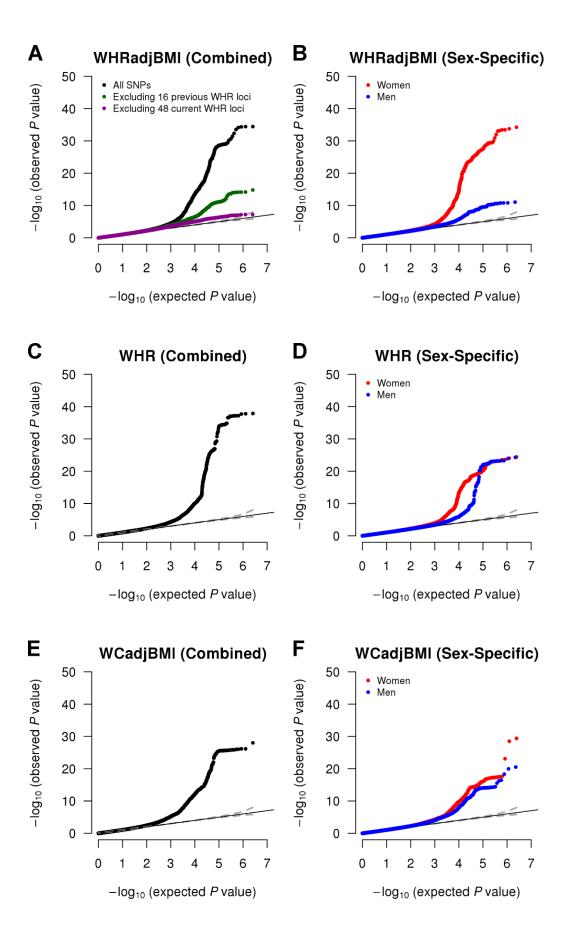
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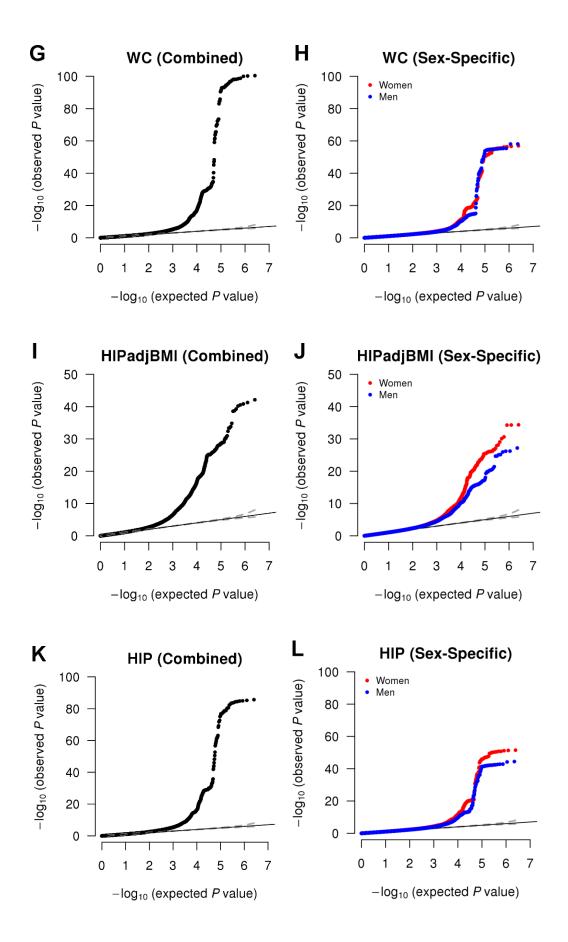
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# **Supplementary Figures**

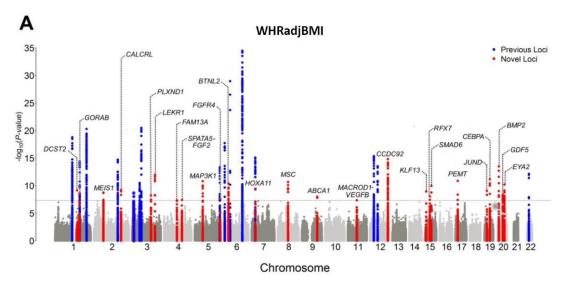
This document contains **Supplementary Figures 1, 2, 3, 4**, and **5**.

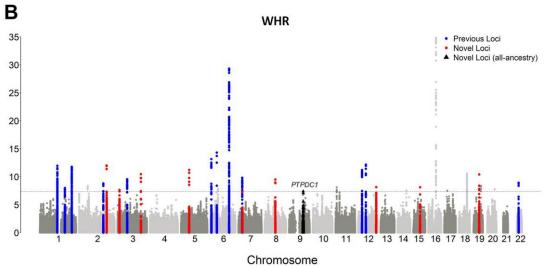
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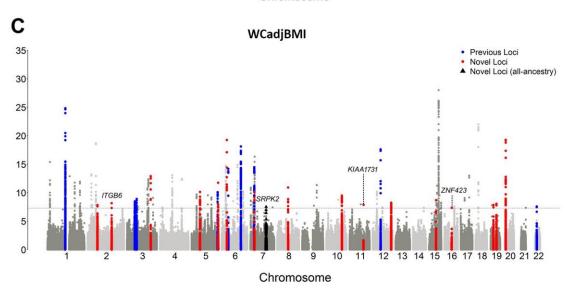


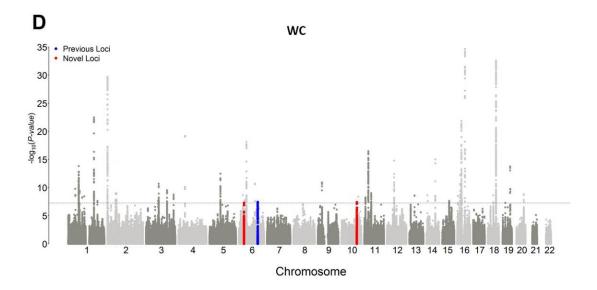


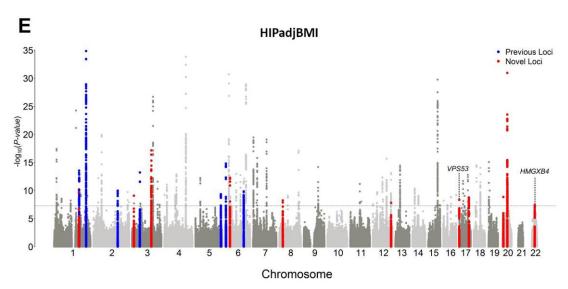
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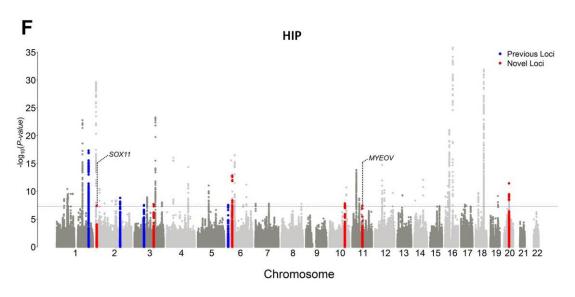






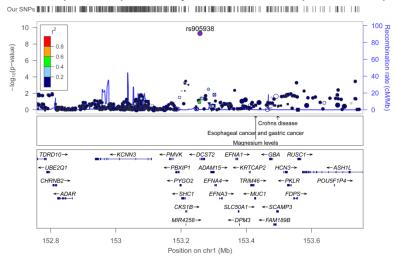




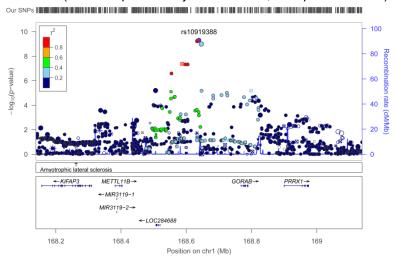


Supplementary Figure 3 | Regional association plots for 68 novel loci achieving genomewide evidence of association with six waist-related traits. The signals shown for waist-hip ratio (WHR), waist circumference, and hip circumference, adjusted and not adjusted for body mass index (BMI) do not overlap with association signals with height or BMI. Plots are arranged in the same order as **Tables 1** and **3**. In the plot of the *HOXA11* locus, the eponymous gene was automatically omitted by LocusZoom for space; it is located just to the left of *HOXA13* (upstream with respect to the genome).

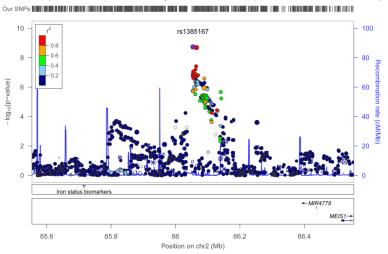
### DCST2 (Waist-Hip Ratio adjusted for BMI, European Women)



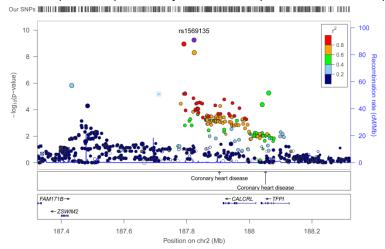
### GORAB (Waist-Hip Ratio adjusted for BMI, European Women)



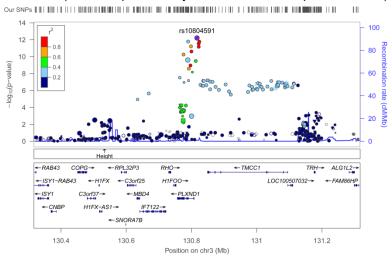
# MEIS1 (Waist-Hip Ratio adj. for BMI, European Sex-Combined)



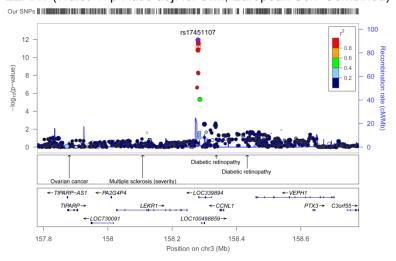
### CALCRL (Waist-Hip Ratio adj. for BMI, European Sex-Combined)



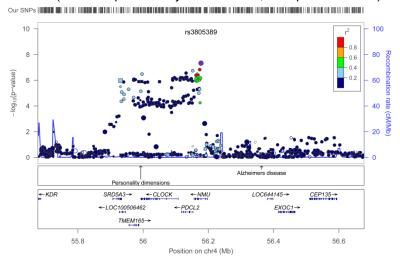
# PLXND1 (Waist-Hip Ratio adjusted for BMI, European Women)



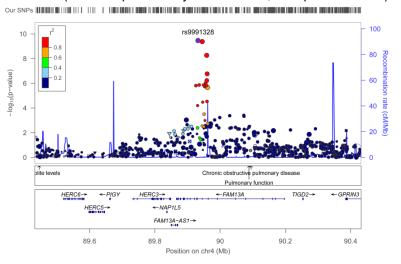
### LEKR1 (Waist-Hip Ratio adj. for BMI, European Sex-Combined)



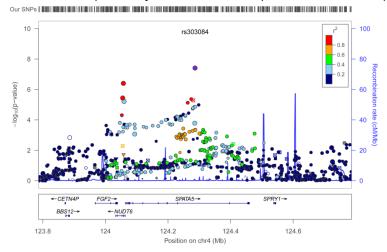
# NMU (Waist-Hip Ratio adjusted for BMI, European Women)



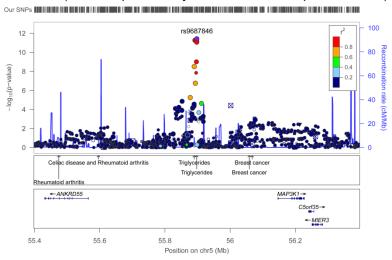
# FAM13A (Waist-Hip Ratio adjusted for BMI, European Women)



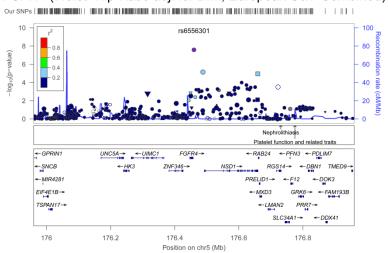
### SPATA5-FGF2 (WHR adjusted for BMI, European Sex-Combined)



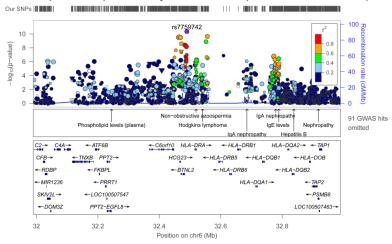
# MAP3K1 (Waist-Hip Ratio adjusted for BMI, European Women)



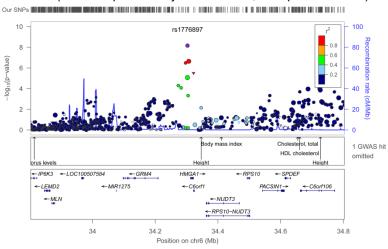
### FGFR4 (Waist-Hip Ratio adj. for BMI, European Sex-Combined)



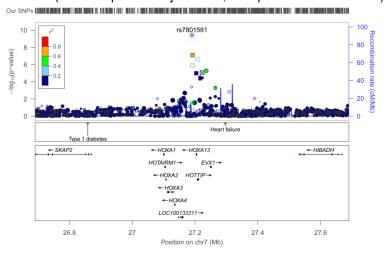
### BTNL2 (Waist-Hip Ratio adj. for BMI, European Sex-Combined)



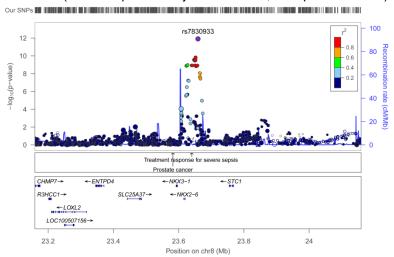
### HMGA1 (Waist-Hip Ratio adjusted for BMI, European Women)



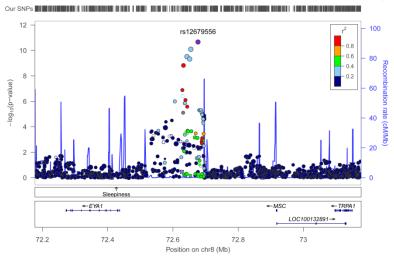
### HOXA11 (Waist-Hip Ratio adj. for BMI, European Sex-Combined)



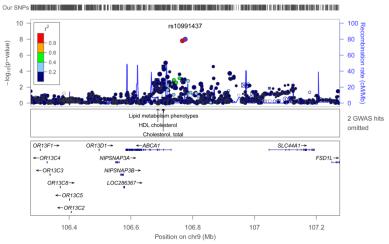
### NKX2-6 (Waist-Hip Ratio adjusted for BMI, European Women)



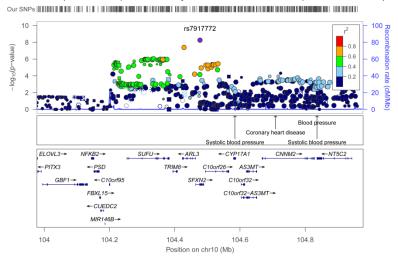
### MSC (Waist-Hip Ratio adj. for BMI, European Sex-Combined)



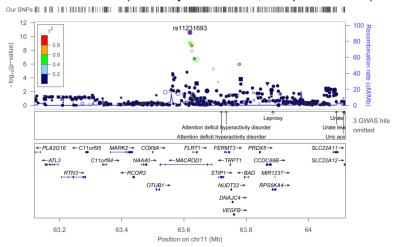
### ABCA1 (Waist-Hip Ratio adj. for BMI, European Sex-Combined)



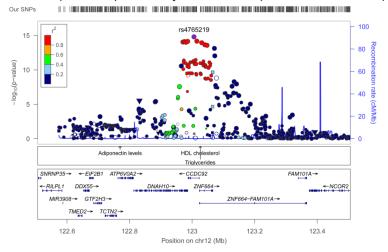
### SFXN2 (Waist-Hip Ratio adjusted for BMI, European Women)



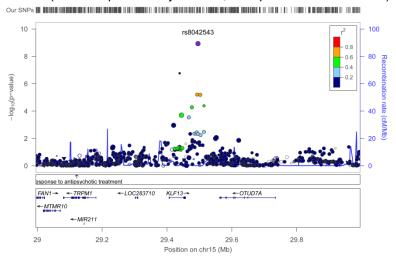
#### MACROD1-VEGFB (WHR adjusted for BMI, European Women)



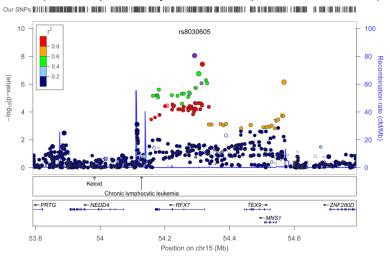
### CCDC92 (Waist-Hip Ratio adj. for BMI, European Sex-Combined)



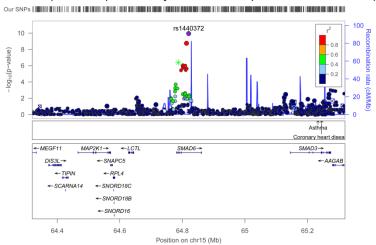
### KLF13 (Waist-Hip Ratio adj. for BMI, European Sex-Combined)



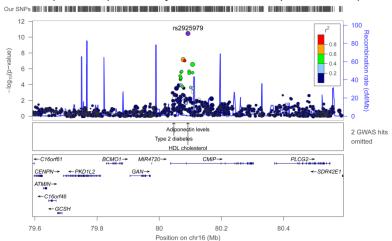
# RFX7 (Waist-Hip Ratio adj. for BMI, European Sex-Combined)



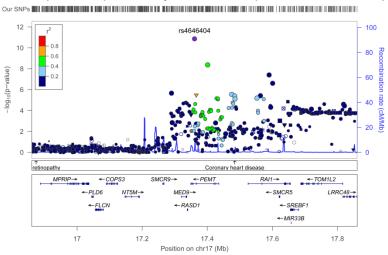
# SMAD6 (Waist-Hip Ratio adj. for BMI, European Sex-Combined)



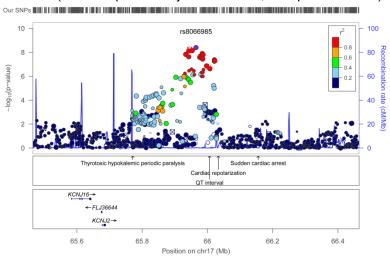
### CMIP (Waist-Hip Ratio adjusted for BMI, European Women)



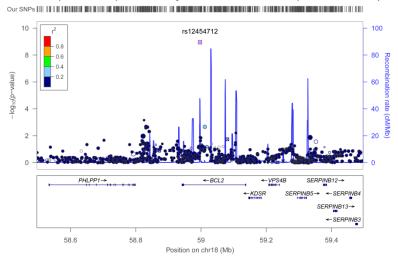
# PEMT (Waist-Hip Ratio adj. for BMI, European Sex-Combined)



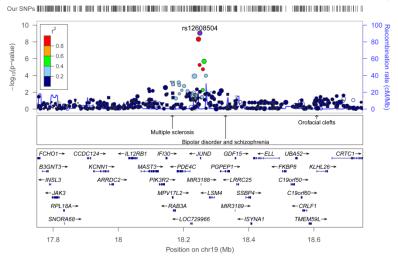
### KCNJ2 (Waist-Hip Ratio adjusted for BMI, European Women)



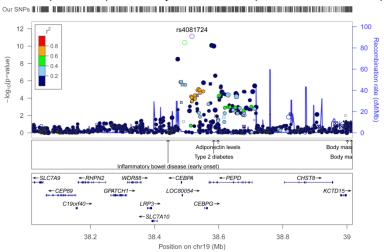
### BCL2 (Waist-Hip Ratio adjusted for BMI, European Women)



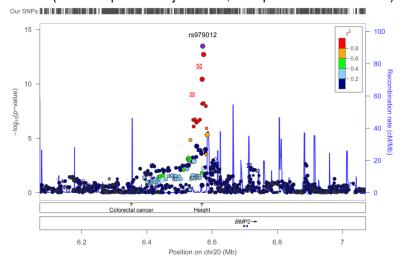
# JUND (Waist-Hip Ratio adj. for BMI, European Sex-Combined)



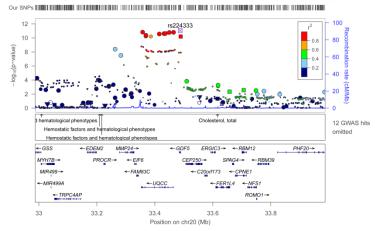
### CEBPA (Waist-Hip Ratio adj. for BMI, European Sex-Combined)



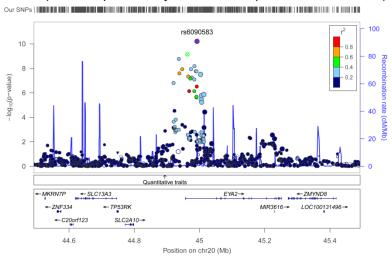
### BMP2 (Waist-Hip Ratio adj. for BMI, European Sex-Combined)



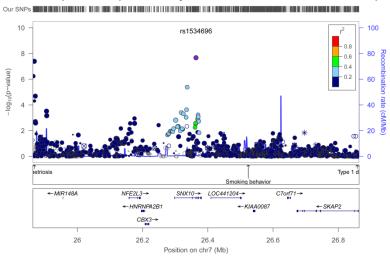
### GDF5 (Waist-Hip Ratio adjusted for BMI, European Men)



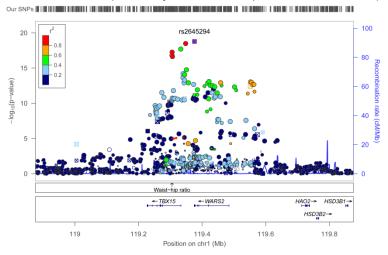
### EYA2 (Waist-Hip Ratio adj. for BMI, European Sex-Combined)



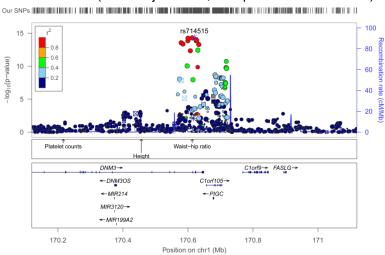
### SNX10 (Waist-Hip Ratio adj. for BMI, All Ancestries Women)



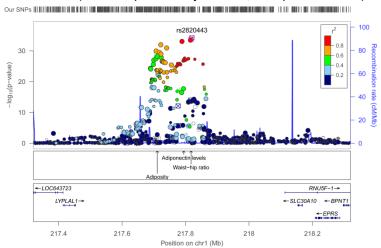
# TBX15-WARS2 (WHR adj. for BMI, European Sex-Combined)



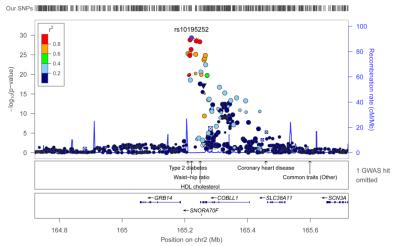
### DMN3-PIGC (WHR adj. for BMI, European Sex-Combined)



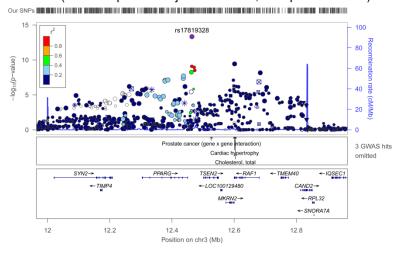
### LYPLAL1 (Waist-Hip Ratio adj. for BMI, European Women)



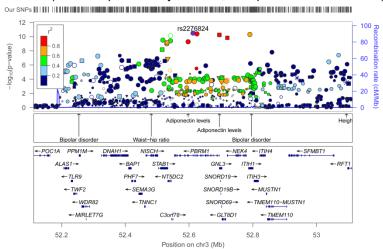
# GRB14-COBLL1 (WHR adj. for BMI, European Women)



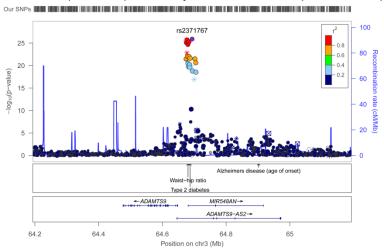
### PPARG (Waist-Hip Ratio adjusted for BMI, European Women)



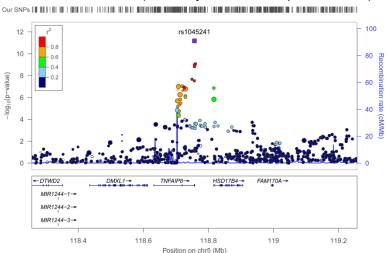
# PBRM1 (Waist-Hip Ratio adj. for BMI, European Sex-Combined)



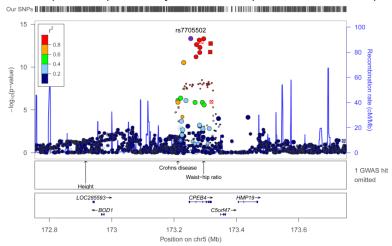
### ADAMTS9 (Waist-Hip Ratio adjusted for BMI, European Women)



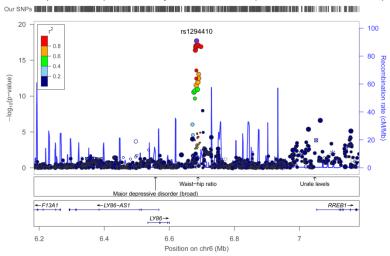
### TNFAIP3-HSD17B4 (WHR adj. for BMI, European Women)



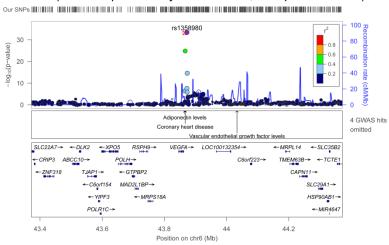
### CPEB4 (Waist-Hip Ratio adj. for BMI, European Sex-Combined)



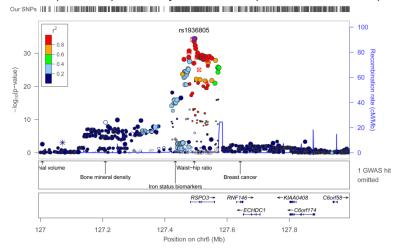
# LY86 (Waist-Hip Ratio adj. for BMI, European Sex-Combined)



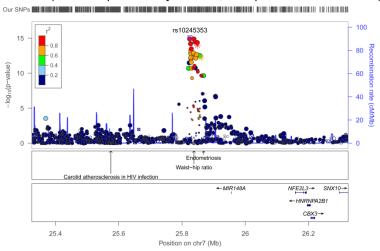
### VEGFA (Waist-Hip Ratio adjusted for BMI, European Women)



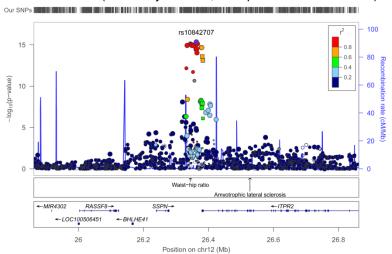
# RSPO3 (Waist-Hip Ratio adj. for BMI, European Sex-Combined)



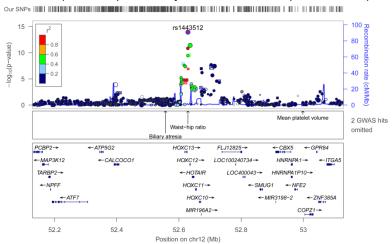
# NFE2L3 (Waist-Hip Ratio adj. for BMI, European Sex-Combined)



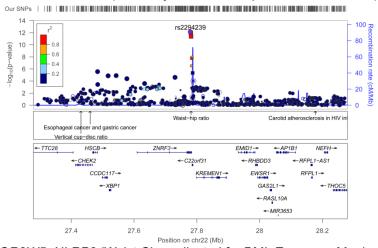
# ITPR2-SSPN (WHR adj. for BMI, European Sex-Combined)



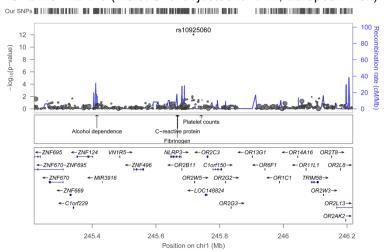
### HOXC13 (Waist-Hip Ratio adjusted for BMI, European Women)



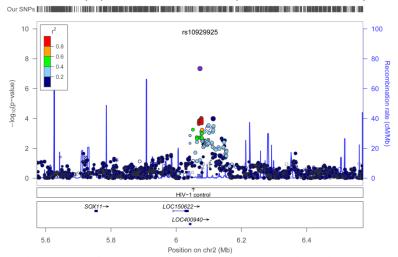
### ZNFR3-KREMEN1 (WHR adj. for BMI, European Sex-Combined)



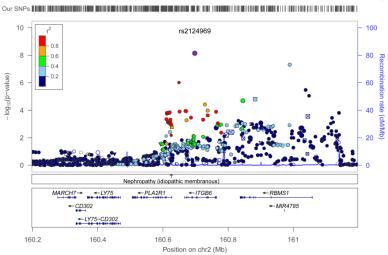
### OR2W5-NLRP3 (Waist Circ. adjusted for BMI, European Men)



