Additional file 1

High-throughput phenotyping of plant resistance to aphids by automated video tracking

Karen J. Kloth^{1,2,3}, Cindy J.M. ten Broeke¹, Manus P.M. Thoen^{1,2,3}, Marianne Hanhart-van den Brink¹, Gerrie L. Wiegers^{1,3}, Olga E. Krips⁴, Lucas P.J.J. Noldus⁴, Marcel Dicke¹ and Maarten A. Jongsma³



Figure S1 Arena settings in EthoVision XT.

Trial and control settings



Figure S2 Trial Control Settings in EthoVision XT for measuring the start time and duration of probes. Zone 1 is the leaf disc. 'Not moving' and 'Moving (1)' were defined by a start velocity of 0.3 mm/s and a stop velocity of 0.02 mm/s, averaged over 5 samples (0.2 seconds). The condition 'Moving (2)' was defined by a start velocity of 0.3 mm/s for *M. persicae*, resp. 0.35 mm/s for *N. ribsinigri*, a stop velocity of 0.1 mm/s, and a minimum duration of 2 seconds. This procedure was repeated indefinitely during data acquisition. Velocities were calculated by taking the average velocity of 5 frames, with a frame rate of 5 frames per second.

Table S1 Subject detection settings in EthoVision XT for *M. persicae*.

Parameters	Settings
Detection method	Grey scaling, centre point detection
Grey scale range	6-200
Subject size	85-600 pixels
Contour erosion (first)	2 pixels
Contour dilation	2 pixels
Sample rate	5 samples per second
Pixel smoothing	Medium
Track noise reduction	Off
Track smoothing profile	Lowess, 10 samples before and after

Table S2 Absolute differences between EPG recording and automated video tracking. Response variables are compared between (1) EPGs on intact plants, (2) EPGs on leaf discs, and (3) automated video tracking. Related variables, such as duration of phloem ingestion in EPGs and duration of long probes in video tracking, were tested for absolute differences (Kruskal-Wallis test, pairwise comparison with Mann-Whitney U test, alpha=0.05). On Arabidopsis accessions Co-2 and Sanna-2, probing behaviour of *M. persicae* was recorded, on lettuce cultivars Corbana and Terlana, probing behaviour of *N. ribisnigri*. Within each plant-aphid combination, different colours represent significant differences between the screening methods. Dark colours represent significantly higher values than lighter colours. Blocks with a cross were not significantly different from other blocks within the plant-aphid combination.

	Co-2	Sanna-2	Corbana	Terlana
	EPG intact plant EPG leaf disc Video leaf disc	EPG intact plant EPG leaf disc Video leaf disc	EPG intact plant Video leaf disc	EPG intact plant Video leaf disc
Total duration not penetrating/not probing				
Total duration phloem feeding /long probes				
Total duration sustained philoeni reeding/long profes				
Number of non-penetrations				
Number of phloem feeding events/long probes				
Number of short probes				
Mean duration phloem feeding events/long probes				
Latency to phloem feeding events/long probes				

Plant - aphid	Duration	Trait (min)	R	S
Arabidopsis -	8h	Mean duration long probes	62.8 ± 31.1	90.0 ± 40.4
Myzus		Total duration sustained probes	276 ± 120	353 ± 77.0
Arabidopsis -	6h	Mean duration long probes	62.6 ± 31.9	84.9 ± 32.0
Myzus		Total duration sustained probes	208 ± 91.9	263 ± 62.5
Lettuce - Nasonovia	8h	Mean duration long probes	72.4 ± 43.6	99.8 ± 57.3
		Total duration sustained probes	132 ± 127	260 ± 133
Lettuce - Nasonovia	4h	Mean duration long probes	53.0 ± 29.9	87.9 ± 49.4
		Total duration sustained probes	44.1 ± 60.8	125 ± 77.2
Plant - aphid	Duration	Trait (min)	Col-0	
Arabidopsis -	8h	Mean duration long probes	55.6 ± 30.9	
Myzus		Total duration sustained probes	170 ± 110	

Table S3 Video tracking variables used to generate simulations.

Mean \pm standard deviation, R= resistant plant line (Co-2, resp. Corbana), S= susceptible plant line (Sanna-2, resp. Terlana).