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Supplemental Data

**TFIIH Subunit Alterations Causing Xeroderma Pigmentosum
and Trichothiodystrophy Specifically Disturb Several Steps
during Transcription**

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Figure S1

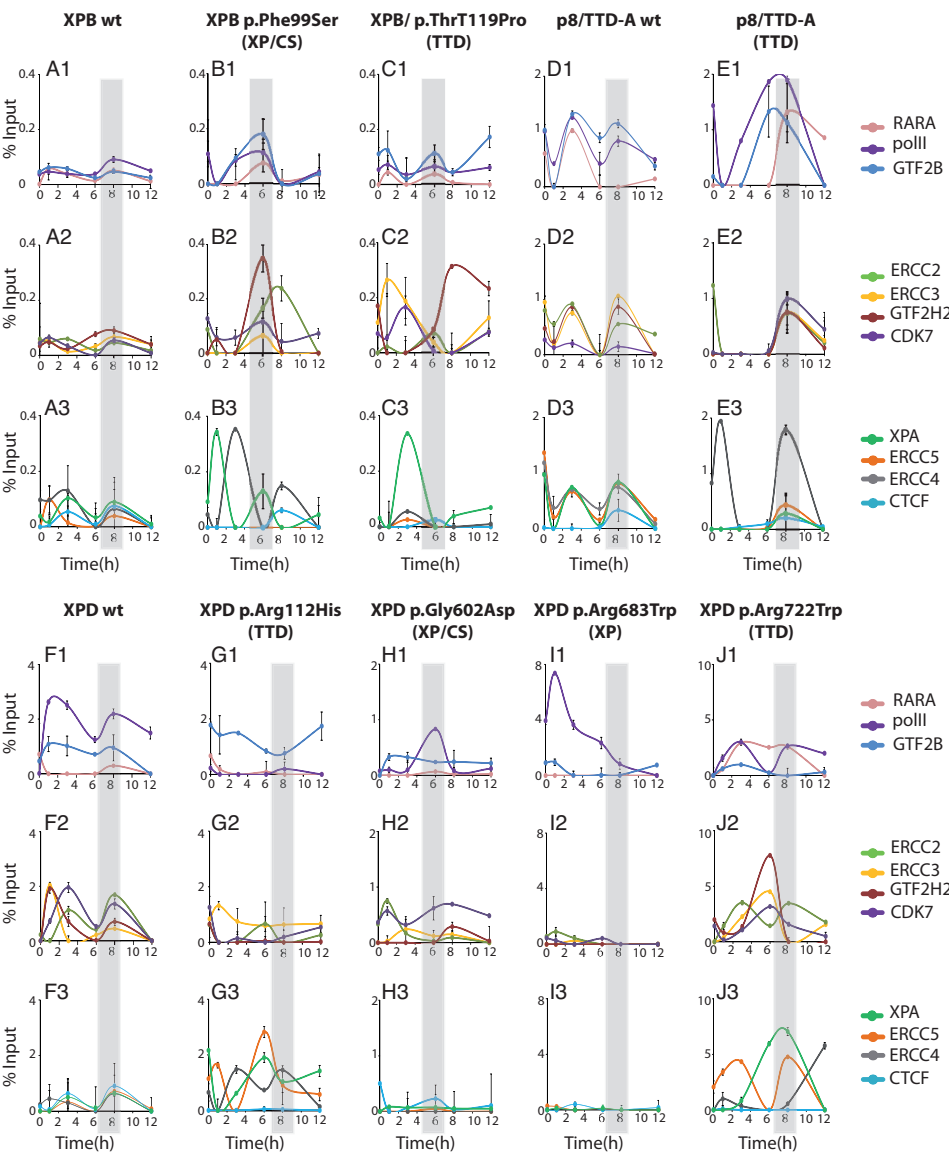


Figure S2

DNA break at terminator region

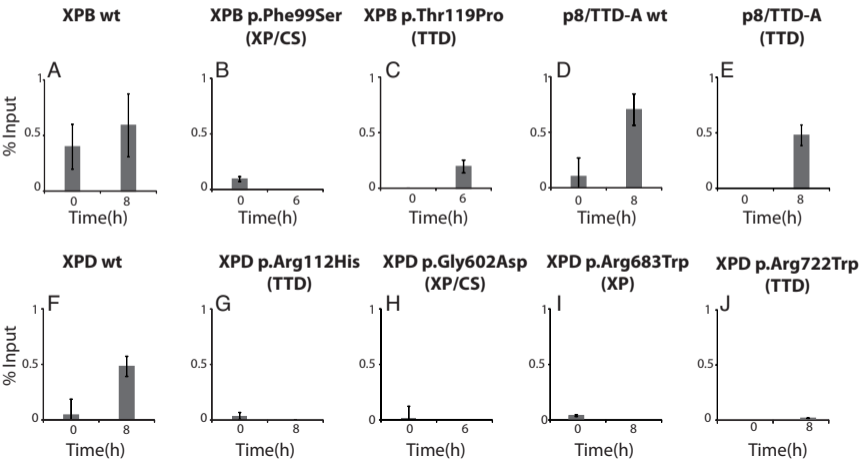
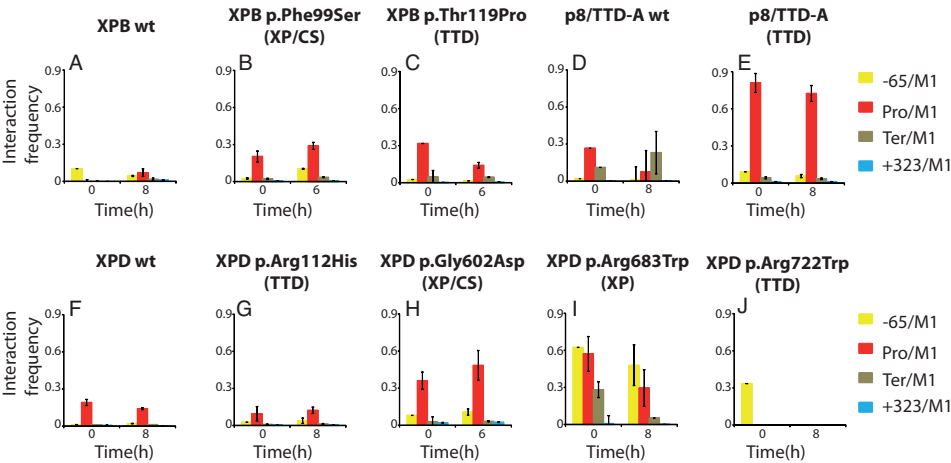
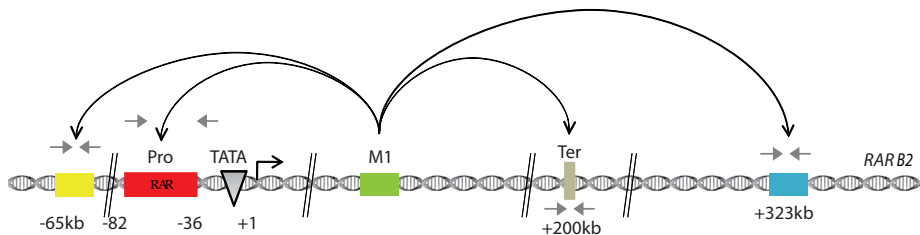


