

Fig. S1

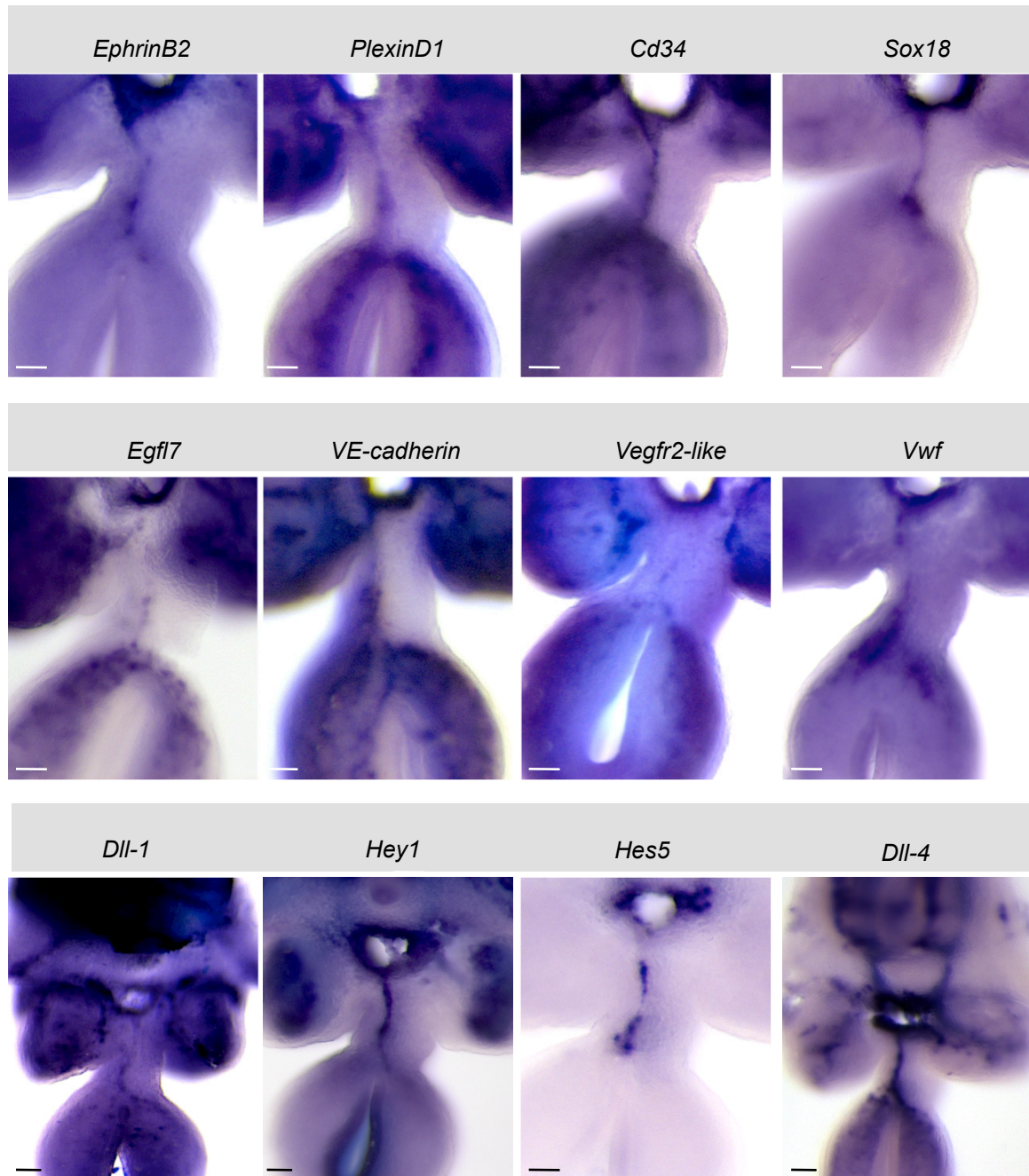


Fig. S2

