CS	CS	CS	CS	
Sex	F	F	M			F	F	M	F	F	F	F	F	F	
Age	48y	48y	9y										36y	58y	
CNS															
Macrocephaly	X		X	X	X										
LDD															
Benign Tumour															
Malignant Tumour															
MR/DD	X														
THYROID															
Benign (adenoma, goitre)		X				X	X	X						X	
Malignant					X										
BREASTS															
Benign							X	X					X	X	
Malignant Tumour		X				X			X				X	X	
SKIN/MUCOSA															
Trichilemmona		X													
MCL, FP, AK, PPK, F		X					X						X	X	
OP		X													
SP			X												
Lipomas															
Malignant Tumour															
GASTROINTESTINAL TRACT															
GI polyps															
Benign (hamartoma)			X				X								
Malignant Tumour															
OTHER BENIGN															
OTHER CANCERS															
SI SCORE	1	4	4	2	3	1	4	4	4	5	3	2	3	4	
REFERENCES	Buxbaum et al., 2007	Sawada et al., 2000	Lachlan et al., 2007	Bubien et al., 2013	Bubien et al., 2013	Lynch et al., 1997	Lynch et al., 1997	Lynch et al., 1997	Lynch et al., 1997	Lynch et al., 1997	Lynch et al., 1997	Lynch et al., 1997	Lynch et al., 1997	Ngeow et al., 2014	Buschet et al., 2013

NE

Abbreviations

AK	Acral Keratosis
CNS	Central Nervous System
CRC	Colorectal Cancer
DGCC	Dysplastic Gangliocytoma of the Cerebellum
ENCA	Endometrial Cancer
F	Fibromas
FBD	Fibrocystic disease
FP	Facial papules
GgNa	Ganglioneuroma
GI	Gastrointestinal
GU les	Genitourinary lesions
H	Hemangioma
H reflux	Hernia reflux
HN	Hydronephrosis
JPC	Juvenile Polyposis Coli
L	Lymphoma
LDD	Lhermitte-Duclos disease
LGTC	Lower Genital Tract Cancer
LH	Lymphoid Hyperplasia
LM	Leiomyoma
LPH	Lipomatous Hemangioma
MCL	Mucocutaneous Lesions
MnG	Meningioma
MR/DD	Mental Retardation/Developmental Delay
MT	Multiple Tumours
NE	Neuroma
NP	Nephrolithiasis
OC	Ovaric Cysts
OD	Oligodendrioma
OP	Oral mucosal Papillomatosis
OVCA	Ovarian cancer
PGT	Papillary Carcinoma
PLS	Proteus-Like Syndrome
PPK	Palmoplantar Keratoses
RC	Renal Carcinoma
RH	Renal Hamartoma
SP	Speckled Penis
UBC	Urinary Bladder Cancer
UF	Uterin Fibroids
UNPUB	Unpublished
UTLM	Uterine Leiomyoma
VASC les	Vascular lesions

Supporting Materials and Methods

Expression vectors and recombinant PTEN protein expression

Plasmid and lentiviral expression vectors for untagged and glutathione S-transferase (GST)-tagged human PTEN have been previously described [58 59]. The mutant expression vectors derived from pGEX6P1 PTEN and pHR-SIN PTEN were prepared by site-directed mutagenesis of the corresponding PTEN WT constructs with primers containing missense mutations. Recombinant wild-type and mutant forms of PTEN were expressed in *Escherichia coli* and purified by glutathione-affinity chromatography.

Immunoblotting

Protein gel electrophoresis was conducted with 10 µg of total soluble protein per sample using NuPage Bis-Tris 4-12% gradient polyacrylamide gels (Invitrogen Life Technologies) following manufacturers protocols. Proteins were transferred onto PVDF membrane (Perkin-Elmer) and membranes blocked in 5% milk powder/TBST for 1 hour at room temperature (RT). Blocked membranes were incubated overnight with primary antibodies. Antibodies used were: anti-P(Ser473)Akt and anti-P(Thr308)Akt (Cat. n.9271L and 9275L, Cell Signaling Technology), anti-PTEN (A2B1 Cat. n. sc-7974, Santa Cruz Biotechnology and PTEN 138G6, Cat. n. 9559S, Cell Signaling Technology), anti-GAPDH (Cat. n. AB2302, Millipore), anti-Akt (Cat. n. 9272, Cell Signaling Technology), anti-P(Thr246)PRAS40 (Cat. n. 29975, Cell Signaling Technology), anti-PRAS40 (Cat. n. 2691S, Cell Signaling Technology). Antibody complexes were detected by 1 hour incubation at RT with HRP conjugated secondary antibodies (Vector Labs). Blots were developed with ECL plus (Millipore)

and chemiluminescence imaged directly using an ImageQuant LAS4000 imaging system.

