1	SUPPLEMENTARY INFORMATION FOR
2	
3	Crystal structure of VmoLac, a tentative quorum quenching
4	lactonase from the extremophilic crenarchaeon Vulcanisaeta
5	moutnovskia
6	Structure of the lactonase VmoLac
7	Julien HIBLOT ^{1*} , Janek BZDRENGA ^{2*} , Charlotte CHAMPION ² , Eric CHABRIERE ^{2#} , Mikael
8	ELIAS ^{2#}
9	
10	

Figure S1: Biochemical analysis of *Vmo*Lac enzyme Figure S2: Chemical structure of phosphoesters (I-V), esters (VI-VIII) and lactones (IX-XXIII) Figure S3: Structural superposition of several PLL with *Vmo*Lac Figure S4: Surface electrostatic potential of *Vmo*Lac

- Figure S5: Surface salt bridge network example in the VmoLac structure 17
- Figure S6: Contacts between *Vmo*Lac active site in the packing contact 18
- Figure S7: Superposition of *Vmo*Lac structures solved in P64 (green) and P622 space groups. 19
- Figure S8: Anomalous X-ray scattering characterization of the VmoLac bimetallic center 20
- 21 Figure S9: Fourier difference maps of the bound ligands in *Vmo*Lac structures
- Figure S10: Superposition of monomers from homodimers of VmoLac bound to a fatty acid 22
- (A.) and to 3-oxo-C10 AHL (B.) 23
- Figure S11: Superposition of the structure bound to a fatty acid with the structure bound to the 24
- AHL. 25

11

12

13

14

15

16

- **Table S1:** Accession numbers of the sequences used in the phylogeny study
 26
- Table S2: Protein sequence identity between PLLs and *Bd*PTE 27
- 28
 Table S3:
 Anomalous X-ray data collection
- 29
- 30
- 31

SUPPLEMENTARY INFORMATION



34 **Figure S1:** Biochemical analysis of the *Vmo*Lac enzyme

35 The protein thermostability has been evaluated using different concentration of guanidinium

36 chloride (4.5, 5, 5.5 and 6 M). The melting-temperature (T_m) of *Vmo*Lac has been extrapolated

at a guanidine concentration equal to 0 using a linear regression and was evaluated to 128 \pm

38 7° C.



40 **Figure S2:** Chemical structure of phosphoesters (I-VI), esters (VII-IX) and lactones (X-XXIV)

41 Chemical structure of ethyl-paraoxon (I), methyl-paraoxon (II), ethyl-parathion (III), methyl-

42 parathion (IV), malathion (V), CMP-Coumarin (VI) phenyl-acetate (VII), pNP-acetate (VIII),

43 pNP-decanoate (IX), dihydrocoumarin (X), C4-AHL (XI), C6-AHL (XII), 3-oxo-C6-AHL

44 (XIII), C8-AHL (XIV), 3-oxo-C8-AHL (XV), 3-oxo-C10-AHL (XVI), γ-caprolactone (XVII),

45 γ -heptanolide (XVIII), Nonanoic- γ -lactone (XIX), Undecanoic- γ -lactone (XX), Dodenanoic- γ -

46 lactone (XXI) Nonanoic-δ-lactone (XXII), Undecanoic-δ-lactone (XXIII) and Dodenanoic-δ-

47 lactone (XXIV)



49 **Figure S3:** Structural superposition of several PLLs with *Vmo*Lac

- 50 Structures of VmoLac (green), SsoPox (cyan, 2VC5), SisLac (yellow, 4G2D), DrOPH (grey,
- 51 2ZC1) and GkL (blue, 3OJG) are represented as smoothed ribbon. The bimetallic center is
- represented by spheres: light pink for cobalt, orange for iron and blue for zinc.
- 53





56 The surface electrostatic potential of a monomer is represented on face (A) and face (B).

57 Negatively charged areas are depicted in red, whereas positively charged areas are in blue and

uncharged areas are in white. The active site crevice is indicated by the presence of the bound

59 fatty acid (green sticks, face (A))



Figure S5: Surface salt bridge network example in the VmoLac structure

The charge groups of involved residues are shown as spheres. Ionic interactions are indicated by black dashes.