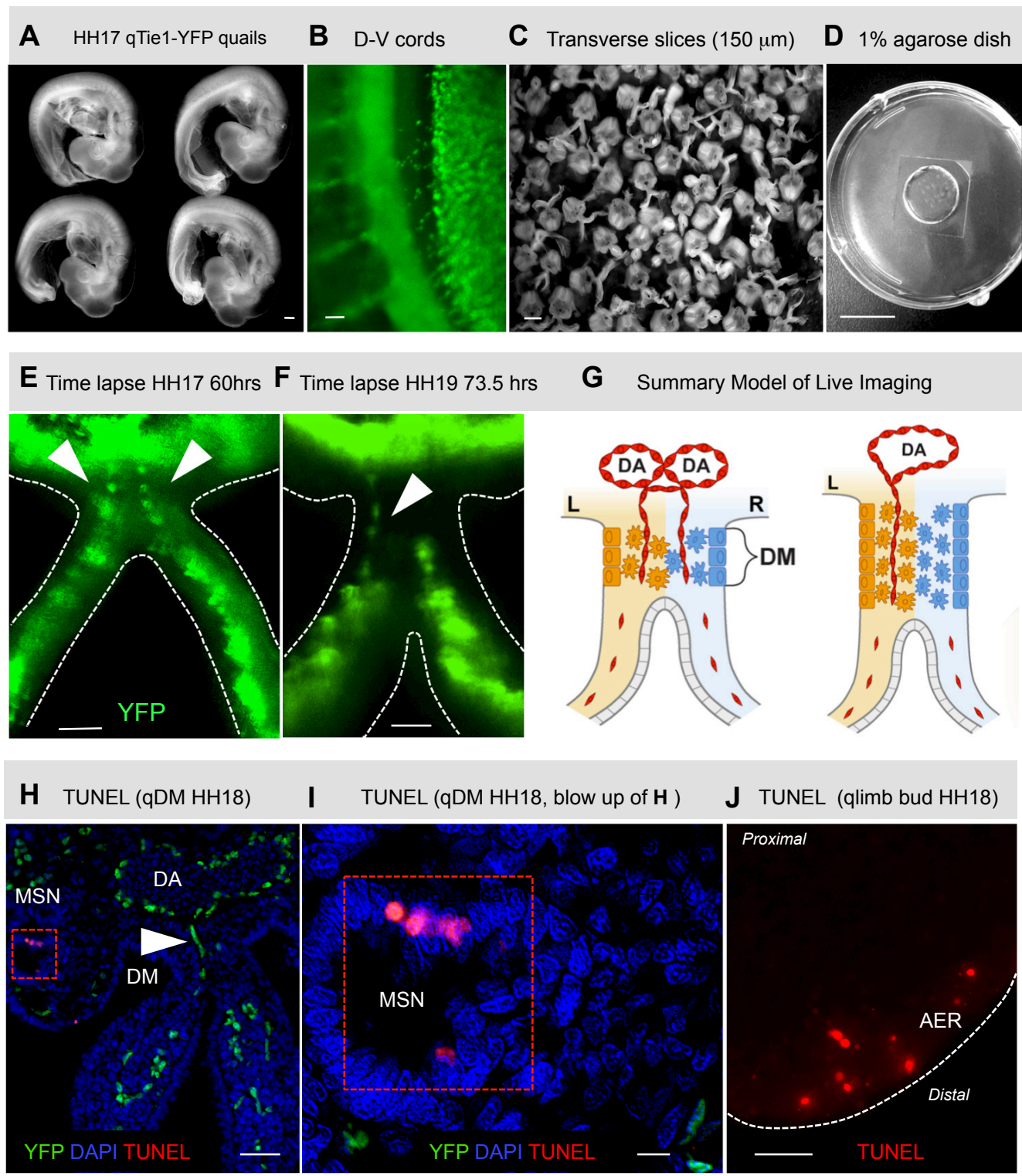


Fig. S3

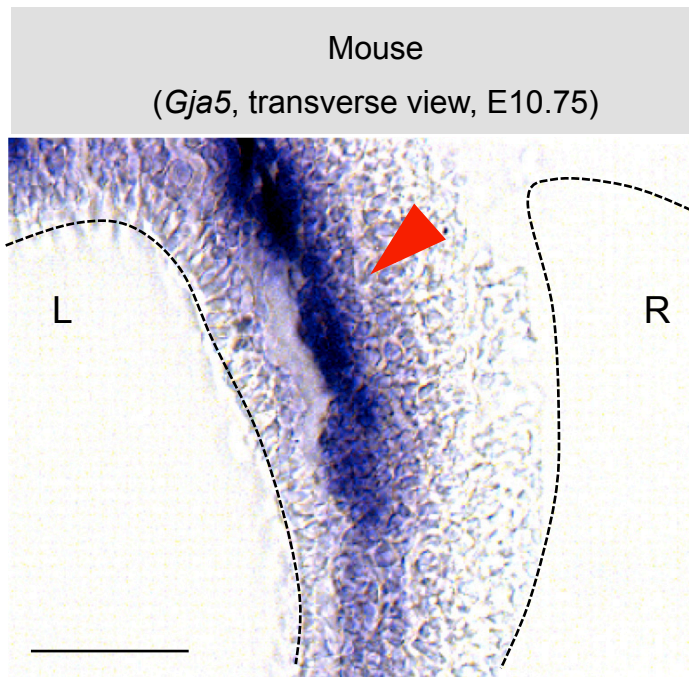


Fig. S4

