

Supplementary Methods:

Cell cycle analysis

Cells were plated and incubated at 37°C in complete DMEM medium. 24 hours later, they were treated with 0,2 μ M MLN4924 for 1, 2 or 24 hrs, as indicated. After collecting the cells with trypsin, they were resuspended in 1 ml PBS and washed twice in 1 ml PBS. Then, cells were fixed in 5 ml 70% ethanol and incubated at 4°C for 2 hrs, washed twice with PBS, resuspended in 0,2 μ l of 1 mg/ml of propidium iodide and RNase 0,2 mg/ml and incubated a 37°C for 30 minutes. At least 10000 cells were counted from each sample.

Supplementary legends

Figure S1: An example of cells positive (solid arrows) or negative (empty arrows) for RPA foci.

Figure S2: Cell cycle distribution of U2OS cells treated with DMSO or MLN4924 for the indicated length of time. Cell cycle distribution was determined by FACS.

Figure S3: Protein depletions. A, Protein extracts were blotted with an anti-NEDD8 antibody to analyze the overall neddylation status in the cells. The signal of the whole lane, normalized to the loading control and relative to the transfection with the empty plasmid was calculated. The average and standard deviation of three independent experiments is shown. B, CtIP levels upon shRNA-mediated downregulation of CtIP. C, siRNA-dependent depletion of RNF111 using two alternative siRNAs compared with control. D, same as C, but depletion of UBE2M. An asterisk marks an unspecific band.

Figure S4: Effect of NEDD8 overexpression on the NHEJ/HR balance. Cells harboring the SSR system were transfected with HA-NEDD8 or an empty HA vector. The NHEJ/HR ratio was calculated as described and normalized to the cells with the empty vector.

Supplementary tables

Table S1: Inhibitors and concentrations used in the SSR screening

Inhibitor	Concentration	Supplier	Reference
Thrichostatin A (TSA)	1.3 μ M	Sigma	T8552
Aphidicolin	1 μ M	Sigma	A0781
Mirin	25 μ M	Tocris	3190
PARPi	1 μ M	AstraZeneca	Olaparib
MNL4924	1 μ M	Vitro	I-502-01M

Table S2: siRNAs used in this study

Target gene	Description	Supplier	Reference
Control	ON-Targetplus Non-targeting pool	Dharmac on	D-001810- 10-20
RNF111-1	GCGCUUCCAUAACAAUUC	Dharmac on	J-007002-07- 0010
RNF111-2	GAGUUGAGAUGAUUAAUAG	Dharmac on	J-007002-08- 0010
hUBE2M-1	CAGAGGUCCUGCAGAACAA	Sigma	(7)
hUBE2M-2	GGGCUUCUACAAGAGUGGGAAGUUU	Sigma	(14)

Table S3: shRNAs used in this study

Target gene	Description	Supplier	Reference
Scramble	pLKO.1-puro Non-Target shRNA Control Plasmid DNA	Sigma	SHC016
CtIP	TRCN0000005403	Sigma	NM_0022894.1- 3008s1c1

Table S4: Primary Antibodies used in this study. IF, immunofluorescence. WB Western blotting. IP, Immunoprecipitation. SMART, Single Molecule Analysis of Resection Tracks