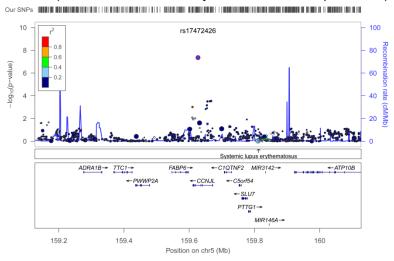
# SOX11 (Hip Circumference, European Sex-Combined)



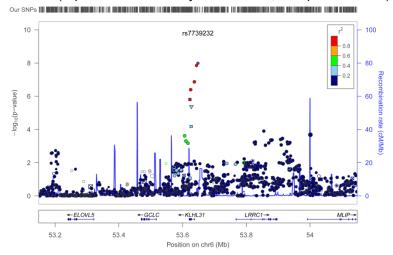
### ITGB6 (Waist Circ. adjusted for BMI, European Sex-Combined)



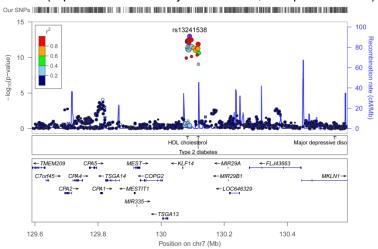
### CCNJL (Waist Circumference adjusted for BMI, European Men)



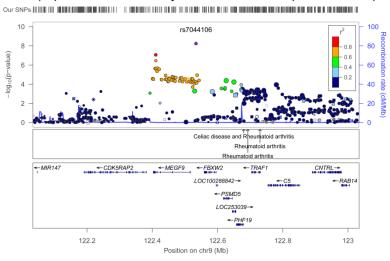
### KLHL31 (Hip Circumference adjusted for BMI, European Women)



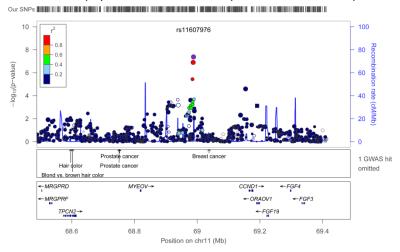
### KLF14 (Hip Circumference adjusted for BMI, European Women)



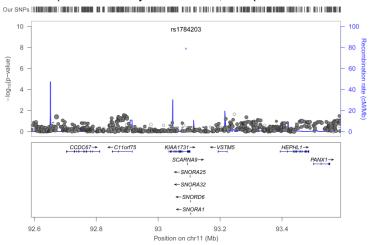
# C5 (Hip Circumference adjusted for BMI, European Women)



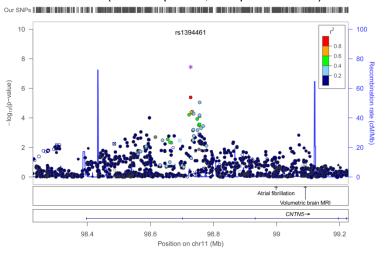
# MYEOV (Hip Circumference, European Sex-Combined)



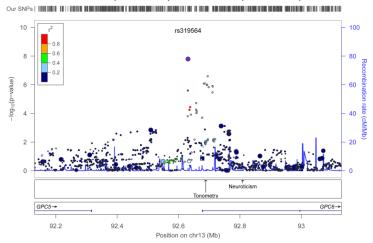
# KIAA1731 (Waist Circ. adjusted for BMI, European Sex-Combined)



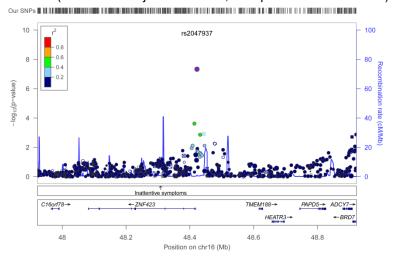
# CNTN5 (Waist-Hip Ratio, European Women)



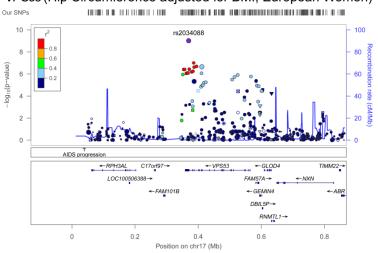
### GPC6 (Waist-Hip Ratio, European Men)



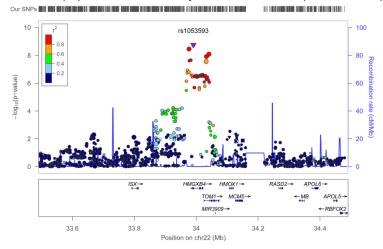
### ZNF423 (Waist Circ. adjusted for BMI, European Sex-Combined)



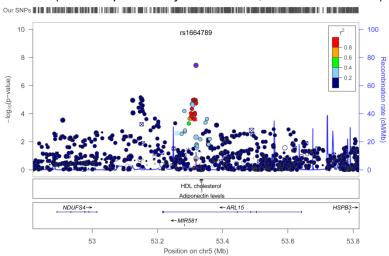
### VPS53 (Hip Circumference adjusted for BMI, European Women)



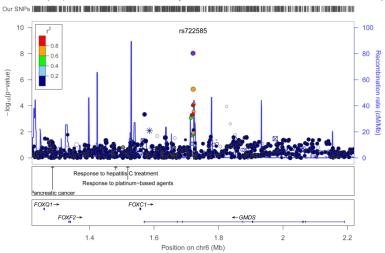
# HMGXB4 (Hip Circumference adjusted for BMI, European Women)



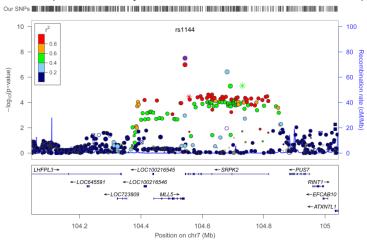
### ARL15 (Waist-Hip Ratio adjusted for BMI, All Ancestries Men)



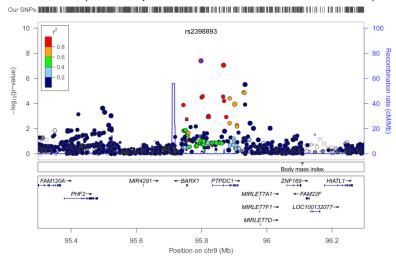
### GMDS (Hip Circumference adjusted for BMI, All Ancestries Men)



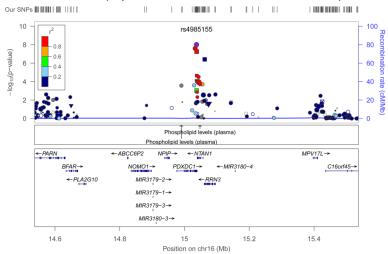
#### SRPK2 (Waist Circ. adj. for BMI, All Ancestries Sex-Combined)



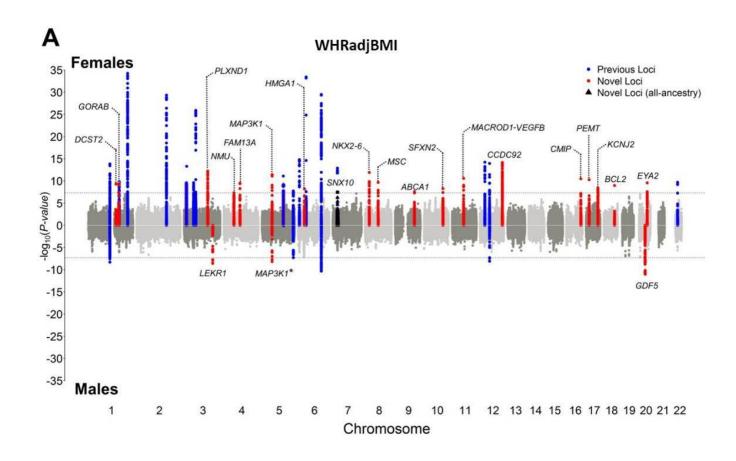
# PTPDC1 (Waist-Hip Ratio, All Ancestries Sex-Combined)

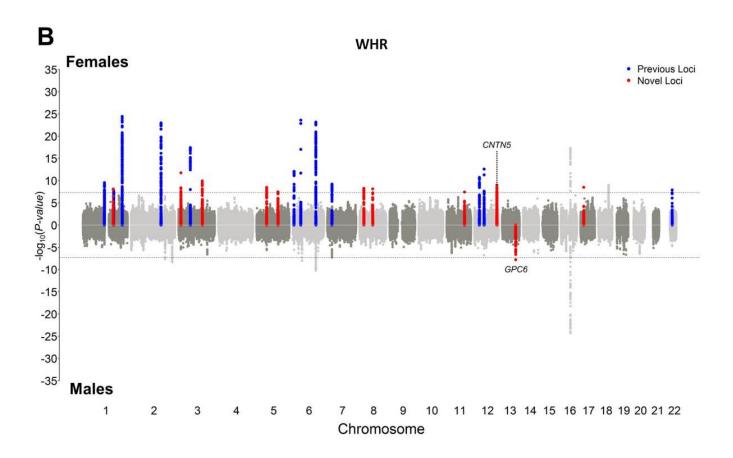


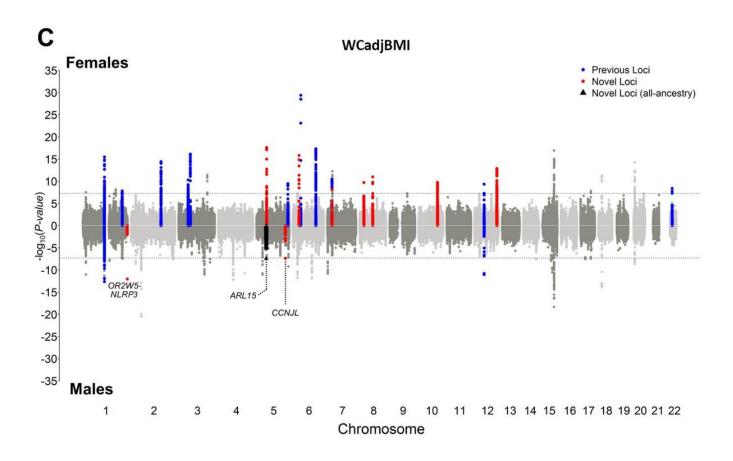
# PDXDC1 (Hip Circumference, All Ancestries Men)

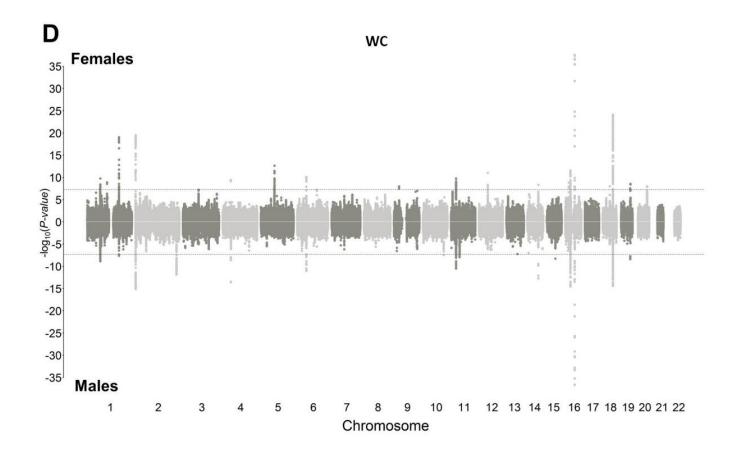


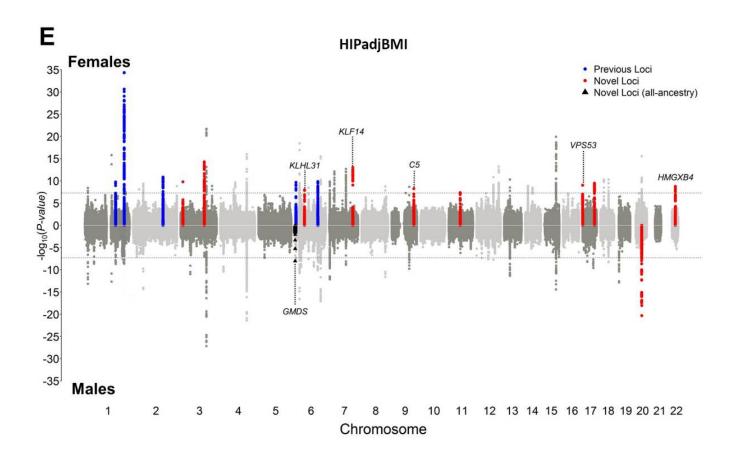
Supplementary Figure 4 | Chicago plots of sex-specific SNP associations for six waistrelated traits. Sex-specific SNP associations are shown for waist-hip ratio (WHR), waist circumference (WC), and hip circumference (HIP), with and without adjustment for body mass index (adiBMI). Associations in women are shown above the x-axis as -log<sub>10</sub>(P values), and associations in men are shown below as  $loq_{10}(P \text{ values})$ . Only SNP results with N > 50,000samples are shown. Dashed gray lines mark statistical significance at the genome-wide level (P = 5 × 10<sup>-8</sup>). Novel loci achieving genome-wide significance in sex-stratified WHR association analysis in Europeans are highlighted in red on all figures (A-F) and annotated on figure A. One additional novel locus achieving genome-wide significance when all ancestries were analyzed is marked as black triangles and annotated on figure A. Novel loci achieving genome-wide significance in Europeans in other waist-related traits (*B-F*) are highlighted in red and annotated only on the relevant figure. Previously established loci are highlighted in blue (A-F). Additional novel loci achieving genome-wide significance when all ancestries were analyzed in other waistrelated traits (B-F) are marked as black triangles and annotated. SNP association signals that achieve genome-wide significance and are previously established height or BMI loci are shown in light or dark grey. In figure A, the asterisk indicates that different lead SNPs were identified in women and men. Detailed information about the loci is presented in Tables 1 and 3 and Supplementary Tables 4 and 25.

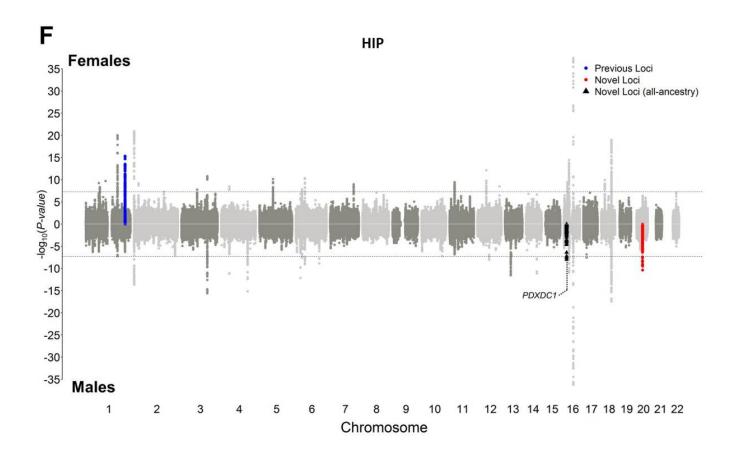






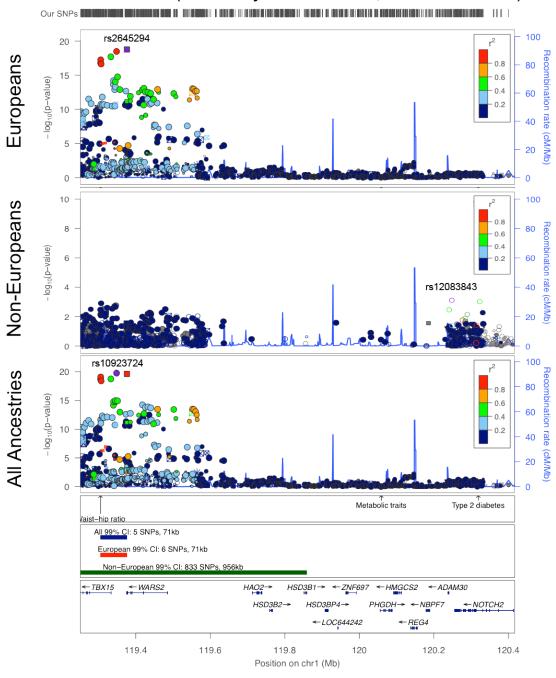




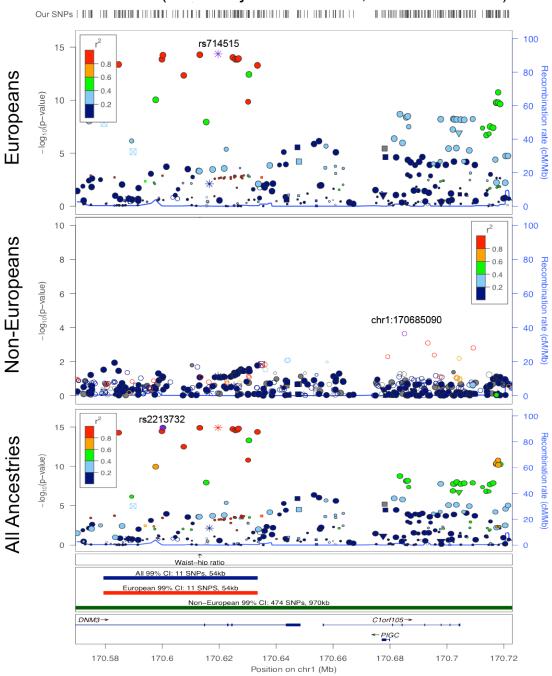


Supplementary Figure 5 | Regional association plots of WHRadjBMI signals covered with fine-mapping density on the Metabochip. Plots of 17 waist-hip ratio adjusted for body mass index (WHRadjBMI) signals from Table 1 are arranged in chromosomal order.

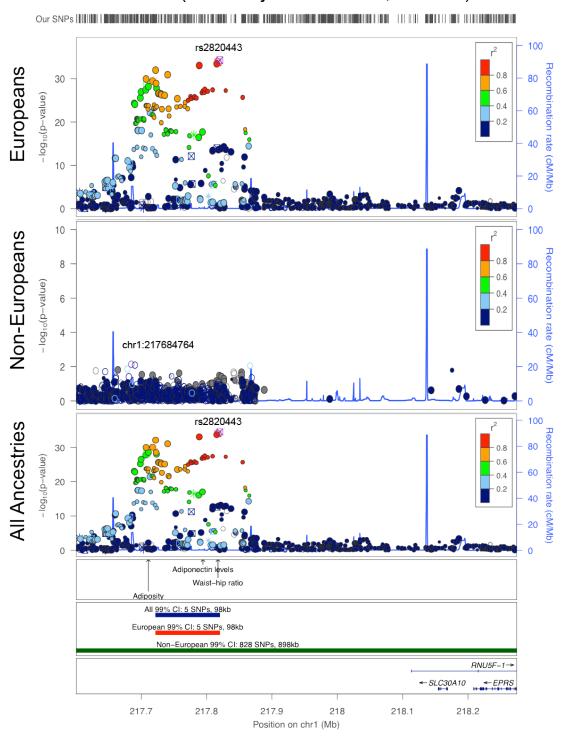
### TBX15-WARS2 (WHR adjusted for BMI, Sex-Combined)



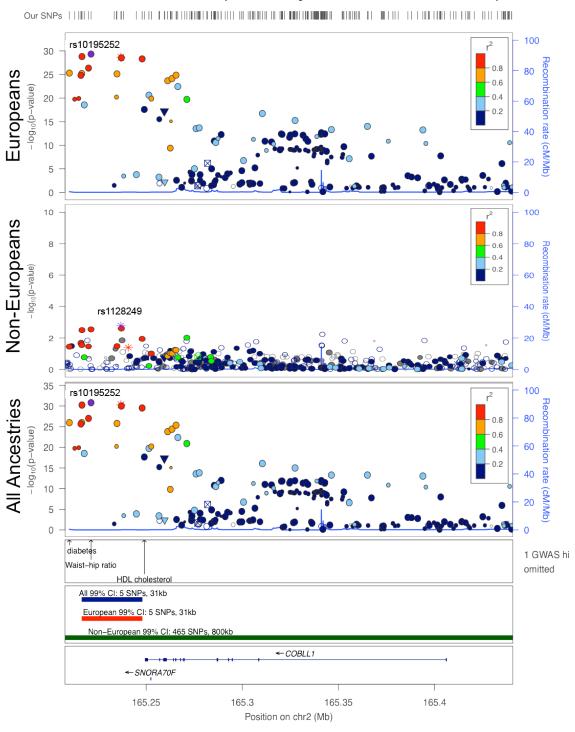
## DNM3-PIGC (WHR adjusted for BMI, Sex-Combined)



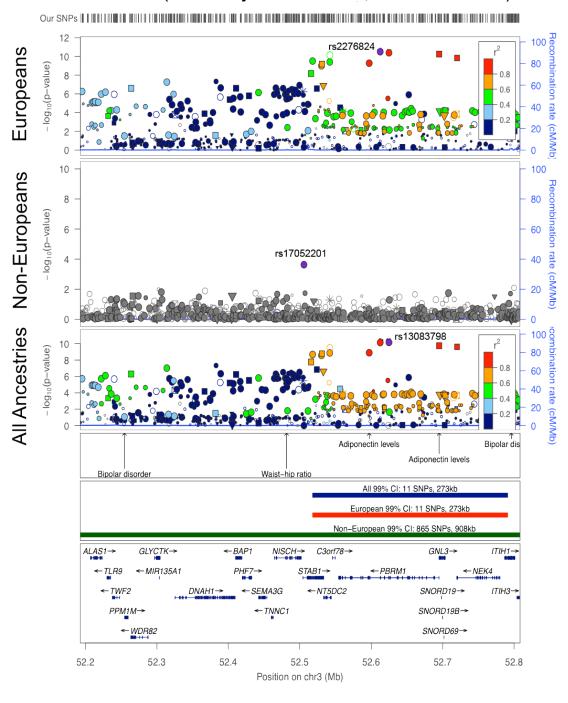
## LYPLAL1 (WHR adjusted for BMI, Women)



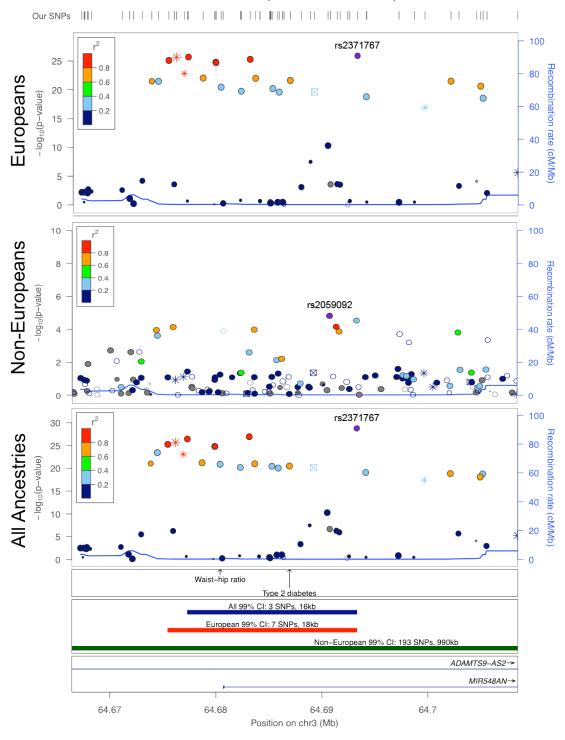
## GRB14-COBLL1 (WHR adjusted for BMI, Women)



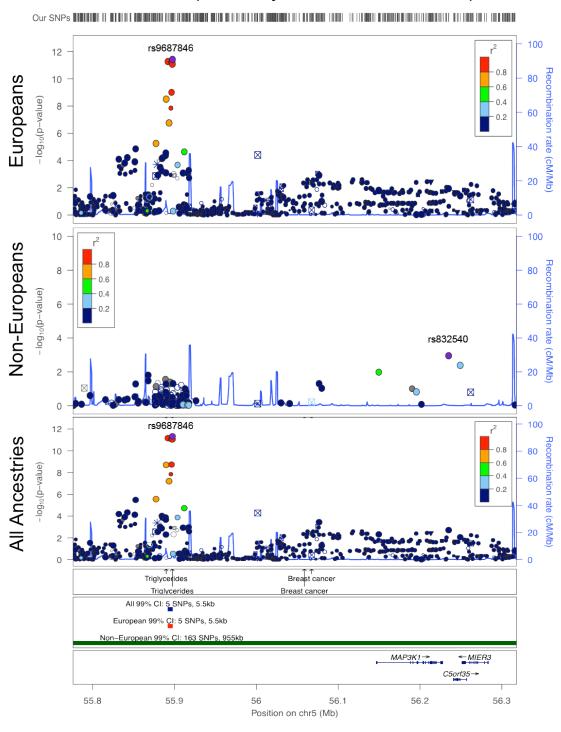
### PBRM1 (WHR adjusted for BMI, Sex-Combined)

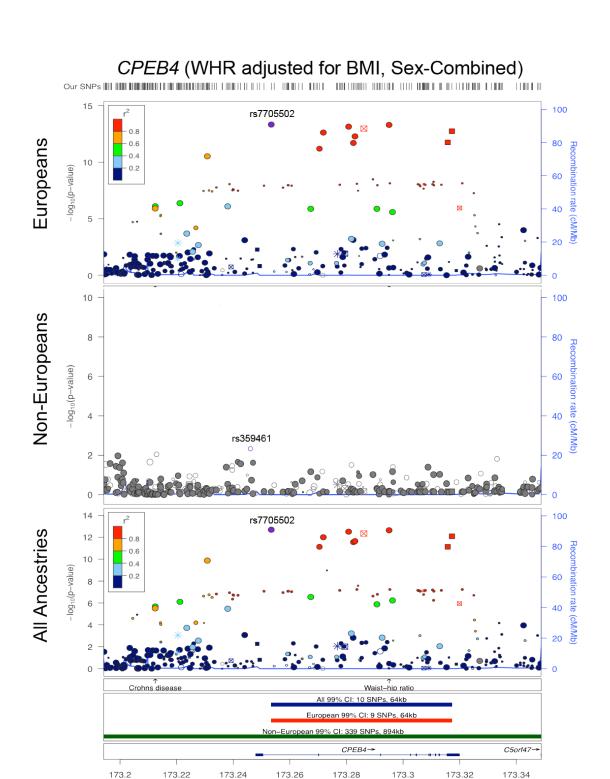


### ADAMTS9 (WHR, Women)



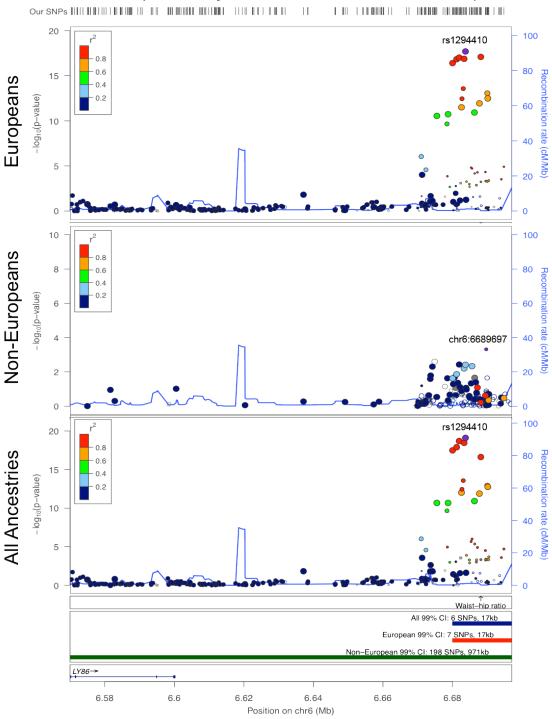
## MAP3K1 (WHR adjusted for BMI, Women)



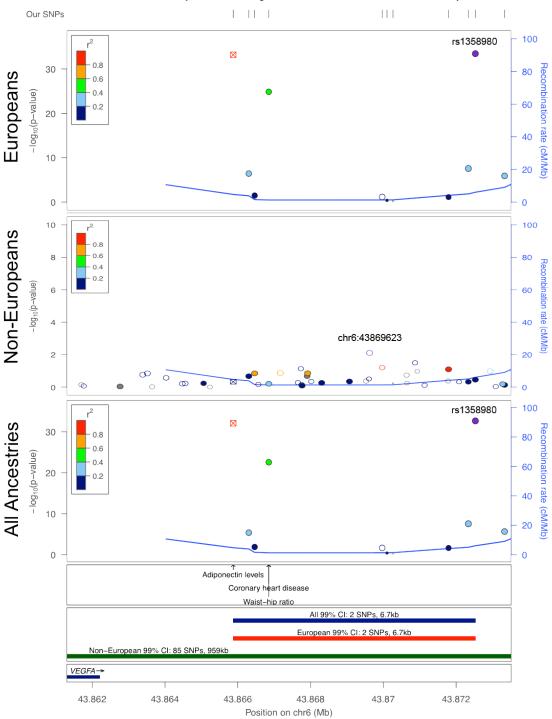


Position on chr5 (Mb)