- **Figure S6:** Contacts between *Vmo*Lac active site in the packing contact
- *Vmo*Lac molecules are shown in cartoon (green and blue) at the crystallographic interface. The
- active site channels of both monomers (grey surface) are connected. The two active site metal





89



Active site superposition of *Vmo*Lac monomers from P6₄ (green) and P622 (dark blue) space group crystals. Colors are conserved as previously. Active site residues are represented as sticks.



95 **Figure S8:** Anomalous X-ray scattering characterization of the *Vmo*Lac bimetallic center.

96 The Bijvoet difference Fourier maps from data collections at the arsenic atom peak (energy = 97 7.725kEV (**B**)) or remote (7.7kEV (**A**)) are contoured at 5 σ . The high anomalous peaks (14 and 98 11.5 σ for α and β sites, respectively) at the cobalt atom anomalous edge and the absence of 99 anomalous signal at slightly lower energy indicate the presence of cobalt in the protein's active 100 site.

101



Figure S9: Fourier difference omit maps of the bound ligands in *Vmo*Lac structures.

106 (A) *Vmo*Lac structure (blue sticks) bound to 3-oxo-C10 AHL (cyan sticks). The omit F_{obs} - F_{calc} 107 is contoured at 2.5 σ (green mesh). (B) *Vmo*Lac structure (blue sticks) bound to a fatty acid, 108 modelled as myristic acid (pink sticks). The omit F_{obs} - F_{calc} is contoured at 3 σ (green mesh). 109 Metal cations and the putative catalytic water molecule are shown as spheres (pink and red, 100 respectively).



143 Monomers A and B for each of the two structures were superposed. Each monomers are

represented in green and cyan with respective bound molecule in pink and orange. Cobalt

- atoms are represented as light pink balls and residues as sticks.
- 146





148 Figure S11: Superposition of the structure bound to a fatty acid with the structure bound

149 to the AHL.

- 150 Comparison of the binding mode of the fatty acid (modeled as myristic acid (orange sticks)
- bount to *Vmo*Lac (green sticks and cartoon)) and 3-oxo-C10 AHL (light pink sticks, bound to
- 152 VmoLac (cyan sticks and cartoon)). Cobalt cations and the bridging water molecule are
- represented by light pink and red spheres, respectively.