Lipid phosphatase assay

The preparation of 3-³³P labelled phosphoinositide substrate has been described previously ([58 59]. To test the lipid phosphatase activity of PTEN against PtdIns(3,4,5)P₃, lipid vesicles were prepared by sonication containing 100 μM phosphatidylcholine (PC), 1 μM unlabelled diC16 PtdIns(3,4,5)P₃ and a volume of ³³P-PtdIns(3,4,5)P₃ to give 100,000 cpm per assay. PTEN was immunoprecipitated using anti-PTEN A2B1 antibody in lysis buffer (50 mM Tris-HCL pH 7.4, 150 mM NaCl, 1 mM EDTA, 1 mM EGTA, 5 mM Sodium pyrophosphate, 10 mM β-glycerophosphate, 50 mM Sodium fluoride, 1% NP-40 and protease inhibitors) and washed twice with high salt (300 mM NaCl) lysis buffer and then with reaction buffer (50 mM Hepes pH 7.4, 1 mM EGTA, 10 mM dithiothreitol (DTT) and 150 mM NaCl). Substrate vesicles were incubated in the reaction buffer with PTEN immune complexes or with 500 ng of recombinant PTEN protein. After 1 hour of incubation at 37°C, the reactions were stopped through the addition of 10 μl of Bovine Serum Albumin (essentially fatty acid free- Roche) (10 mg/ml) and 500 μl of 1M ice cold perchloric acid (PCA). The samples were mixed and incubated for 30 minutes on ice, followed by centrifugation at 14,000 rpm for 10 minutes at 4°C to remove lipid and protein. 10% (w/v) ammonium molybdate was added to the supernatant to allow the inorganic phosphate to partition into the organic phase. After incubation of 10 minutes at room temperature, 1 ml Toluene:Isobutyl alcohol (1:1 v/v) was added. A two-phase mix was formed, and the upper organic phase, containing the phosphate complex, was then removed, mixed to liquid scintillation cocktail (Scint Safe 3, Fisher Chemical) and radioactivity was counted using a Beckman scintillation counter.

Reverse Transcriptase quantitative PCR (qPCR)

Total cellular RNA was isolated from U87MG cells expressing PTEN WT and mutants by using TRIzol (Life Technologies - Invitrogen) and RNeasy Mini kit (Qiagen), reversed transcribed into random primed cDNA (RNA to cDNA EcoDry Premix – Random Hexamers Clontech) and used as a template in quantitative PCR reactions to measure PTEN mRNA levels. GAPDH was used as internal control and Power Sybr green as the fluorescent reporter (Applied Biosystems). All reactions were performed according to the manufacturer's instructions on a Step One Plus real time PCR instrument (Life Technologies).

Primers were designed to be able to measure PTEN mRNA levels only and not its pseudogene. PTEN Primers: F: 5'-CCCGGGGGGGATCCACTAG; R: 5'-GGATAAATATAGGTCAAG. GAPDH Primers: F: 5'-GTGAAGGTCGGAGTCAACGG; R: 5'-GAGGGATCTCGCTCCTGGAA.

Immunocytochemistry

Neuronal cultures treated as described above were fixed using 4% paraformaldehyde for 30 minutes and then blocked with 3% BSA and 0.1% Triton x100 for 1 hour. Fixed neurons were incubated with anti-GFP antibody (1:1,000, Abcam), anti-PTEN antibody (1:500, PTEN138G6 Cell Signaling Technology) or anti-RFP (1:1,000, Cat. n. ab62341 Abcam) antibody diluted in 3% BSA, 1% goat serum and 1% sodium azide followed by incubation with Alexa488, Alexa568 and Alexa647 conjugated secondary antibodies (1:1,000, Invitrogen). Cells were stained with DAPI (1 µg/ml) for 10 minutes prior to mounting. The soma of the neurons, density and length of the spines were measured using ImageJ and were represented in µm.