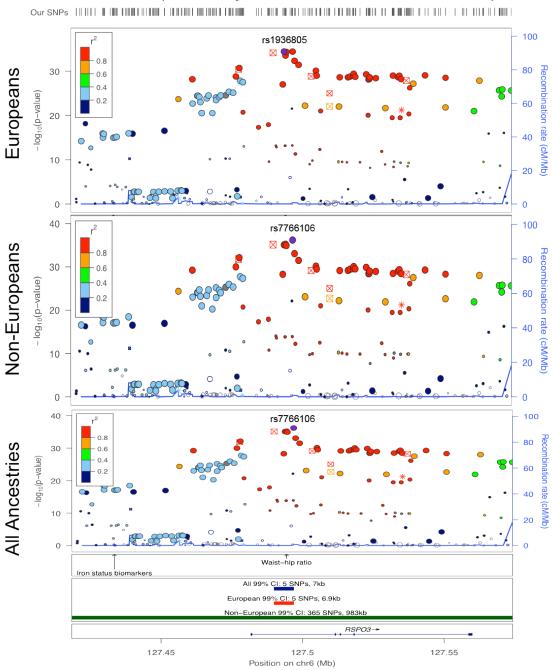
## LY86 (WHR adjusted for BMI, Sex-Combined)



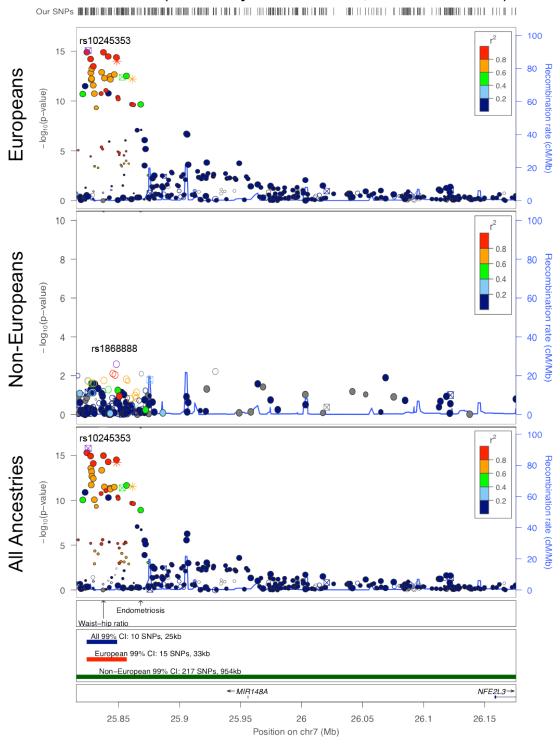
## VEGFA (WHR adjusted for BMI, Women)



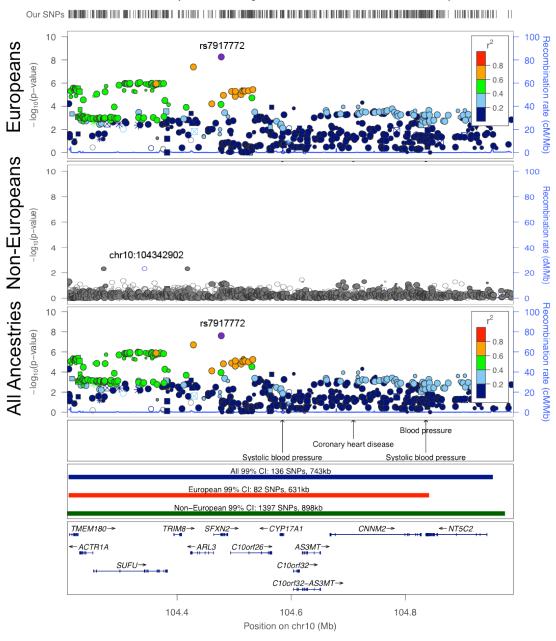
## RSPO3 (WHR adjusted for BMI, Sex-Combined)



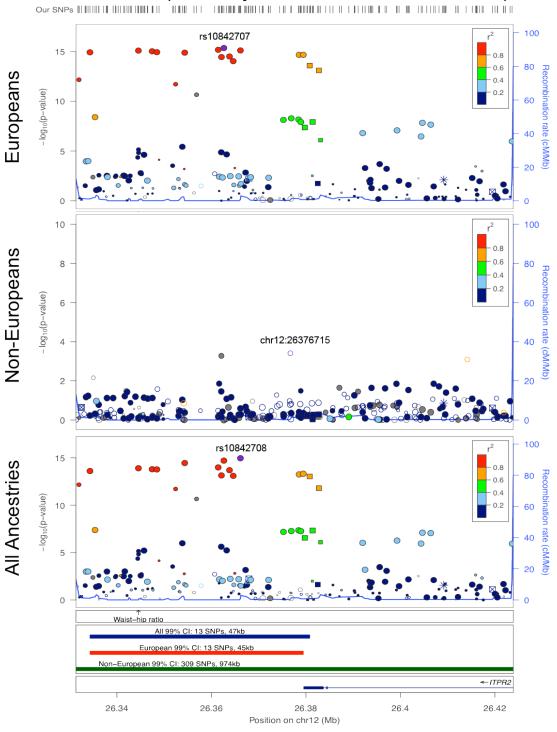
## NFE2L3 (WHR adjusted for BMI, Sex-Combined)

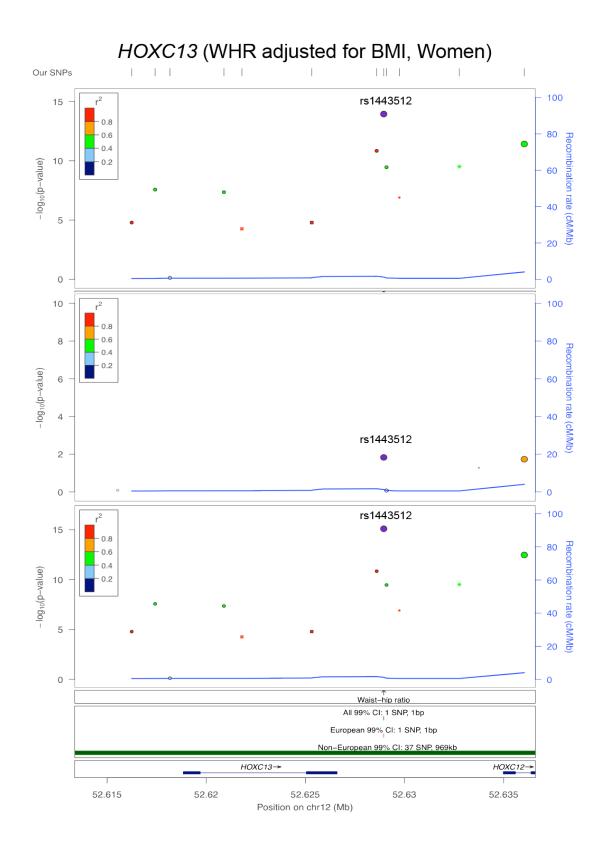


### SFXN2 (WHR adjusted for BMI, Women)

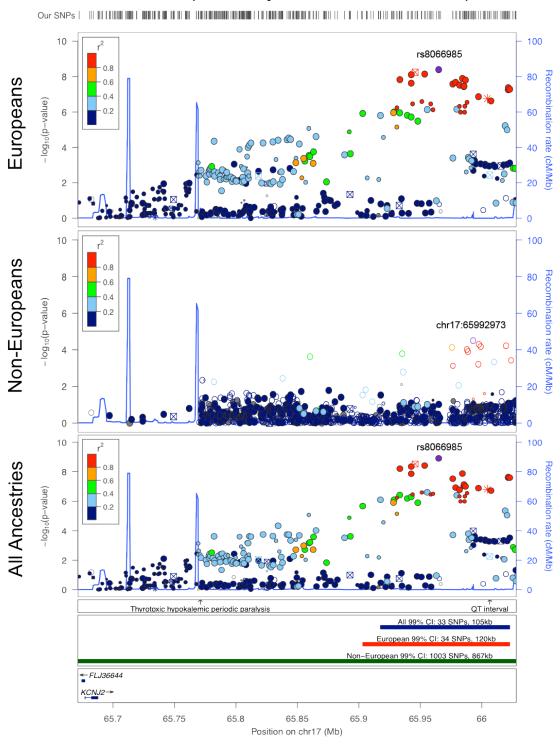


# ITPR2-SSPN (WHR adjusted for BMI, Sex-Combined)

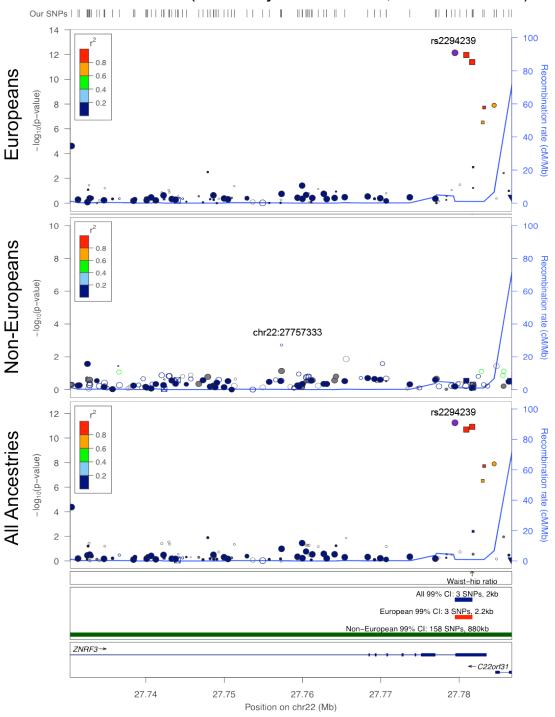




### KCNJ2 (WHR adjusted for BMI, Women)



## ZNFR3-KREMEN1 (WHR adjusted for BMI, Sex-Combined)



## **Supplementary Tables**

This document contains Supplementary Tables 5, 6, 7, 9, 10, 12, 13, 14, 17, 20, and 24.

Supplementary Table 5 | Estimated narrow-sense age-adjusted heritability  $(h^2)$  for waist-related traits, height, weight, and body mass index

			Wo	men						Men			Sex D	fference P
	Framir	ngham He	eart Study		TWINGEN	E	Fram	ingham He	art Study		TWINGENE		FHS	TWINGENE
Traits	n	h² (SE)	Р	n	<i>h</i> <sup>2</sup> (95% CI)	P	n	h² (SE)	P	n	<i>h</i> ² (95% CI)	P	P	<i>P</i> value
WHRadjBMI	1,595	0.463 (0.10)	6.00E-07	6,471	0.558 (0.441, 0.595)	1.46E-14	1,446	0.188 (0.12)	5.11E-02	5,362	0.317 (0.145, 0.381)	8.23E-04	3.67E-03	1.04E-03
WHR	1,599	0.388 (0.10)	4.39E-05	6,704	0.577 (0.488, 0.612)	3.15E-18	1,452	0.315 (0.11)	1.22E-03	5,541	0.473 (0.289, 0.524)	5.10E-07	1.87E-01	4.30E-02
ВМІ	4,177	0.492 (0.04)	8.50E-47	6,515	0.702 (0.576, 0.730)	2.83E-93	3,528	0.586 (0.04)	1.56E-48	5,446	0.662 (0.595, 0.701)	4.65E-32	3.88E-01	4.28E-01
Height	4,181	0.999 (0.03)	1.25E-193	6,515	0.908 (0.811, 0.921)	2.31E-105	3,528	0.965 (0.03)	3.11E-136	5,446	0.885 (0.610, 0.901)	2.11E-57	3.23E-03	7.00E-03
Weight	4,177	0.550 (0.04)	9.56E-60	6,515	0.718 (0.688, 0.745)	3.14E-169	3,528	0.596 (0.04)	3.81E-51	5,446	0.711 (0.674, 0.744)	1.99E-118	5.64E-02	5.68E-01
WC	3,615	0.473 (0.04)	2.36E-38	6,738	0.661 (0.627, 0.693)	3.56E-28	3,233	0.549 (0.05)	8.62E-40	5,608	0.651 (0.601, 0.690)	4.60E-23	6.95E-02	6.79E-01
WCadjBMI	3,610	0.505 (0.04)	5.20E-43	6,496	0.595 (0.363, 0.633)	5.45E-13	3,227	0.572 (0.05)	3.39E-41	5,420	0.359 (0.195, 0.419)	5.59E-05	3.86E-02	1.82E-10
HIP	1,601	0.447 (0.09)	3.00E-07	6,718	0.675 (0.595, 0.706)	3.50E-32	1,455	0.416 (0.10)	1.10E-05	5,553	0.602 (0.521, 0.645)	2.60E-18	1.20E-02	8.49E-03
HIPadjBMI	1,597	0.495 (0.09)	6.37E-09	6,476	0.592 (0.519, 0.631)	2.61E-20	1,449	0.528 (0.10)	2.52E-08	5,369	0.451 (0.251, 0.505)	1.36E-06	9.05E-01	6.71E-05

Narrow-sense heritability estimated separately from the Framingham Heart Study and the TWINGENE study. SE, standard error; CI, confidence interval; WHR, waist-to-hip ratio; WC, waist circumference; HIP, hip circumference; adjBMI, adjusted for body mass index.

Supplementary Table 6 | Variance in WHRadjBMI explained by all current loci compared to only previously established loci

Loci Source	Sex-combined $R^2$	Male-specific $R^2$	Female-specific $R^2$	Data set	Phenotype Transformation
14 Heid <i>et al.</i> loci	1.03% <sup>a</sup>	0.46% <sup>a</sup>	1.34% <sup>a</sup>	Heid <i>et al.</i> (Follow-up studies)	Raw
14 Heid <i>et al.</i> loci	0.60% <sup>b</sup>	0.31% <sup>b</sup>	1.03% <sup>b</sup>	Heid <i>et al.</i> (Follow-up studies)	Inverse-normalized
2 Randall <i>et al.</i> loci (additional to Heid <i>et al.</i> )	0.05% <sup>c</sup>	0.02% <sup>c</sup>	0.12% <sup>c</sup>	Randall <i>et al.</i> (Follow-up studies)	Inverse-normalized
16 previously reported loci (14 Heid <i>et al.</i> + 2 Randall <i>et al.</i> )	0.66% <sup>d</sup>	0.28% <sup>d</sup>	1.27% <sup>d</sup>	This paper; all studies	Inverse-normalized
49 current loci	1.36% <sup>e</sup>	0.82% <sup>e</sup>	2.40% <sup>e</sup>	This paper; all studies	Inverse-normalized

The variance explained by all waist-hip ratio adjusted for body mass index (WHRadjBMI) loci reported in this paper compared to WHRadjBMI loci in two previous reports. <sup>a</sup>The 14 previously reported loci from Heid *et al.* [PMID: 20935629] as presented there based on meta-analyzed beta-estimates of follow-up studies (*N* > 110,000) using "raw" WHRadjBMI (i.e., residuals of WHR adjusted for BMI, age, age<sup>2</sup>, and study-specific covariates, stratified by sex but not inverse normal transformed) using a model variance of that trait as observed in the population-based cross-sectional study KORA S3 (*N* = 3,996). <sup>b</sup>The 14 previously reported loci from Heid *et al.* [PMID: 20935629] using the same data as in (a) using the meta-analyzed beta estimates of the inverse normal transformed values of WHRadjBMI (thus Var(Y)=1). <sup>c</sup>The two additional reported loci from Randall *et al.* [PMID:23754948] (*PPARG* and *HSD17B4*) based on meta-analyzed beta-estimates of follow-up studies (*N* > 105,000) using inverse normal transformed values of WHRadjBMI (thus Var(Y)=1). <sup>d</sup>The 16 previously reported loci using the here presented data (*N* > 210,000) based on meta-analyzed beta-estimates of the inverse normal transformed values of WHRadjBMI (thus Var(Y)=1). <sup>e</sup>The 49 reported loci using the same data as in (c) also on inverse normal transformed values of WHRadjBMI (as in (c)).

Supplementary Table 7 | 99% credible intervals for fine-mapped waist-related loci

						Europe	eans only	Non-E	uropeans	All an	cestries
Trait	Analysis	Gene	Index SNP	Chr	Position (bp) NCBI build 36	# of SNPs	Distance (bp)	# of SNPs	Distance (bp)	# of SNPs	Distance (bp)
WHRadjBMI	Overall	TBX15- WARS2	rs2645294	1	119,376,110	6	70,745	833	956,454	5	70,745
WHRadjBMI	Overall	DNM3-PIGC	rs714515	1	170,619,613	11	54,038	474	969,517	11	54,038
WHRadjBMI	Overall	PBRM1	rs2276824	3	52,612,526	11	272,987	865	908,219	11	272,987
WHRadjBMI	Overall	CPEB4	rs7705502	5	173,253,421	9	63,849	339	894,027	10	63,849
WHRadjBMI	Overall	LY86	rs1294410	6	6,683,751	7	17,038	198	971,095	6	17,038
WHRadjBMI	Overall	RSPO3	rs1936805	6	127,493,809	5	6,890	365	982,689	5	6,890
WHRadjBMI	Overall	NFE2L3	rs10245353	7	25,825,139	15	32,536	217	954,173	10	24,650
WHRadjBMI	Overall	ITPR2-SSPN	rs10842707	12	26,362,631	13	45,153	309	974,009	13	46,515
WHRadjBMI	Overall	ZNRF3- KREMEN1	rs2294239	22	27,779,477	3	2,195	158	879,614	3	2,195
WHRadjBMI	Women	LYPLAL1	rs2820443	1	217,820,132	5	98,141	828	897,911	5	98,141
WHRadjBMI	Women	COBLL1	rs10195252	2	165,221,337	5	31,273	465	800,236	5	31,273
WHRadjBMI	Women	ADAMTS9	rs2371767	3	64,693,298	7	17,834	193	989,524	3	15,984
WHRadjBMI	Women	MAP3K1	rs9687846	5	55,897,651	5	5,520	163	954,728	5	5,520
WHRadjBMI	Women	VEGFA	rs1358980	6	43,872,529	2	6,656	85	959,317	2	6,656
WHRadjBMI	Women	SFXN2	rs7917772	10	104,477,433	82	631,096	1397	898,492	136	742,751
WHRadjBMI	Women	HOXC13	rs1443512	12	52,628,951	1	1	37	969,387	1	1
WHRadjBMI	Women	KCNJ2	rs8066985	17	65,964,940	34	119,579	1003	866,939	33	105,152
HIPadjBMI	Women	KLF14	rs13241538	7	130,090,402	22	34,374	130	968,985	22	34,374

Fine-mapping analysis was performed at loci with high-density coverage on the Metabochip, which included 17 waist-hip ratio adjusted for body mass index (WHRadjBMI) loci and one hip circumference adjusted for body mass index (HIPadjBMI) locus. Association summary statistics were used to define credible sets of variants with a high probability of containing the likely functional variant (see Methods). The number of SNPs in the credible sets and the distance in kilobases spanned by those variants are shown. The Metabochip does not include all known SNPs. Previously reported genome-wide associated loci from the Heid *et al.* (2010) paper are marked in bold.

# Supplementary Table 9 | Genome-wide significant associations of waist-related SNPs with metabolic and anthropometric traits

Trait	WHRadjBMI SNPs with look-up P < 5 × 10 <sup>-8</sup>	Corresponding WHRadjBMI locus names	Other waist and hip trait SNPs with look-up $P < 5 \times 10^{-8}$	Corresponding other waist and hip trait locus names
Type 2 diabetes (T2D)	1	GRB14-COBLL1	0	-
Fasting glucose (FG)	0	-	0	-
Fasting insulin adjusted for BMI (FladjBMI)	2	LYPLAL1,GRB14- COBLL1	0	-
2-hour glucose (G120)	0	-	0	-
Diastolic blood pressure (DBP)	0	-	0	-
Systolic blood pressure (SBP)	0	-	0	-
Body mass index (BMI)	0	-	0	-
Height	7	FGFR4, HMGA1, SFXN2, SMAD6, BMP2, GDF5, NFE2L3 FAM13A,	0	-
High-density lipoprotein cholesterol (HDL-C)	7	MAP3K1, CCDC92, CMIP, GRB14-COBLL1, VEGFA, RSPO3	1	KLF14
Low-density lipoprotein cholesterol (LDL-C)	2	GRB14-COBLL1, PPARG MAP3K1,	0	-
Triglycerides (TG)	5	CCDC92, GRB14-COBLL1, PPARG, VEGFA	1	KLF14
Adiponectin adjusted for BMI	3	CCDC92, CMIP, PBRM1	0	-
Nephropathy (in Chinese subjects)	0	-	0	-
Nephropathy (in Italian subjects)	0	-	0	-
Estimated glomerular filtration rate of creatinine (eGFRcrea)	0	-	0	-
Chronic kidney disease (CKD)	0	-	0	-
Urine albumin-to-creatinine ratio (UACR)	0	-	0	-
Endometriosis	0	-	0	-
Coronary artery disease (CAD)	0	-	0	-
Menopause	0	-	0	-
Menarche	0	-	0	-
Femoral neck bone mineral density (FN-BMD)	0	-	0	-

Trait	WHRadjBMI SNPs with look-up P < 5 × 10 <sup>-8</sup>	Corresponding WHRadjBMI locus names	Other waist and hip trait SNPs with look-up P < 5 × 10 <sup>-8</sup>	Corresponding other waist and hip trait locus names
Lumbar spine bone mineral density (LS-BMD)	0	-	0	-

This table summarizes the cross-trait association data as provided by other consortia. Waist-hip ratio adjusted for body mass index (WHRadjBMI) SNPs and locus names refer to SNPs presented on **Table 1**. Other waist and hip trait SNPs refer to the SNPs presented on **Table 3**. *KLF14* is a hip circumference adjusted for body mass index (HIPadjBMI) locus.

Supplementary Table 10 | Joint associations of 39 WHRadjBMI SNPs with metabolic and anthropometric traits

Trait	# of SNPs	β	SE	P
Type 2 diabetes (T2D)	39	0.502	0.077	6.3E-11
Fasting glucose (FG)	39	0.061	0.014	6.1E-06
Fasting insulin adjusted for BMI (FladjBMI)	39	0.226	0.013	1.7E-64
2-hour glucose (G120)	39	0.221	0.079	5.0E-03
Diastolic blood pressure (DBP)	39	1.450	0.405	3.6E-04
Systolic blood pressure (SBP)	39	2.470	0.639	1.1E-04
Body mass index (BMI)	39	-0.217	0.020	2.0E-28
Height	39	-0.010	0.019	0.62
High-density lipoprotein cholesterol (HDL-C)	39	-0.356	0.023	1.1E-55
Low-density lipoprotein cholesterol (LDL-C)	39	0.157	0.024	1.1E-10
Total cholesterol	39	0.131	0.024	4.7E-08
Triglycerides (TG)	39	0.366	0.022	2.7E-65
Adiponectin	39	-0.351	0.029	1.5E-34
Endometriosis	36	-0.284	0.291	0.33
Nephropathy (in Chinese subjects)	35	0.192	0.445	0.67
Nephropathy (in Italian subjects)	34	0.264	0.398	0.51
Estimated glomerular filtration rate of creatinine (eGFRcrea)	39	0.010	0.007	0.14
Chronic kidney disease (CKD)	39	-0.122	0.135	0.37
Urine albumin-to-creatinine ratio (UACR)	39	-0.026	0.059	0.67
Menopause	39	0.211	0.193	0.28
Menarche	39	0.030	0.048	0.53
Coronary artery disease (CAD)	38	0.041	0.058	0.48
Femoral neck bone mineral density (FN-BMD)	39	-0.002	0.053	0.98
Lumbar spine bone mineral density (LS-BMD)	39	-0.011	0.057	0.85

This table shows the results of a meta-regression of beta ( $\beta$ ) estimates of the 39 waist-hip ratio adjusted for body mass index (WHRadjBMI)-increasing alleles that exhibit genome-wide significant association in the sex-combined analysis with beta estimates of metabolic traits from other consortia (DIAGRAM, MAGIC, ICBP, GLGC, ADIPOgen, IEC, IgAN-Chinese, IgAN-Italian, CKDgen, ReproGEN and CARDIOGRAM). Significant associations (P < 0.05 / 24) are marked in bold.

#### **Supplementary Table 12 | Candidate functional nonsynonymous variants**

Trait	Index SNP	Proxy SNP	r²	Effect allele	Non-effect allele	Distance (bp)	Coding impact	Gene	Gene/transcript annotation
WHRadjBMI	rs224333	rs224331	0.96	Α	С	1,575	nonsynonymous	GDF5	NM_000557p.S276A
HIPadjBMI	rs1053593	rs1053593	-	G	Т	0	nonsynonymous	HMGXB4	NM_001003681:p.G165V
WCadjBMI	rs1664789	rs1135999	0.89	Α	G	2,503	nonsynonymous	NTAN1	NM_173474:p.S287P
WCadjBMI	rs1664789	rs1136001	0.89	G	Т	2,515	nonsynonymous	NTAN1	NM_173474p.H283N

Variants identified in 1000G CEU individuals annotated as nonsynonymous and in linkage disequilibrium ( $r^2$ ) with index SNPs. WHR, waist-to-hip ratio; WC, waist circumference; HIP, hip circumference; adjBMI, adjusted for body mass index.

Supplementary Table 13 | CNV-tagging variants associated with waist traits in sex-combined European-ancestry metaanalyses

Trait	CNV- tagging SNP	Chr	Position (bp)	Nearby Gene	CNV name	SNP-trait association sex-combined <i>P</i> <sup>a</sup>	Nearest index SNP from waist trait association results	r <sup>2</sup> between CNV tagging SNP and index SNP <sup>b</sup>
WHRadjBMI	rs1294421	6	6,688,148	LY86	CNVR2760.1	8.16E-18	rs1294410	0.83
WHRadjBMI	rs3733034	3	52,927,473	SFMBT1	RR_CNV_269	1.75E-06	rs2276824	0.11
WHRadjBMI	rs1150753	6	32,167,845	TNXB	CNVR2843.1	4.45E-06	rs7759742	-
HIPadjBMI	rs1543302	22	33,976,861	HMGXB4	CNVR8147.1	1.13E-07	rs1053593	0.86

A total of 6,200 copy number variant (CNV)-tagging SNPs were looked up in sex-combined SNP association results with waist traits. Positions are shown in NCBI build 36. <sup>a</sup>The SNP-trait P value achieving genome-wide significance is shown in **bold** and was described previously in Heid *et al.* (2010). The other P values are significant at P < 0.05 / 6,200. <sup>b</sup>Linkage disequilbirium (LD)  $r^2$  values were based on 1000G Pilot1 CEU dataset. CNVs in the *SFMBT1* and *TNXB* gene regions are in low LD with index SNPs from main results and may represent independent or partially independent signals from these regions. Waist-hip ratio adjusted for body mass index (WHRadjBMI); hip circumference adjusted for body mass index (HIPadjBMI).

### Supplementary Table 14 | Candidate variants at transcription start sites

217		<b>-</b>	Distance to	
SNP	Chr	Position (bp)	nearest TSS (bp)	Nearest transcript
New WHRadjBMI Lo				
rs62310884	4	56,458,400	-21	PDCL2
rs5007262	6	32,379,011	500	BTNL2
rs5007261	6	32,379,031	480	BTNL2
rs5007260	6	32,379,047	464	BTNL2
rs5007259	6	32,379,101	410	BTNL2
rs5007258	6	32,379,239	272	BTNL2
rs6906730	6	32,379,445	66	BTNL2
rs6911383	6	32,379,682	-171	BTNL2
rs7801581	7	27,223,771	-365	HOXA11-AS
rs17427875	7	27,225,558	406	HOXA11-AS
rs1550279	8	23,600,854	-299	RP11-213G6.2
rs11680316	2	188,135,298	233	U6
rs11057396	12	124,419,062	469	DNAH10OS
rs11057397	12	124,419,728	-197	DNAH10OS
rs3186071	12	124,429,279	-363	CCDC92
rs11835839	12	124,431,049	271	CCDC92
rs34180676	12	124,431,519	-199	CCDC92
rs7966192	12	124,431,701	-381	CCDC92
rs12823740	12	124,458,002	161	ZNF664
rs1075403	19	18,384,950	369	KIAA1683
rs7247222	19	18,392,873	-441	JUND
rs6088816	20	33,982,435	-461	UQCC
rs6142379	20	33,999,267	499	UQCC
rs143384	20	34,025,756	267	GDF5
rs143383	20	34,025,983	40	GDF5
Previously Establish	ned WHRadjBN	//I Loci		
rs4846302	1	219,730,586	53	RP11-95P13.2
rs4846303	1	219,730,799	266	RP11-95P13.2
rs6753142	2	165,544,071	216	SNORA70F
rs6738627	2	165,544,450	-163	SNORA70F
chr2:165544572	2	165,544,572	-285	SNORA70F
chr2:165544573	2	165,544,573	-286	SNORA70F
rs10933	3	52,719,816	36	PBRM1
rs1108842	3	52,720,080	49	GNL3
rs2302417	3	52,814,256	320	ITIH1
chr5:173315416	5	173,315,416	134	CPEB4
rs55946741	5	173,345,023	-195	CPEB4
rs2800709	6	127,439,297	-193 -451	RSPO3
197000109	Ü	127,439,297	- <del>4</del> 31	NOFUS

			Distance to	
SNP	Chr	Position (bp)	nearest TSS (bp)	Nearest transcript
rs4963979	12	26,468,964	317	RP11-612B6.1
rs10842705	12	26,469,016	369	RP11-612B6.1
rs7132434	12	26,472,562	91	RP11-283G6.5
rs1049380	12	26,489,544	339	RP11-612B6.2
chr12:54348554	12	54,348,554	-63	HOXC12
rs10876529	12	54,421,810	-331	HOXC6
chr12:54422043	12	54,422,043	-98	HOXC6
rs2071449	12	54,428,011	278	MIR615
Secondary Signals				
rs4282054	3	52,566,065	465	NT5DC2
rs7636227	3	52,566,682	-152	NT5DC2
rs7614981	3	52,566,914	-384	NT5DC2
rs12489828	3	52,567,014	-484	NT5DC2
rs66782572	3	52,567,617	176	NT5DC2
rs7639267	3	52,568,805	265	C3orf78

Variants at novel and previously known waist-hip ratio adjusted for body mass index (WHRadjBMI)-associated loci located within 500 bp of the nearest transcription start (TSS) site are displayed. Negative distance from nearest GENCODE v12 TSS indicates the variant is 5' of the TSS.

