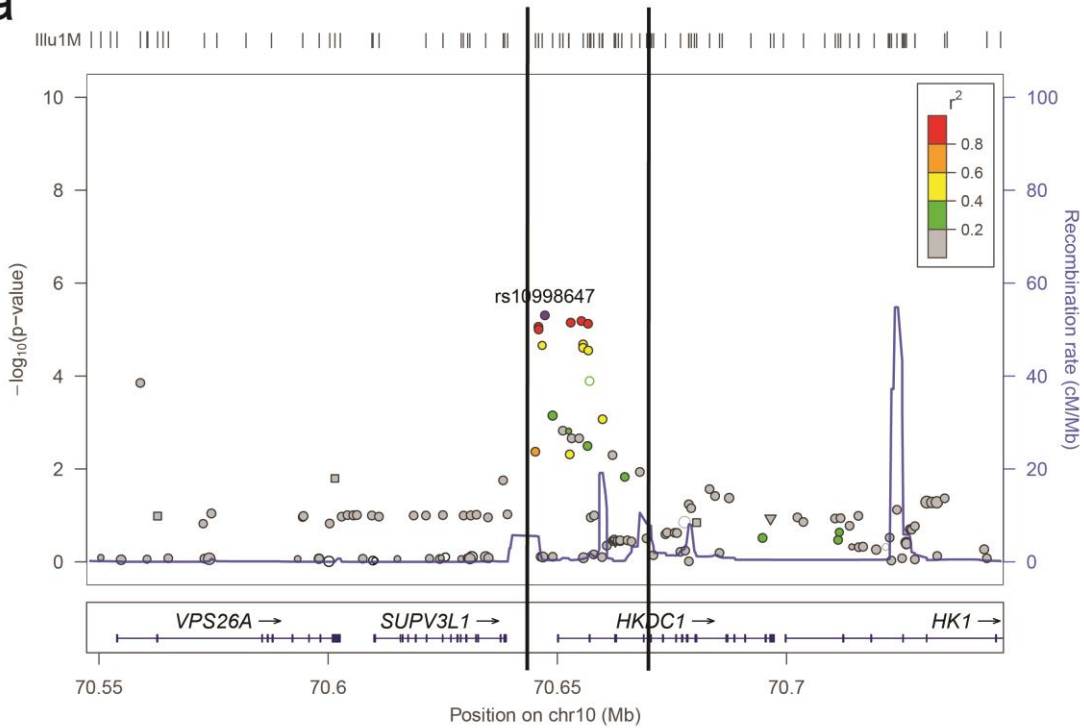
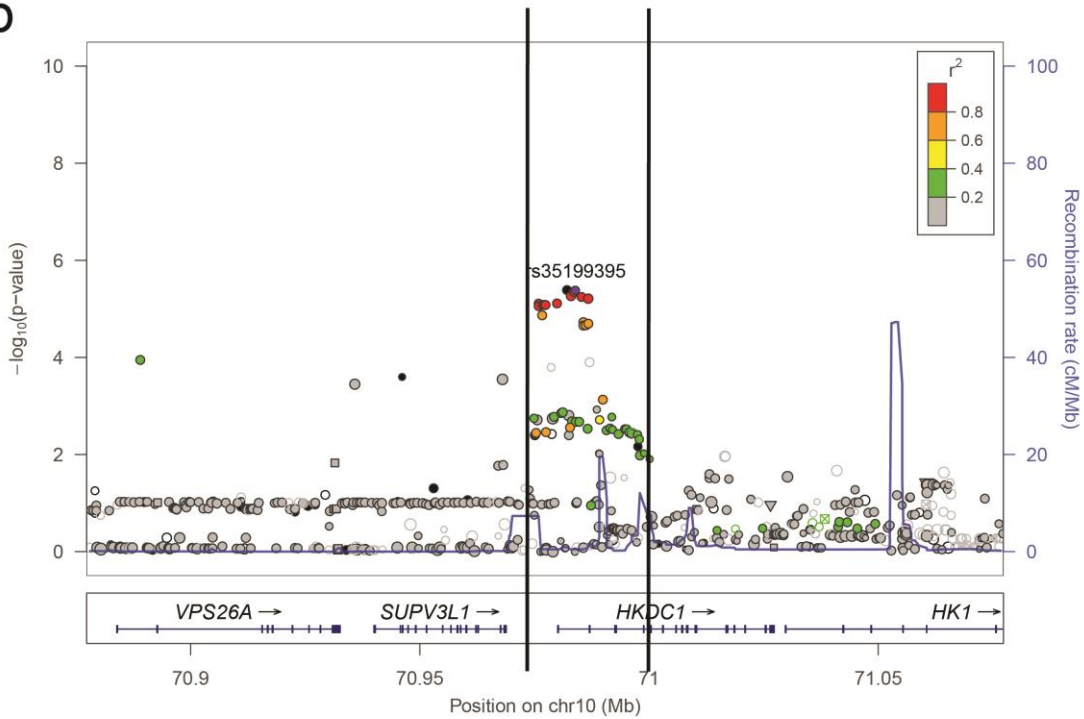