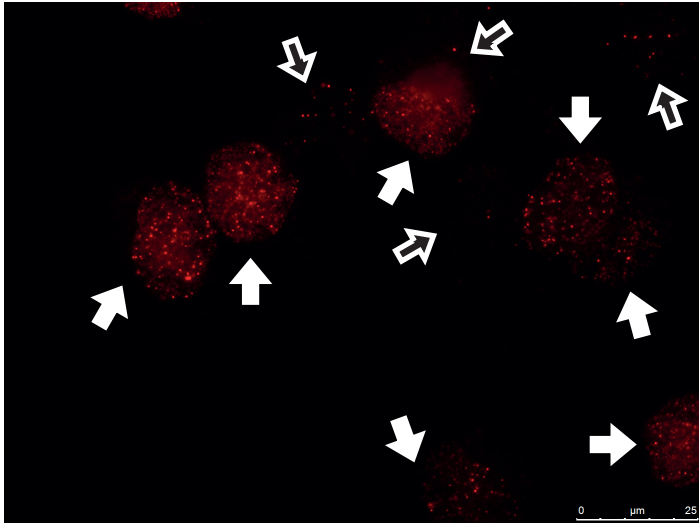
Target protein	Application	Supplier	Reference
NEDD8	IP	GeneTex	GTX61205
RNF111	WB	Abnova	H00054778-M05
BRCA1	IP	Santa Cruz	sc-642
BRCA1	IP, WB	Santa Cruz	sc-6954
HSP70	WB	Santa Cruz	sc-24
Tubulin	WB	Sigma	T9026
RPA32	IF	Abcam	ab2175
RIF1	IF	Bethyl Laboratories	A300-568A
Non related Rabbit IgG	IP	Sigma	I8140

Non related Mouse IgG	IP	Sigma	I8765
CtIP	WB	R.Baer	
UBE2M	WB	Bostom Biochem	A-655
BrdU	SMART	Sigma	B5002-100MG

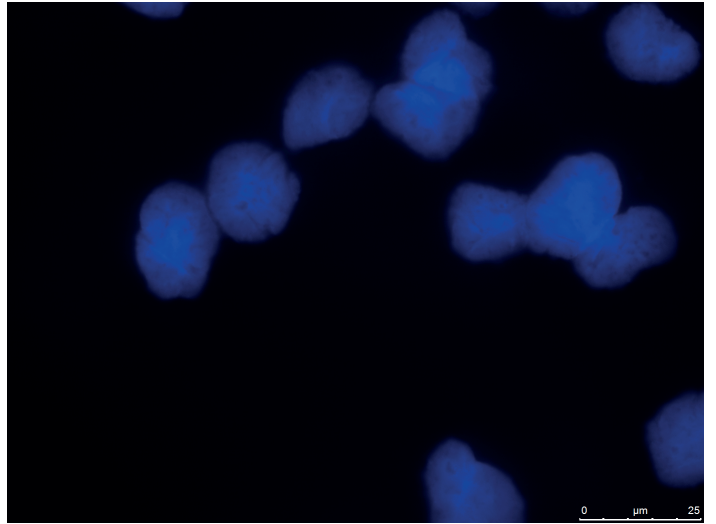
Table S5: Secondary antibodies used in this study. IF, immunofluorescence. WB Western blotting.

Antibody	Source	Application
Alexa Fluor 594 goat anti-mouse	Invitrogen	IF
Alexa Fluor 488 goat anti-rabbit	Invitrogen	IF
Alexa Fluor 647 goat anti-mouse IgG	Invitrogen	IF
Alexa Fluor 568 goat anti-rabbit	Invitrogen	IF
IR-Dye 680 RD goat anti-mouse IgG (H+L)	Li-Cor	WB
IR-Dye 800CW goat anti-rabbit IgG (H+L)	Li-Cor	WB

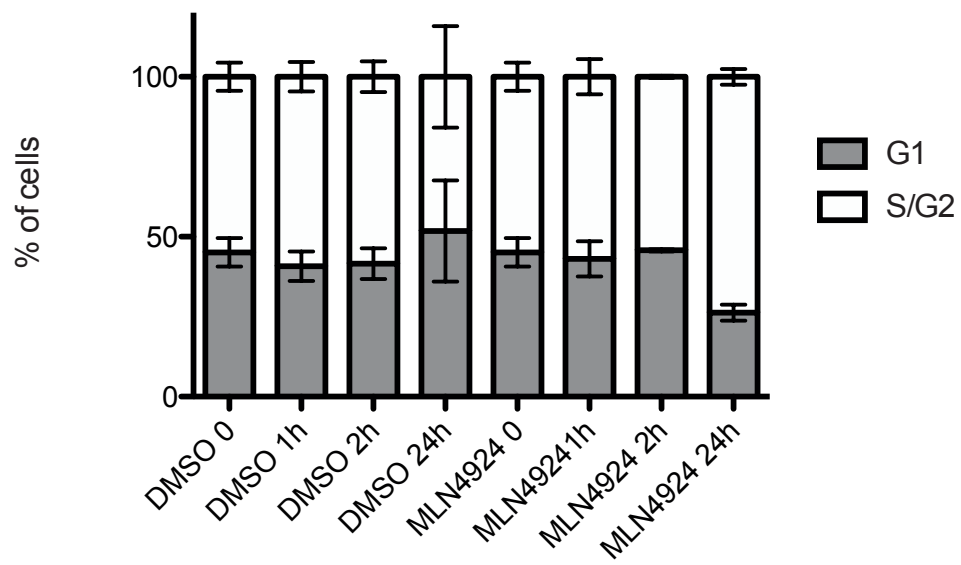
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DAPI