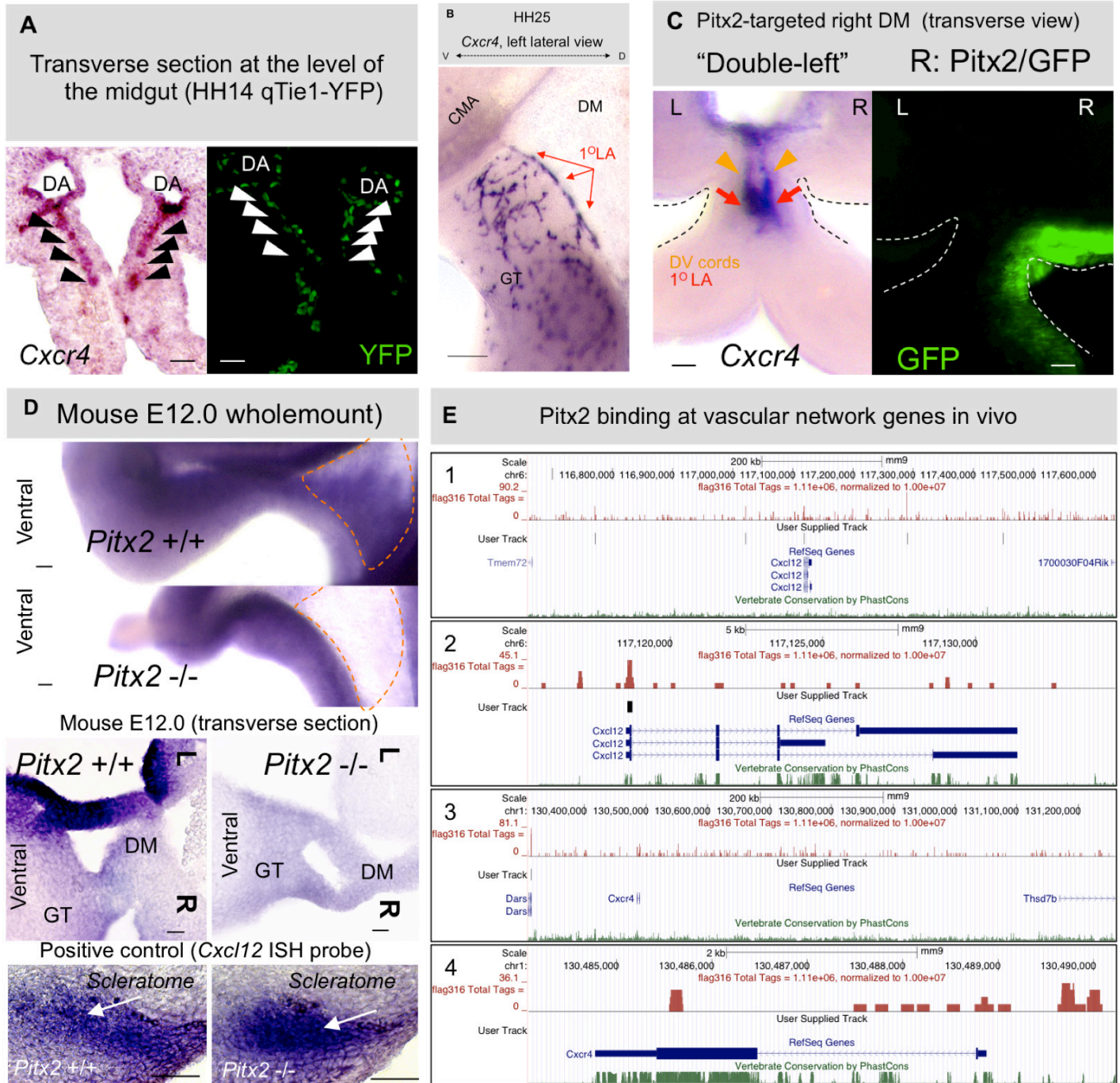


Fig. S5

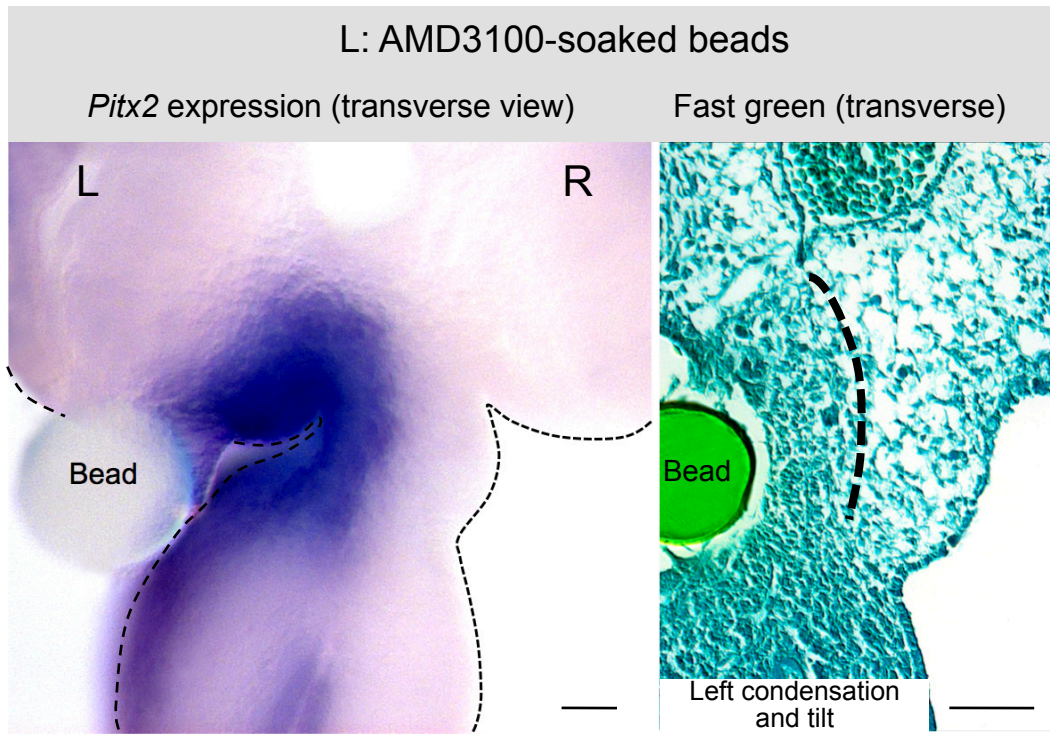