Supplementary Figures

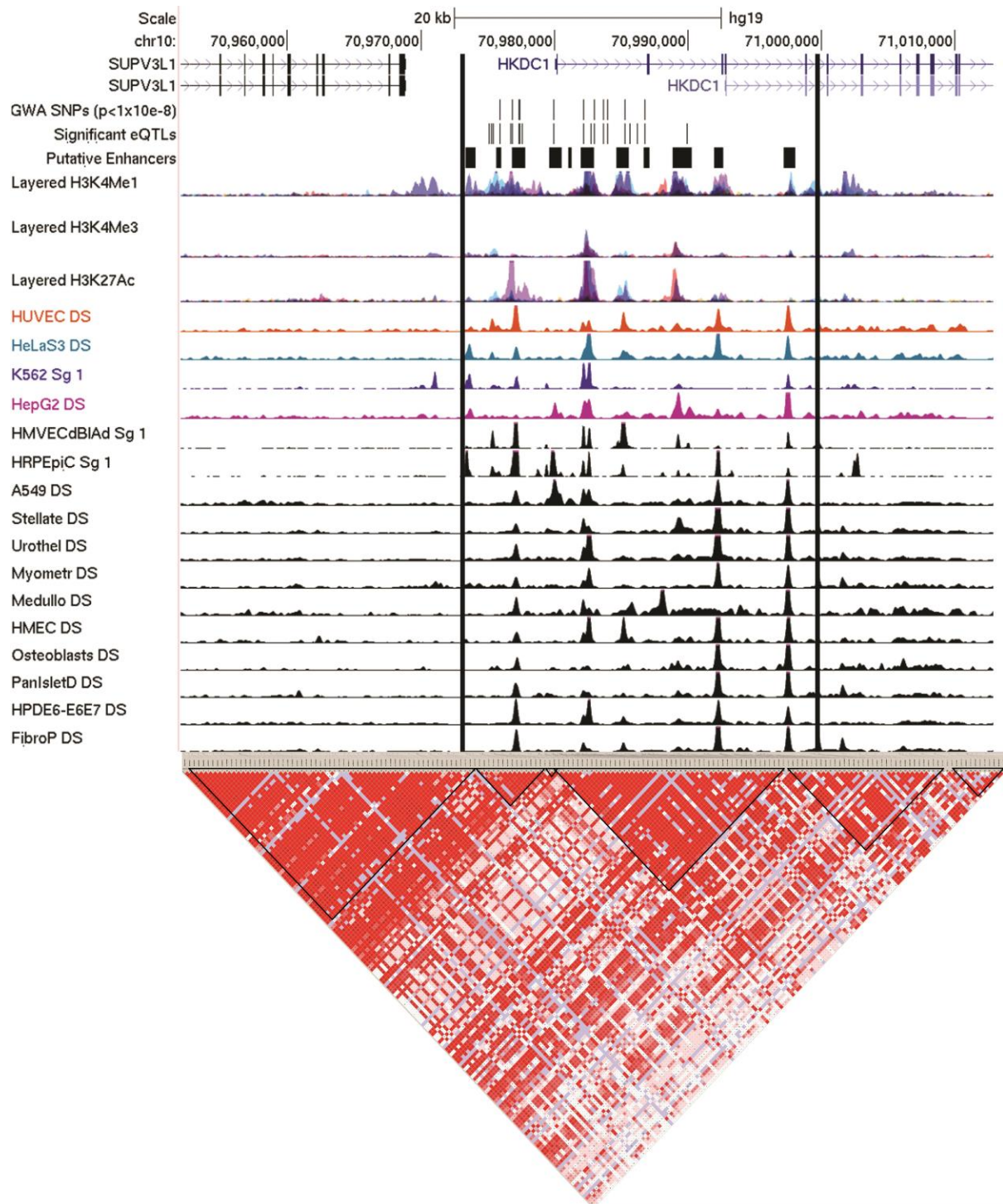
a



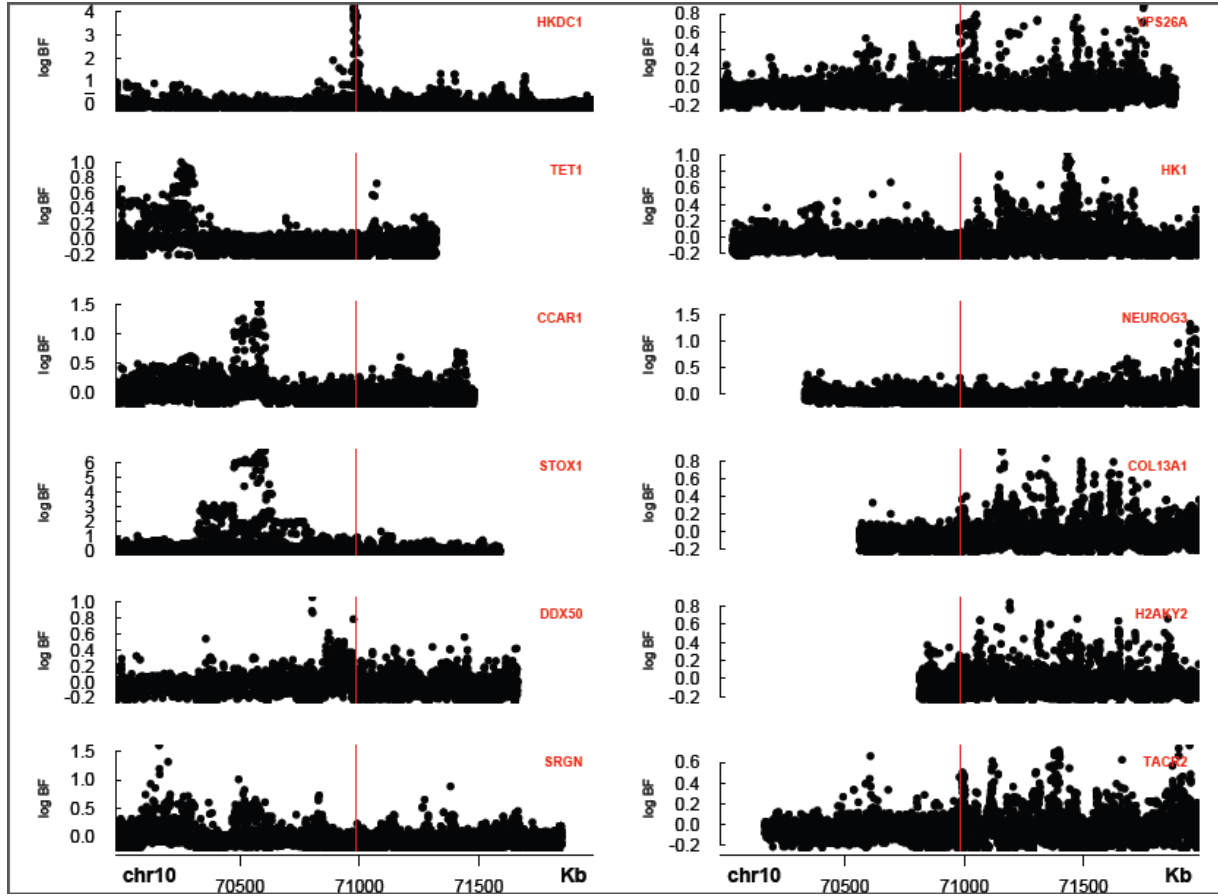
b



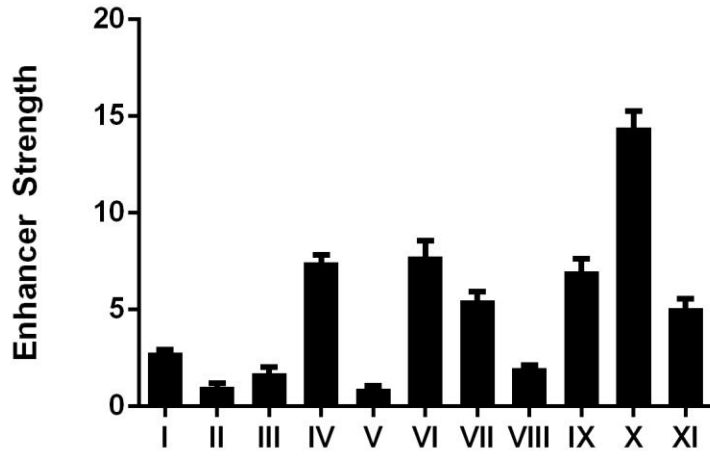
Supplementary Figure 1: Loci associated with 2 hr glucose during pregnancy imputed to HAPMAP **(a)** and 1000 Genomes **(b)**. Peak of association is in the first intron of HKDC1. Black bars delineate a 30kb block with the strongest association (F-test, $p < 1 \times 10^{-5}$, $n = 1,367$) and LD ($r^2 > 0.3$).



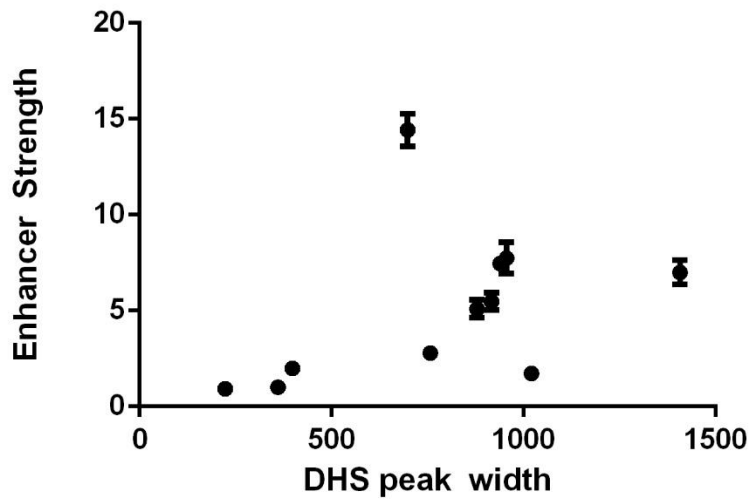
Supplementary Figure 2: Significant HKDC1 eQTLs [$\log_{10}(\text{Bayes Factor}) > 2.5$] and GWA SNPs (F-test, $p < 1 \times 10^{-8}$) in the maternal 2 hr glucose level associated locus. An enrichment of open chromatin is located in a 30kb block between the 1st and 5th exons of HKDC1, flanked