Animal procedures

All animal procedures (breeding and sacrifice) were conducted in accordance with local ethical guidelines and approved animal care protocols (Berlin: Institutional Animal Care and Use Committee (IACUC) and the Landesamt für Gesundheit und Soziales (LAGeSO) – license T 0347/11).

Supplemental References

1. Hobert JA, Embacher R, Mester JL, et al. Biochemical screening and PTEN mutation analysis in individuals with autism spectrum disorders and macrocephaly. *European journal of human genetics : EJHG* 2014;**22**(2):273-6 doi: 10.1038/ejhg.2013.114[published Online First: Epub Date]].
2. Vanderver A, Tonduti D, Kahn I, et al. Characteristic brain magnetic resonance imaging pattern in patients with macrocephaly and PTEN mutations. *American journal of medical genetics Part A* 2014;**164A**(3):627-33 doi: 10.1002/ajmg.a.36309[published Online First: Epub Date]].
3. Bubien V, Bonnet F, Brouste V, et al. High cumulative risks of cancer in patients with PTEN hamartoma tumour syndrome. *J Med Genet* 2013;**50**(4):255-63 doi: 10.1136/jmedgenet-2012-101339[published Online First: Epub Date]].
4. Frazier TW, Embacher R, Tilot AK, et al. Molecular and phenotypic abnormalities in individuals with germline heterozygous PTEN mutations and autism. *Molecular psychiatry* 2014 doi: 10.1038/mp.2014.125[published Online First: Epub Date]].
5. Nagy R, Ganapathi S, Comeras I, et al. Frequency of germline PTEN mutations in differentiated thyroid cancer. *Thyroid* 2011;**21**(5):505-10 doi: 10.1089/thy.2010.0365[published Online First: Epub Date]].
6. Ngeow J, Stanuch K, Mester JL, et al. Second malignant neoplasms in patients with Cowden syndrome with underlying germline PTEN mutations. *Journal of clinical oncology : official journal of the American Society of Clinical Oncology* 2014;**32**(17):1818-24 doi: 10.1200/JCO.2013.53.6656[published Online First: Epub Date]].
7. Buxbaum JD, Cai G, Chaste P, et al. Mutation screening of the PTEN gene in patients with autism spectrum disorders and macrocephaly. *Am J Med Genet B Neuropsychiatr Genet* 2007;**144B**(4):484-91 doi: 10.1002/ajmg.b.30493[published Online First: Epub Date]].
8. Celebi JT, Tsou HC, Chen FF, et al. Phenotypic findings of Cowden syndrome and Bannayan-Zonana syndrome in a family associated with a single germline mutation in PTEN. *J Med Genet* 1999;**36**(5):360-4
9. Mester J, Eng C. Estimate of de novo mutation frequency in probands with PTEN hamartoma tumor syndrome. *Genetics in medicine : official journal of the American College of Medical Genetics* 2012;**14**(9):819-22 doi: 10.1038/gim.2012.51[published Online First: Epub Date]].
10. Marsh DJ, Kum JB, Lunetta KL, et al. PTEN mutation spectrum and genotype-phenotype correlations in Bannayan-Riley-Ruvalcaba syndrome suggest a single entity with Cowden syndrome. *Human molecular genetics* 1999;**8**(8):1461-72
11. Olschwang S, Serova-Sinilnikova OM, Lenoir GM, et al. PTEN germ-line mutations in juvenile polyposis coli. *Nat Genet* 1998;**18**(1):12-4 doi: 10.1038/ng0198-12[published Online First: Epub Date]].
12. Zhou X, Hampel H, Thiele H, et al. Association of germline mutation in the PTEN tumour suppressor gene and Proteus and Proteus-like syndromes. *Lancet* 2001;**358**(9277):210-1
13. Celebi JT, Ping XL, Zhang H, et al. Germline PTEN mutations in three families with Cowden syndrome. *Exp Dermatol* 2000;**9**(2):152-6
14. Klein S, Sharifi-Hannauer P, Martinez-Agosto JA. Macrocephaly as a clinical indicator of genetic subtypes in autism. *Autism research : official journal of the International Society for Autism Research* 2013;**6**(1):51-6 doi: 10.1002/aur.1266[published Online First: Epub Date]].
15. Tate G, Suzuki T, Endo Y, et al. A novel mutation of the PTEN gene in a Japanese patient with Cowden syndrome and bilateral breast cancer. *Cancer Genet Cytogenet* 2008;**184**(1):67-71 doi: 10.1016/j.cancergencyto.2008.03.013[published Online First: Epub Date]].
16. Varga EA, Pastore M, Prior T, et al. The prevalence of PTEN mutations in a clinical pediatric cohort with autism spectrum disorders, developmental delay, and macrocephaly. *Genetics in*