Fig. S6

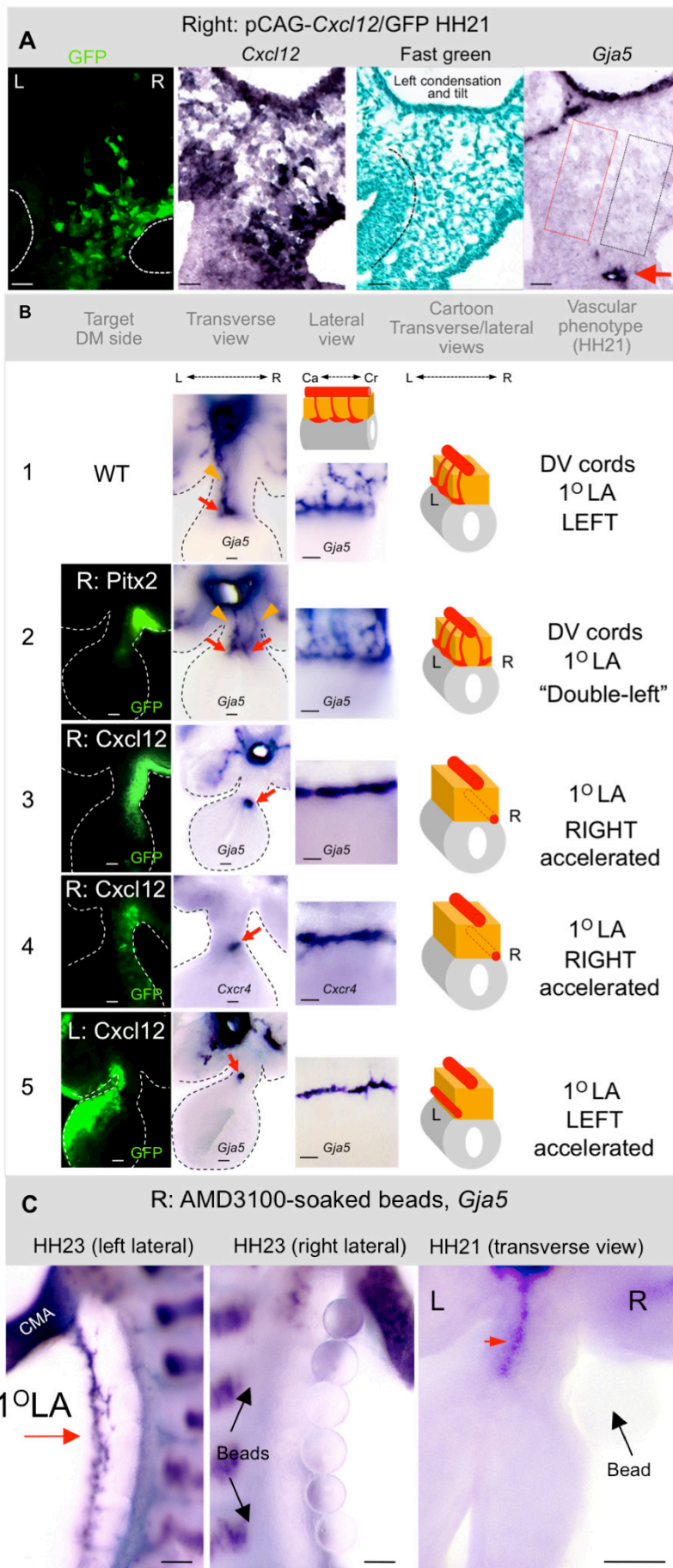


Fig. S7