by two regulatory “deserts” on either side. Open chromatin for each cell line is denoted by DNaseI hypersensitivity (ENCODE). Putative enhancer marks are marked by H3KMe1 and H3K27AC. Black bars delineate the same 30 kb block in Supplementary Figure 1.



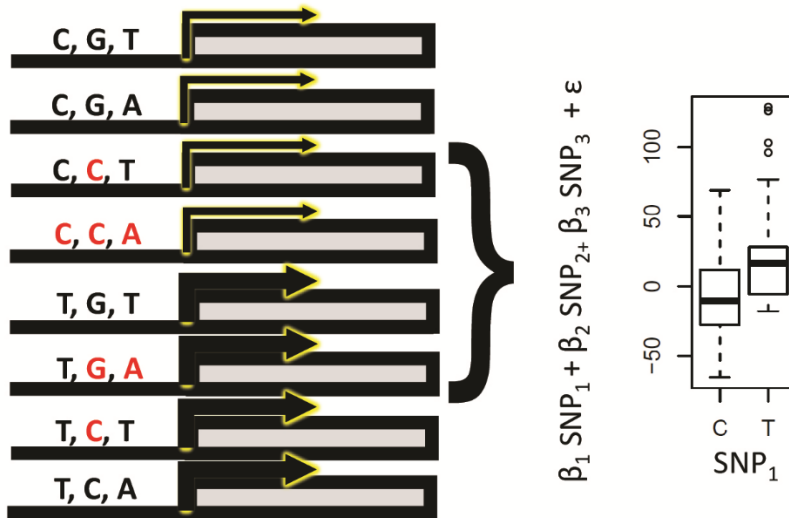
Supplementary Figure 3: eQTL analysis in primary liver for all genes within 500 kb of HKDC1. The red line represents the lead GWAS SNP rs4746822.



Supplementary Figure 4: Enhancer strength of selected regulatory elements. Firefly luciferase intensity was normalized by dividing by the *Renilla* luciferase intensity. Enhancer strength was determined by dividing the normalized luciferase intensity of each construct by the normalized intensity of the empty vector. Error bars show s.d (n = 8 to 19).