Figure S3



Supplemental Figure Legends

Figure S1. Transcriptional machinery and NER factors recruitment on the *RARB2* terminator are disturbed in cells bearing mutations in genes encoding TFIIH subunits

ChIP monitoring the t-RA-dependent recruitment of RARA, pol II, GTF2B (panels A1-J1), ERCC3, ERCC2, GTF2H2, CDK7 subunits of TFIIH (panels A2-J2) and XPA, ERCC5, ERCC4, CTCF (panels A3-J3) on the *RARB2* terminator; Each series of ChIP is representative of at least two independent experiments. Values are expressed as percentage of the input. Error bars represent standard deviation.

Figure S2. Mutations in genes encoding TFIIH subunits dysregulated DNA breaks on the *RARB2* terminator

(A-J) Detection of DNA breaks at *RARB2* terminator at 0 and either at 6 or 8 hours post t-RA treatment depending on the formation of the transcriptional machinery corresponding to RNA expression peak (see Figure 2 shadowed areas). Each series of BioChIP is representative of three independent experiments and values are expressed as percentage of the input. Error bars represent standard deviation.

Figure S3. 3C Controls for TFIIH mutated cells

(Upper panel) Schematic representation of the quantitative chromatin conformation capture (q3C). One probe was designed at the *RARB2* intronic region (M1) to investigate the associations between the different elements including upstream (-65 kb), promoter (Pro), terminator (Ter), and downstream (+323 kb) regions as indicated by the black arrows.

(Lower panels, A-J) q3C assays were performed using crosslinked and HindIII-digested chromatin from all the cells as indicated at 0 and either at 6 or 8 hours post t-RA treatment depending on the formation of the transcriptional machinery corresponding to RNA expression peak (see Figure 2 shadowed areas). The bar chart (y axis) shows the enrichment of PCR product (%) normalized to the enrichment within human *ERCC3* (=100%). Each PCR was performed at least three times. Signals were normalized to the total amount of DNA used, estimated with an amplicon located within a HindIII fragment in *RARB2*. Error bars represent standard deviation.

Table S1. List of the primers used in the study.

Primers	Forward	Reverse
mRNA		
<i>GAPDH</i>	AGCTCACTGGCATGGCCTTC	ACGCCTGCTTCACCACCTTC
<i>RARβ2</i>	CCAGCAAGCCTCCATGTTC	TACACGCTCTGCACCTTTAGC
ChIP		
<i>RARβ2</i> Promoter	TGGTGATGTCAGACTAGTTGGGTC	GCTCACTTCCTACTACTTCTGTAC
<i>RARβ2</i> Terminator	TGTTTGTGCTCTTTGGGCACT	CGGTCCGGGCTAGGAAACAAGTAAA
3C primers		
-65	CCTGGCAATTGAAACATGAAAGT	
Pro	TCCAAAGATGCCTATTAAGTTGTAAGAG	
M1	AGCAGCAAAATGCAGGCTTTA	TGACACCAGTGAAAAGGAAGCA
Ter	AAGATGCAGTTTGAGAGCATC	CTGGGCAACATGAAATAAAAAGATG
323	CCAAACAATTTTCTTCATGGTCATT	
<i>RARβ2</i> promoter	CAGACTAGTTGGGTCATTTGAAGGT	TTGAATTGCCTAATATATGCGAGTGA
<i>XPB</i>	CGGTGAGGTGAGTTTGTGGAAT	AGGATCTCTGTTTAATGGAAAAGCTT
3C Probes		
Ter probe	6[FAM]TTGCTCTTTCTGATGCTCTCAAA[TAM]	
M1 probe	6[FAM]CAGTACAGTCAAGGTGGCCCGTCT[TAM]	
<i>RARβ2</i> probe	6[FAM]AGCCCGGGTAGGGTTCACCGAAAG[TAM]	
<i>XPB</i> probe	6[FAM]AAGGATGAAGGCGTGATCCGACTCTG[TAM]	