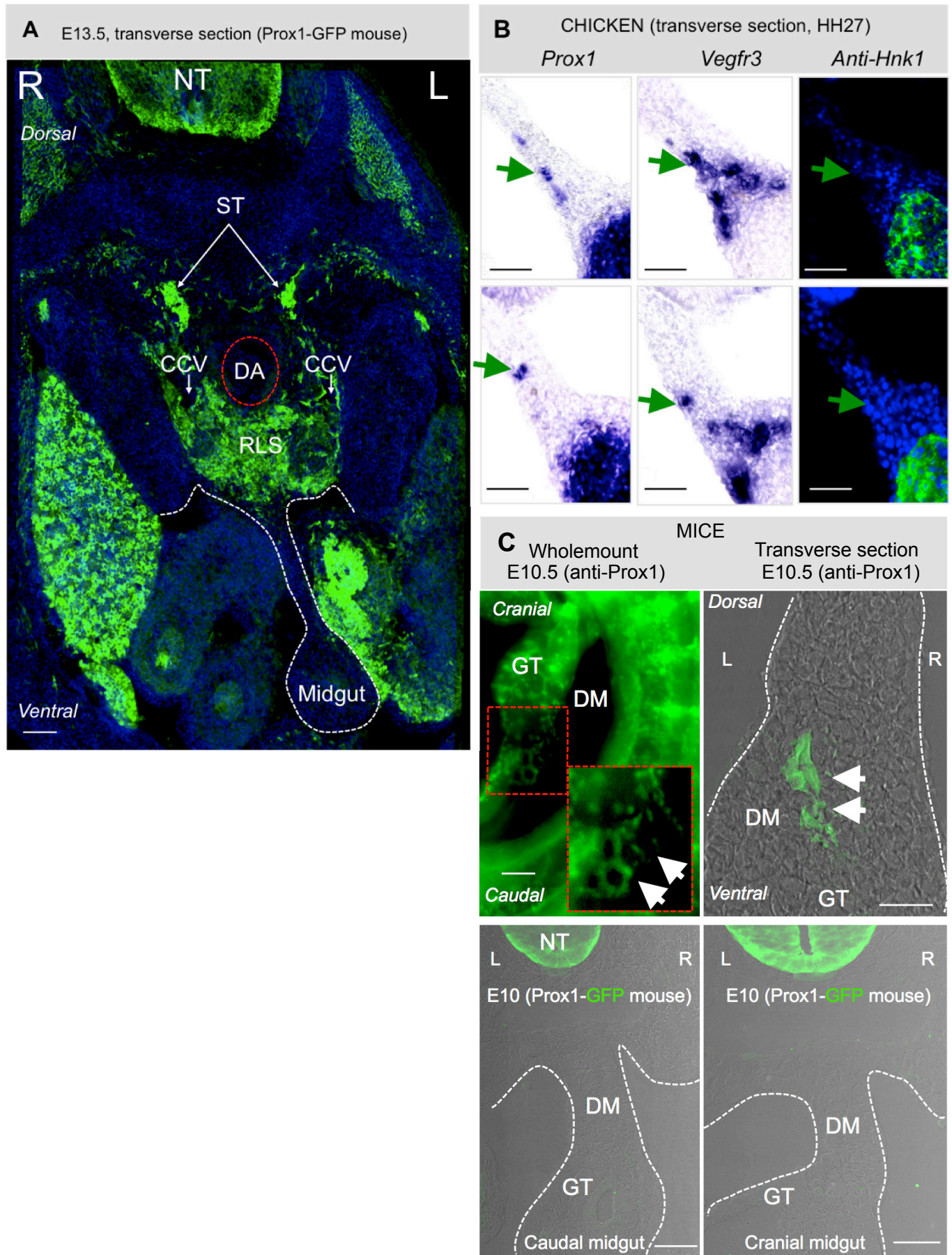
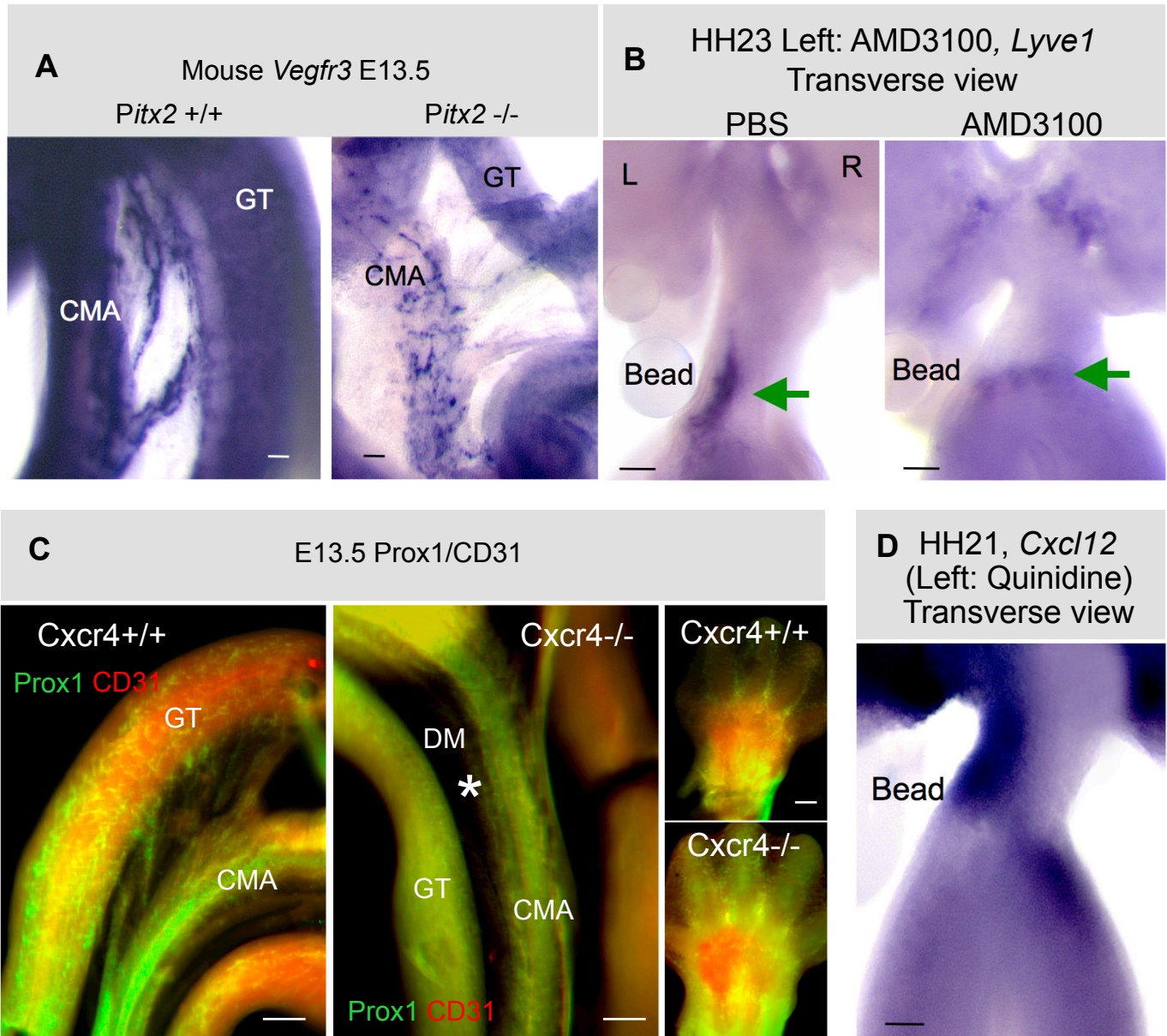