#### Supplementary Table 17 | Sources of data sets used for regulatory annotation

Sample	Tissue	DNase	H3K4me1	H3K27ac	H3K4me3	H3K9ac	FAIRE	H3K4me2	TF
Adipose Nuclei	Adipose	-	IDR	MACS2	IDR	IDR	-	-	-
Anterior Caudate	Brain	-	IDR	MACS2	IDR	MACS2	-	-	-
Mid Frontal Lobe	Brain	-	IDR	MACS2	IDR	MACS2	-	-	-
Substantia Nigra	Brain	-	IDR	-	IDR	MACS2	-	-	-
Adult Liver	Liver	-	IDR	-	IDR	IDR	-	-	-
Skeletal Muscle	Muscle	-	IDR	MACS2	IDR	IDR	-	-	-
Pancreatic Islet	Pancreatic Islet	-	MACS2	-	MACS2	MACS2	-	-	-

Sample	Tissue	DNase	H3K4me1	H3K27ac	H3K4me3	H3K9ac	FAIRE	H3K4me2	TF
GM12878	Blood	Integrative	Integrative	Integrative	Integrative (2)	Integrative	Integrative	Integrative	Integrative (75)
Osteoblasts	Bone	Integrative	Integrative	Integrative	Original	-	-	Integrative	Integrative (1)
Astrocytes	Brain	Integrative	Original	Integrative	Integrative	-	Integrative	-	Integrative (1)
Cerebellum	Brain	Original	-	-	-	-	-	-	-
Cerebrum Frontal	Brain	Original	-	-	-	-	-	-	-
Frontal Cortex	Brain	Original	-	-	-	-	Integrative		-
HUVEC	Endothelial	Integrative	Integrative	Integrative	Integrative (2)	Integrative	Integrative	Integrative	Integrative (11)
HepG2	Liver	Integrative	Integrative	Integrative	Integrative (2)	Integrative	Integrative	Integrative	Integrative (61)
Hepatocytes	Liver	Integrative	-	-	-	-	-	-	-
Huh-7	Liver	Integrative	-	-	-	-	-	-	-
Myocyte	Muscle	Integrative	Integrative	Integrative	Integrative	Integrative	-	Integrative	Integrative (1)
PSOAS Muscle	Muscle	Original	-	-	-	-	-	-	-
Pancreatic Islet	Pancreatic Islet	Integrative	-	-	_	-	Integrative	-	=

Tables indicate the source of regulatory data used to annotate associated SNPs. Peaks were identified in Roadmap Epigenomics data using Irreproducible Discovery Rate methods (IDR; multiple replicates) or MACS2 alone (single replicate). For ENCODE data, peak calls were obtained from the Integrative analysis (when available) or the original analyses. Numbers in parentheses indicate the number of datasets when more than one is available. TF, Transcription Factor Binding; -, Dataset not available.

# Supplementary Table 20 | Significant gene sets based on the WHRadjBMI sex-combined GWA analysis results using the MAGENTA gene set enrichment method

		Effective gene set	# of expected genes (> 95 <sup>th</sup> percentile	# of observed genes (> 95 <sup>th</sup> percentile		False- discovery	
Database	Gene set	size	cutoff)	cutoff)	P value	rate	Nominally significant genes
Ingenuity	VEGF Signaling	15	1	6	0.0001	0.0006	SHC1, RAF1, BAD, BCL2, PXN,PTK2B
Ingenuity	PTEN Signaling	22	1	7	0.0002	0.0008	SHC1, RAF1, BAD, CDKN1B, PTEN, BCL2, BCL2L11
Ingenuity	ERK MAPK Signaling	22	1	6	0.0006	0.0076	SHC1, RAF1, PPARG, BAD, PXN, PTK2B
Ingenuity	Neuregulin Signaling	25	1	6	0.0007	0.0082	SHC1, RAF1, ADAM17, BAD, CDKN1B, PTEN
Ingenuity	BMP Signaling pathway	24	1	6	0.0009	0.0097	HOXC9, SMAD6, RAF1, ZNF423, SMAD7, NOG
Ingenuity	PI3K AKT Signaling	27	1	6	0.0015	0.0117	SHC1, RAF1, BAD, CDKN1B, PTEN, BCL2
Ingenuity	Ceramide Signaling	15	1	4	0.0051	0.0148	RAF1, BAD, DIABLO, BCL2, CNKSR1
Ingenuity	RAR Activation	46	2	8	0.0014	0.0153	VEGFA, NSD1, IGFBP3, PML, RELA, PTEN, RAC1, PSMC5
Ingenuity	Insulin Receptor Signaling	31	2	6	0.0035	0.0208	SHC1, RAF1, BAD, SOCS3, INSR, PTEN
Ingenuity	PPAR Signaling	17	1	4	0.0082	0.0265	SHC1, RAF1, PPARG, INSR
Ingenuity	IGF-1 Signaling	18	1	4	0.0095	0.0275	SHC1, RAF1, BAD, PXN, NEDD4
Ingenuity	Tight Junction Signaling	35	2	6	0.0076	0.0314	CEBPA, RHOA, PTEN, LLGL1, RAC1, VASP, PARD6A
Ingenuity	Integrin Signaling	37	2	6	0.0087	0.0346	SHC1, RAF1, MAP3K11, PTEN, PXN, VASP
Ingenuity	Glucocorticoid Receptor Signaling	92	5	11	0.0048	0.0351	CEBPA, SHC1, POMC, RAF1, RELA, HMG1L1, SCGB1A1, IL3, CSF2, BCL2, IFNG, RAC1, PBX1
Ingenuity	TGF-beta Signaling	28	1	5	0.0124	0.0352	HOXC9, SMAD6, RAF1, ZNF423, SMAD7, AMH
Ingenuity	Erythropoietin Signaling	13	1	3	0.0213	0.0377	SHC1, RAF1, SOCS3
Ingenuity	Chemokine Signaling	21	1	4	0.0181	0.0400	RAF1, RHOA, ROCK2, PTK2B
PANTHER Molecular Function	Hydroxylase	17	1	5	0.0006	0.0469	PH-4, P4HA2, ASPH, DBH, MUS81

Database	Gene set	Effective gene set size	# of expected genes (> 95 <sup>th</sup> percentile cutoff)	# of observed genes (> 95 <sup>th</sup> percentile cutoff)	<i>P</i> value	False- discovery rate	Nominally significant genes
KEGG	Focal Adhesion	179	9	22	0.0001	0.0496	VEGFA, SHC1, RAF1, LAMB1, COL5A3, VEGFC, BAD, LAMB2, VEGFB, LAMB3, COL6A1, COL6A2, RHOA, PTEN, BCL2, ITGB3, PXN, ROCK2, RAC1, ITGA9, VAV3, ITGB6, VASP, ITGB8, FN1, CDC42

The table lists waist-hip ratio adjusted for body mass index (WHRadjBMI) gene sets that reached significance (false-discovery rate < 0.05) using MAGENTA and its 95-percentile cutoff model. Using the  $75^{th}$ -percentile cutoff, 'PTEN Signaling' was the only gene set that reached significance ( $P = 3.2 \times 10^{-3}$ ; false-discovery rate =  $3.7 \times 10^{-2}$ ). The 'Nominally Significant Genes' column lists genes that were part of a given gene set and exhibited nominally significant MAGENTA gene P values.

Supplementary Table 24 | Phenotypic correlations among waist-related traits, height, weight, and body mass index

	ВМІ	WHR	HEIGHT	WEIGHT	HIP	wc	WHRadjBMI	WCadjBMI	HIPadjBMI
BMI: KORA		0.56	-0.07	0.87	0.59	0.88	0.00	0.00	0.00
EGCUT		0.46	0.00	0.90	0.73	0.82	NA	NA	NA
TWINGENE	-	0.41	-0.01	NA	0.81	0.78	-0.06	0.00	0.22
FRAMINGHAM		0.57	-0.02	0.88	0.83	0.88	0.01	-0.02	-0.03
WHR: KORA	0.44		-0.04	0.49	0.18	0.76	0.83	0.56	-0.19
EGCUT	0.40		0.03	0.43	0.10	0.70	0.89	0.56	-0.36
TWINGENE	0.41	-	0.42	NA	0.14	0.79	0.89	0.76	-0.23
FRAMINGHAM	0.48		-0.04	0.49	0.30	0.75	0.82	0.52	-0.31
HEIGHT: KORA	-0.13	-0.05		0.42	0.35	0.16	0.00	0.48	0.49
EGCUT	-0.06	-0.03		0.42	0.26	0.20	0.04	0.35	0.38
TWINGENE	-0.02	0.42	-	NA	0.14	0.38	0.47	0.64	0.24
FRAMINGHAM	-0.08	-0.02		0.45	0.34	0.21	-0.04	0.48	0.62
WEIGHT: KORA	0.92	0.41	0.28		0.71	0.88	0.00	0.24	0.24
EGCUT	0.93	0.38	0.30		0.77	0.82	0.02	0.15	0.16
TWINGENE	NA	NA	NA	-	NA	NA	NA	NA	NA
FRAMINGHAM	0.94	0.46	0.26		0.89	0.88	-0.01	0.21	0.27
HIP: KORA	0.67	0.11	0.21	0.73		0.58	-0.18	0.14	0.81
EGCUT	0.82	0.14	0.16	0.84		0.78	-0.27	0.31	0.68
TWINGENE	0.81	0.14	0.14	NA	-	0.71	-0.26	0.12	0.75
FRAMINGHAM	0.92	0.33	0.12	0.93		0.85	-0.20	0.25	0.54
WC: KORA	0.88	0.72	0.05	0.87	0.60		0.32	0.47	0.08
EGCUT	0.83	0.71	0.09	0.82	0.80		0.36	0.58	0.26
TWINGENE	0.78	0.79	0.38	NA	0.71	-	0.47	0.62	0.29
FRAMINGHAM	0.89	0.75	0.07	0.89	0.86		0.31	0.45	0.19
WHRadjBMI:KORA	0.00	0.90	0.01	0.01	-0.20	0.37		0.68	-0.22
EGCUT	NA	0.92	0.00	0.00	-0.21	0.41		0.63	-0.40
TWINGENE	0.10	0.95	0.46	NA	-0.13	0.59	-	0.84	-0.37
FRAMINGHAM	0.01	0.88	0.02	0.02	-0.12	0.38		0.65	-0.36
WCadjBMI: KORA	0.00	0.70	0.35	0.14	0.03	0.47	0.78		0.18
EGCUT	NA	0.66	0.25	0.10	0.21	0.56	0.72		0.45
TWINGENE	0.06	0.79	0.63	NA	0.17	0.67	0.83	-	0.19
FRAMINGHAM	0.01	0.73	0.31	0.12	0.11	0.47	0.83		0.47
HIPadjBMI: KORA	0.00	-0.24	0.39	0.16	0.75	0.02	-0.27	0.05	
EGCUT	NA	-0.33	0.37	0.13	0.57	0.21	-0.36	0.37	
TWINGENE	-0.14	-0.38	0.25	NA	0.47	0.02	-0.36	0.19	-
FRAMINGHAM	0.01	-0.28	0.49	0.17	0.40	0.12	-0.32	0.25	

Phenotypic trait correlations were evaluated in four studies (KORA, EGCUT, TWINGENE and FRAMINGHAM, see Online Methods). Trait correlations in men are listed in the upper triangle and correlations in women are listed in the lower triangle. Correlations > 0.75 are marked in bold. Missing correlations are marked with "NA". BMI, body mass index; WHR, waist-to-hip ratio; HIP, hip circumference; WC, waist circumference; adjBMI, adjusted for body mass index

# **Supplementary Note and Methods**

# Candidate genes at new loci for WHRadjBMI achieving genome-wide significance

# 1. **Chromosome 1q21.3-q22:** *DCST2*, DC-STAMP domain containing 2

*DCST2* encodes dendritic cell-specific transmembrane protein domain containing 2, a multimembrane spanning protein that contains a domain similar to that found in dendritic cells. DC-STAMP proteins have been implicated in skewing hematopoietic differentiation of bone marrow cells toward the myeloid lineage, and in cell fusion during osteoclastogenesis and giant cell formation<sup>97</sup>. A nearby gene is *ZBTB7B*, zinc finger and BTB domain containing 7B, also known as ThPOK, which encodes a zinc finger transcription factor that is critical to CD4+ T cell development in CD4/CD8 lineage commitment, and suppresses CD8-lineage gene expression<sup>98,99</sup>. ZBTB7B has been shown to function as a transcriptional repressor of fibronectin and alpha1 collagen genes<sup>100</sup>.

#### 2. Chromosome 1q24.2: GORAB, golgin, RAB6-interacting

*GORAB* encodes a member of the golgin family, and is a coiled-coil protein localized to the Golgi apparatus. This protein family may play a role in Rab6-regulated membrane-tethering events<sup>101</sup>.

#### 3. Chromosome 2p14: MEIS1, Meis homeobox 1

The lead WHRadjBMI-associated SNP is located ~500 kb from *MEIS1*, which encodes a transcription factor that is a member of the three-amino-acid loop extension family of homeobox-containing proteins. Meis1 is essential for hematopoiesis and vascular patterning in the mouse embryo<sup>102</sup> and regulates vascular development in zebrafish<sup>103</sup>. Dysregulation of *MEIS1* expression has been linked to a variety of leukemias<sup>104-106</sup>. The lead SNP is also <400 kb from *miR4778*.

#### 4. Chromosome 2q32.1: CALCRL, calcitonin receptor-like

*CALCRL* encodes calcitonin receptor-like protein receptor, involved in G-protein coupled receptor-like signaling. Calcitonin receptor-like receptor, CRLR, along with receptor activity-modifying protein-2, RAMP2, is a receptor for adrenomedullin. Adrenomedullin and CLRL/RAMP2 levels were increased in epididymal, mesenteric, and retroperitoneal adipose tissue in rats fed a high-fat diet compared to rats fed a normal diet<sup>107</sup>. *CRLR* mRNA levels were decreased in epicardial white adipose tissue compared to subcutaneous white adipose tissue from human biopsies<sup>108</sup>. A nearby gene, *TFPI*, encodes a protease inhibitor that regulates the tissue factor (TF)-dependent pathway of blood coagulation. The encoded protein is predominantly found in the vascular endothelium and plasma in both free forms and in complexes with plasma lipoproteins.

#### 5. Chromosome 3q22.1: PLXND1, plexin D1

*PLXND1* encodes plexin D1 protein, a co-receptor for semaphorin proteins<sup>109</sup>. *Plxnd1* is expressed in cells from the central nervous system and vascular endothelium in mouse embryos<sup>110</sup>. Plexin D1 plays a role in vascular patterning; *plxnD1*-deficient zebrafish embryos show defects in segmental artery development such as premature and ectopic sprouting and improper blood vessel branching<sup>111</sup>. Semaphorin-plexinD1 signaling antagonizes the proangiogenic activity of vascular endothelial growth factor, VEGF<sup>34</sup>.

# 6. **Chromosome 3q25.31:** *LEKR1*, leucine, glutamate and lysine rich 1 protein

LEKR1 encodes leucine, glutamate and lysine rich 1 protein with unknown function. The lead WHRadjBMI-associated SNP is also located near *CCNL1*, encoding cyclin L1, and two uncharacterized noncoding RNAs, LINC00880 and LINC00881. Also nearby, *TIPARP* encodes a poly(ADP-ribose) polymerase superfamily member, which catalyzes the transfer of multiple ADP-ribose groups from nicotinamide-adenine dinucleotide (NAD) onto protein targets, and *VEPH1* encodes ventricular zone expressed PH domain-containing 1.

#### 7. Chromosome 4q12: NMU, neuromedin U

*NMU* encodes neuromedin U, a highly conserved neuropeptide. NMU is found at highest levels in the gastrointestinal tract and pituitary, and has been implicated in the regulation of smooth muscle contraction, blood pressure and local blood flow, ion transport in the gut, stress responses, cancer, gastric acid secretion, and feeding behavior<sup>112</sup>. *Nmu* knockout mice are hyperphagic and obese<sup>36</sup>. Rare coding variants in NMU have been found to be associated with obesity<sup>113</sup>.

# 8. Chromosome 4q22.1: FAM13A, family with sequence similarity 13,

member A

*FAM13A* has a putative role in signal transduction, however is poorly described. SNPs in this gene region were found to be associated with chronic obstructive pulmonary disease and lung function<sup>114,115</sup>. Other nearby genes include *HERC3*, *NAP1L5*, *PIGY* (phosphatidylinositol-glycan biosynthesis class Y protein), and *TIGD2*.

9. Chromosome 4q28.1: SPATA5, spermatogenesis associated 5 – FGF2, fibroblast growth factor 2 SPATA5 belongs to the AAA ATPase family and AFG2 subfamily, and may be involved in mitochondrial transformation during spermatogenesis. SNPs at SPATA5 have been associated with alopecia areata<sup>116</sup>. Other nearby genes include FGF2, NUDT6, and SPRY1. FGF2 enhanced vascularization for human adipose tissue engineering<sup>117</sup>. NUDT6 (nudix-type motif 6) is an antisense gene to FGF2 that showed associations with fat deposition related traits in pigs<sup>118</sup>. Conditional Spry1 (sprouty homolog 1) expression

in mouse adipose tissue protected against high-fat diet-induced obesity, bone loss, and metabolic dysfunction<sup>119</sup>.

# 10. **Chromosome 5q11.2**: *MAP3K1*, mitogen-activated protein kinase kinase kinase 1, E3 ubiquitin protein ligase

The lead SNP is located within the intron of an uncharacterized transcript **AC022431**. Located 250 kb away, *MAP3K1*, also known as *MEKK1*, encodes a protein in the MAPK group of serine/threonine protein kinases. The protein contains a PHD plant homeodomain that exhibits E3 ubiquitin ligase activity toward ERK1/2<sup>120</sup>. MAP3K1 also activates the JNK signaling pathway and plays a role in apoptosis<sup>121</sup> and wound healing<sup>122</sup>. Along with IL-1beta, MAP3K1 inhibited basal and membrane depolarization and cAMP-induced transcription of the insulin gene in a hamster beta cell line<sup>123</sup>.

# 11. Chromosome 5q35.2: FGFR4, fibroblast growth factor receptor 4

*FGFR4* is a member of the receptor tyrosine kinase family<sup>124</sup>. FGFR4 is expressed mainly in lung, kidney, pancreas, spleen and developing muscle<sup>125</sup>. FGFR4-deficient mice on a normal diet displayed increased mass of white adipose tissue, hyperlipidemia, glucose intolerance, insulin resistance and hypercholesterolemia<sup>37</sup>.

# 12. Chromosome 6p21.32: BTNL2, butyrophilin-like 2 (MHC class II associated)

Located 30 kb from the HLA cluster, *BTNL2* is an MHC class II gene-linked butyrophilin family member that inhibits T-cell activation<sup>126</sup>. Variants in *BTNL2* are associated with inflammatory diseases<sup>127,128</sup>. Other nearby genes include *HLA-DRA*, *HLA-DRB5*, *HLA-DRB1*, *HLA-DRB6*, *HLA-DRB1*, *HLA-DRB1*, *HLA-DQA1*, *HLA-DQB1*. These HLA genes belong to the HLA class II proteins, which are expressed in antigen presenting cells, such as B lymphocytes, macrophages, and dendritic cells.

#### 13. Chromosome 6p21.31: HMGA1, high mobility group AT-hook 1

HMGA1 encodes a protein that binds to the minor groove of stretches of A-T-rich DNA<sup>129</sup>. *HMGA1* is a downstream nuclear target of the insulin receptor signaling pathway<sup>130</sup>, and *Hmga1* knockout mice showed decreased insulin receptor expression, impaired insulin signaling and reduced insulin secretion<sup>38</sup>.

#### 14. Chromosome 7p15.2: HOXA11, homeobox A11

There are 12 *HOXA* genes at this locus, as well as several antisense transcripts. HOX genes encode conserved transcription factors containing a homeodomain that regulate body and axis development and organogenesis<sup>131</sup>. *HOXA11* is necessary for female fertility and regulates embryonic uterine and endometrium development<sup>132,133</sup>. *HOXA11* mutations were found in individuals affected with the blood disease amegakaryocytic thrombocytopenia and the skeletal defect radio-ulnar synostosis<sup>134</sup>.

#### 15. Chromosome 8p21.2: NKX2-6, NK2 homeobox 6

*NKX2-6* encodes a homeobox-containing protein that is a homolog of Drosophila tinman<sup>135</sup>. At early stages of mouse embryogenesis, *NKX2-6* is expressed in the pharyngeal endoderm, developing gut endoderm, cardiac progenitors, and heart<sup>136,137</sup>. Nearby *NKX3-1* is also a homeobox gene that is involved in prostate epithelium development during embryogenesis<sup>138</sup> and is androgen-regulated<sup>139</sup>. *STC1* encodes a secreted, homodimeric glycoprotein that is expressed in a wide variety of tissues and is upregulated by VEGFD. STC1 may play a role in the regulation of renal and intestinal calcium and phosphate transport, cell metabolism, and angiogenesis.

#### 16. Chromosome 8q13.3: MSC, musculin

*MSC* encodes a basic helix-loop-helix transcription factor expressed in developing skeletal muscle<sup>140</sup> and mouse embryonic ectoderm<sup>141</sup>. *EYA1* encodes eyes absent homolog1, a protein phosphatase and co-activator for the transcription factor SIX1, which regulates skeletal muscle fiber-type and development<sup>142</sup>. Mutations in *EYA1* cause Branchio-oto-renal syndrome and Branchiootic syndrome, which are characterized by hearing loss, branchial arch defects and renal abnormalities<sup>143</sup>. EYA protein phosphatase activity promotes angiogenesis<sup>144</sup>.

# 17. Chromosome 9q31.1: ABCA1, ATP-binding cassette, sub-family A (ABC1), member 1

This gene encodes an ATP-binding cassette transporter. Mutations in *ABCA1* have been found to be associated with Tangier's disease and familial high-density lipoprotein deficiency<sup>145</sup>. Adipose tissue abundantly expresses *ABCA1*, and adipose tissue *ABCA1*-dependent cholesterol efflux and nascent HDL particle formation contribute to systemic HDL biogenesis<sup>146</sup>.

#### 18. Chromosome 10q24.32: SFXN2, sideroflexin 2

*SFXN2* encodes a mitochondrial transmembrane protein that may facilitate transport of pyridoxine or enzyme cofactors involved in heme synthesis into the mitochondria. The gene is widely expressed, and is expressed at particularly high levels in adult kidney and liver<sup>147</sup>. *Sfxn2* was found upregulated in pancreatic islets from streptozotocin-induced diabetic rats compared to normal rats<sup>148</sup>.

# 19. **Chromosome 11q13.1:** *MACROD1*, MACRO domain containing 1, *VEGFB*, vascular endothelial growth factor B

Macrodomains are known to bind ADP-ribose derivatives<sup>149</sup>. Also known as LRP16, MACROD1 was found to play a role in estrogen signaling by interacting with estrogen receptor alpha and enhancing the receptor's transcriptional activity<sup>150</sup>. It has also been found to bind to the androgen receptor via its macro domain and amplifies the transactivation of androgen receptor in response to androgen<sup>151</sup>. LRP16 regulated insulin content and glucose-stimulated insulin secretion in MIN6 cells, and overexpression of this

gene protected MIN6 cells from fatty acid-induced apoptosis<sup>152</sup>. Diabetic db/db Vegfb knockout mice had ectopic lipid deposition, increased muscle glucose uptake and maintained normoglycemia, and treatment of db/db mice with a VEGF-B antibody enhanced glucose tolerance, preserved pancreatic islet architecture, improved  $\beta$ -cell function and improved dyslipidemia<sup>153</sup>. The index SNP is located ~6 kb from *FLRT1*, fibronectin leucine rich transmembrane protein 1, involved in cell adhesion and fibroblast growth factor mediated signaling<sup>154</sup>.

# 20. Chromosome 12q24.31: CCDC92, coiled-coil domain containing 92 protein

The closest genes to the index variant are not obvious candidate genes. *CCDC92* encodes a protein with unknown function that was found to be upregulated in human B lymphoblastoid cells treated with a polychlorinated biphenyl pollutant<sup>10</sup>. *DNAH10* encodes dynein, axonemal, heavy chain 10, which may a play a role in cilia or flagella. *ZNF664* encodes zinc finger protein 664; coding variants in ZNF664 have been implicated in myopia<sup>155</sup>.

#### 21. Chromosome 15q13.3: KLF13, Kruppel-like factor 13

*KLF13* encodes Kruppel-like factor 13, which belongs to the Sp1-like family of transcription factors that contain 3 C-terminal zinc finger DNA-binding domains, and bind to GC-rich sequences<sup>156</sup>. KLF13 is a regulator of heart development<sup>157</sup>, and was also found to bind and repress the low density lipoprotein receptor promoter<sup>158</sup>. A nearby gene, *OTUD7A*, belongs to a deubiquitinating enzyme subfamily characterized by an ovarian tumor (OTU) domain. This gene encodes a protease that cleaves ubiquitin linkages.

# 22. Chromosome 15q21.3: RFX7, regulatory factor X, 7

*RFX7* encodes a member of the regulatory factor X family of transcription factors. It is a winged-helix transcription factor and contains a well-conserved RFX DNA binding domain. It has high ubiquitous expression, particularly in brain<sup>159</sup>. *TEX9*, encoding testis-expressed sequence 9, is poorly described. Another nearby gene, *NEDD4*, encodes neural precursor cell expressed, developmentally down-regulated 4, an E3 ligase. Overexpression of *Nedd4* suppressed BMP-induced osteoblast transdifferentiation process of mouse premyoblast C2C12 cells, and *NEDD4* was also found to be an important modulator of phospho-Smad1 in both BMP-2 and TGF-β1 action<sup>160</sup>.

#### 23. Chromosome 15q22.31: SMAD6, SMAD family member 6

SMAD6 belongs to the SMAD family of proteins, which are related to *Drosophila* 'mother's against decapentaplegic' and *C elegans* Sma. SMAD proteins are signal transducers of the TGF-β superfamily and are involved in cell growth, morphogenesis, development and immune responses<sup>161</sup>. SMAD6 inhibits the

Bone morphogenetic protein/Smad1 signaling pathway<sup>162</sup>. 3T3-F442A mouse pre-adipocytes overexpressing *Smad6* show increased TGF-β signaling and decreased adipocyte differentiation<sup>163</sup>.

# 24. Chromosome 16q23.3: CMIP, c-MAF inducing protein

This gene encodes C-maf inducing protein, which interacts with phosphatidylinositol 3-kinase complex and plays a role in ERK signaling<sup>164</sup>. *CMIP* is expressed in peripheral blood mononuclear cells, kidney, fetal liver, and adult brain and liver<sup>165</sup>. A nearby gene, *PLCG2*, encodes phospholipase C, gamma 2 (phosphatidylinositol-specific), which hydrolyzes phosphatidyl inositol 4,5-biphosphate (PIP<sub>2</sub>) to inositol-1,4,5-triphosphate (IP<sub>3</sub>), resulting in an increase in intracellular calcium levels<sup>166</sup>.

#### 25. **Chromosome 17p11.2: PEMT**, phosphatidylethanolamine N-methyltransferase

This gene encodes a liver enzyme that converts phosphatidylethanolamine to the phospholipid phosphatidylcholine by methylation in the liver. The protein localizes to the endoplasmic reticulum and mitochondria-associated membranes. *Pemt* knockout mice on a high fat diet show adipocyte hypotrophy<sup>167</sup>. *Pemt* mRNA and protein increase upon adipocyte differentiation in 3T3-L1 cells<sup>168</sup>.