- medicine : official journal of the American College of Medical Genetics 2009;**11**(2):111-7 doi: 10.1097/GIM.0b013e31818fd762[published Online First: Epub Date]].
17. Vega A, Torres J, Torres M, et al. A novel loss-of-function mutation (N48K) in the PTEN gene in a Spanish patient with Cowden disease. *J Invest Dermatol* 2003;**121**(6):1356-9 doi: 10.1111/j.1523-1747.2003.12638.x[published Online First: Epub Date]].
 18. Reardon W, Zhou XP, Eng C. A novel germline mutation of the PTEN gene in a patient with macrocephaly, ventricular dilatation, and features of VATER association. *J Med Genet* 2001;**38**(12):820-3
 19. Loffeld A, McLellan NJ, Cole T, et al. Epidermal naevus in Proteus syndrome showing loss of heterozygosity for an inherited PTEN mutation. *Br J Dermatol* 2006;**154**(6):1194-8 doi: 10.1111/j.1365-2133.2006.07196.x[published Online First: Epub Date]].
 20. Marsh DJ, Dahia PL, Caron S, et al. Germline PTEN mutations in Cowden syndrome-like families. *J Med Genet* 1998;**35**(11):881-5
 21. Lobo GP, Waite KA, Planchon SM, et al. Germline and somatic cancer-associated mutations in the ATP-binding motifs of PTEN influence its subcellular localization and tumor suppressive function. *Human molecular genetics* 2009;**18**(15):2851-62 doi: 10.1093/hmg/ddp220[published Online First: Epub Date]].
 22. Schaaf CP, Sabo A, Sakai Y, et al. Oligogenic heterozygosity in individuals with high-functioning autism spectrum disorders. *Human molecular genetics* 2011;**20**(17):3366-75 doi: 10.1093/hmg/ddr243[published Online First: Epub Date]].
 23. Figer A, Kaplan A, Frydman M, et al. Germline mutations in the PTEN gene in Israeli patients with Bannayan-Riley-Ruvalcaba syndrome and women with familial breast cancer. *Clinical genetics* 2002;**62**(4):298-302
 24. Kohno T, Takahashi M, Fukutomi T, et al. Germline mutations of the PTEN/MMAC1 gene in Japanese patients with Cowden disease. *Jpn J Cancer Res* 1998;**89**(5):471-4
 25. Butler MG, Dasouki MJ, Zhou XP, et al. Subset of individuals with autism spectrum disorders and extreme macrocephaly associated with germline PTEN tumour suppressor gene mutations. *J Med Genet* 2005;**42**(4):318-21 doi: 10.1136/jmg.2004.024646[published Online First: Epub Date]].
 26. Bussaglia E, Pujol RM, Gil MJ, et al. PTEN mutations in eight Spanish families and one Brazilian family with Cowden syndrome. *J Invest Dermatol* 2002;**118**(4):639-44 doi: 10.1046/j.1523-1747.2002.01728.x[published Online First: Epub Date]].
 27. Busch RM, Chapin JS, Mester J, et al. Cognitive characteristics of PTEN hamartoma tumor syndromes. *Genetics in medicine : official journal of the American College of Medical Genetics* 2013;**15**(7):548-53 doi: 10.1038/gim.2013.1[published Online First: Epub Date]].
 28. Tan WH, Baris HN, Burrows PE, et al. The spectrum of vascular anomalies in patients with PTEN mutations: implications for diagnosis and management. *J Med Genet* 2007;**44**(9):594-602 doi: 10.1136/jmg.2007.048934[published Online First: Epub Date]].
 29. Ngeow J, Heald B, Rybicki LA, et al. Prevalence of germline PTEN, BMPR1A, SMAD4, STK11, and ENG mutations in patients with moderate-load colorectal polyps. *Gastroenterology* 2013;**144**(7):1402-9, 09 e1-5 doi: 10.1053/j.gastro.2013.02.001[published Online First: Epub Date]].
 30. Tsou HC, Ping XL, Xie XX, et al. The genetic basis of Cowden's syndrome: three novel mutations in PTEN/MMAC1/TEP1. *Hum Genet* 1998;**102**(4):467-73
 31. Orrico A, Galli L, Buoni S, et al. Novel PTEN mutations in neurodevelopmental disorders and macrocephaly. *Clinical genetics* 2009;**75**(2):195-8 doi: 10.1111/j.1399-0004.2008.01074.x[published Online First: Epub Date]].
 32. De Vivo I, Gertig DM, Nagase S, et al. Novel germline mutations in the PTEN tumour suppressor gene found in women with multiple cancers. *J Med Genet* 2000;**37**(5):336-41
 33. McBride KL, Varga EA, Pastore MT, et al. Confirmation study of PTEN mutations among individuals with autism or developmental delays/mental retardation and macrocephaly.