Fig. S8



Supplemental Figure Legends

Fig. S1. Validation of laser capture microdissection transcriptional analyses of the chicken DM (related to Fig. 1). In situ hybridization at HH21, using indicated probes. Scale bars: 50 μm .

Fig. S2. Live imaging setup for time lapse of transient D-V cords and TUNEL assay (related to Fig. 2). **A** Tie-1 H2B-YFP transgenic quails with wholemount view (**B**) of D-V endothelial cords. **C** Embryos are sliced into 150 μm transverse sections and then (**D**) embedded in low melting point agarose for imaging on glass-bottom culture plates. **E** Example of YFP fluorescent tissue slice, showing bilateral (left panel) and left-sided (right panel) endothelial cords; these panels are simplified in a cartoon model (**G**). (**H-J**) TUNEL assay of Tie-1 H2B-YFP transgenic quails demonstrating in **H** the absence of TUNEL-positive cells (red) in the DM and in Tie-1 H2B-YFP expressing endothelial cells (green, arrowhead); TUNEL-positive cells in the apoptotic mesonephros (MSN, boxed region of **H** is magnified in **I**) and in the limb bud AER (apical ectodermal ridge, **J**, from the same embryo as **H**). Scale bars: **A** 500 μm ; **B,C** 200 μm ; **D** 1.5 cm; **E,F,H** 50 μm ; **I** 10 μm ; **J** 20 μm .