# 26. **Chromosome 17q24.3**: *KCNJ2*, potassium inwardly-rectifying channel, subfamily J, member 2 Inwardly rectifying K+ channels control the resting K+ conductance and stabilize the resting potential in many cells<sup>169</sup>. *KCNJ2* was upregulated during myoblast differentiation into skeletal muscle<sup>170</sup> and was expressed in smooth muscle<sup>171</sup> and cardiomyocytes<sup>172</sup>.

#### 27. Chromosome 18q21.33: BCL2, B-cell CLL/lymphoma 2

B-cell CLL/lymphoma 2 encodes an anti-apoptotic protein that binds the BH3 domain of pro-apoptotic factors and regulates permeabilization of the outer mitochondrial membrane, a critical step in apoptosis 173,174. *Bcl2* was upregulated and apoptosis was reduced in rat pancreatic beta-cells treated with leptin 175.

#### 28. Chromosome 19p13.11: JUND, jun D proto-oncogene

JUND is a component of the Activating protein 1 transcription factor; AP-1 is a dimeric transcription factor with basic leucine zipper domains<sup>176</sup>. JunD dimerizes with DeltaFosB and binds to the *IL-11* gene promoter. Suppression of osteoblast differentiation by aging involved decreased JunD binding to the *IL-11* promoter and reduced *IL-11* transcription<sup>177</sup>. IL-11 inhibits the accumulation of adipose in human long-term bone marrow culture stromal layers<sup>178</sup>. Other nearby genes include *KIAA1683*, *LSM4 PIK3R2*, *PDE4C*, and *miR3188*.

# 29. Chromosome 19q13.11: CEBPA, CCAAT/enhancer binding protein alpha

C/EBP alpha is a basic leucine zipper transcription factor that is highly expressed in liver and adipose tissue, and is required for differentiation of white adipose tissue<sup>179</sup>. *C/ebp alpha* knockout mice have defects in gluconeogenesis, are hypoglycemic, and die shortly after birth<sup>180</sup>. Additionally, C/EBP alpha also binds to the leptin promoter, a gene that plays an important role in body weight homeostasis<sup>181</sup>. Other nearby genes include *C/EBPG*, encoding C/EBP gamma, which forms heterodimers with C/EBP beta<sup>182</sup>, and *PEPD*, encoding peptidase D.

#### 30. Chromosome 20p12.3: BMP2, bone morphogenetic protein 2

*BMP2* belongs to the transforming growth factor beta (TGF-β) superfamily of genes<sup>183</sup>. BMPs signal through transmembrane serine/threonine kinase receptors and stimulate Smad, MAPK and Akt signaling pathways<sup>184</sup>. High levels of BMP2 induce chondrogenesis or osteogenesis, while low levels of BMP2 promote adipogenesis<sup>185</sup>. BMP2 stimulates commitment of C3H10T1/2 pluripotent stem cells into adipocytes<sup>186</sup>. BMP2, along with IGF-1, induces differentiation of adipose-derived mesenchymal stem cells into cartilage cells<sup>187</sup>.

# 31. Chromosome 20q11.22: GDF5, growth differentiation factor 5

*GDF5* is a member of the bone morphogenetic protein BMP family and the transforming growth factor-beta superfamily<sup>188</sup>. GDF5 promoted osteogenic differentiation of rat fat-derived stromal cells and may promote angiogenic activity of stromal cells by increasing vascular endothelial growth factor gene expression in vitro<sup>189</sup>. GDF5 also induced chondrogenesis in rat adipose-derived stem cells<sup>190</sup>. Human mesenchymal stem cells that overexpressed GDF5 displayed osteogenic differentiation<sup>191</sup>. *UQCC* is a nearby gene, which encodes ubiquinol-cytochrome c reductase complex chaperone, a ZIC-binding protein repressed by basic fibroblast growth factor<sup>192</sup>.

#### 32. Chromosome 20q13.12: EYA2, eyes absent homolog 2

This gene encodes a member of the eyes absent, EYA, family of proteins. EYA2 is a transcriptional co-activator and protein phosphatase. *Eya2* acts synergistically with both *Dach2* and *Six1* to regulate myogenic differentiation and development<sup>193</sup>. Eya2 also prevents adverse cardiac remodeling under pressure overload<sup>194</sup>. Nearby, *SLC2A10* encodes solute carrier family 2 (facilitated glucose transporter) member 10.

#### 33. Chromosome 7p15.2: SNX10, sorting nexin 10

SNX10 encodes a nexin family protein involved in intracellular trafficking. SNX10 has been shown to cause osteopetrosis, a rare disorder resulting from osteoclast dysfunction<sup>195</sup>, and to regulate ciliogenesis<sup>196</sup> and endosome homeostasis<sup>197</sup>.

# Candidate genes at new loci for five additional waist and hip traits

 Chromosome 1q44: OR2W5 olfactory receptor family 2, subfamily W, member 5 and NLRP3, NLR family, pyrin domain containing 3

*OR2W5* encodes an olfactory receptor. *NLRP3* regulates inflammation, immune response, and apoptosis, and is associated with several inflammatory and autoimmune disorders. Other variants near *NLRP3* are associated with C-reactive protein levels<sup>198</sup>. NLRP3-containing inflammasome and proinflammatory T cell populations in adipose tissue contribute to inflammation and in insulin resistance<sup>42</sup>. Other nearby genes include *OR2C3*, encoding an olfactory receptor, and *GCSAML-AS1* antisense non-coding RNA.

#### 2. Chromosome 2p25: SOX11, SRY (sex determining region Y)-box 11

This intronless gene encodes a member of the SOX (SRY-related HMG-box) family of transcription factors involved in the regulation of embryonic development and in the determination of the cell fate. SOX11 plays a role in the embryonic development of the central nervous system (CNS) and is expressed in the adult immature neuron. Knockdown of *SOX11* with siRNA decreased the proliferation and osteogenic differentiation potential of mesenchymal stem cells<sup>199</sup>. SOX11 has tumor suppressor function in hematopoietic malignancies<sup>200</sup>, and prevents tumorigenesis of glioma initiating cells by inducing neuronal differentiation<sup>201</sup>.

# 3. Chromosome 2q24.2: ITGB6, Integrin, beta 6

*ITGB6* encodes a heterodimeric cell surface receptor, which is absent from the normal epithelium but is expressed in wound-edge keratinocytes during re-epithelialization. ITGB6 is involved in tumor growth and metastasis and may serve a protective role in re-epithelialization of diabetic wounds<sup>202</sup>. Other nearby genes encode *PLA2R1*, phospholipase A2 receptor, and *RBMS1*, a protein that binds single-stranded DNA and RNA.

#### 4. Chromosome 5q11.2: ARL15, ADP-ribosylation factor-like 15

ARL15 encodes an ADP-ribosylation factor-like (ARL) protein. ARL proteins are small GTPases that regulate the affinity of ARLs for binding other proteins, lipids, or membranes<sup>203</sup>. *ARL15* is expressed in insulin-responsive tissues, including adipose tissue and skeletal muscle<sup>204</sup>. Other SNPs at *ARL15* have previously been associated with adiponectin levels<sup>65,204</sup>, HDL levels<sup>205</sup> and replicated CNVs in childhood obesity<sup>206</sup>.

#### 5. Chromosome 5q33.3: CCNJL, cyclin J-like

This gene encodes a protein that belongs to the cyclin family, cyclin J subfamily, which regulates cyclin dependent kinases<sup>207</sup>. A nearby gene is *FABP6*, encoding fatty acid binding protein 6, which binds fatty acids and is involved in fatty acid uptake, transport and metabolism<sup>208</sup>. *Fabp6* is necessary for absorption and transport of bile acids in mouse small intestine<sup>209</sup>. Other nearby genes include *PWWP2A*, encoding PWWP domain containing 2A; and *C1QTNF2*, encoding C1q and tumor necrosis factor related protein 2.

#### 6. Chromosome 6p25: GMDS, GDP-mannose 4,6-dehydratase

*GMDS* catalyzes the first step of GDP-fucose synthesis from GDP-mannose and can inhibit apoptosis in colon cancers<sup>210</sup>. Other nearby genes include *FOXC1* (forkhead box C1), *FOXQ1* (forkhead box Q1), *FOXF2* (forkhead box F2), all of which are DNA-binding proteins involved in cell growth, apoptosis, migration and differentiation<sup>211</sup>. *FOXQ1* is negatively regulated by Oct4 in adipose tissue stromal cells<sup>212</sup>.

# 7. Chromosome 6p12.1: KLHL31, kelch-like 31

*KLHL31* regulates transcription in the MAPK/JNK pathway<sup>213</sup>. In chicken, *Klhl31* was found to be highly expressed in the somite myotome, heart, and in differentiated myocardium and skeletal muscle<sup>214</sup>. Nearby gene *GCLC*, encodes the glutamate-cysteine ligase catalytic subunit, which plays a regulatory role in glutathione synthesis, and may play a role in growth and development<sup>215</sup>. Another nearby gene, *ELVOL*, encodes fatty acid elongase 5, which is involved in fatty acid synthesis and elongation. Increased expression of *Elvol* has been shown to restore glucose homeostasis and decrease insulin localization in hyperglycemic mice fed a high-fat diet<sup>216</sup>.

#### 8. Chromosome 7q22.3: SRPK2, Serine/arginine-rich splicing factor protein kinase 2

*SRPK2* encodes a non-small nuclear ribonucleoprotein particle that regulates the intracellular storage of splicing factors<sup>217,218</sup>. Knockdown of *SRPK2* by RNAi in HeLa cells demonstrated that this gene essential for cell viability<sup>219</sup>. Nearby genes include *LHFPL3* (lipoma HMGIC fusion partner-like 3), *MLL5* (myeloid/lymphoid or mixed-lineage leukemia 5), which is suggested to have a role in chromatin remodeling and cellular growth suppression<sup>220</sup>, and several non-coding RNAs.

#### 9. Chromosome 7q32: KLF14, Kruppel-like factor 14

*KLF14* encodes an imprinted developmental transcription factor exhibiting maternal allelic expression induced by TGF-beta. *KLF14* has been shown to be a master trans-regulator affecting multiple metabolic phenotypes<sup>221</sup>. Other nearby genes include *CPA4* (carboxypeptidase A4), *CPA2* (carboxypeptidase 2), *MEST* (mesoderm specific transcript homolog), and *COPG2* (coatomer protein complex, subunit gamma 2).

#### 10. Chromosome 9q22.32: PTPDC1, protein tyrosine phosphatase domain containing 1

*PTPDC1* is a member of a protein family known to play roles in molecular signaling in a wide variety of biological processes<sup>222,223</sup>. Mouse *Ptpcd1* was suggested to play a role in centriole duplication and cytokinesis<sup>224</sup> and depletion has been shown to correlate with cilia elongation<sup>225</sup>. Nearby genes include *BARX1*, encoding BARX homeobox transcription factor, implicated in dentition and cleft lip syndrome<sup>226</sup>, and *ZNF169*, which encodes zinc finger protein 169 transcription factor. A near genome-wide significant association has been found between a nearby SNP (rs10993160, *P*=5.5×10<sup>-7</sup>) and BMI in East Asians<sup>47,227</sup>.

#### 11. Chromosome 9q33-q34: C5, complement component 5 (also known as CPAMD4)

C5 encodes the fifth component of complement, which plays an important role in host defense and inflammatory processes. Mutations in C5 cause a propensity for severe recurrent infections. Complement component 5 contributes to poor disease outcome in humans and mice with pneumococcal meningitis<sup>228</sup>. Defects in this gene have also been linked to susceptibility to liver fibrosis and rheumatoid arthritis<sup>229</sup>. Other nearby genes include **PSMD5**, encoding 26S proteasome non-ATPase regulatory subunit 5, **FBXW2**, encoding F-box and WD repeat domain containing 2, and **TRAF1**, TNF receptor-associated factor 1.

# 12. Chromosome 11q13: MYEOV, myeloma overexpressed

*MYEOV*, encoding myeloma overexpressed (in a subset of t(11;14) positive multiple myelomas) has been implicated in multiple myeloma, as well as some other cancer types<sup>230</sup>. Nearby are members of the fibroblast growth factor family *FGF19*, *FGF4* and *FGF3*. FGFs play important roles in multiple physiologic functions, including angiogenesis, mitogenesis, pattern formation, cellular differentiation, metabolic regulation, tissue repair, and oncogenesis. *FGF19* has been shown to activate an insulin-independent endocrine pathway that regulates hepatic protein and glycogen metabolism<sup>231</sup>. Other nearby genes include *CCND1* (cyclin D1) and *ORAOV1* (oral cancer overexpressed 1).

#### 13. Chromosome 11q21: KIAA1731

RNAi analyses suggest that *KIAA1731* encodes a centrosomal protein responsible for centriole formation/stability<sup>232</sup>. Other nearby genes include *TAF1D* (TATA box binding protein associated factor, RNA polymerase I), which plays a role in RNA polymerase I transcription<sup>233,234</sup>, *MED17* (mediator complex subunit 17), *C11orf54*, *C11orf54*, *SCARNA9* and *VSTM5*.

#### 14. **Chromosome 11q22.1**: *CNTN5*, contactin 5

CNTN5 is a glycosylphosphatidylinositol (GPI)-anchored neuronal membrane protein that functions as a cell adhesion molecule. CNTN5 may play a role in the formation of axon connections in the developing nervous system<sup>235</sup>. Other nearby genes include *PGR*, encoding the progesterone receptor, and *TMEM133*, encoding transmembrane protein 133.

#### 15. Chromosome 13q31.3, GPC6, glypican 6

*GPC6* is a member of a family of glycosylphosphatidylinositol-anchored heparan sulfate proteoglycans that are ubiquitously expressed in most fetal and adult tissues<sup>236</sup>. GPC6 may influence cellular growth control and differentiation during development, and mutations in this gene have been shown to cause the rare skeletal dysplasia autosomal recessive generalized omodysplasia<sup>237</sup>.

# 16. Chromosome 16p13.11: PDXDC1, pyridoxal-dependent decarboxylase domain containing 1

PDXDC1 has been predicted to belong to the family of group II pyridoxal-dependent decarboxylases, which includes enzymes that decarboxylate glutamate, histidine, tyrosine and tryptophan<sup>238</sup>. Nearby gene *PLA2G10* (phospholipase A2, group X), is important for the breakdown of phospholipids and cholesterol into fatty acids<sup>239-242</sup>. Nearby gene *NTAN1* (N-terminal asparagine amidase) is an integral part of the N-end rule pathway; disruption of this pathway by knocking out the *Ubr1* gene resulted in mice with decreased body weight due to reduced skeletal muscle and adipose tissue<sup>243</sup>.

#### 17. Chromosome 16q12: ZNF423, zinc finger protein 423

*ZNF423* encodes a zinc finger transcription factor that associates with RARalpha/RXRalpha nuclear receptor complex and is critical for retinoic acid-induced differentiation<sup>244</sup>. Delayed induction of preadipocyte transcription factor *ZNF423* in fibroblasts resulted in delayed adipogenesis<sup>41</sup>. A nearby gene, *CNEP1R1*, encoding CTD nuclear envelope phosphatase 1 regulatory subunit 1 is involved in the conversion of phosphatidic acid to diacylglycerols and may indirectly modulate the lipid composition of nuclear and/or endoplasmic reticulum membranes and to regulate the production of lipid droplets and triacylglycerol<sup>245</sup>.

#### 18. Chromosome 17p13.3: VPS53, Vacuolar protein sorting 53 homolog (S. cerevisiae)

The VPS53 protein is a component of the Golgi-associated retrograde protein complex, and is required to maintain the cycling of mannose 6-phosphate receptors between the trans-Golgi network and endosomes<sup>246,247</sup>. Other nearby genes include *FAM101B*, which encodes an actin regulator that stabilizes perinuclear actin filament bundles<sup>248</sup>.

19. **Chromosome 22q12.3:** *HMGXB4*, high mobility group (HMG) box domain containing 4 *HMGXB4* encodes a DNA-binding protein responsible for repression of smooth muscle differentiation<sup>249</sup>. *HMGXB4* was previously named *HMG2L1*. Nearby genes include *TOM1* (target of myb1), *HMOX1* (heme oxygenase 1), *ISX* (intestine-specific homeobox), and *MCM5* (minichromosome maintenance complex component 5).

# Comparison of ARIC and PIVUS as reference panels for GCTA

To evaluate robustness of the GCTA results, we compared results using reference datasets from PIVUS (949 individuals with GWAS and Metabochip data) and ARIC (6,654 individuals with GWAS data, see **Online Methods**). Although the sets of SNPs selected by GCTA as independently associated with waist-hip ratio adjusted for body mass index (WHRadjBMI) when using either reference dataset were very similar, with the estimated effect sizes in the joint association model highly correlated, a few differences were observed. Given that ARIC includes only GWAS genotype data, while our combined European ancestry meta-analysis includes both GWAS and Metabochip SNPs, any Metabochip SNP in the meta-analysis for which ARIC does not have genotype data was excluded from the GCTA search for independent association signals. These missing reference dataset genotypes explained the majority of the differences observed between the two analyses, including the larger number of loci with multiple association signals identified when estimating the correlation between the variants from PIVUS. In addition, a small number of discrepancies between the two analyses were the result of minor differences between the estimated association p-value for the joint model, with some SNPs reaching the  $P < 5 \times 10^{-8}$  threshold when using one dataset as reference, and therefore being selected by GCTA, while they did not reach that threshold when the correlation between SNPs was estimated from the other dataset.

In our particular setting, the choice of the preferable reference dataset is equivalent to the choice to give preference to the larger sample size provided by ARIC or to the larger SNP coverage obtained when using PIVUS. Given the observations, and in this particular case, we believe that we could achieve more insights into the genetic basis of body fat distribution by having a more dense coverage of the SNPs in our meta-analysis as that provided by PIVUS.

#### Genetic risk score comparison of high versus average genetic susceptibility

We further used the genetic risk score to compare high genetic susceptibility with the average population. We used the linear regression estimates (see Main text) to calculate the difference in WHR units between the 95<sup>th</sup> percentile and the median of the sex-combined score (median: 46; 95<sup>th</sup> percentile: 53), the women-specific score (median: 45; 95<sup>th</sup> percentile: 52) and the men-specific score (median: 31; 95<sup>th</sup> percentile: 37).

The difference between individuals at the 50<sup>th</sup> percentile and at the 95<sup>th</sup> percentile genetic susceptibility risk score groups was 0.007 WHRadjBMI units overall, 0.014 in women and 0.004 in men. These results would imply, for example, that two people from the 50<sup>th</sup> and 95<sup>th</sup> percentiles of this risk score distribution and of the same sex, BMI, age, and hip circumference of 100 cm would exhibit a 0.7 cm difference in waist circumference because of differences in their genotypes at these genetic variants (1.4 cm in women and 0.4 cm in men).

# Directional consistency of effects in GWAS and Metabochip meta-analyses

To investigate whether additional common variants may contribute to the phenotypic variance of WHRadjBMI, we compared directional consistency in sex-combined allelic effects between GWAS and Metabochip studies in the European-ancestry meta-analysis. We considered the distribution of association Z-scores from the Metabochip European ancestry sex-combined meta-analysis for WHRadjBMI, aligned to the trait-increasing allele from the GWA meta-analysis, at a subset of 1,343 independent WHRadjBMI "replication" variants on Metabochip<sup>13</sup> (CEU  $r^2 < 0.1$ ), excluding SNPs within 500 kb of the lead SNPs at identified WHRadjBMI loci. We counted the number of SNPs with the same direction of effect in both GWAs and Metabochip meta-analysis, and performed a one-sided binomial test for enrichment in concordance over that expected by chance (50%). For comparison, we repeated this process by obtaining a subset of 775 independent QT-interval "replication" variants<sup>13</sup> (CEU  $r^2 < 0.1$  with each other and >300 kb from any WHRadjBMI "replication" variants) not expected to be associated with anthropometric traits.

Among the 1,343 SNPs included on the Metabochip array based on nominal significance for WHRadjBMI<sup>13</sup>, we observed 797 (59%) directionally consistent SNPs compared to 671.5 expected by chance ( $P_{binomial} = 3.9 \times 10^{-12}$ ). The set of 775 SNPs selected for the array on the basis of QT interval<sup>13</sup> did not show such enrichment (372 SNPs, or 48%, compared to 388 expected,  $P_{binomial} = 0.87$ ). These results suggest that additional common WHRadjBMI variants may be found to be reproducible with larger samples.

#### Copy-number variant analysis

To investigate the associations with copy number variants (CNVs), we used a list of SNPs that are known to be robust tags of CNVs due to high linkage disequilibrium (LD) in European cohorts. Altogether four different CNV-tagging SNPs were genome-wide significant in sex-combined analysis. In **Supplementary Table 13**, all CNV-tagging SNP results are given from the 49 identified loci, which remained significant after multiple testing correction.

In the WHRadjBMI analysis, the marker rs1294421 ( $P = 8.2 \times 10^{-18}$ ), which is in LD with CNVR2760.1 near LY86 gene, was strongly associated. The same association was described in the previous GIANT analysis<sup>9</sup>. The same CNV tagging SNP was found to be genome-wide significant in the WHR analysis without BMI correction ( $P = 6.9 \times 10^{-14}$ ). Additionally we were able to detect statistically significant results after multiple testing correction at three additional loci: *SFMBT1* (WHRadjBMI: rs3733034 has  $P = 1.75 \times 10^{-6}$ ); *TNXB* (WHRadjBMI: rs1150753 has  $P = 4.45 \times 10^{-6}$ ), and *HMGXB4* (HIPadjBMI: rs1543302 has  $P = 1.13 \times 10^{-7}$ ).

#### Comparison of results from MAGENTA, DEPICT, and GRAIL analyses

Overlap of gene sets for WHRadjBMI that were significantly prioritized by the MAGENTA and DEPICT pathway methods. The Data-driven Enrichment-Prioritized Integration for Complex Traits (DEPICT) assesses for enrichment of 14,462 reconstituted gene sets, while MAGENTA assesses for enrichment of 3,216 gene sets. Consequently, there may be gene sets with different gene IDs that represent similar molecular functions or pathways (e.g., the BMP.Signaling.pathway in MAGENTA may represent similar biological pathways as the BMP4, BMP6, and BMPR1B protein complexes). To compare overlap of significantly enriched gene sets, we manually identified reconstituted DEPICT gene sets (false discovery rate (FDR) < 0.05) with gene set IDs similar to the enriched MAGENTA gene sets (FDR < 0.05). Among the 19 WHRadjBMI gene sets significantly prioritized by MAGENTA, 9 highly similar gene sets were prioritized by DEPICT (Supplementary Table 22).

Overlap of predicted genes for WHRadjBMI identified by both the GRAIL and DEPICT pathway methods. The following 14 genes were significantly predicted by GRAIL (adjusted P < 0.05) and DEPICT (FDR < 0.05): TBX15, EYA2, HOXA11, GDF5, WNT4, BMP2, CITED2, SMAD6, VEGFA, LAMB1, PPARG, RSPO3, DNMT3A, and CDC42EP3.

#### **Evaluation of potential sources of heterogeneity**

We tested for heterogeneity of effects to determine if the locus discovered through all ancestries metaanalysis in women (*SNX10*) was the result of increased sample size or due to heterogeneity. We used effect estimates from non-European women (Metabochip meta-analysis) and European descent-only women (GWAS+Metabochip meta-analysis) in the method outlined in Randall et al.<sup>10</sup>, and determined there was no evidence for heterogeneity.

To address the effects of study ascertainment for specific diseases or phenotypes, we compared effects in seven subsets of our study sample using population-based studies as described in the Methods. We evaluated significance for heterogeneity tests within each comparison using a Bonferroni-corrected p-value of  $0.05/49 = 0.05/49 = 1.02 \times 10^{-3}$  as well as an FDR threshold of 5%.

# Sources of data for expression QTL analyses

Our aim was to discover *cis*-acting expression quantitative trait loci (eQTL) in multiple tissues for our lead SNPs at loci that were associated with waist-related traits (**Tables 1** and **3**). We performed look-ups in previously published eQTL data from multiple biologically-relevant tissues.

In the MuTHER study<sup>250</sup>, expression profiling was performed using the Illumina Human HT-12 V3 BeadChips in lymphoblastoid cell lines (LCLs, n = 778), subcutaneous adipose tissue (SAT, n = 776) and skin (n = 667) biopsies from monozygotic and dizygotic female twins from the United Kingdom. Genotyping was done with a combination of Illumina arrays (HumanHap300, HumanHap610Q, 1M-Duo and 1.2MDuo 1M) followed by imputation into HapMap II. Association tests between genotypes and gene expression within 1 Mb windows were performed with the GenABEL/ProbABEL packages using the polygenic linear model incorporating a kinship matrix.

In the MolOBB study, expression profiling in abdominal and gluteal adipose tissue biopsies from 73 individuals (29 with and 44 without metabolic syndrome) were performed with the Affymetrix hgu133plus2 array, as described previously in detail<sup>251</sup>. Genotyping was done with the Illumina 317K array, and *cis* associations between genotypes and expression values were tested using linear regression models assuming additive genetic effects.

Expression data in liver (n = 955), SAT (n = 610), and omental fat tissue (n = 740) from the Massachusetts General Hospital collection<sup>252</sup> was obtained using a custom Agilent 44,000 feature microarray in gastric bypass surgery patients. Genotyping was done using Illumina HumanHap650Y and Affymetrix 500K genotyping arrays followed by imputation into HapMap II. Association analyses within 1 Mb windows were performed using linear regression under an additive genetic model.

For whole blood (n = 743) and SAT biopsies (n = 603) from deCODE, expression profiling of 23,720 transcripts was done using custom arrays, as previously described in detail<sup>253</sup>. *Cis* associations within 1 Mb windows between each SNP (Illumina 317K or 370K chips were used for genotyping followed by imputation to HapMap II) and expression data were tested were tested separately in men and women assuming additive genetic effects using linear regression models accounting for family structure.

Expression data in LCLs from the family asthma study (MRC-A) $^{254}$  was obtained with Affymetrix HG-U133 Plus 2.0 chip (n = 405 siblings) and Illumina Human6V1 array (n = 550 siblings). Genotyping was done using Illumina arrays (Human1M and HumanHap300K) and cis associations between genotypes and expression values were tested using linear regression models assuming additive genetic effects.

For peripheral blood mononuclear cells (PBMCs), gene expression data was available in the integrated dataset of 1,469 healthy controls and patient samples from the United Kingdom and the Netherlands (Fehrmann-HT12v3 and Fehrmann-H8v2), and 891 individuals from Estonia (EGCUT). In Fehrmann-HT12v3 (*n* =1,240) expression profiling was performed with the Illumina HumanHT-12 array and in Fehrmann-H8v2 (*n*=229) with the Illumina HumanRef-8 v2 array as described in detail previously<sup>255</sup>. Genotyping was done using the Illumina HumanHap300, HumanHap370 or 610 Quad platform followed by imputation to HapMap II. In EGCUT, expression profiling was performed using Illumina HumanHT12v3 array while genotyping was performed with Illumina Human370CNV-duo chip followed by imputation to HapMap II as described previously<sup>256</sup>. Associations between genotype dosages and gene expression values were tested using linear regression models assuming additive genetic effects within 1 Mb windows. All 2,360 peripheral blood samples from three studies were then meta-analyzed using a z-score method, weighted for the sample size of each dataset.