Autism research : official journal of the International Society for Autism Research
2010;**3**(3):137-41 doi: 10.1002/aur.132[published Online First: Epub Date]].

34. Kersseboom R, Dubbink HJ, Corver WE, et al. PTEN in colorectal cancer: a report on two Cowden syndrome patients. *Clinical genetics* 2012;**81**(6):555-62 doi: 10.1111/j.1399-0004.2011.01639.x[published Online First: Epub Date]].
35. Nelen MR, van Staveren WC, Peeters EA, et al. Germline mutations in the PTEN/MMAC1 gene in patients with Cowden disease. *Human molecular genetics* 1997;**6**(8):1383-7
36. Elia M, Amato C, Bottitta M, et al. An atypical patient with Cowden syndrome and PTEN gene mutation presenting with cortical malformation and focal epilepsy. *Brain Dev* 2012;**34**(10):873-6 doi: 10.1016/j.braindev.2012.03.005[published Online First: Epub Date]].
37. Liaw D, Marsh DJ, Li J, et al. Germline mutations of the PTEN gene in Cowden disease, an inherited breast and thyroid cancer syndrome. *Nature Genetics* 1997;**16**(1):64-67
38. Kurose K, Araki T, Matsunaka T, et al. Variant manifestation of Cowden disease in Japan: hamartomatous polyposis of the digestive tract with mutation of the PTEN gene. *Am J Hum Genet* 1999;**64**(1):308-10 doi: 10.1086/302207[published Online First: Epub Date]].
39. Heindl M, Handel N, Ngeow J, et al. Autoimmunity, intestinal lymphoid hyperplasia, and defects in mucosal B-cell homeostasis in patients with PTEN hamartoma tumor syndrome. *Gastroenterology* 2012;**142**(5):1093-96 e6 doi: 10.1053/j.gastro.2012.01.011[published Online First: Epub Date]].
40. O'Roak BJ, Vives L, Fu W, et al. Multiplex targeted sequencing identifies recurrently mutated genes in autism spectrum disorders. *Science* 2012;**338**(6114):1619-22 doi: 10.1126/science.1227764[published Online First: Epub Date]].
41. Derrey S, Proust F, Debono B, et al. Association between Cowden syndrome and Lhermitte-Duclos disease: report of two cases and review of the literature. *Surg Neurol* 2004;**61**(5):447-54; discussion 54 doi: 10.1016/S0090-3019(03)00576-7[published Online First: Epub Date]].
42. Tekin M, Hisimi BO, Fitoz S, et al. A germline PTEN mutation with manifestations of prenatal onset and verrucous epidermal nevus. *American journal of medical genetics Part A* 2006;**140**(13):1472-5 doi: 10.1002/ajmg.a.31273[published Online First: Epub Date]].
43. Busa T, Chabrol B, Perret O, et al. Novel PTEN germline mutation in a family with mild phenotype: difficulties in genetic counseling. *Gene* 2013;**512**(2):194-7 doi: 10.1016/j.gene.2012.09.134[published Online First: Epub Date]].
44. Boccone L, Dessi V, Zappu A, et al. Bannayan-Riley-Ruvalcaba syndrome with reactive nodular lymphoid hyperplasia and autism and a PTEN mutation. *American journal of medical genetics Part A* 2006;**140**(18):1965-9 doi: 10.1002/ajmg.a.31396[published Online First: Epub Date]].
45. Kubo Y, Urano Y, Hida Y, et al. A novel PTEN mutation in a Japanese patient with Cowden disease. *Br J Dermatol* 2000;**142**(6):1100-5
46. Gicquel JJ, Vabres P, Bonneau D, et al. Retinal angioma in a patient with Cowden disease. *Am J Ophthalmol* 2003;**135**(3):400-2
47. Nelen MR, Kremer H, Konings IB, et al. Novel PTEN mutations in patients with Cowden disease: absence of clear genotype-phenotype correlations. *European journal of human genetics : EJHG* 1999;**7**(3):267-73 doi: 10.1038/sj.ejhg.5200289[published Online First: Epub Date]].
48. Banneau G, Guedj M, MacGrogan G, et al. Molecular apocrine differentiation is a common feature of breast cancer in patients with germline PTEN mutations. *Breast cancer research : BCR* 2010;**12**(4):R63 doi: 10.1186/bcr2626[published Online First: Epub Date]].
49. Marsh DJ, Dahia PL, Zheng Z, et al. Germline mutations in PTEN are present in Bannayan-Zonana syndrome [letter]. *Nat Genet* 1997;**16**(4):333-4
50. Lachlan KL, Lucassen AM, Bunyan D, et al. Cowden syndrome and Bannayan Riley Ruvalcaba syndrome represent one condition with variable expression and age-related penetrance: results of a clinical study of PTEN mutation carriers. *J Med Genet* 2007;**44**(9):579-85 doi: 10.1136/jmg.2007.049981[published Online First: Epub Date]].