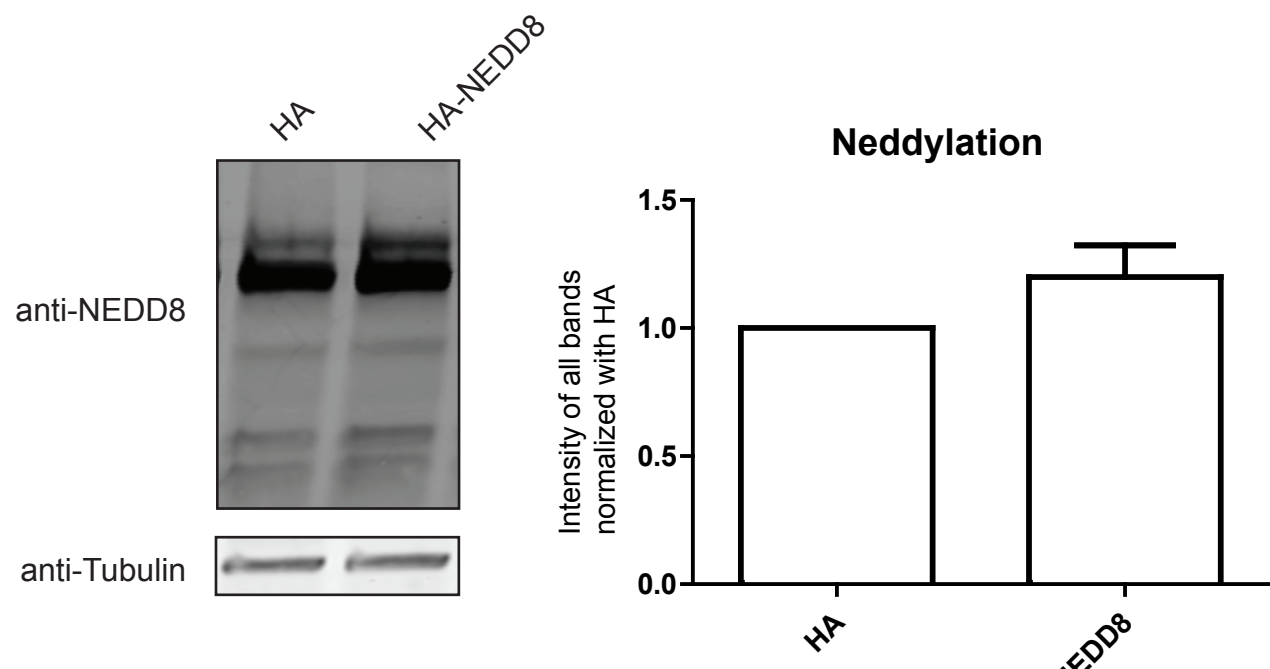


Jimeno *et.al.* Figure S1

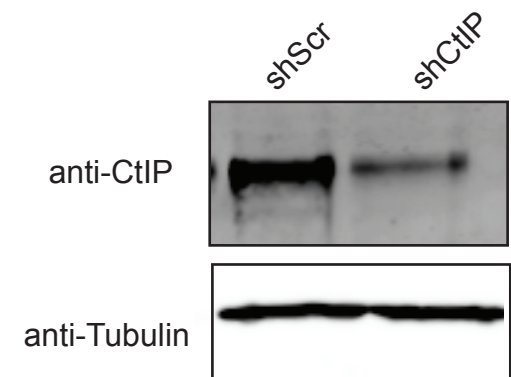


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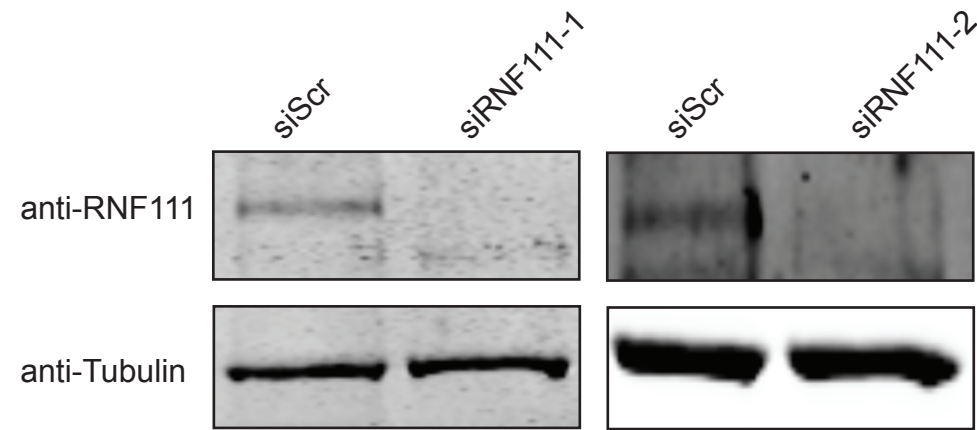