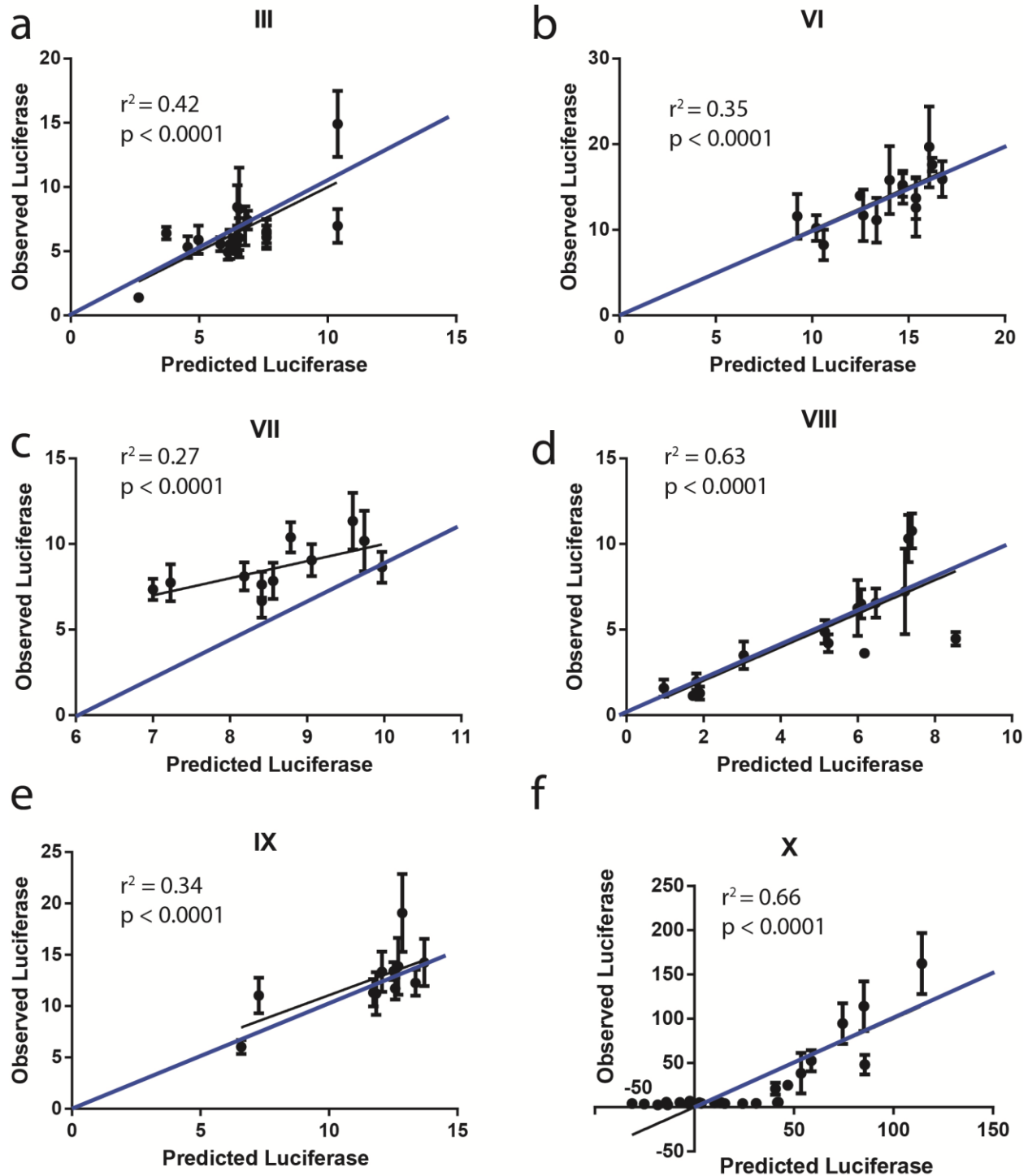


Supplementary Figure 5: Enhancer strength vs DHS peak width. Spearman $\rho = 0.5$, $p = 0.117$ ($n = 8$ to 19). Error bars show s.d.