Fig. S3. Analysis of arterial vasculogenesis in chicken and mice (related to Fig. 3). Asymmetric formation of D-V cords in mice at E.10.75 (*Gja5*). Scale bar: 50 μm .

Fig. S4. Analysis of Cxcr4/Cxcl12 expression downstream of Pitx2 (related to Fig. 4). **A** ISH at HH14 (chicken) reveals *Cxcr4* expression (**left panel**) in Tie1-expressing (H2B-YFP, **right panel**) endothelial cells of the bilateral intervening plexus (black arrowheads). **B** Later, *Cxcr4*

expression is found both in the 1⁰LA and gut plexus, but not the CMA. **C**, Misexpression of Pitx2/GFP in the right DM (right panel GFP marks electroporated cells) causes left isomerism (“double-left” phenotype) including double *Cxcr4*-positive arterial cords (orange arrowheads) and duplicated 1⁰LA (red arrows). **D** Loss of *Cxcl12* expression in the DM of *Pitx2*^{-/-} mouse (E12.0), with left lateral (**top two rows**) and transverse views shown (**third row**). Positive control tissue from matched embryos (**bottom row**) reveals *Cxcl12* staining in the scleratome. **E** Pitx2 binding at vascular network genes in vivo. **(1-2)** 5 enriched Pitx2 binding peaks surround *Cxcl12* **(1)**, one of which directly overlaps with exon 1 on *Cxcl12* **(2)**. **(3-4)** No significant Pitx2 binding is observed at the *Cxcr4* locus, the promoter of the adjacent gene *Dars* contains an enriched peak **(3)** but no peaks are observed in the promoter proximal region of *Cxcr4* **(4)**. ChIP-seq data derive from published transgenic FLAG-tagged Pitx2 binding in 12-week mouse cardiac tissue (NCBI Gene Expression Omnibus (GEO) data repository accession GSE50401): top green track shows normalized ChIP-seq tag numbers (y-axis) and the user supplied track below shows peaks with at least 4-fold enrichment over input control indicated as vertical black bars. RefSeq gene models are indicated in blue, while bottom track shows PhastCons vertebrate conservation score in green. Scale bars: **A** 20 μm; **B** 200 μm; **C,D** 50 μm.

Fig. S5. Side-specific effects of AMD3100 inhibitor-soaked beads on *Pitx2* expression and DM morphology (related to Fig. 5). AMD3100 has no effect on normal left-sided *Pitx2* expression (**left panel**) or cellular morphology (**right panel**) of the DM. Scale bars: 50 μm.

Fig. S6. *Cxcl12* is not sufficient to drive vessel formation in the absence of *Pitx2* (related to Fig. 6). **A** Misexpression of *Cxcl12* (pCAG-*Cxcl12*/pCAG-GFP) on the right side via co-

electroporation produces normal L-R DM cellular morphology (fast green), results in ectopic levels of Cxcl12 that exceed those on the left side (ISH, *Cxcl12*), and is not sufficient to promote formation of ectopic D-V cords in the right DM (ISH, *Gja5*, black dashed box). **B** Results of left-sided and right-sided targeting of Cxcl12 in the DM at HH21 (ISH, *Gja5*, model cartoon included). Transverse views shown to appreciate L-R situs and lateral view (Cranial-Caudal) to appreciate the extent of 1⁰LA formation. (1) Wild type DM showing left-sided D-V cord (orange arrowhead) and 1⁰LA formation (red arrow). (2) ‘Double-left’ phenotype with right-sided Pitx2 electroporation (GFP) showing left- and right-sided D-V cords and two 1⁰LA. (3) Right-sided Cxcl12 electroporation (GFP) showing ectopic 1⁰LA forming on the right side as a result of accelerated remodeling of the left-sided, Cxcr4-positive (4) D-V cords (see also Fig. 6). This result is in contrast to (2) highlighting an important functional difference between Pitx2 and Cxcl12 in the DM. (5) Left-sided Cxcl12 electroporation (GFP) showing accelerated formation of 1⁰LA as a result of accelerated remodeling of the left-sided D-V cords. **C** The diffusion range of AMD3100 inhibitor does not allow it to cross the midline boundary of the DM: inhibitor beads placed on the right side (**middle panel**, right lateral view) have no effect on the normal left-sided assembly of endothelial cords (**right panel**, arrowhead) and 1⁰LA (**left panel**, arrow). Bead positions are indicated. Scale bars: **A-B** 50 μm ; **C** 100 μm .