# Author contributions Steering Committee Overseeing the Consortium

Goncalo R. Abecasis, Themistocles L. Assimes, Inês Barroso, Sonja I. Berndt, Michael Boehnke, Ingrid B. Borecki, Panos Deloukas, Caroline S. Fox, Timothy M. Frayling, Leif Groop, Iris M. Heid, Joel N. Hirschhorn, David J. Hunter, Erik Ingelsson, Robert C. Kaplan, Ruth J.F. Loos, Mark I. McCarthy, Karen L. Mohlke, Kari E. North, Jeffrey R. O'Connell, David Schlessinger, David P. Strachan, Unnur Thorsteinsdottir, Cornelia M. van Duijn

# **Writing Group**

Dmitry Shungin, Thomas W. Winkler, Damien C. Croteau-Chonka, Teresa Ferreira, Adam E. Locke, Reedik Mägi, Rona J. Strawbridge, Tune H. Pers, Anne E. Justice, Martin L. Buchkovich, Tamara S. Roman, Alexander W. Drong, M. Carola Zillikens, Mark I. McCarthy, Elizabeth K. Speliotes, Kari E. North, Caroline S. Fox, Inês Barroso, Paul W. Franks, Erik Ingelsson, Iris M. Heid, Ruth J.F. Loos, L. Adrienne Cupples, Andrew P. Morris, Cecilia M. Lindgren (co-chair), Karen L. Mohlke (co-chair)

# **Data Cleaning and Preparation**

Damien C. Croteau-Chonka, Felix R. Day, Tõnu Esko, Tove Fall, Teresa Ferreira, Stefan Gustafsson, Iris M Heid, Zoltán Kutalik, Cecilia M Lindgren, Adam E. Locke, Ruth J.F. Loos, Jian'an Luan, Reedik Mägi, Joshua C. Randall, Andre Scherag, Elizabeth K. Speliotes, Sailaja Vedantam, Thomas W. Winker, Andrew R. Wood, Tsegaselassie Workalemahu

# **GWAS** and Metabochip Meta-analyses

Damien C. Croteau-Chonka, L. Adrienne Cupples, Teresa Ferreira, Krista Fischer, Paul W. Franks, Iris M. Heid, Cecilia M. Lindgren, Adam E. Locke, Reedik Magi, Karen L. Mohlke, Lu Qi, Dmitry Shungin, Rona J. Strawbridge, Thomas W. Winkler, Tsegaselassie Workalemahu, Joseph M.W. Wu

# Biological, Enrichment, and Pathway analyses

Inês Barroso, Damien C. Croteau-Chonka, Alexander W. Drong, Teresa Ferreira, Caroline S. Fox, Stefan Gustafsson, Erik Ingelsson, Anne E. Justice, Bratati Kahili, Cecilia M. Lindgren, Adam E. Locke, Reedik Mägi, Karen L. Mohlke, Kari E. North, Tune H. Pers, Lu Qi, Tamara S. Roman, Dmitry Shungin, Rona J. Strawbridge

#### Conditional Analysis, Transethnic Meta-analysis and Fine-mapping

Damien C. Croteau-Chonka, L. Adrienne Cupples, Teresa Ferreira, Adam E. Locke, Cecilia M. Lindgren, Karen L. Mohlke, Andrew P. Morris, Kari E. North, Peter M. Visscher, Jian Yang

# Gene Expression (eQTL) Analyses

(Brain eQTL) Ruth J.F. Loos, Jing Hua Zhao; (PBMC) Tonu Esko, Lude Franke, Andres Metspalu, Eva Reinmaa, Harm-Jan Westra; (eQTL Liver/Omental/Subq eSNPs) Eric E. Schadt; (Lymphocytes) Jinyan Huang, Liming Liang, Baoshan Ma, Miriam F. Moffatt; (MolOBB) Alexander W. Drong, Cecilia M. Lindgren, Mark I. McCarthy, Fredrik Karpe, Josine L. Min, George Nicholson; (MuTHER) Åsa K. Hedman, Sarah Keildson, MuTHER Consortium; (ASAP) Per Eriksson, Lasse Folkersen, Anders Franco-Cereceda, Christian Olsson

# **Other Analyses and Contributions**

(DEPICT analysis) Rudolf Fehrmann, Lude Franke, Juha Karjalainen, Tune H. Pers, Joel Hirschhorn; (ENCODE analysis) Martin L. Buchkovich, Jin Chen, Karen L. Mohlke, Ellen M. Schmidt, Cristen J. Willer; (Heritability, phenotypic and genetic correlation analysis) Nancy L. Heard-Costa, Ci Song, Erik Ingelsson, L. Adrienne Cupples, Krista Fischer, Thomas W. Winkler, Iris M. Heid; (Heterogeneity and interaction analyses) Dmitry Shungin, Thomas W. Winkler, Adam E. Locke, Anne E. Justice, Andrew R. Wood, Ching-Ti Liu, Kari E North, Iris M. Heid, L. Adrienne Cupples

#### **GWAS Look-ups in Other Consortia**

(Adiponectin) Marie-France Hivert, ADIPOGen Consortium; (Blood pressure) ICBP (The International Consortium for Blood Pressure Genome-Wide Association Studies); (Bone mineral density) Edgar E. Vallejo, GEFOS Consortium; (Coronary artery disease) CARDIOGRAMplusC4D, Panos Deloukas, Stavroula Kanoni, Ruth McPherson; (Creatinine traits/kidney disease) Caroline S. Fox, CDKGen consortium; (Endometriosis) Grant W. Montgomery, Dale R. Nyholt, Krina T. Zondervan, International Endogene Consortium; (Glucose and insulin traits) Robert A. Scott, MAGIC (Meta-Analyses of Glucose and Insulin-Related Traits Consortium) investigators; (IgA Nephropathy) Murim Choi, Ali G. Gharavi, Krzysztof Kiryluk, Richard P. Lifton; (Lipids) Global Lipids Genetics Consortium; (Menarche and menopause) Joanne M. Murabito, John R.B. Perry, Lisette Stolk, ReproGen Consortium; (Nephropathy) Niina Sandholm, Eoin P. Brennan, Amy J. McKnight, Rany M. Salem, GENIE Consortium; (Type 2 diabetes) Andrew P. Morris

# **Project Design, Management and Coordination of Contributing Studies**

#### **METABOCHIP STUDIES**

(ADVANCE) Themistocles L Assimes, Thomas Quertermous; (ARIC Metabochip) Kari E North; (B1958C) Elina Hypponen, Chris Power; (BHS MC) John Beilby, Jennie Hui; (CLHNS) Linda S Adair, Karen L Mohlke; (DESIR) Stéphane Cauchi, Philippe Froguel; (DIAGEN) Stefan R Bornstein, Peter EH Schwarz; (DILGOM) Pekka Jousilahti, Antti M Jula, Satu Männistö, Markus Perola, Veikko Salomaa; (DPS) Matti Uusitupa; (DR's EXTRA) Timo A Lakka, Rainer Rauramaa; (Dundee - GoDarts) Colin NA Palmer; (EGCUT) Andres Metspalu; (ELY) Nita G Forouhi, Claudia Langenberg, Ruth JF Loos, Ken K Ong, Robert A Scott, Nicholas J Wareham; (EMIL (SWABIA)) Bernhard O Boehm; (EPIC-Norfolk) Nita G Forouhi, Claudia Langenberg, Ruth JF Loos, Ken K Ong, Robert A Scott, Nicholas J Wareham; (FBPP) Aravinda Chakravarti, Richard S Cooper, Steven C Hunt; (Fenland) Nita G Forouhi, Claudia Langenberg, Ruth JF Loos, Ken K Ong, Robert A Scott, Nicholas J Wareham; (FIN-D2D 2007) Sirkka M Keinanen-Kiukaanniemi, Timo E Saaristo; (FUSION stage 2) Francis S Collins, Jouko Saramies, Jaakko Tuomilehto; (GLACIER) Paul W Franks; (GxE) Richard S Cooper, Joel N Hirschhorn, Colin A McKenzie; (HNR) Raimund Erbel, Karl-Heinz Jöckel, Stefan Möhlenkamp; (HUNT 2) Kristian Hveem; (IMPROVE) Ulf de Faire, Anders Hamsten, Steve E Humphries, Elena Tremoli; (KORA S3 (MetaboChip)) Iris M Heid; (KORA S4 (MetaboChip)) Annette Peters, Konstantin Strauch, H-Erich Wichmann; (Leipzig adults) Michael Stumvoll; (LURIC) Winfried März; (METSIM) Johanna Kuusisto, Markku Laakso; (MORGAM) Philippe Amouyel, Dominique Arveiler, Jean Ferrières, Frank Kee, Kari Kuulasmaa, Giovanni Veronesi; (NSHD) Diana Kuh; (PIVUS) Erik Ingelsson; (PROMIS) John Danesh, Panos Deloukas, Danish Saleheen; (SardiNIA) Goncalo R Abecasis, David Schlessinger; (ScarfSheep) Ulf de Faire, Anders Hamsten; (SPT) Richard S Cooper, Joel N Hirschhorn, Colin A McKenzie; (STR) Erik Ingelsson; (Tandem) Murielle Bochud, Michel Burnier; (THISEAS) George Dedoussis,

Panos Deloukas; (Tromsø) Inger Njølstad; (ULSAM) Erik Ingelsson; (WHI Metabochip) Charles Kooperberg, Loic Le Marchand, Ulrike Peters; (Whitehall) Aroon D Hingorani, Mika Kivimaki, Nicholas J Wareham; (WTCCC-T2D) Cecilia M Lindgren, Mark I McCarthy; (DGE DietGeneExpression) Berit Johansen

#### **NEW GWAS**

(BLSA) Luigi Ferrucci; (DESIR) Stéphane Cauchi, Philippe Froguel; (EGCUT) Andres Metspalu, (ERF) Ben A Oostra, Cornelia M van Duijn; (FamHS) Ingrid B Borecki; (Health ABC) Tamara B Harris, Yongmei Liu; (HERITAGE Family Study) Claude Bouchard, Tuomo Rankinen, DC Rao,; (InCHIANTI) Timothy M Frayling, Luigi Ferrucci; (LifeLines) The LifeLines Cohort Study; (LLS) P Eline Slagboom; (LOLIPOP) John C Chambers, Jaspal S Kooner; (PREVEND) Pim van der Harst; (PROCARDIS) Anders Hamsten; (QFS) Claude Bouchard, André Marette, Louis Pérusse, Angelo Tremblay, Marie-Claude Vohl; (RISC) Timothy M Frayling, Ele Ferrannini, Mark Walker; (RSIII) Oscar H Franco, Albert Hofman, Fernando Rivadeneira, André G Uitterlinden, Cornelia M van Duijn, Jacqueline C Witteman, M Carola Zillikens; (SHIP-TREND) Hans-Jörgen Grabe; (TRAILS) Albertine J Oldehinkel, Harold Snieder; (TWINGENE) Erik Ingelsson; (TwinsUK) Tim D Spector; (BHS, Busselton Health Study) Alan L James, Arthur W Musk, Lyle J Palmer; (COROGENE) Markus Perola, Juha Sinisalo; (GOOD) Claes Ohlsson; (HBCS) Johan G Eriksson; (RSII) Oscar H Franco, Albert Hofman, Fernando Rivadeneira, André G Uitterlinden, Cornelia M van Duijn, Jacqueline C Witteman, M Carola Zillikens; (Sorbs) Anke Tönjes; (WGHS) Paul M Ridker; (Young Finns Study (YFS)) Terho Lehtimäki, Olli T Raitakari

#### PREVIOUS GWAS

(AGES) Vilmundur Gudnason, Tamara B Harris; (Amish) Alan R Shuldiner; (ARIC GWAS) Kari E North; (B58C T1D CONTROLS) David P Strachan; (B58C WTCCC) David P Strachan; (BRIGHT) Morris J Brown, Mark Caulfield, Patricia Munroe, Nilesh J Samani; (COLAUS) Peter Vollenweider; (CROATIA-Vis) Igor Rudan; (deCODE) Kari Stefansson, Unnur Thorsteinsdottir; (DGI) Leif C Groop; (EGCUT) Andres Metspalu; (EPIC-Norfolk) Nicholas J Wareham; (Fenland) Nicholas J Wareham; (Finnish Twin Cohort) Jaakko Kaprio; (FRAM) L Adrienne Cupples, Caroline S Fox; (FUSION (GWAS)) Richard N Bergman, Michael Boehnke; (H2000) Markku Heliövaara; (HPFS) David J. Hunter; (KORA S4 (GWA)) Christian Gieger; (MICROS) Andrew A Hicks, Peter P Pramstaller; (NFBC66) Marjo-Riitta Jarvelin; (NTRNESDA) Brenda W Penninx; (ORCADES) James F Wilson; (RSI) Oscar H Franco, Albert Hofman, Fernando Rivadeneira, André G Uitterlinden, Cornelia M van Duijn, Jacqueline C Witteman, M Carola Zillikens; (SHIP) Hans-Jörgen Grabe; (WTCCC-T2D) Cecilia M Lindgren, Mark I McCarthy

# **Genotyping of Contributing Studies**

#### **METABOCHIP STUDIES**

(ARIC Metabochip) Kari E North; (ADVANCE) Devin Absher, Themistocles L Assime, Thomas Quertermous; (B1958C) Christopher J Groves, Thorhildur Juliusdottir, Neil R Robertson; (BHS MC) Gillian M Arscott, Jennie Hui; (CLHNS) Damien C Croteau-Chonka; (DESIR) Elodie Eury, Stéphane Lobbens; (DIAGEN) Narisu Narisu; (Dundee - Go darts) Amanda J Bennett, Colin NA Palmer, Nigel W Rayner; (EGCUT) Tonu Esko, Lili Milani; (ELY) Claudia Langenberg, Ruth JF Loos, Ken K Ong, Nicholas J Wareham; (EMIL (SWABIA)) Bernhard O Boehm; (EPIC-Norfolk)

Claudia Langenberg, Ruth JF Loos, Ken K Ong, Nicholas J Wareham; (FBPP) Aravinda Chakravarti; (Fenland) Claudia Langenberg, Ruth JF Loos, Ken K Ong, Nicholas J Wareham; (FIN-D2D 2007) Peter S Chines; (FUSION stage 2) Leena Kinnunen; (GLACIER) Inês Barroso; (HNR) Thomas W Mühleisen; (HUNT 2) Amy J Swift; (KORA S4 (MetaboChip)) Harald Grallert, Peter Lichtner; (Leipzig adults) Yvonne Böttcher, Peter Kovacs; (LURIC) Marcus E Kleber; (NSHD) Diana Kuh, Ken K Ong, Andrew Wong; (PIVUS) Christian Berne, Erik Ingelsson, Lars Lind, Johan Sundström; (PROMIS) Kathleen Stirrups; (SardiNIA) Ramaiah Nagaraja, Serena Sanna; (ScarfSheep) Bruna Gigante; (STR) Nancy L Pedersen; (Tandem) Georg B Ehret, François Mach; (Tromsø) Michael R Erdos; (ULSAM) Erik Ingelsson, Ann-Christine Syvänen, Johan Ärnlöv; (WHI Metabochip) Charles Kooperberg, Ulrike Peters; (Whitehall) Claudia Langenberg; (WTCCC-T2D) Andrew T Hattersley, Mark I McCarthy; (DGE DietGeneExpression) Berit Johansen

#### **NEW GWAS**

(DESIR) Elodie Eury, Stéphane Lobbens, (EGCUT) Tonu Esko, Lili Milani; (ERF) Aaron Isaacs, Ben A Oostra, Cornelia M van Duijn; (FamHS) Ingrid B Borecki, E Warwick Daw, Mary F Feitosa, Aldi T Kraja, Mary K Wojczynski, Qunyuan Zhang; (Health ABC) Yongmei Liu; (HERITAGE Family Study) Tuomo Rankinen; (HYPERGENES) Chiara Troffa; (LifeLines) Morris A Swertz, The LifeLines Cohort Study; (LLS) Joris Deelen, Quinta Helmer; (LOLIPOP) John C Chambers, Jaspal S Kooner; (PREVEND) Irene Mateo Leach, Pim van der Harst; (PROCARDIS) Martin Farrall, Hugh Watkins; (QFS) Claire Bellis, John Blangero; (RSIII) Karol Estrada, Fernando Rivadeneira, André G Uitterlinde; (SHIP-TREND) Nele Friedrich, Georg Homuth, Uwe Völker; (TRAILS) Marcel Bruinenberg, Catharina A Hartman; (TWINGENE) Anders Hamsten, Nancy L Pedersen; (TwinsUK) Massimo Mangino, Alireza Moayyeri; (GOOD) Mattias Lorentzon, Claes Ohlsson; (RSII) Karol Estrada, Fernando Rivadeneira, André G Uitterlinden; (WGHS) Daniel I Chasman, Lynda M Rose; (Young Finns Study (YFS)) Terho Lehtimäki, Olli T Raitakari

#### **PREVIOUS GWAS**

(AGES) Albert Vernon Smith; (Amish) Jeffrey R O'Connell; (B58C T1D CONTROLS) Wendy L McArdle; (B58C WTCCC) Wendy L McArdle; (CROATIA-Vis) Caroline Hayward; (EGCUT) Mari Nelis; (EPIC-Norfolk) Nicholas J Wareham; (Fenland) Nicholas J Wareham; (Finnish Twin Cohort) Jaakko Kaprio; (KORA S3 (GWA)) Thomas Illig; (KORA S4 (GWA)) Martina Müller-Nurasyid; (MICROS) Andrew A Hicks; (NFBC66) Marjo-Riitta Jarvelin; (ORCADES) Harry Campbell; (RSI) Karol Estrada, Fernando Rivadeneira, André G Uitterlinden; (SHIP) Nele Friedrich, Georg Homuth, Uwe Völker; (WTCCC-T2D) Andrew T Hattersley, Mark I McCarthy

# **Phenotype Coordination of Contributing Studies**

#### **METABOCHIP STUDIES**

(ADVANCE) Alan S Go, Thomas Quertermous; (B1958C) Elina Hypponen, Chris Power; (BHS MC) Alan L James, Arthur W Musk; (CLHNS) Delia B Carba, Nanette R Lee; (DESIR) Fabrice Bonnet; (DIAGEN) Jürgen Gräßler, Gabriele Müller; (DPS) Jaana Lindström; (DR's EXTRA) Maija Hassinen; (Dundee - Go darts) Alex SF Doney, Andrew D Morris, Colin NA Palmer; (EGCUT) Tonu Esko, Andres Metspalu; (ELY) Nita G Forouhi, Nicholas J Wareham; (EMIL (SWABIA)) Bernhard O Boehm, Simone Claudi-Boehm, Wolfgang Kratzer, Sigrun Merger, Thomas

Seufferlein, Roman Wennauer; (EPIC-Norfolk) Nita G Forouhi, Nicholas J Wareham; (FBPP) Richard S Cooper, Steven C Hunt; (Fenland) Nita G Forouhi, Nicholas J Wareham; (GLACIER) Goran Hallmans, Ingegerd Johansson; (GxE)Terrence Forrester, Bamidele O Tayo; (HNR) Raimund Erbel, Karl-Heinz Jöckel, Stefan Möhlenkamp; (HUNT 2) Oddgeir Holmen; (KORA S3 (MetaboChip)) Wolfgang Koenig, Barbara Thorand; (KORA S4 (MetaboChip)) Annette Peters, H-Erich Wichmann; (Leipzig adults) Matthias Blüher; (METSIM) Heather M Stringham; (NSHD) Diana Kuh; (PIVUS) Christian Berne, Erik Ingelsson, Lars Lind, Johan Sundström; (PROMIS) Danish Saleheen; (SardinNIA) Antonella Mulas; (ScarfSheep) Karin Leander; (SPT) Terrence Forrester, Bamidele O Tayo; (STR) Nancy L Pedersen; (Tandem) Murielle Bochud, Michel Burnier; (THISEAS) Ioanna-Panagiota Kalafati; (Tromsø) Tom Wilsgaard; (ULSAM) Vilmantas Giedraitis, Erik Ingelsson, Johan Ärnlöv; (WHI Metabochip) Charles Kooperberg, Ulrike Peters; (Whitehall) Meena Kumari; (WTCCC-T2D) Amy Barrett, Andrew T Hattersley; (DGE DietGeneExpression) Ida H Caspersen, Berit Johansen

#### **NEW GWAS**

(DESIR) Fabrice Bonnet; (EGCUT) Tonu Esko, Andres Metspalu; (ERF) Ben A Oostra, Cornelia M van Duijn; (FamHS) Ingrid B Borecki, Mary F Feitosa; (Health ABC) Melissa E Garcia, Tamara B Harris, Michael A Nalls; (HERITAGE Family Study) Claude Bouchard, Tuomo Rankinen; (HYPERGENES) Chiara Troffa; (InCHIANTI) Luigi Ferrucci, Luigi Ferrucci; (LifeLines) Salome Scholtens, Morris A Swertz, Judith M Vonk, The LifeLines Cohort Study; (LLS) Simon P Mooijaart; (LOLIPOP) John C Chambers, Jagvir Grewal, Jaspal S Kooner, Ishminder K Kooner, Rebecca Mills; (PREVEND) Stephan JL Bakker, Ron T Gansevoort, Hans L Hillege; (PROCARDIS) Robert Clarke, Bengt Sennblad; (QFS) Claude Bouchard, Angelo Tremblay; (RSIII) Oscar H Franco, Albert Hofman, Fernando Rivadeneira, André G Uitterlinden, Cornelia M van Duijn, Jacqueline C Witteman; (SHIP-TREND) Marcus Dörr, Wolfgang Hoffmann, Till Ittermann; (TRAILS) Catharina A Hartman, Ronald P Stolk, Floor VA van Oort; (TWINGENE) Patrik KE Magnusson, Nancy L Pedersen; (TwinsUK) Massimo Mangin, Cristina Menni; (Busselton Health Study) Alan L James, Arthur W Musk; (GOOD) Mattias Lorentzon, Claes Ohlsson; (HBCS) Johan G Eriksson; (HYPERGENES) Nicola Glorioso, Jan A Staessen; (RSII) Oscar H Franco, Albert Hofman, Fernando Rivadeneira, André G Uitterlinden, Cornelia M van Duijn, Jacqueline C Witteman; (Sorbs) Anke Tönjes; (WGHS) Daniel I Chasman, Lynda M Rose; (Young Finns Study (YFS)) Terho Lehtimäki, Olli T Raitakari

#### **PREVIOUS GWAS**

(B58C T1D CONTROLS) David P Strachan; (Amish) Alan R Shuldiner; (B58C WTCCC) David P Strachan; (BRIGHT) Morris J Brown, Nilesh J Samani; (COLAUS) Peter Vollenweider; (CROATIA-Vis) Igor Rudan; (DGI) Valeriya Lyssenko; (EGCUT) Andres Metspalu; (EPIC-Norfolk) Nicholas J Wareham; (Fenland) Nicholas J Wareham; (Finnish Twin Cohort) Jaakko Kaprio; (FRAM) Caroline S Fox; (NFBC66) Karl-Heinz Herzig, Marjo-Riitta Jarvelin; (NTRNESDA) Eco JC de Geus; (ORCADES) Harry Campbell; (RSI) Oscar H Franco, Albert Hofman, Fernando Rivadeneira, André G Uitterlinden, Cornelia M van Duijn, Jacqueline C Witteman; (SHIP) Marcus Dörr, Wolfgang Hoffmann, Till Ittermann; (WTCCC-T2D) Amy Barrett, Andrew T Hattersley

# **Data Analysis**

#### **METABOCHIP STUDIES**

(ADVANCE) Devin Absher, Themistocles L Assimes, Lindsay L Waite; (ARIC Metabochip) Steven Buyske, Anne E Justice, Kari E North; (B1958C) Teresa Ferreira; (BHS MC) Denise Anderson; (CLHNS) Damien C Croteau-Chonka; (DESIR) Stéphane Cauchi, Loïc Yengo; (DIAGEN) Anne U Jackson, Gabriele Müller; (DILGOM) Kati Kristiansson; (Dundee - Go darts) Teresa Ferreira; (EGCUT) Tonu Esko, Krista Fischer, Evelin Mihailov; (ELY) Jian'an Luan; (EMIL (SWABIA)) Bernhard O Boehm; (EPIC-Norfolk) Jian'an Luan; (FBPP) Aravinda Chakravarti, Georg B Ehret; (Fenland) Jian'an Luan; (GLACIER) Frida Renstrom, Dmitry Shungin; (GxE) Cameron D Palmer; (HNR) Sonali Pechlivanis, André Scherag; (IMPROVE) Lasse Folkersen, Rona J Strawbridge; (KORA S3 (MetaboChip)) Matthias Olden, Janina S Ried, Thomas W Winkler; (KORA S4 (MetaboChip)) Eva Albrecht; (Leipzig adults) Anubha Mahajan, Inga Prokopenko; (LURIC) Graciela Delgado de Moissl, Tanja B Grammer, Marcus E Kleber, Hubert Scharnagl, Andreas Tomaschitz; (METSIM) Alena Stančáková, (NSHD) Jian'an Luan, Andrew Wong; (PIVUS) Stefan Gustafsson, Erik Ingelsson; (PROMIS) Stavroula Kanoni; (SardiNIA) Jennifer L Bragg-Gresham; (ScarfSheep) Lasse Folkersen, Rona J Strawbridge; (SPT) Cameron D Palmer; (STR) Stefan Gustafsson, Erik Ingelsson; (Tandem) Georg B Ehret, François Mach; (ULSAM) Stefan Gustafsson, Erik Ingelsson; (Whitehall) Jian'an Luan; (WHI Metabochip) Ewa Deelman, Marylyn Ritchie; (WTCCC-T2D) Teresa Ferreira, Anubha Mahajan, Andrew P Morris, Reedik Mägi; (DGE DietGeneExpression) Ida H Caspersen

#### **NEW GWAS**

(BLSA) Toshiko Tanaka; (DESIR) Stéphane Cauchi, Loïc Yengo; (EGCUT) Tonu Esko, Krista Fischer, Toomas Haller, Reedik Mägi; (ERF) Najaf Amin, Ayse Demirkan; (FamHS) Mary F Feitosa; (Health ABC) Michael A Nalls; (HERITAGE Family Study) Claude Bouchard, Tuomo Rankinen, DC Rao, Treva K Rice, Mark A Sarzynski, Yun Ju Sung; (InCHIANTI) Dorota Pasko, Toshiko Tanaka, Andrew R Wood; (LifeLines) Ilja M Nolte, Jana V Van Vliet-Ostaptchouk; (LLS) Marian Beekman, Stefan Böhringer; (LOLIPOP) Weihua Zhang; (PREVEND) Irene Mateo Leach, Pim van der Harst, Niek Verweij; (PROCARDIS) Anuj Goel; (QFS) John Blangero, Louis Pérusse; (RISC) Alain Golay, Dorota Pasko, Andrew R Wood; (RSIII) Karol Estrada, Carolina Medina-Gomez, Marjolein J Peters, Fernando Rivadeneira, André G Uitterlinden; (SHIP-TREND) Alexander Teumer, (TRAILS) Harold Snieder; (TWINGENE) Stefan Gustafsson, Erik Ingelsson; (TwinsUK) Massimo Mangino; (GOOD) Mattias Lorentzon, Claes Ohlsson; (HBCS) Niina Eklund; (RSII) Karol Estrada, Carolina Medina-Gomez, Marjolein J Peters, Fernando Rivadeneira, André G Uitterlinden; (Sorbs) Reedik Mägi; (WGHS) Daniel I Chasman, Lynda M Rose;

#### **PREVIOUS GWAS**

(AGES) Albert Vernon Smith; (Amish) Jeffrey R O'Connell; (ARIC GWAS) Keri L Monda, Kari E North; (B58C T1D CONTROLS) David P Strachan; (B58C WTCCC) David P Strachan; (BRIGHT) Patricia Munroe; (CHS) Barbara McKnight, Colleen M Sitlani; (COLAUS) Toby Johnson; (CROATIA-Vis) Caroline Hayward; (deCODE) Valgerdur Steinthorsdottir, Gudmar Thorleifsson; (EGCUT) Mari Nelis; (EPIC-Norfolk) Jing Hua Zhao; (Fenland) Jian'an Luan; (Finnish Twin Cohort) Kauko Heikkilä; (FRAM) L Adrienne Cupples, Nancy L Heard-Costa; (H2000) Niina Eklund; (HPFS) Lu Qi; (KORA S3 (GWA)) Claudia Lamina; (RSI) Karol Estrada, Carolina Medina-Gomez, Marjolein J Peters, Fernando Rivadeneira, André G Uitterlinden; (SHIP) Alexander Teumer; (WTCCC-T2D) Teresa Ferreira, Anubha Mahajan, Andrew P Morris, Reedik Mägi

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# **Contributing consortia**

## The ADIPOGen Consortium

Zari Dastani, 1\* Marie-France Hivert, 2,3\* Nicholas Timpson, 4\* John R.B Perry, 5,6\* Xin Yuan, 7\* Robert A. Scott, 8\* Peter Henneman, 9\* Iris M. Heid, 10\* Jorge R. Kizer, 11\* Leo-Pekka Lyytikainen, 12\* Christian Fuchsberger, <sup>13</sup>\* Toshiko Tanaka, <sup>14</sup> Andrew P. Morris, <sup>5</sup> Kerrin Small, <sup>15,16</sup> Aaron Isaacs, <sup>17,18</sup> Marian Beekman, <sup>19</sup> Stefan Coassin,<sup>20</sup> Kurt Lohman,<sup>21</sup> Lu Qi,<sup>22</sup> Stavroula Kanoni,<sup>16</sup> James S. Pankow,<sup>23</sup> Hae-Won Uh,<sup>24</sup> Ying Wu,<sup>25</sup> Aurelian Bidulescu, 26 Laura J. Rasmussen-Torvik, 27 Celia M.T. Greenwood, 28 Martin Ladouceur, 29 Jonna Grimsby, 3,30 Alisa K. Manning, 1 Ching-Ti Liu, 1 Jaspal Kooner, 2 Vincent E. Mooser, Peter Vollenweider, 33 Karen A. Kapur,<sup>34</sup> John Chambers,<sup>35</sup> Nicholas J. Wareham,<sup>8</sup> Claudia Langenberg,<sup>8</sup> Rune Frants,<sup>9</sup> Ko WillemsvanDijk, Ben A. Oostra, 18,36 Sara M. Willems, Claudia Lamina, Thomas Winkler, Bruce M. Psaty, 7,38 Russell P. Tracy,<sup>39</sup> Jennifer Brody,<sup>40</sup> Ida Chen,<sup>41</sup> Jorma Viikari,<sup>42</sup> Mika Kähönen,<sup>43</sup> Peter P. Pramstaller,<sup>44-46</sup> David M. Evans, <sup>4</sup> Beate St Pourcain, <sup>47</sup> Naveed Sattar, <sup>48</sup> Andy Wood, <sup>6</sup> Stefania Bandinelli, <sup>49</sup> Olga D. Carlson, <sup>50</sup> Josephine M. Egan,<sup>50</sup> Stefan Böhringer,<sup>51</sup> Diana van Heemst,<sup>52</sup> Lyudmyla Kedenko,<sup>53</sup> Kati Kristiansson,<sup>54</sup> Marja-Liisa Nuotio,<sup>54</sup> Britt-Marie Loo,<sup>55</sup> Tamara Harris,<sup>56</sup> Melissa Garcia,<sup>56</sup> Alka Kanaya,<sup>57</sup> Margot Haun,<sup>20</sup> Norman Klopp,<sup>58</sup> H. Erich Wichmann,<sup>58-60</sup> Panos Deloukas,<sup>16</sup> Efi Katsareli,<sup>61</sup> David J. Couper,<sup>62</sup> Bruce B. Duncan, 63,64 Margreet Kloppenburg, 65 Linda S. Adair, 66 Judith B. Borja, 67 DIAGRAM+ Consortium, MAGIC Consortium, GLGC Investigators, MuTHER Consortium, James G. Wilson, 68 Solomon Musani, 69 Xiuging Guo, 70 Toby Johnson, <sup>34,71,72</sup> Robert Semple, <sup>73</sup> Tanya M. Teslovich, <sup>13</sup> Matthew A. Allison, <sup>74</sup> Susan Redline, <sup>75</sup> Sarah G. Buxbaum, <sup>76</sup> Karen L. Mohlke, <sup>25</sup> Ingrid Meulenbelt, <sup>77</sup> Christie M. Ballantyne, <sup>78</sup> George V. Dedoussis, <sup>61</sup> Frank B. Hu,<sup>22</sup> Yongmei Liu,<sup>21</sup> Bernhard Paulweber,<sup>53</sup> Timothy D. Spector,<sup>15</sup> P. Eline Slagboom,<sup>19</sup> Luigi Ferrucci,<sup>14</sup> Antti Jula, 55 Markus Perola, 54 Olli Raitakari, 79 Jose C. Florez, 30,80-82 Veikko Salomaa, 83 Johan G. Eriksson, 84 Timothy M. Frayling, Andrew A Hicks, 44 Terho Lehtimäki, 2 George Davey Smith, David S. Siscovick, 85 Florian Kronenberg, <sup>20</sup> Cornelia van Duijn, <sup>17,18</sup> Ruth J.F. Loos, <sup>8</sup> Dawn M. Waterworth, <sup>7</sup> James B. Meigs, <sup>3,30</sup> Josee Dupuis, 31,86 John Brent Richards. 15,87

- 1. Department of Epidemiology, Biostatistics and Occupational Health. Lady Davis Institute, Jewish General Hospital, McGill University, Montreal, Quebec H3T 1E2, Canada.
- 2. Department of Medicine, Université de Sherbrooke, Sherbrooke, Québec, Canada.
- 3. General Medicine Division, Massachusetts General Hospital, Boston, MA, USA.
- 4. MRC CAiTE Centre & School of Social and Community and Medicine, University of Bristol, Bristol, UK, Oakfield House, Oakfield Grove, Bristol, BS8
- 5. Wellcome Trust Centre for Human Genetics, University of Oxford, Oxford OX3 7BN, UK.
- 6. Genetics of Complex Traits, Peninsula Medical School, University of Exeter, UK.
- 7. Genetics, GlaxoSmithKline, King of Prussia, PA, USA.
- 8. MRC Epidemiology Unit, Institute of Metabolic Science, Addenbrooke's Hospital, Cambridge CB2 0QQ, UK.
- 9. Department of Human Genetics, Leiden University Medical Center, Leiden, The Netherlands.
- 10. Department of Epidemiology and Preventive Medicine, Regensburg University Medical Center, 93053 Regensburg, Germany.