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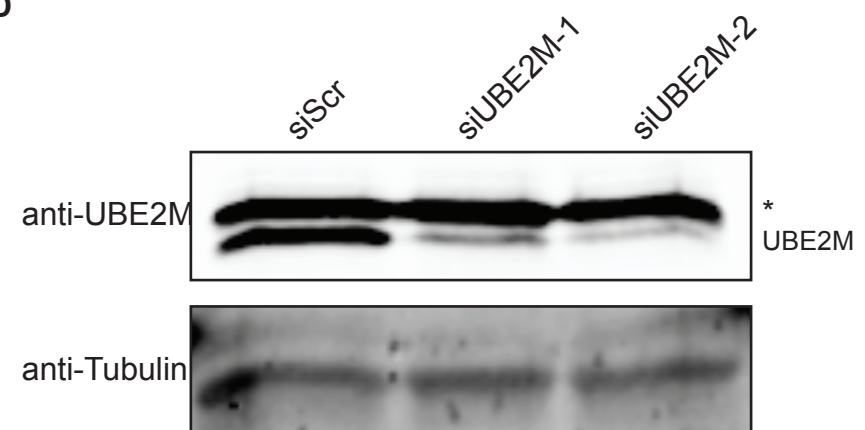
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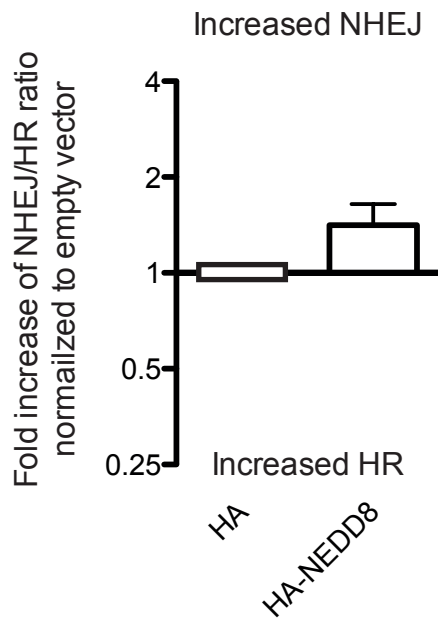


C



D





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