51. Kirches E, Steiner J, Schneider T, et al. Lhermitte-Duclos disease caused by a novel germline PTEN mutation R173P in a patient presenting with psychosis. *Neuropathol Appl Neurobiol* 2010;**36**(1):86-9 doi: 10.1111/j.1365-2990.2009.01041.x[published Online First: Epub Date]].
52. Thiffault I, Schwartz CE, Der Kaloustian V, et al. Mutation analysis of the tumor suppressor PTEN and the glypican 3 (GPC3) gene in patients diagnosed with Proteus syndrome. *American journal of medical genetics Part A* 2004;**130A**(2):123-7 doi: 10.1002/ajmg.a.30335[published Online First: Epub Date]].
53. Kim DK, Myung SJ, Yang SK, et al. Analysis of PTEN gene mutations in Korean patients with Cowden syndrome and polyposis syndrome. *Dis Colon Rectum* 2005;**48**(9):1714-22 doi: 10.1007/s10350-005-0130-9[published Online First: Epub Date]].
54. Staal FJ, van der Luijt RB, Baert MR, et al. A novel germline mutation of PTEN associated with brain tumours of multiple lineages. *Br J Cancer* 2002;**86**(10):1586-91 doi: 10.1038/sj.bjc.6600206[published Online First: Epub Date]].
55. Chi SG, Kim HJ, Park BJ, et al. Mutational abrogation of the PTEN/MMAC1 gene in gastrointestinal polyps in patients with Cowden disease. *Gastroenterology* 1998;**115**(5):1084-9
56. Sawada T, Hamano N, Satoh H, et al. Mutation analysis of the PTEN / MMAC1 gene in Japanese patients with Cowden disease. *Jpn J Cancer Res* 2000;**91**(7):700-5
57. Lynch ED, Ostermeyer EA, Lee MK, et al. Inherited mutations in PTEN that are associated with breast cancer, cowden disease, and juvenile polyposis. *Am J Hum Genet* 1997;**61**(6):1254-60 doi: 10.1086/301639[published Online First: Epub Date]].
58. Davidson L, Maccario H, Perera NM, et al. Suppression of cellular proliferation and invasion by the concerted lipid and protein phosphatase activities of PTEN. *Oncogene* 2010;**29**(5):687-97 doi: onc2009384 [pii] 10.1038/onc.2009.384[published Online First: Epub Date]].
59. McConnachie G, Pass I, Walker SM, et al. Interfacial kinetic analysis of the tumour suppressor phosphatase, PTEN: evidence for activation by anionic phospholipids. *Biochem J* 2003;**371**(Pt 3):947-55