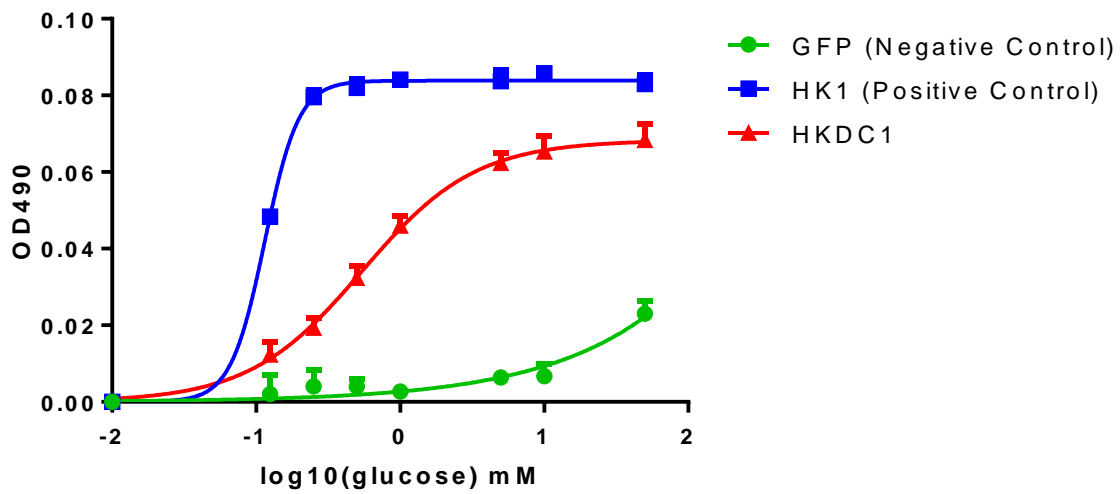


SNP: 1, 2, 3

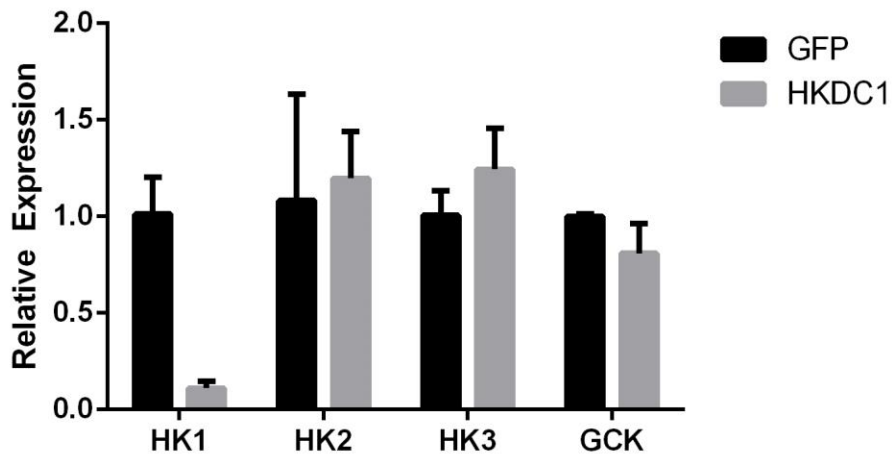
Supplementary Figure 6: Experimental design for luciferase reporter assays. Multiple haplotypes of each regulatory element were cloned upstream of the promoter and site directed mutagenesis was used to segregate alleles which do not segregate naturally in the population. A linear regression model was used to determine the effect of each SNP on luciferase expression. The bottom and top boxes are the first and third quartiles, and the band inside the box is the median. The ends of the whiskers represent the lowest and highest data points within 1.5 interquartile range of the lower and upper quartiles. Circles represent outliers defined as 1.5 times the interquartile range above the upper quartile or below the lower quartile.



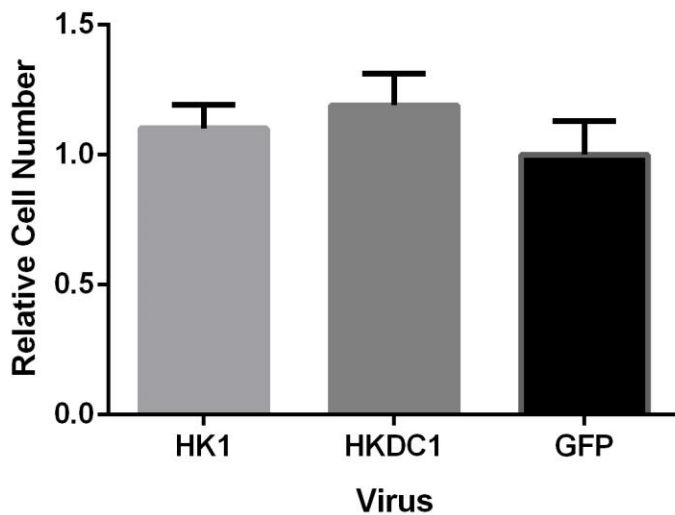
Supplementary Figure 7: Plots of predicted luciferase values versus observed luciferase values. In **a-f**, predicted luciferase values are positively and significantly associated with observed luciferase values, suggesting that the effects of individual variants may combine additively within haplotypes. The blue line is $y = x$ and the black line is the regression line. Error bars show s.d.



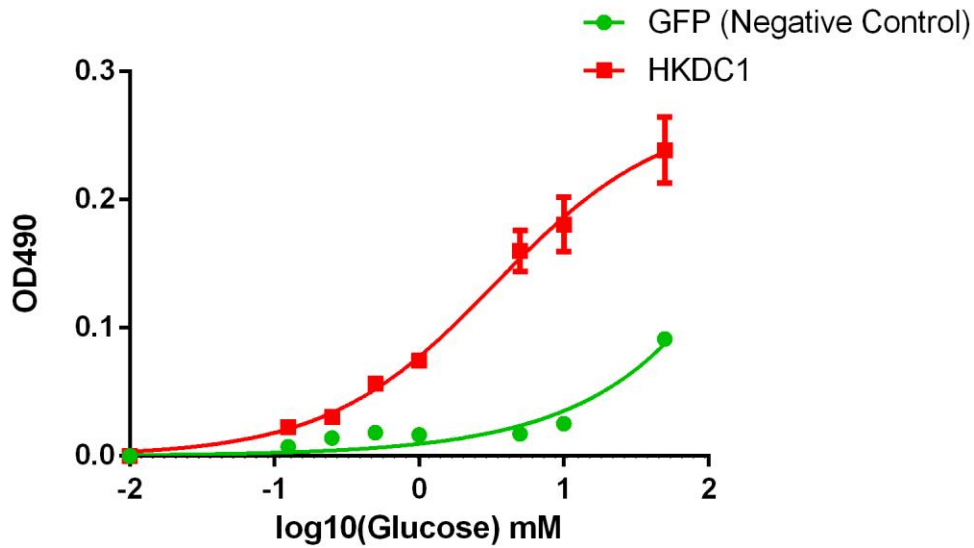
Supplementary Figure 9: Un-normalized HKDC1 virus transduction results. HKDC1 overexpression in INS-1 cells via adenovirus shows increased HK activity between a range of 0-50mM glucose. Error bars show s.d.



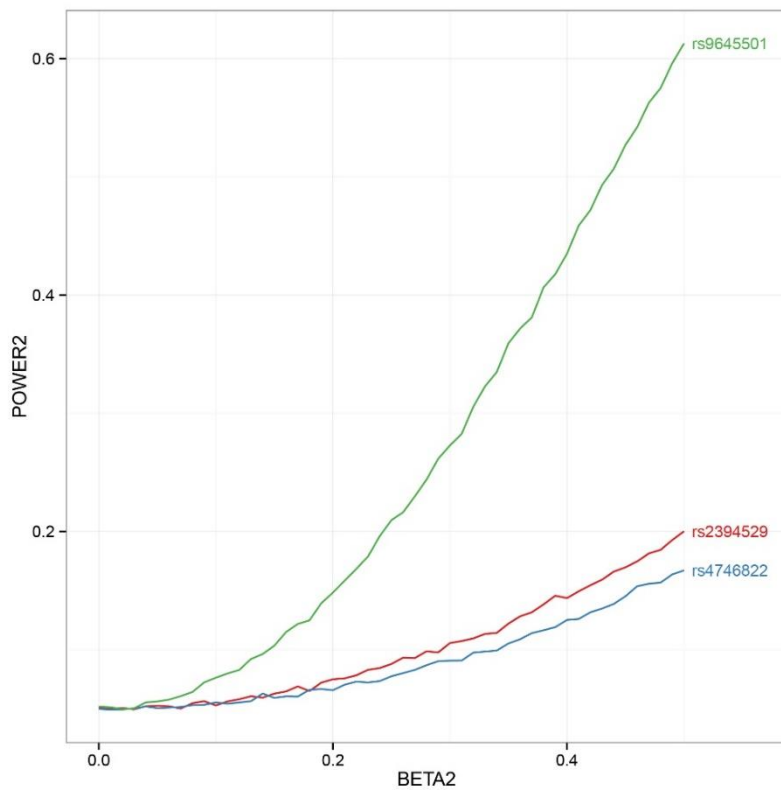
Supplementary Figure 10: Expression of other HK mRNAs in INS-1 cells after either GFP or HKDC1 adenovirus transduction. Expression was quantified via qPCR and expression was normalized using the $\Delta\Delta$ CT method (n = 3). Error bars show s.d.