Table S1: Accession numbers of the sequences used in the phylogeny study

Sequence	NCBI Accession number
GsP (Geobacillus thermodenitrificans)	<u>YP_001125472.1</u>
GkL (Geobacillus kaustophilus)	<u>YP_147359.1</u>
DrOPH (Deinococcus radiodurans)	<u>NP_294654.1</u>
1HZY PTE (<i>Pseudomonas diminuta</i>)	<u>GI:13786715</u>
PTEflavob (Flavobacterium sp.)	<u>AAV39527.1</u>
PTEAgrobac (Agrobacterium tumefaciens)	<u>AAK85308.1</u>
2R1M opd (Agrobacterium radibacter)	<u>GI:167744959</u>
PLLBreviba (Brevibacterium mcbrellneri)	<u>WP 005881372.1</u>
PLLDermaco (Dermacoccus sp.)	<u>WP_006945246.1</u>
AhlA (Rhodococcus erythropolis)	<u>WP_003943005.1</u>
QsdA (<i>Rhodococcus erythropolis</i>)	<u>ABQ42704.1</u>
PLLRhodoco (Rhodococcus jostii)	<u>YP_701486.1</u>
PLLStrepto (Streptosprangium roseum)	<u>YP_003338238.1</u>
MCP (Mycobacterium avium subsp. paratuberculosis K-10)	<u>NP_962602.1</u>
PPH (Mycobacterium tuberculosis)	<u>NP_214744.1</u>
PLLMycobCD (Mycobacterium tuberculosis)	<u>NP_214744.1</u>
PLLMycbovi (Mycobacterium bovis)	<u>NP 853900.1</u>
SacPox (Sulfolobus acidocaldarius)	<u>YP_256726.1</u>
SisLac (Sulfolobus islandicus)	<u>YP_002828495.1</u>
SsoPox (Sulfolobus solfataricus)	<u>NP_343863.1</u>
VmoLac (Vulcanisaeta moutnovskia)	YP_004245953
Symbact (Symbiobacterium thermophilum)	<u>YP_074383.1</u>
PHP_E (Escherichia coli)	<u>YP_001723339.1</u>
PHP_P (<i>Photorhabdus asymbitoca</i>)	<u>YP_003041416.1</u>
PHP_Y (Yersinia aldovae)	<u>WP_004702383.1</u>
PHP_X (Xenorhabdus bovienii)	<u>YP_003466084.1</u>
RTX_K1 (Klebsellia pneumoniae)	<u>YP_001335395.1</u>
RTX_K2 (Klebsellia variicola)	<u>YP_003439523.1</u>
RTX_P (Pseudomonas syringiae)	EGH93047.1
RTX_3K (Rhodobacter sphaeroides)	<u>YP_001045290.1</u>

	Bd PTE	<i>Sso</i> Pox	Sis Lac	<i>Sac</i> Pox	VmoLac	PPH	MCP	AhIA/QsdA	DrOPH	<i>Gk</i> L	GsP
Bd PTE	-	32.8	32.2	33.8	29	34.8	35.7	28.9	30.34	27.2	27.2
<i>Sso</i> Pox		-	91.4	76.1	52	39.2	37.6	37.9	28.34	33.8	32.9
<i>Sis</i> Lac			-	76.1	51.27	37.6	36.31	36.6	27.4	33.8	32.2
<i>Sac</i> Pox				-	51.9	39.8	39.2	38.5	30.6	33.1	32.2
VmoLac					-	41.1	40.8	36	29	29.6	28.3
РРН						-	92	59	31.6	32.7	32.4
МСР							-	58.7	31.6	32.7	32.7
AhlA/QsdA								-	32	30.8	31.1
Dr OPH									-	58.8	60.7
<i>Gk</i> L										-	90.4

	Data collection			
Dataset	Co-K edge high	Co-K edge low		
Beamline	ID29	ID29		
Wavelength (Å)	1.6049 Å	1.6101 Å		
Detector	PILATUS 6M	PILATUS 6M		
Oscillation (°)	0.1	0.1		
Number of frames	3600	3600		
Resolution (Å) (last bin)	1.8 (1.9-1.8)	1.7 (1.8-1.7)		
Space group	P64	P64		
Unit-cell parameters (Å)	a = 174.87, b = 174.87, c = 62.07, α = 90, β = 90, γ = 120	a = 174.85, b = 174.85, = 62.06, α = 90, β = 90, = 120		
No. of observed reflections (last bin)	1 940 202 (258 680)	2 242 020 (321 007)		
No. of unique reflections (last bin)	196 781 (29 433)	232 940 (36 350)		
Completeness (%)(last bin)	100 (100)	99.8 (98.7)		
R _{meas} (%) (last bin)	7.3 (53.2)	9.5 (74.1)		
I/σ(I) (last bin)	21.74 (4.12)	16.25 (2.89)		
Redundancy (last bin)	9.86 (8.79)	9.62 (8.83)		
CC (1/2) (%)(last bin)	99.9 (90.9)	99.9 (81.3)		
Anomalous peak (σ) [#]	α metal = 14 B metal = 11 5	N/D		

<u>Table S3: Anomalous X-ray data collection</u>