Fig. S7 Lymphatic marker expression in tissues surrounding the DM (related to Fig. 7). **A** Prox1-GFP-marked lymphatic anlagen in the retroperitoneal lymph sac (RLS), caudal cardinal veins (CCV) and non-lymphatic staining in the sympathetic trunk (ST) and neural tube (NT). **B** Adjacent transverse sections showing that location of lymphatic anlagen (green arrows) in three different HH27 chicken embryos, marked by *Prox1* (**left column**) and *Vegfr3* (**middle column**)

is spatially distinct from *Prox1*-expressing cells of the avian-specific Nerve of Remak (Hnk1-positive, **right column**). **C Top row** Wholemount (left) and transverse views (right) of wild type mouse DM stained with anti-Prox1; lymphatic plexus is first visible ventrally (white arrows, higher magnification inset) that extends dorsally. **Bottom row** Absence of Prox1-GFP reporter signal in mouse cranial or caudal DM prior to E10.5, revealed by transverse section of E10 Prox1 transgenic reporter mice. Scale bars: **A** 100 μm ; **B** 50 μm ; **C** (top left panel) 100 μm (top right panel) 20 μm (bottom panels) 50 μm .

Fig. S8 Local lymphangiogenesis in the left DM requires the preceding Pitx2-driven arterial program (related to Fig. 8). **A** *Vegfr3* expression (ISH) is absent from *Pitx2*^{-/-} mutant DM at E13.5 but remains unchanged along the CMA. **B**, Cxcr4/Cxcl12-inhibiting AMD3100 beads (**right panel**) on the left side of the chicken DM abrogate the *Lyve1*-positive network (HH23), compared to PBS beads (**left panel**). **C** Wholemount double immunohistochemistry for Prox1 (green) and PECAM/CD31 (red) reveals loss of Prox1 staining (asterisk) in the DM of Cxcr4^{-/-} mutant mouse embryos at E13.5 (**middle panel**) compared to wild type littermates (**left panel**). Peripheral limb lymphatics are unchanged in matched embryos (**right column**). **D** Arteriogenesis-inhibiting Quinidine beads on the left side of the chicken DM have no effect on *Cxcl12* expression (HH21). Scale bars: **A,B,D** 50 μm ; **C** 100 μm .

Movie S1. Right-sided endothelial cord regression in midgut explants of Tie-1-H2B-eYFP quail embryos (related to Fig. 2 and Fig. S2). **Left panel:** Select nuclei of endothelial cells of right DM were manually tracked with dragon tails (Bitplane Imaris software). **Right panel:** Tracking of select endothelial nuclei of the left cord (green), right cord (red) and dorsal aorta

(blue). At time $T=0$ (HH17, 60 hours), bilateral endothelial cords are present in the DM. At $T=4.5$ hours (HH18, 65 hours), cells of the right side endothelial cords have significantly regressed and by $T=6.5$ hours (HH19, 70.5 hours) the regression is complete.

Movie S2. Right to left endothelial cell migration in midgut explants of Tie-1-H2B-eYFP quail embryos (related to Fig. 2 and Fig. S2). **Left panel:** Select nuclei of endothelial cells of right DM were manually tracked with dragon tails (Bitplane Imaris software). **Right panel:** Tracking of select endothelial nuclei of the left cord (green), right cord (red) and dorsal aorta (blue). Starting at HH19, time-lapse imaging reveals that cells within right-sided endothelial cords regress, migrate and anastomose with the left endothelial cords giving rise to vasculature asymmetries in the DM.

Movie S3: Complete regression and endothelial cell emigration of right-sided arterial cord (related to Fig. 2 and Fig. S2). **Left panel:** Time-lapse of Tie-1 H2B-eYFP quail embryo midgut explants shows remodeling of the intervening endothelial plexus to yield the left DV arterial cord. At time $T=0$ (HH17), bilateral endothelial cords are present in the DM. At $T=5$ hours (HH18), cells of the right side endothelial cords have significantly regressed and by $T=10.5$ hours (HH19) regression is complete. By $T=13.5$ hours (HH20) we find that regressed cells from the right migrate left and contribute to the left endothelial cord. **Right panel:** Select nuclei of endothelial cells of the left and right cord, and left and right dorsal aortae are labeled with green, red, purple and blue spots, respectively; tracking of these spots models the cell movements in the left panel movie.