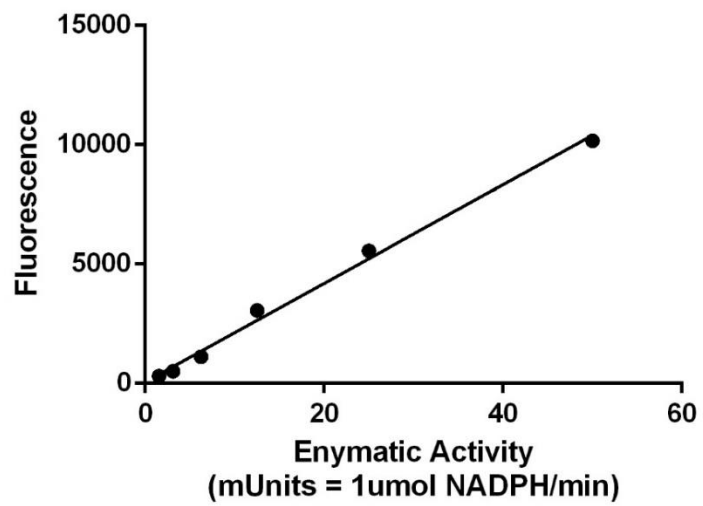
Supplementary Figure 11: Cell number is consistent between groups of INS-1 cells treated with adenovirus. Cell number was quantified using the Cell-Titer Glo assay (Promega) (n = 3). Error bars show s.d.



Supplementary Figure 12: Replicate of HKDC1 activity dose response curve. This experiment is a duplicate experiment represented in Supplementary Figure 4 (n = 3). Error bars show s.d.



Supplementary Figure 13: Power analysis to detect independent additive effects of regulatory variants. The beta of SNP1 (rs10762264) = 0.48 and the MAF = 0.4. R^2 values between the 4 SNPs range from 0.8-0.99.



Supplementary Figure 14: Standard curve for specific activity versus fluorescence (n = 3).

Supplementary Tables

Supplementary Table 1. Putative regulatory element chromatin marks

Region	Coordinates	DHS Width (bp)	DNaseI HS	H3Kme1	H3K27Ac
I	chr10:70973319-70974076	757	HeLa, K562, HepG2, HRPEpiC	K562	K562
II	chr10:70975656-70976016	360	HUVEC, HeLa, k562, HMVEC, HRPE	HUVEC, K562, NHEK, NHLF	HUVEC, NHEK, K562
III	chr10:70976807-70977827	1020	HUVEC, HeLa, K562, HMVEC, HRPE, A549, Stellate, Urothel, Myometr, Medullo, HMEC, Osteoblasts, PanIsletD, HPDE6, FibroP	HUVEC, K562, NHEK, NHLF	HSMM, HUVEC, NHEK, NHLF
IV	chr10:70979612-70980551	939	HepG2, HRPE, A549, K562	K562	H1-hESC
V	chr10:70981067-70981290	223	HepG2, A549	K562, NHEK	GM12878, H1-hESC, NHEK
VI	chr10:70982000-70982955	955	HUVEC, HeLa, K562, HepG2, HMVEC, HRPE, A549, Stellate, Urothel, Myometr, Medullo, HMEC, HMEC, Osteoblast, PanIsletD, HPDE6, FibroP	GM12878, H1-hESC, HSMM, HUVEC, K562, NHEK, NHLF	GM12878, H1-hESC, HSMM, HUVEC, K562, NHEK
VII	chr10:70984649-70985565	916	HUVEC, HeLa, HepG2, HMVEC, HRPE, Medullo, HMEC, HPDE6, FibroP	GM12878, HSMM, HUVEC, K562, NHEK	GM12878, HUVEC, NHEK
VIII	chr10:70986715-70987113	398	HepG2, Medullo		
IX	chr10:70988887-70990294	1407	HUVEC, K562, HepG2, HMVEC, HRPE, A549, Stellate, Urothel, Medullo, HMEC, Osteoblast, PanIslet, FibroP	GM12878, H1-hESC, HSMM, HUVEC, K562, NHEK, NHLF	GM12878, K562
X	chr10:70991933-70992631	698	HUVEC, HeLa, HepG2, HMVEC, HRPE, A549, Stellate, Urothel, Myometr, Medullo, HMEC, HMEC, Osteoblast, PanIsletD, HPDE6, FibroP	H1-hESC, HSMM, HUVEC, NHEK, NHLF	HSMM, NHEK, NHLF
XI	chr10:70997194-70998073	879	HUVEC, HeLa, K562, HMVEC, HRPE, A549, Stellate, Urothel, Myometr, Medullo, HMEC, Osteoblasts, PanIsletD, HPDE6, FibroP	GM12878, H1-hESC, HSMM, HUVEC, K562, NHEK, NHLF	