- 11. Departments of Medicine and Public Health, Weill Cornell Medical College, New York, NY, USA.
- 12. Department of Clinical Chemistry, University of Tampere and Tampere University Hospital, Tampere 33521, Finland.
- 13. Center for Statistical Genetics, Department of Biostatistics, University of Michigan, Ann Arbor, MI 48109, USA.
- 14. Clinical Research Branch, National Institute on Aging, Baltimore, MD 21250, USA.
- 15. Department of Twin Research and Genetic Epidemiology, King's College London, London SE1 7EH, UK.
- 16. Wellcome Trust Sanger Institute, Wellcome Trust Genome Campus, Hinxton, UK.
- 17. Genetic Epidemiology Unit, Department of Epidemiology, Erasmus Medical Center, Rotterdam, The Netherlands.
- 18. Centre for Medical Systems Biology, Leiden, the Netherlands.
- 19. Molecular Epidemiology, Leiden University Medical Center, Leiden, 2300 RC, The Netherlands.
- 20. Division of Genetic Epidemiology, Innsbruck Medical University, 6020 Innsbruck, Austria.
- 21. Wake Forest University School of Medicine, Winston-Salem, North Carolina 27157, USA.
- 22. Harvard School of Public Health, Boston, MA 02115, USA.
- 23. Division of Epidemiology and Community Health, University of Minnesota, Minneapolis, MN, USA.
- 24. Department of Medical Statistics and Bioinformatics, Leiden, 2333 ZC, The Netherlands.
- 25. Department of Genetics, University of North Carolina, Chapel Hill, NC, USA.
- 26. Cardiovascular Research Institute, Morehouse School of Medicine, Atlanta, GA 30310-1495, USA.
- 27. Department of Preventive Medicine, Chicago, IL, USA.
- 28. Lady Davis Institute for Medical Research, Department of Oncology, McGill University, Montreal, Quebec H3T 1E2, Canada.
- 29. Department of Human genetics, McGill University, Montreal, Quebec H3T 1E2, Canada.
- 30. Department of Medicine, Harvard Medical School, Boston, MA, USA.
- 31. Department of Biostatistics, Boston University School of Public Health, Boston, MA, USA.
- 32. Cardiology, Ealing Hospital National Health Service (NHS) Trust, London, UK.
- 33. Department of Internal Medicine, 1011 Lausanne, Switzerland.
- 34. Department of Medical Genetics, University of Lausanne, 1005 Lausanne, Switzerland.
- 35. Epidemiology and Biostatistics, Imperial College London, London, UK.
- 36. Deptartment of Clinical Genetics and Department of Epidemiology, Erasmus Medical Center, Rotterdam, The Netherlands.
- 37. Cardiovascular Health Research Unit, Departments of Medicine and Epidemiology, University of Washington, Seattle, WA, USA.
- 38. Group Health Research Institute, Group Health Cooperative, Seattle, WA.
- 39. Departments of Pathology and Biochemistry, University of Vermont, Burlington, VT, USA.
- 40. Cardiovascular Health Research Unit, Seattle, WA, USA.
- 41. Medical Genetics Research Institute, Cedars Sinai Medical Center, Los Angeles, CA, USA.
- 42. Department of Medicine, University of Turku and Turku University Hospital, Turku 20521, Finland.
- 43. Department of Clinical Physiology, University of Tampere and Tampere University Hospital, Tampere 33521, Finland.
- 44. Institute of Genetic Medicine, European Academy Bozen/Bolzano (EURAC), Bolzano, Italy- Affiliated Institute of the University of Lübeck, Lübeck, Germany.
- 45. Department of Neurology, General Central Hospital, Bolzano, Italy.
- 46. Department of Neurology, University of Lübeck, Lübeck, Germany.
- 47. School of Social and community medicine, University of Bristol, UK, Oakfield House, Oakfield Grove, Bristol, BS8 2BN, UK.
- 48. British Heart Foundation Glasgow Cardiovascular Research Centre, University of Glasgow, Glasgow, United Kingdom. Wolfson Medical School Building, University Avenue, Glasgow, G12 8QQ, UK.
- 49. Geriatric Unit, Azienda Sanitaria Firenze (ASF), Florence, Italy.
- 50. Laboratory of Clinical Investigation, National Institute of Aging, Baltimore, MD, USA.
- 51. Medical Statistics and Bioinformatics, Leiden University Medical Center, Leiden, 2333 ZC, The Netherlands.
- 52. Gerontology and Geriatrics, Leiden University Medical Center, Leiden, 2300 RC, The Netherlands.
- 53. First Department of Internal Medicine, St. Johann Spital, Paracelsus Private Medical University Salzburg, 5020 Salzburg, Austria.
- 54. Public Health Genomics Unit, Department of Chronic Disease Prevention, National Institute for Health and Welfare, Helsinki, Finland, Institute for Molecular Medicine Finland FIMM, University of Helsinki, Finland.

- 55. Population Studies Unit, Department of Chronic Disease Prevention, National Institute for Health and Welfare, Turku, Finland.
- 56. Intramural Research Program, Laboratory of Epidemiology, Demography, and Biometry, National Institute on Aging, NIH.
- 57. Division of General Internal Medicine, Women's Health Clinical Research Center, University of California, San Francisco, California, USA.
- 58. Institute of Epidemiology, Helmholtz Zentrum München, German Research Center for Environmental Health, Germany.
- 59. Institute of Medical Informatics, Biometry and Epidemiology, Ludwig-Maximilians-Universität, Munich, Germany.
- 60. Klinikum Großhadern, Munich, Germany.
- 61. Harokopio University, Athens, Greece.
- 62. Collaborative Studies Coordinating Center, Department of Biostatistics, University of North Carolina at Chapel Hill, Chapel Hill, NC, USA.
- 63. School of Medicine, Federal University of Rio Grande do Sul, Porto Alegre, Brazil.
- 64. Department of Epidemiology, University of North Carolina at Chapel Hill, Chapel Hill, NC, USA.
- 65. Department of Rheumatology and Department of Clinical Epidemiology, Leiden, 2300 RC, The Netherlands.
- 66. Department of Nutrition, University of North Carolina, Chapel Hill, NC, USA.
- 67. Office of Population Studies Foundation, University of San Carlos, Cebu City, Philippines.
- 68. Department of Physiology and Biophysics, University of Mississippi Medical Center, Jackson, MS 39216, USA.
- 69. Department of Medicine, University of Mississippi Medical Center, Jackson, MS 39213, USA.
- 70. Medical Genetics Institute, Los Angeles, CA, USA.
- 71. University Institute of Social and Preventative Medicine, Centre Hospitalier Universitaire Vaudois (CHUV) and University of Lausanne, Lausanne, Switzerland.
- 72. Swiss Institute of Bioinformatics, Lausanne, Switzerland.
- 73. Metabolic Research Laboratories, Institute of Metabolic Science, University of Cambridge, Addenbrooke's Hospital, Cambridge, United Kingdom.
- 74. Department of Family and Preventive Medicine, La Jolla, CA, USA.
- 75. Brigham and Women's Hospital, Boston, MA 02115, USA.
- 76. Jackson Heart Study Coordinating Center, Jackson State University, Jackson, MS 39213, USA.
- 77. Section of Molecular Epidemiology, Leiden University Medical Center & The Netherlands Genomics Initiative-Sponsored by the Netherlands Consortium for Healthy Aging, Leiden, 2333 ZC, The Netherlands.
- 78. Baylor College of Medicine and Methodist DeBakey Heart and Vascular Center, Houston, TX, USA.
- 79. Research Centre of Applied and Preventive Cardiovascular Medicine, University of Turku and the Department of Clinical Physiology, Turku University Hospital, Turku 20521, Finland.
- 80. Chronic Disease Epidemiology and Prevention Unit, Department of Chronic Disease Prevention, National Institute for Health and Welfare, Helsinki, Finland.
- 81. Diabetes Prevention Unit, Department of Chronic Disease Prevention, National Institute for Health and Welfare, Helsinki, Finland, Unit of General Practice, Helsinki University Central Hospital, Helsinki, Finland, Folkhalsan Research Centre, Helsinki, Finland, Vaasa Central Hospital, Vasa, Finland, Department of General Practice and Primary Health Care, University of Helsinki, Finland.
- 82. University of Washington, Seattle, WA, USA.
- 83. Program in Medical and Population Genetics, Broad Institute, Cambridge, MA, USA.
- 84. Center for Human Genetic Research, Massachusetts General Hospital, Boston, MA, USA.
- 85. Diabetes Research Center, Diabetes Unit, Massachusetts General Hospital, Boston, MA, USA.
- 86. National Heart, Lung, and Blood Institute's Framingham Heart Study, Framingham, MA, USA.
- 87. Departments of Medicine, Human Genetics, Epidemiology and Biostatistics. Lady Davis Institute, Jewish General Hospital, McGill University Montreal, Quebec H3T 1E2, Canada.

## The CARDIoGRAMplusC4D Consortium

Panos Deloukas<sup>1</sup>, Stavroula Kanoni<sup>1</sup>, Christina Willenborg<sup>2</sup>, Martin Farrall<sup>3,4</sup>, Themistocles L Assimes<sup>5</sup>, John R Thompson<sup>6</sup>, Erik Ingelsson<sup>7</sup>, Danish Saleheen<sup>8-10</sup>, Jeanette Erdmann<sup>2</sup>, Benjamin A Goldstein<sup>5</sup>, Kathleen Stirrups<sup>1</sup>, Inke R König<sup>11</sup>, Jean-Baptiste Cazier<sup>4</sup>, Åsa Johansson<sup>12</sup>, Alistair S Hall<sup>13</sup>, Jong-Young Lee<sup>14</sup>, Cristen J Willer<sup>15,16</sup>, John C Chambers<sup>17</sup>, Tõnu Esko<sup>18,19</sup>, Lasse Folkersen<sup>20,21</sup>, Anuj Goel<sup>3,4</sup>, Elin Grundberg<sup>22</sup>, Aki S Havulinna<sup>23</sup>, Weang K Ho<sup>10</sup>, Jemma C Hopewell<sup>24,25</sup>, Niclas Eriksson<sup>12</sup>, Marcus E Kleber<sup>26,27</sup>, Kati Kristiansson<sup>23</sup>, Per Lundmark<sup>28</sup>, Leo-Pekka Lyytikäinen<sup>29,30</sup>, Suzanne Rafelt<sup>31</sup>, Dmitry Shungin<sup>32–34</sup>, Rona J Strawbridge<sup>20,21</sup>, Gudmar Thorleifsson<sup>35</sup>, Emmi Tikkanen<sup>36,37</sup>, Natalie Van Zuydam<sup>38</sup>, Benjamin F Voight<sup>39</sup>, Lindsay L Waite<sup>40</sup>, Weihua Zhang<sup>17</sup>, Andreas Ziegler<sup>11</sup>, Devin Absher<sup>40</sup>, David Altshuler<sup>41-</sup> <sup>44</sup>, Anthony J Balmforth<sup>45</sup>, Inês Barroso<sup>1,46</sup>, Peter S Braund<sup>31,47</sup>, Christof Burgdorf<sup>48</sup>, Simone Claudi-Boehm<sup>49</sup>, David Cox<sup>50</sup>, Maria Dimitriou<sup>51</sup>, Ron Do<sup>41,43</sup>, CARDIOGENICS Consortium<sup>52</sup>, DIAGRAM Consortium<sup>52</sup>, Alex S F Doney<sup>38</sup>, NourEddine El Mokhtari<sup>53</sup>, Per Eriksson<sup>20,21</sup>, Krista Fischer<sup>18</sup>, Pierre Fontanillas<sup>41</sup>, Anders Franco-Cereceda<sup>54</sup>, Bruna Gigante<sup>55</sup>, Leif Groop<sup>56</sup>, Stefan Gustafsson<sup>7</sup>, Jörg Hager<sup>57</sup>, Göran Hallmans<sup>58</sup>, Bok-Ghee Han<sup>14</sup>, Sarah E Hunt<sup>1</sup>, Hyun M Kang<sup>59</sup>, Thomas Illig<sup>60</sup>, Thorsten Kessler<sup>48</sup>, Joshua W Knowles<sup>5</sup>, Genovefa Kolovou<sup>61</sup>, Johanna Kuusisto<sup>62</sup>, Claudia Langenberg<sup>63</sup>, Cordelia Langford<sup>1</sup>, Karin Leander<sup>55</sup>, Marja-Liisa Lokki<sup>64</sup>, Anders Lundmark<sup>28</sup>, Mark I McCarthy<sup>3,65,66</sup>, Christa Meisinger<sup>67</sup>, Olle Melander<sup>56</sup>, Evelin Mihailov<sup>19</sup>, Seraya Maouche<sup>68</sup>, Andrew D Morris<sup>38</sup>, Martina Müller-Nurasyid<sup>69–72</sup>, MuTHER Consortium<sup>52</sup>, Kjell Nikus<sup>73</sup>, John F Peden<sup>3</sup>, N William Rayner<sup>3</sup>, Asif Rasheed<sup>9</sup>, Silke Rosinger<sup>74</sup>, Diana Rubin<sup>53</sup>, Moritz P Rumpf<sup>48</sup>, Arne Schäfer<sup>75</sup>, Mohan Sivananthan<sup>76,77</sup>, Ci Song<sup>7</sup>, Alexandre F R Stewart<sup>78,79</sup>, Sian-Tsung Tan<sup>80</sup>, Gudmundur Thorgeirsson<sup>81,82</sup>, C Ellen van der Schoot<sup>83</sup>, Peter J Wagner<sup>36,37</sup>, Wellcome Trust Case Control Consortium<sup>52</sup>, George A Wells<sup>78,79</sup>, Philipp S Wild<sup>84,85</sup>, Tsun-Po Yang<sup>1</sup>, Philippe Amouyel<sup>86</sup>, Dominique Arveiler<sup>87</sup>, Hanneke Basart<sup>88</sup>, Michael Boehnke<sup>59</sup>, Eric Boerwinkle<sup>89</sup>, Paolo Brambilla<sup>90</sup>, Francois Cambien<sup>68</sup>, Adrienne L Cupples<sup>91,92</sup>, Ulf de Faire<sup>55</sup>, Abbas Dehghan<sup>93</sup>, Patrick Diemert<sup>94</sup>, Stephen E Epstein<sup>95</sup>, Alun Evan<sup>896</sup>, Marco M Ferrario<sup>97</sup>, Jean Ferrières<sup>98</sup>, Dominique Gauguier<sup>3,99</sup>, Alan S Go<sup>100</sup>, Alison H Goodall<sup>31,47</sup>, Villi Gudnason81,101, Stanley L Hazen<sup>102</sup>, Hilma Holm<sup>35</sup>, Carlos Iribarren<sup>100</sup>, Yangsoo Jang<sup>103</sup>, Mika Kähönen<sup>104</sup>, Frank Kee<sup>105</sup>, Hyo-Soo Kim<sup>106</sup>, Norman Klopp<sup>60</sup>, Wolfgang Koenig<sup>107</sup>, Wolfgang Kratzer<sup>108</sup>, Kari Kuulasmaa<sup>23</sup>, Markku Laakso<sup>62</sup>, Reijo Laaksonen<sup>108</sup>, Ji-Young Lee<sup>14</sup>, Lars Lind<sup>28</sup>, Willem H Ouwehand<sup>1,109,110</sup>, Sarah Parish<sup>24,25</sup>, Jeong E Park<sup>111</sup>, Nancy L Pedersen<sup>7</sup>, Annette Peters<sup>67,112</sup>, Thomas Quertermous<sup>5</sup>, Daniel J Rader<sup>113</sup>, Veikko Salomaa<sup>23</sup>, Eric Schadt<sup>114</sup>, Svati H Shah<sup>115,116</sup>, Juha Sinisalo<sup>117</sup>, Klaus Stark<sup>118</sup>, Kari Stefansson<sup>35,81</sup>, David-Alexandre Trégouët<sup>68</sup>, Jarmo Virtamo<sup>23</sup>, Lars Wallentin<sup>12</sup>, Nicholas Wareham<sup>63</sup>, Martina E Zimmermann<sup>118</sup>, Markku S Nieminen<sup>117</sup>, Christian Hengstenberg<sup>118</sup>, Manjinder S Sandhu<sup>1,63</sup>, Tomi Pastinen<sup>119</sup>, Ann-Christine Syvänen<sup>28</sup>, G Kees Hovingh<sup>88</sup>, George Dedoussis<sup>51</sup>, Paul W Franks<sup>32–34,120</sup>, Terho Lehtimäki<sup>29,30</sup>, Andres Metspalu<sup>18,19</sup>, Pierre A Zalloua<sup>121</sup>, Agneta Siegbahn<sup>12</sup>, Stefan Schreiber<sup>94</sup>, Samuli Ripatti<sup>1,37</sup>, Stefan S Blankenberg<sup>74</sup>, Markus Perola<sup>23</sup>, Robert Clarke<sup>24,25</sup>, Bernhard O Boehm<sup>74</sup>, Christopher O'Donnell<sup>93</sup>, Muredach P Reilly<sup>122</sup>, Winfried

März<sup>26,123</sup>, Rory Collins<sup>24,25</sup>, Sekar Kathiresan<sup>41,124,125</sup>, Anders Hamsten<sup>20,21</sup>, Jaspal S Kooner<sup>80</sup>, Unnur Thorsteinsdottir<sup>35,81</sup>, John Danesh<sup>9</sup>, Colin N A Palmer<sup>38</sup>, Robert Roberts<sup>78,79</sup>, Hugh Watkins<sup>3,4</sup>, Heribert Schunkert<sup>2</sup> & Nilesh J Samani<sup>31,47</sup>

- 1. Wellcome Trust Sanger Institute, Hinxton, Cambridge, UK.
- 2. Institut für Integrative und Experimentelle Genomik, Universität zu Lübeck, Lübeck, Germany.
- 3. Wellcome Trust Centre for Human Genetics, University of Oxford, Oxford, UK.
- 4. Cardiovascular Medicine, Radcliffe Department of Medicine, University of Oxford, Oxford, UK.
- 5. Department of Medicine, Stanford University School of Medicine, Stanford, California, USA.
- 6. Department of Health Sciences, University of Leicester, Leicester, UK.
- 7. Department of Medical Epidemiology and Biostatistics, Karolinska Institutet, Stockholm, Sweden.
- 8. Center for Non-Communicable Diseases, Karachi, Pakistan.
- 9. Department of Public Health and Primary Care, University of Cambridge, Cambridge, UK.
- 10. Department of Medicine, University of Pennsylvania, Philadelphia, Pennsylvania, USA.
- 11. Institut für Medizinische Biometrie und Statistik, Universität zu Lübeck, Lübeck, Germany.
- 12. Uppsala Clinical Research Center, Uppsala University, Uppsala, Sweden.
- 13. Division of Cardiovascular and Neuronal Remodelling, Multidisciplinary Cardiovascular Research Centre, Leeds Institute of Genetics, Health and Therapeutics, University of Leeds, Leeds, UK.
- 14. Center for Genome Science, Korea National Institute of Health, Korea Center for Disease Control and Prevention, Yeonje-ri, Chungwon-gun, Chungcheongbuk-do, Korea.
- 15. Division of Cardiovascular Medicine, Department of Internal Medicine, University of Michigan, Ann Arbor, Michigan, USA.
- 16. Department of Human Genetics, University of Michigan, Ann Arbor, Michigan, USA.
- 17. Department of Epidemiology and Biostatistics, Imperial College London, London, UK.
- 18. Estonian Genome Center, University of Tartu, Tartu, Estonia.
- 19. Institute of Molecular and Cell Biology, University of Tartu, Tartu, Estonia.
- 20. Atherosclerosis Research Unit, Department of Medicine, Karolinska Institutet, Stockholm, Sweden.
- 21. Center for Molecular Medicine, Karolinska University Hospital, Stockholm, Sweden.
- 22. Department of Twin Research and Genetic Epidemiology, King's College London, London, UK.
- 23. Department of Chronic Disease Prevention, National Institute for Health and Welfare, Helsinki, Finland.
- 24. Clinical Trial Service Unit, University of Oxford, Oxford, UK.
- 25. Epidemiological Studies Unit, University of Oxford, Oxford, UK.
- 26. Mannheim Institute of Public Health, Social and Preventive Medicine, Medical Faculty of Mannheim, University of Heidelberg, Mannheim, Germany.
- 27. Ludwigshafen Risk and Cardiovascular Health (LURIC) Study, Freiburg, Germany.
- 28. Department of Medical Sciences, Uppsala University, Uppsala, Sweden.
- 29. Department of Clinical Chemistry, Fimlab Laboratories, Tampere University Hospital, Tampere, Finland.
- 30. Department of Clinical Chemistry, University of Tampere School of Medicine, Tampere, Finland
- 31. Department of Cardiovascular Sciences, University of Leicester, Glenfield Hospital, Leicester, UK.
- 32. Genetic & Molecular Epidemiology Unit, Department of Clinical Sciences, Lund University Diabetes Center, Skåne University Hospital, Malmö, Sweden.
- 33. Department of Public Health & Clinical Medicine, Genetic Epidemiology & Clinical Research Group, Section for Medicine, Umeå University, Umeå, Sweden
- 34. Department of Odontology, Umeå University, Umeå, Sweden.
- 35. deCODE Genetics, Reykjavik, Iceland.
- 36. Institute for Molecular Medicine FIMM, University of Helsinki, Helsinki, Finland.
- 37. Public Health Genomics Unit, National Institute for Health and Welfare, Helsinki, Finland.
- 38. Medical Research Institute, University of Dundee, Ninewells Hospital and Medical School, Dundee, UK

- 39. Department of Pharmacology, University of Pennsylvania, Philadelphia, Pennsylvania, USA.
- 40. HudsonAlpha Institute for Biotechnology, Huntsville, Alabama, USA.
- 41. Broad Institute of Harvard and MIT, Cambridge, Massachusetts, USA.
- 42. Department of Molecular Biology, Massachusetts General Hospital, Boston, Massachusetts, USA.
- 43. Center for Human Genetic Research, Massachusetts General Hospital, Boston, Massachusetts, USA.
- 44. Department of Genetics, Harvard Medical School, Boston, Massachusetts, USA.
- 45. Division of Cardiovascular and Diabetes Research, Multidisciplinary Cardiovascular Research Centre, Leeds Institute of Genetics, Health and Therapeutics, University of Leeds, Leeds, UK.
- 46. University of Cambridge Metabolic Research Laboratories, Institute of Metabolic Science, Addenbrooke's Hospital, Cambridge, UK.
- 47. National Institute for Health Research (NIHR) Leicester Cardiovascular Biomedical Research Unit, Glenfield Hospital, Leicester, UK.
- 48. Deutsches Herzzentrum München, Technische Universität München, Munich, Germany.
- 49. Practice of Gynecology, Ulm University Medical Centre, Ulm, Germany.
- 50. Biotherapeutics and Bioinnovation Center, Pfizer, South San Francisco, California, USA.
- 51. Department of Dietetics-Nutrition, Harokopio University, Athens, Greece.
- 52. A list of members and affiliations appears in the Supplementary Note.
- 53. Klinik für Innere Medizin, Kreiskrankenhaus Rendsburg, Rendsburg, Germany.
- 54. Cardiothoracic Surgery Unit, Department of Molecular Medicine and Surgery, Karolinska Institutet, Stockholm, Sweden.
- 55. Division of Cardiovascular Epidemiology, Institute of Environmental Medicine, Karolinska Institutet, Stockholm, Sweden.
- 56. Department of Clinical Sciences, Diabetes and Endocrinology, Lund University, University Hospital Malmö, Malmö, Sweden.
- 57. CEA-Genomics Institute, National Genotyping Centre, Paris, France. Commissariat à l'énergie atomique et aux energies alternatives]
- 58. Department of Public Health & Clinical Medicine, Section for Nutritional Research, Umeå University, Umeå, Sweden.
- 59. Department of Biostatistics, Center for Statistical Genetics, University of Michigan, Ann Arbor, Michigan USA.
- 60. Hannover Unified Biobank, Hannover Medical School, Hannover, Germany.
- 61. First Cardiology Department, Onassis Cardiac Surgery Center 356, Athens, Greece.
- 62. Department of Medicine, University of Eastern Finland and Kuopio University Hospital, Kuopio, Finland.
- 63. Medical Research Council (MRC) Epidemiology Unit, Institute of Metabolic Science, Addenbrooke's Hospital, Cambridge, UK.
- 64. Transplantation Laboratory, Haartman Institute, University of Helsinki, Helsinki, Finland.
- 65. Oxford Centre for Diabetes, Endocrinology and Metabolism, University of Oxford, Oxford, UK.
- 66. Oxford NIHR Biomedical Research Centre, Churchill Hospital, Oxford, UK.
- 67. Institute of Epidemiology II, Helmholtz Zentrum München-German Research Center for Environmental Health, Neuherberg, Germany.
- 68. Institut National de la Santé et la Recherche Médicale (INSERM) Unité Mixte de Recherche (UMR) S937, Institute for Cardiometabolism and Nutrition (ICAN), Pierre and Marie Curie (Paris 6) University, Paris, France.
- 69. Department of Medicine I, University Hospital Grosshadern, Ludwig-Maximilians-Universität, Munich, Germany.
- 70. Chair of Epidemiology, Institute of Medical Informatics, Biometry and Epidemiology, Ludwig-Maximilians-Universität, Munich, Germany.
- 71. Chair of Genetic Epidemiology, Institute of Medical Informatics, Biometry and Epidemiology, Ludwig-Maximilians-Universität, Munich, Germany.
- 72. Institute of Genetic Epidemiology, Helmholtz Zentrum München-German Research Center for Environmental Health, Neuherberg, Germany.
- 73. Heart Centre, Department of Cardiology, Tampere University Hospital, Tampere, Finland.
- 74. Division of Endocrinology and Diabetes, Department of Internal Medicine, Ulm University Medical Centre, Ulm, Germany.
- 75. Institut für Klinische Molekularbiologie, Christian-Albrechts Universität, Kiel, Germany.
- 76. Division of Epidemiology, Multidisciplinary Cardiovascular Research Centre (MCRC) University of Leeds, Leeds, UK.
- 77. Leeds Institute of Genetics, Health and Therapeutics, University of Leeds, Leeds, UK.
- 78. University of Ottawa Heart Institute, Cardiovascular Research Methods Centre Ontario, Ottawa, Ontario, Canada.
- 79. Ruddy Canadian Cardiovascular Genetics Centre, Ottawa, Ontario, Canada.
- 80. National Heart and Lung Institute (NHLI), Imperial College London, Hammersmith Hospital, London, UK.
- 81. Faculty of Medicine, University of Iceland, Reykjavik, Iceland.
- 82. Department of Medicine, Landspitali University Hospital, Reykjavik, Iceland.
- 83. Department of Experimental Immunohematology, Sanquin, Amsterdam, The Netherlands.