Supplementary Table 2. Haplotype frequencies across the regulatory elements

Region	Coordinates	SNPs (common)	Common Haplotypes (>1%)	Rare Haplotypes (<1%)	Total Haplotypes
I	chr10:70973319- 70974076	9(4)	5	6	11
II	chr10: 70975656- 70976016	4(2)	2	3	5
III	chr10:70976807- 70977827	11(8)	5	6	11
IV	chr10:70979612- 70980551	13(2)	3	12	15
V	chr10:70981067- 70981290	2(1)	2	1	3
VI	chr10:70982000- 70982955	21(9)	8	19	27
VII	chr10:70984649- 70985565	15(4)	5	14	19
VIII	chr10:70986715- 70987113	7(4)	4	4	8
IX	chr10:70988887- 70990294	26(12)	13	55	68
X	chr10:70991933- 70992631	15(7)	6	9	15
XI	chr10:70997194- 70998073	13(7)	7	14	21
	Total	136(60)	60	143	203

Supplementary Table 3. 1000 Genomes individual IDs and ancestries

Catalog ID	Ancestry	Sex
NA19399	LUHYA IN WEBUYE, KENYA	Female
NA19428	LUHYA IN WEBUYE, KENYA	Male
NA18557	YORUBA IN IBADAN, NIGERIA	Male
NA19007	JAPANESE IN TOKYO, JAPAN	Male
HG01133	COLOMBIAN IN MEDELLIN, COLOMBIA	Male
HG00346	FINNISH IN FINLAND	Female
NA19473	LUHYA IN WEBUYE, KENYA	Female
NA18861	YORUBA IN IBADAN, NIGERIA	Female
NA19346	LUHYA IN WEBUYE, KENYA	Male
NA19457	LUHYA IN WEBUYE, KENYA	Female
HG01437	COLOMBIAN IN MEDELLIN, COLOMBIA	Male
NA19438	LUHYA IN WEBUYE, KENYA	Female
HG00189	FINNISH IN FINLAND	Male
NA19150	YORUBA IN IBADAN, NIGERIA	Male
HG01626	IBERIAN POPULATIONS IN SPAIN	Female
NA19445	LUHYA IN WEBUYE, KENYA	Female
HG01079	PUERTO RICAN IN PUERTO RICO	Male
GM19707	AFRICAN ANCESTRY IN SOUTHWEST USA	Female
GM12144	CEPH/UTAH	Male

Supplementary Table 4. Pre- and post-mutagenesis rare allele frequencies

SNPId	Pre-mutagenesis	Post-mutagenesis
rs7089312	0.125	0.2857
rs5030941	0.125	0.2857
rs2394529	0.1667	0.2727
rs141651118	0.2	0.3
rs147449838	0.2	0.4
rs55755107	0.1667	0.4286
rs4072135	0.1667	0.35714
rs11813186	0.1667	0.21429
rs5030945	0.125	0.38889
rs144643300	0.125	0.16667
rs12246517	0.25	0.5
rs4746829	0.25	0.5

Supplementary Table 5. Table of significant regulatory variants

Region	SNP	Allele A	Allele B	MAF	β Luciferase	P-Value
III	rs10762264	G	A	0.48	-3.13	5.05E-05
III	rs12241136	A	T	0.1	2.77	4.14E-08
VI	rs78983061	C	A	0.06	2.21	0.003
VI	rs7089277	T	G	0.07	-8.59	8.90E-11
VI	rs4746822	C	T	0.49	-6.7	8.00E-08
VII	rs4746824	C	A	0.22	1.18	-2.00E-06
VII	rs75405157	T	C	0.06	1.55	3.08E-08
VII	rs2394529	G	C	0.5	-1.41	1.06E-04
VIII	rs9645501	G	A	0.34	-2.08	9.04E-08
VIII	rs147449838	G	A	<.01	-1.23	4.35E-04
VIII	rs200216341	G	A	<.01	-1.31	1.71E-04
IX	rs1983128	G	A	<.01	-4.45	9.49E-09
X	rs5030945	T	C	0.45	-28.14	2.19E-10
X	rs874557	A	G	0.45	-31.58	1.54E-06

Supplementary Table 6. Concordant SNP effects in four different cell lines

Region	SNP	Allele A	Allele B	MAF	P-value	β Luc A549	β Luc K562	β Luc Fibroblast	β Luc HepG2
III	rs10762264	G	A	0.48	5.05E-05	-2.368	N.E.	-8.9581	-3.13
VI	rs4746822	C	T	0.49	8.00E-08	-2.724	N.E.	-3.86	-6.7
VII	rs2394529	G	C	0.5	1.06E-04	N.E.	N.E.	3.137	-1.41
VIII	rs9645501	G	A	0.34	9.04E-08	-29.078	-115.44	-15.29	-2.08

Supplementary Table 7. Discordant SNP effects in four different cell lines

Region	SNP	Allele A	Allele B	MAF	β Luc A549	β Luc K562	β Luc Fibroblast	β Luc HepG2
X	rs5030945	T	C	0.45	5.04	66.89	-3.13	-28.14
X	rs874557	A	G	0.45	28.37	62.46	-2.3	-31.58

Supplementary Table 8. RPKM for primary hepatocytes and HepG2 cells

	Hepatocytes (RPKM)	HepG2 (RPKM)
HK1	6.5	0.7
HK2	1.3	27.2
HK3	6.6	0
GCK	1	0.1
HKDC1	1.2	28.3

Supplementary Table 9. Amino acid percent identity matrix

	GCK	HK3	HKDC1	HK1	HK2
GCK	100	52.41	51.52	53.13	54.64
HK3	52.41	100	52.97	53.23	55.93
HKDC1	51.52	52.97	100	70.8	67.79
HK1	53.13	53.23	70.8	100	72.71
HK2	54.64	55.93	67.79	72.71	100

Supplementary Table 10. DNA percent identity matrix

	GCK	HK3	HK1	HKDC1	HK2
GCK	100	53.18	50.35	48.77	49.46
HK3	53.18	100	57.31	57.86	60.38
HK1	50.35	57.31	100	63.44	63.09
HKDC1	48.77	57.86	63.44	100	64.12
HK2	49.46	60.38	63.09	64.12	100

Supplementary Table 11. Primer sequences for amplifying regulatory elements

Region	Forward	Reverse
I	CGGTACCTGAGCTCGAAGGCCACAGG TTTTTAGATCTCCTT	TATCCTCGAGGCTAGTGGTCCTGGCG CATCTTGTA
II	CGGTACCTGAGCTCGTCCTGAGCATT CTTGTCTTCG	TATCCTCGAGGCTAGGGAACAGTTGG GAATTTTATGGAAA
III	CGGTACCTGAGCTCGCCAGATGGGTG ATACGCTTT	TATCCTCGAGGCTAGCCTAATTCCCCC ACTAGTTGATT
IV	CGGTACCTGAGCTCGAATGGAAGAATC TGCCCCAACA	TATCCTCGAGGCTAGTGGCCTTTCCGA GCAAACAT
V	CGGTACCTGAGCTCGTTGTCACTTTGC TCAGGGAAGTTG	TATCCTCGAGGCTAGGACCTTGGCTAA GTCCTCCTGCT
VI	CGGTACCTGAGCTCGAGAGGAAGTGT GGCCCCTTACA	TATCCTCGAGGCTAGGCCATGAAAAAT ACACACTGAAAACC
VII	CGGTACCTGAGCTCGCCCTTAGTGCCT GGGACGTG	TATCCTCGAGGCTAGTCCCCAGGTGA GAGGTGAGG
VIII	CGGTACCTGAGCTCGTCTGCAGCCCTC TCACCTCA	TATCCTCGAGGCTAGGAATGGCCCTG ACGAAGGTG
IX	CGGTACCTGAGCTCGTCCCATGTCAGG GTCCAGT	TATCCTCGAGGCTAGCCCTCCATGAAC TGCCTTGG
X	CGGTACCTGAGCTCGTCCTCAGATTCT GCCCTTCAGA	TATCCTCGAGGCTAGGAATGGCCCTG ACGAAGGTG
XI	CGGTACCTGAGCTCGTCCCACCACTAG AATTGTGGGTTT	TATCCTCGAGGCTAGCATGGTTTGCCA GGTCCACA

Supplementary Table 12. Primer sequences for site directed mutagenesis

SNP ID	Forward	Reverse	Edit
kg0002	CTTGTCTTCGGTTCA <u>C</u> AATAAACCTGT AAAA	TTTTACAGGTTTATTGTGAACCGAAGA CAAG	G>C
rs50309 37	TACCAGCAGGTCACA <u>C</u> TGATTTATGTT ATGA	TCATAACATAAATCAGTGTGACCTGCT GGTA	T>C
rs50309 38	TTTATGTTATGACGA <u>C</u> TTAACCTTTGGA CTTA	TAAGTCCAAAGTTAAGTCGTCATAACA TAAA	T>C
rs96455 01	ACCTTCTCAGCCCTC <u>A</u> TTCATTCTGC TTTC	GAAAGCAGAATGGAATGAGGGCTGAG AAGGT	G>A
rs14744 9838	CACATGCGGCTCTCC <u>A</u> ATGACACCCT TTTGG	CCAAAAGGGTGTCATTGGAGAGCCGC ATGTG	G>A
rs20021 6341	AGGCGGTTCCGGGCT <u>A</u> AGATGGAGAA GGGCC	GGCCCTTCTCCATCTTAGCCCGGAAC CGCCT	G>A
kg0001	TGGCTGCTCCTCGCC <u>A</u> CACATCCTGT CTTCC	GGAAGACAGGATGTGTGGCGAGGAG CAGCCA	G>A
rs50309 45	CCTTTATCCGAGTTT <u>T</u> CCCAGGCTGG GAGCA	TGCTCCAGCCTGGGAAAACCTCGGAT AAAGG	C>T
rs50309 46	CCAGGCTGGGAGCAC <u>A</u> AGCTCTCTCT TCCA	TGGGAAGAGAGAGCTTGTGCTCCAG CCTGG	C>A
rs14464 3300	ACCGCAAGGGCTGTG <u>T</u> TACATTTGC CTTCT	AGAAGGCAAATGTGAACACAGCCCTT GCGGT	C>T
rs47468 28	GTGCTCACATTTGCC <u>T</u> TCTCCAGTTGA CATG	CATGTCAACTGGAGAAGGCAAATGTG AGCAC	C>T
rs12241 136	AAAATCCAACACTT <u>T</u> TAAGATTTAAAC CTA	TAGGTTTAAATCTTAAAAGTAGTTGGA TTTT	A>T
rs10998 649	GGCAAGGCTGGTCTC <u>G</u> AACCTCTGAC CTCAGGCAATCTGCC	GGCAGATTGCCTGAGGTCAGGAGTTC GAGACCAGCCTTGCC	T>G
rs79142 56	GGCAAGGCTGGTCTCTAACTCCTGAT <u>I</u> CTCAGGCAATCTGCC	GGCAGATTGCCTGAGATCAGGAGTTA GAGACCAGCCTTGCC	C>T

Supplementary Table 13. Primer sequences for qPCR

	Forward	Reverse
Bactin	GTGGCCATCTCTTGCTGCAAG	GGGAAATCGTGCGTGACATTAAG
HKDC1	GGTCAGGATGCTGCCACCT	CCCAAGATCCAGGGCGAGAA
HK1	TGAAGTCGGCCTGATCATCG	TCCTCCCCTCGTCTCCTTCC
HK2	GGGTCCTGCTGGTCCGTGTT	TCCTGCGGGATGGCGTAGAT
HK3	GAGGAGACCCTGGCCCCATT	CCTTCCGCATCTGTGCCTGA
GCK	TGGATGTGGTGGCAATGGTG	GATCATGCCGACCTCGCACT
ratHK1	TTAACCCGCTTGGGAGTGGA	GGCTGATCGGAAGGAGACGA
ratHK2	AGCGACTTCGCTCCACCATC	GGAGACGCTTGGCAAAATGG
ratHK3	CTCTTCCAGGATGCGCCTGT	TCCCCTCTGTGGATGGTGGT
ratGCK	GGCACTGCCGAGATGCTCTT	GAAGCCCAGGGGCAGTTTCT
ratBactin	CACTGCCGCATCCTCTTCT	GGAACCGCTCATTGCCGATA