- 84. Center for Thrombosis and Hemostasis, University Medical Center Mainz, Johannes Gutenberg University Mainz, Mainz, Germany.
- 85. Department of Medicine 2, University Medical Center Mainz, Johannes Gutenberg University Mainz, Mainz, Germany.
- 86. Institut Pasteur de Lille, INSERM U744, Université Lille Nord de France, Lille, France.
- 87. Department of Epidemiology and Public Health, EA3430, University of Strasbourg, Strasbourg, France.
- 88. Department of Vascular Medicine, Academic Medical Center, Amsterdam, The Netherlands.
- 89. Human Genetics Center, University of Texas Health Science Center, Houston, Texas, USA.
- 90. Department of Experimental Medicine, University of Milano-Bicocca, Monza, Italy.
- 91. Department of Biostatistics, Boston University School of Public Health, Boston, Massachusetts, USA
- 92. National Heart, Lung, and Blood Institute's Framingham Heart Study, Framingham, Massachusetts, USA.
- 93. Department of Epidemiology, Erasmus Medical Center, Rotterdam, The Netherlands.
- 94. Clinic for General and Interventional Cardiology, University Heart Center Hamburg, Hamburg, Germany.
- 95. Cardiovascular Research Institute, Washington Hospital Center, Washington, DC, USA.
- 96. Centre for Public Health, The Queen's University of Belfast, Belfast, UK.
- 97. Research Centre for Epidemiology and Preventive Medicine (EPIMED), Department of Clinical and Experimental Medicine, University of Insubria, Varese, Italy.
- 98. Department of Cardiology, Toulouse University School of Medicine, Rangueil Hospital, Toulouse, France.
- 99. INSERM UMR S872, Cordeliers Research Centre, Paris, France.
- 100. Division of Research, Kaiser Permanente Northern California, Oakland, California, USA.
- 101. Icelandic Heart Association, Kopavogur, Iceland.
- 102. Lerner Research Institute, Cleveland Clinic, Cleveland, Ohio, USA.
- 103. Cardiology Division, Department of Internal Medicine, Cardiovascular Genome Center, Yonsei University, Seoul, Korea.
- 104. Department of Clinical Physiology, Tampere University Hospital and University of Tampere, Tampere, Finland.
- 105. UK Clinical Research Collaboration (UKCRC) Centre of Excellence for Public Health (Northern Ireland), Queen's University of Belfast, Belfast, UK.
- 106. Department of Internal Medicine, Cardiovascular Center, Seoul National University Hospital, Seoul, Korea.
- 107. Department of Internal Medicine II-Cardiology, Ulm University Medical Center, Ulm, Germany.
- 108. Science Center, Tampere University Hospital, Tampere, Finland.
- 109. Department of Haematology, University of Cambridge, Cambridge, UK.
- 110. National Health Service (NHS) Blood and Transplant, Cambridge, UK.
- 111. Division of Cardiology, Samsung Medical Center, Seoul, Korea.
- 112. Munich Heart Alliance, Munich, Germany.
- 113. Division of Translational Medicine and Human Genetics, Department of Medicine, Perelman School of Medicine at the University of Pennsylvania, Philadelphia, Pennsylvania, USA.
- 114. Institute for Genomics and Multiscale Biology, Department of Genetics and Genomic Sciences, Mount Sinai School of Medicine, New York, New York, USA.
- 115. Center for Human Genetics, Department of Medicine, Duke University Medical Center, Durham,

North Carolina, USA.

- 116. Division of Cardiology, Department of Medicine, Duke University Medical Center, Durham, North Carolina, USA.
- 117. Division of Cardiology, Department of Medicine, Helsinki, University Central Hospital (HUCH), Helsinki, Finland.
- 118. Klinik und Poliklinik für Innere Medizin II, Regensburg, Germany.
- 119. Department of Human Genetics, McGill University, Montréal, Québec, Canada.
- 120. Department of Nutrition, Harvard School of Public Health, Boston, Massachusetts, USA.
- 121. Lebanese American University, Chouran, Beirut, Lebanon.
- 122. Cardiovascular Institute, Perelman School of Medicine at the University of Pennsylvania, Philadelphia, Pennsylvania, USA.
- 123. Synlab Academy, Mannheim, Germany.
- 124. Cardiology Division, Center for Human Genetic Research, Massachusetts General Hospital and Harvard Medical School, Boston, Massachusetts, USA.
- 125. Cardiovascular Research Center, Massachusetts General Hospital and Harvard Medical School, Boston, Massachusetts, USA.

# The CKDGen Consortium (glomerular filtration rate of creatinine and chronic kidney disase data)

Cristian Pattaro, Anna Köttgen, Alexander Teumer, Maija Garnaas, Carsten A. Böger, Christian Fuchsberger, Matthias Olden, 8,9 Ming-Huei Chen, 10,11 Adrienne Tin, Daniel Taliun, Man Li, Xiaoyi Gao, 12 Mathias Gorski, 13,14 Qiong Yang, 15 Claudia Hundertmark, 16 Meredith C. Foster, 17 Conall M. O'Seaghdha, 17,18 Nicole Glazer, 19 Aaron Isaacs, 20,21 Ching-Ti Liu, 22 Albert V. Smith, 23,24 Jeffrey R. O'Connell, 25 Maksim Struchalin,<sup>26</sup> Toshiko Tanaka,<sup>27</sup> Guo Li,<sup>28</sup> Andrew D. Johnson,<sup>17</sup> Hinco J. Gierman,<sup>29</sup> Mary Feitosa,<sup>12</sup> Shih-Jen Hwang,<sup>17</sup> Elizabeth J. Atkinson,<sup>30</sup> Kurt Lohman,<sup>31</sup> Marilyn C. Cornelis,<sup>32</sup> Åsa Johansson,<sup>33</sup> Anke Tönjes,<sup>34,35</sup> Abbas Dehghan,<sup>36</sup> Vincent Chouraki,<sup>37</sup> Elizabeth G. Holliday,<sup>38,39</sup> Rossella Sorice,<sup>40</sup> Zoltan Kutalik,<sup>41,42</sup> Terho Lehtimäki, 43 Tõnu Esko, 44,45 Harshal Deshmukh, 46 Sheila Ulivi, 47 Audrey Y. Chu, 48 Federico Murgia, 49 Stella Trompet, 50 Medea Imboden, 51 Barbara Kollerits, 52 Giorgio Pistis, 53 CARDIOGRAM Consortium, ICBP Consortium, CARe Consortium, Wellcome Trust Case Control Consortium 2 (WTCCC2), Tamara B. Harris, 54 Lenore J. Launer,<sup>54</sup> Thor Aspelund,<sup>23,24</sup> Gudny Eiriksdottir,<sup>23</sup> Braxton D. Mitchell,<sup>25</sup> Eric Boerwinkle,<sup>55</sup> Helena Schmidt,<sup>56</sup> Margherita Cavalieri,<sup>57</sup> Madhumathi Rao,<sup>58</sup> Frank B. Hu,<sup>32</sup> Ayse Demirkan,<sup>20</sup> Ben A. Oostra,<sup>20</sup> Mariza de Andrade, 30 Stephen T. Turner, 59 Jingzhong Ding, 60 Jeanette S. Andrews, 61 Barry I. Freedman, 62 Wolfgang Koenig, 63 Thomas Illig, 64 Angela Döring, 14,64 H.-Erich Wichmann, 14,65,66 Ivana Kolcic, 67 Tatijana Zemunik, <sup>67</sup> Mladen Boban, <sup>67</sup> Cosetta Minelli, <sup>1</sup> Heather E. Wheeler, <sup>68,69</sup> Wilmar Igl, <sup>33</sup> Ghazal Zaboli, <sup>33</sup> Sarah H. Wild, <sup>70</sup> Alan F. Wright, <sup>71</sup> Harry Campbell, <sup>70</sup> David Ellinghaus, <sup>72</sup> Ute Nöthlings, <sup>72,73</sup> Gunnar Jacobs, <sup>72,73</sup> Reiner Biffar,<sup>74</sup> Karlhans Endlich,<sup>75</sup> Florian Ernst,<sup>4</sup> Georg Homuth,<sup>4</sup> Heyo K. Kroemer,<sup>76</sup> Matthias Nauck,<sup>77</sup> Sylvia Stracke,<sup>78</sup> Uwe Völker,<sup>4</sup> Henry Völzke,<sup>79</sup> Peter Kovacs,<sup>80</sup> Michael Stumvoll,<sup>34,35</sup> Reedik Mägi,<sup>44,81</sup> Albert Hofman,<sup>36</sup> Andre G. Uitterlinden,<sup>82</sup> Fernando Rivadeneira,<sup>82</sup> Yurii S. Aulchenko,<sup>36</sup> Ozren Polasek,<sup>83</sup> Nick Hastie, 84 Veronique Vitart, 84 Catherine Helmer, 85,86 Jie Jin Wang, 87,88 Daniela Ruggiero, 40 Sven Bergmann, 42 Mika Kähönen, 89 Jorma Viikari, 90 Tiit Nikopensius, 45 Michael Province, 12 Shamika Ketkar, 12 Helen Colhoun, 46 Alex Doney, 91 Antonietta Robino, 92 Franco Giulianini, 48 Bernhard K. Krämer, 93 Laura Portas, 49 Ian Ford, 94 Brendan M. Buckley, <sup>95</sup> Martin Adam, <sup>51</sup> Gian-Andri Thun, <sup>51</sup> Bernhard Paulweber, <sup>96</sup> Margot Haun, <sup>97</sup> Cinzia Sala,<sup>53</sup> Marie Metzger,<sup>98</sup> Paul Mitchell,<sup>87</sup> Marina Ciullo,<sup>40</sup> Stuart K. Kim,<sup>29,68</sup> Peter Vollenweider,<sup>99</sup> Olli Raitakari, 100 Andres Metspalu, 44,45 Colin Palmer, 101 Paolo Gasparini, 92 Mario Pirastu, 49 J. Wouter Jukema, 50,102,103,104 Nicole M. Probst-Hensch, 51 Florian Kronenberg, 52 Daniela Toniolo, 53 Vilmundur Gudnason, 23,24 Alan R. Shuldiner, 25,105 Josef Coresh, 2,106 Reinhold Schmidt, 57 Luigi Ferrucci, 27 David S. Siscovick,<sup>28</sup> Cornelia M. van Duijn,<sup>20</sup> Ingrid Borecki,<sup>12</sup> Sharon L. R. Kardia,<sup>107</sup> Yongmei Liu,<sup>31</sup> Gary C. Curhan, <sup>108</sup> Igor Rudan, <sup>70</sup> Ulf Gyllensten, <sup>33</sup> James F. Wilson, <sup>70</sup> Andre Franke, <sup>72</sup> Peter P. Pramstaller, <sup>1</sup> Rainer Rettig, 109 Inga Prokopenko, 81 Jacqueline C. M. Witteman, 36 Caroline Hayward, 84 Paul Ridker, 48,110 Afshin Parsa, 111 Murielle Bochud, 112 Iris M. Heid, 113,114 Wolfram Goessling, 115,116 Daniel I. Chasman, 48,110 W. H. Linda Kao, 2,106 and Caroline S. Fox 17,117

- 1. Institute of Genetic Medicine, European Academy of Bozen/Bolzano (EURAC) and Affiliated Institute of the University of Lübeck, Bolzano, Italy
- 2. Department of Epidemiology, Johns Hopkins Bloomberg School of Public Health, Baltimore, Maryland, USA
- 3. Renal Division, Freiburg University Clinic, Freiburg, Germany
- 4. Interfaculty Institute for Genetics and Functional Genomics, University of Greifswald, Greifswald, Germany
- 5. Division of Genetics, Department of Medicine, Brigham and Women's Hospital, Harvard Medical School, Boston, Massachusetts, USA
- 6. Department of Internal Medicine II, University Medical Center Regensburg, Regensburg, Germany
- 7. Center for Statistical Genetics, Department of Biostatistics, University of Michigan, Ann Arbor, Michigan, USA
- 8. Department of Internal Medicine II, University Hospital Regensburg, Regensburg, Germany
- 9. Department of Epidemiology and Preventive Medicine, Regensburg University Medical Center, Regensburg, Germany
- 10. Department of Neurology, Boston University School of Medicine, Boston, Massachusetts, USA
- 11. Department of Biostatistics, Boston University School of Public Health, Boston, Massachusetts, USA
- 12. Division of Statistical Genomics, Washington University School of Medicine, St. Louis, Missouri, USA
- 13. Department of Epidemiology and Preventive Medicine, University Hospital Regensburg, Regensburg, Germany
- 14. Institute of Epidemiology I, Helmholtz Zentrum München, German Research Center for Environmental Health, Neuherberg, Germany
- 15. Department of Biostatistics, Boston University School of Public Health, Boston, Massachusetts, USA
- 16. Renal Division, Freiburg University Clinic, Freiburg, Germany
- 17. National Heart, Lung, and Blood Institute's Framingham Heart Study and the Center for Population Studies, Framingham, Massachusetts, USA
- 18. Division of Nephrology, Brigham and Women's Hospital and Harvard Medical School, Boston, Massachusetts, USA
- 19. Section of Preventive Medicine and Epidemiology, Department of Medicine, Boston University School of Medicine, Boston, Massachusetts, USA
- 20. Genetic Epidemiology Unit, Department of Epidemiology, Erasmus University Medical Center, Rotterdam, The Netherlands
- 21. Centre for Medical Systems Biology, Leiden, The Netherlands
- 22. Department of Biostatistics, Boston University School of Public Health, Boston, Massachusetts, USA
- 23. Icelandic Heart Association, Research Institute, Kopavogur, Iceland
- 24. University of Iceland, Reykjavik, Iceland
- 25. Department of Medicine, University of Maryland Medical School, Baltimore, Maryland, USA
- 26. Department of Epidemiology and Biostatistics and Department of Forensic Molecular Biology, Erasmus University Medical Centre, Rotterdam, The Netherlands
- 27. Clinical Research Branch, National Institute of Aging, Baltimore, Maryland, USA
- 28. University of Washington, Seattle, Washington, USA
- 29. Department of Developmental Biology, Stanford University, Stanford, California, USA
- 30. Division of Biomedical Statistics and Informatics, Mayo Clinic, Rochester, Minnesota, USA
- 31. Department of Epidemiology and Prevention, Public Health Sciences, Wake Forest School of Medicine, Winston-Salem, North Carolina, USA
- 32. Department of Nutrition, Harvard School of Public Health, Boston, Massachusetts, USA
- 33. Genetics and Pathology, Rudbeck Laboratory, Uppsala University, Uppsala, Sweden
- 34. Department of Medicine, University of Leipzig, Leipzig, Germany
- 35. IFB Adiposity Diseases, University of Leipzig, Leipzig, Germany
- 36. Department of Epidemiology, Erasmus University Medical Center, Rotterdam, The Netherlands
- 37. Inserm UMR744, Institut Pasteur, Lille, France
- 38. Centre for Clinical Epidemiology and Biostatistics, School of Medicine and Public Health, University of Newcastle, Newcastle, Australia
- 39. Centre for Information-based Medicine, Hunter Medical Research Institute, Newcastle, Australia
- 40. Institute of Genetics and Biophysics "Adriano-Buzzati Traverso"-CNR, Napoli, Italy
- 41. Department of Medical Genetics, University of Lausanne, Lausanne, Switzerland
- 42. Swiss Institute of Bioinformatics, Lausanne, Switzerland
- 43. Department of Clinical Chemistry, University of Tampere and Tampere University Hospital, Centre for Laboratory Medicine Tampere Finn-Medi 2, Tampere, Finland

- 44. Estonian Genome Center of University of Tartu (EGCUT), Tartu, Estonia
- 45. Estonian Biocenter and Institute of Molecular and Cell Biology, University of Tartu, Tartu, Estonia
- 46. Wellcome Trust Centre for Molecular Medicine, Clinical Research Centre, Ninewells Hospital, University of Dundee, Dundee, United Kingdom
- 47. Institute for Maternal and Child Health IRCCS "Burlo Garofolo", Trieste, Italy
- 48. Brigham and Women's Hospital, Boston, Massachusetts, USA
- 49. Institute of Population Genetics CNR, Sassari, Italy
- 50. Department of Cardiology, Leiden University Medical Center, Leiden, The Netherlands
- 51. Unit of Chronic Disease Epidemiology, Swiss Tropical and Public Health Institute, Basel, Switzerland
- 52. Division of Genetic Epidemiology, Innsbruck Medical University, Innsbruck, Austria
- 53. Division of Genetics and Cell Biology, San Raffaele Scientific Institute, Milano, Italy
- 54. Laboratory of Epidemiology, Demography, and Biometry, NIA, Bethesda, Maryland, USA
- 55. Human Genetics Center, University of Texas Health Science Center, Houston, Texas, USA
- 56. Austrian Stroke Prevention Study, Institute of Molecular Biology and Biochemistry and Department of Neurology, Medical University Graz, Graz, Austria
- 57. Austrian Stroke Prevention Study, University Clinic of Neurology, Department of Special Neurology, Medical University Graz, Graz, Austria
- 58. Division of Nephrology/Tufts Evidence Practice Center, Tufts University School of Medicine, Tufts Medical Center, Boston, Massachusetts, USA
- 59. Department of Internal Medicine, Division of Nephrology and Hypertension, Mayo Clinic, Rochester, Minnesota, USA
- 60. Department of Internal Medicine/Geriatrics, Wake Forest School of Medicine, Winston-Salem, North Carolina, USA
- 61. Department of Biostatistical Sciences, Public Health Sciences, Wake Forest School of Medicine, Winston-Salem, North Carolina, USA
- 62. Department of Internal Medicine, Wake Forest School of Medicine, Winston-Salem, North Carolina, USA
- 63. Abteilung Innere II, Universitätsklinikum Ulm, Ulm, Germany
- 64. Institute of Epidemiology II, Helmholtz Zentrum München, German Research Center for Environmental Health, Neuherberg, Germany
- 65. Institute of Medical Informatics, Biometry, and Epidemiology, Ludwig-Maximilians-Universität, Munich, Germany
- 66. Klinikum Grosshadern, Neuherberg, Germany
- 67. Croatian Centre for Global Health, University of Split Medical School, Split, Croatia
- 68. Department of Genetics, Stanford University, Stanford, California, USA
- 69. Department of Medicine, University of Chicago, Chicago, Illinois, USA
- 70. Center for Population Health Sciences, University of Edinburgh Medical School, Edinburgh, United Kingdom
- 71. MRC Human Genetics Unit, Institute of Genetics and Molecular Medicine, Western General Hospital, Edinburgh, United Kingdom
- 72. Institute of Clinical Molecular Biology, Christian-Albrechts University, Kiel, Germany
- 73. Popgen Biobank, University Hospital Schleswig-Holstein, Kiel, Germany
- 74. Clinic for Prosthodontic Dentistry, Gerostomatology, and Material Science, University of Greifswald, Greifswald, Germany
- 75. Institute of Anatomy and Cell Biology, University of Greifswald, Greifswald, Germany
- 76. Institute of Pharmacology, University of Greifswald, Greifswald, Germany
- 77. Institute of Clinical Chemistry and Laboratory Medicine, Ernst-Moritz-Arndt-University Greifswald, Greifswald, Germany
- 78. Clinic for Internal Medicine A, University of Greifswald, Greifswald, Germany
- 79. Institute for Community Medicine, University of Greifswald, Greifswald, Germany
- 80. Department of Medicine, University of Leipzig, Leipzig, Germany
- 81. Wellcome Trust Centre for Human Genetics and Oxford Centre for Diabetes, Endocrinology, and Metabolism, University of Oxford, Oxford, United Kingdom
- 82. Department of Internal Medicine, Erasmus University Medical Center, Rotterdam, The Netherlands
- 83. Croatian Centre for Global Health, Faculty of Medicine, University of Split, Split, Croatia
- 84. MRC Human Genetics Unit, Institute of Genetics and Molecular Medicine, Western General Hospital, Edinburgh, United Kingdom
- 85. INSERM U897, Université Victor Ségalen Bordeaux 2, ISPED, Bordeaux, France
- 86. Université Bordeaux 2 Victor Segalen, Bordeaux, France
- 87. Centre for Vision Research, Westmead Millennium Institute, Westmead Hospital, University of Sydney, Sydney, Australia
- 88. Centre for Eye Research Australia (CERA), University of Melbourne, Melbourne, Australia

- 89. Department of Clinical Physiology, University of Tampere and Tampere University Hospital, Tampere, Finland
- 90. Department of Medicine, University of Turku and Turku University Hospital, Turku, Finland
- 91. NHS Tayside, Wellcome Trust Centre for Molecular Medicine, Clinical Research Centre, Ninewells Hospital, University of Dundee, Dundee, United Kingdom
- 92. Institute for Maternal and Child Health, IRCCS "Burlo Garofolo," University of Trieste, Trieste, Italy
- 93. University Medical Centre Mannheim, 5th Department of Medicine, Mannheim, Germany
- 94. Robertson Centre for Biostatistics, University of Glasgow, Glasgow, United Kingdom
- 95. Department of Pharmacology and Therapeutics, University College Cork, Cork, Ireland
- 96. First Department of Internal Medicine, Paracelsus Medical University, Salzburg, Austria
- 97. Division of Genetic Epidemiology, Innsbruck Medical University, Innsbruck, Austria
- 98. Inserm UMRS 1018, CESP Team 10, Université Paris Sud, Villejuif, France
- 99. Department of Internal Medicine, Centre Hospitalier Universitaire Vaudois, Lausanne, Switzerland
- 100. Research Centre of Applied and Preventive Cardiovascular Medicine, Department of Clinical Physiology, Turku University Hospital, University of Turku, Turku, Finland
- 101. Biomedical Research Institute, Ninewells Hospital and Medical School, University of Dundee, Dundee, United Kingdom
- 102. Interuniversity Cardiology Institute of the Netherlands (ICIN), Utrecht, The Netherlands
- 103. Einthoven Laboratory for Experimental Vascular Medicine, Leiden, The Netherlands
- 104. Durrer Center for Cardiogenetic Research, Amsterdam, The Netherlands
- 105. Geriatric Research and Education Clinical Center, Veterans Administration Medical Center, Baltimore, Maryland, USA
- 106. Welch Center for Prevention, Epidemiology, and Clinical Research, Baltimore, Maryland, USA
- 107. Department of Epidemiology, School of Public Health, University of Michigan, Ann Arbor, Michigan, USA
- 108. Brigham and Women's Hospital and Channing Laboratory, Harvard Medical School, Boston, Massachusetts, USA
- 109. Institute of Physiology, University of Greifswald, Greifswald, Germany
- 110. Harvard Medical School, Boston, Massachusetts, USA
- 111. Division of Nephrology, University of Maryland Medical School, Baltimore, Maryland, USA
- 112. University Institute of Social and Preventive Medicine, Centre Hospitalier Universitaire Vaudois and University of Lausanne, Epalinges, Switzerland
- 113. Department of Epidemiology and Preventive Medicine, University Hospital Regensburg, Regensburg, Germany
- 114. Institute of Epidemiology I, Helmholtz Zentrum München, German Research Center for Environmental Health, Neuherberg, Germany
- 115. Divisions of Genetics and Gastroenterology, Department of Internal Medicine, Brigham and Women's Hospital, Boston, Massachusetts, USA
- 116. Harvard Stem Cell Institute, Harvard University, Cambridge, Massachusetts, USA
- 117. Division of Endocrinology, Brigham and Women's Hospital and Harvard Medical School, Boston, Massachusetts, USA

## The CKDGen Consortium (urine albumin-to-creatinine ratio data)

Carsten A. Böger, Ming-Huei Chen, Adrienne Tin, Matthias Olden, Anna Köttgen, Ian H. de Boer, Christian Fuchsberger, Conall M. O'Seaghdha, Cristian Pattaro, Alexander Teumer, Ching-Ti Liu, Nicole L. Glazer, <sup>11</sup> Man Li, <sup>3</sup> Jeffrey R. O'Connell, <sup>12</sup> Toshiko Tanaka, <sup>13,14</sup> Carmen A. Peralta, <sup>15</sup> Zoltán Kutalik, <sup>16,17</sup> Jian'an Luan, 18 Jing Hua Zhao, 18 Shih-Jen Hwang, 19 Ermeg Akylbekova, 20 Holly Kramer, 21 Pim van der Harst, 22 Albert V. Smith, 23,24 Kurt Lohman, 25 Mariza de Andrade, 26 Caroline Hayward, 27 Barbara Kollerits, 28 Anke Tönjes,<sup>29</sup> Thor Aspelund,<sup>23,24</sup> Erik Ingelsson,<sup>30</sup> Gudny Eiriksdottir,<sup>24</sup> Lenore J. Launer,<sup>31</sup> Tamara B. Harris,<sup>31</sup> Alan R. Shuldiner, <sup>32</sup> Braxton D. Mitchell, <sup>33</sup> Dan E. Arking, <sup>34</sup> Nora Franceschini, <sup>35</sup> Eric Boerwinkle, <sup>36</sup> Josephine Egan,<sup>37</sup> Dena Hernandez,<sup>38</sup> Muredach Reilly,<sup>39</sup> Raymond R. Townsend,<sup>40</sup> Thomas Lumley,<sup>11</sup> David S. Siscovick, 41 Bruce M. Psaty, 42 Bryan Kestenbaum, 6 Talin Haritunians, 43 Sven Bergmann, 16,17 Peter Vollenweider, 44 Gerard Waeber, 44 Vincent Mooser, 45 Dawn Waterworth, 45 Andrew D. Johnson, 19 Jose C. Florez, 46 James B. Meigs, 47 Xiaoning Lu, 10 Stephen T. Turner, 48 Elizabeth J. Atkinson, 26 Tennille S. Leak, 49 Knut Aasarød, 50,51 Frank Skorpen, 51 Ann-Christine Syvänen, 52 Thomas Illig, 53 Jens Baumert, 53 Wolfgang Koenig,<sup>54</sup> Bernhard K. Krämer,<sup>55</sup> Olivier Devuyst,<sup>56</sup> Josyf C. Mychaleckyj,<sup>57</sup> Cosetta Minelli,<sup>7</sup> Stephan J.L. Bakker,<sup>58</sup> Lyudmyla Kedenko,<sup>59</sup> Bernhard Paulweber,<sup>59</sup> Stefan Coassin,<sup>28</sup> Karlhans Endlich,<sup>60</sup> Heyo K. Kroemer, 61 Reiner Biffar, 62 Sylvia Stracke, 63 Henry Völzke, 64 Michael Stumvoll, 29 Reedik Mägi, 65 Harry Campbell, <sup>66</sup> Veronique Vitart, <sup>27</sup> Nicholas D. Hastie, <sup>27</sup> Vilmundur Gudnason, <sup>23,24</sup> Sharon L.R. Kardia, <sup>67</sup> Yongmei Liu,<sup>25</sup> Ozren Polasek,<sup>68</sup> Gary Curhan,<sup>69</sup> Florian Kronenberg,<sup>28</sup> Inga Prokopenko,<sup>65</sup> Igor Rudan,<sup>70</sup> Johan Ärnlöv, 71 Stein Hallan, 50,51 Gerjan Navis, 58 the CKDGen Consortium, Afshin Parsa, 72 Luigi Ferrucci, 14 Josef Coresh, 73 Michael G. Shlipak, 74 Shelley B. Bull, 75 Andrew D. Paterson, 76 on behalf of DCCT/EDIC, H.-Erich Wichmann, 53,77,78 Nicholas J. Wareham, 18 Ruth J.F. Loos, 18 Jerome I. Rotter, 43 Peter P. Pramstaller, L. Adrienne Cupples, 10 Jacques S. Beckmann, 29 Qiong Yang, 80 Iris M. Heid, 4,53 Rainer Rettig, 81 Albert W. Dreisbach, 82 Murielle Bochud, 83 Caroline S. Fox, 19,84 and W.H.L. Kao<sup>3</sup>

- 1. Department of Internal Medicine II, University Medical Center Regensburg, Regensburg, Germany;
- 2. Department of Neurology, Boston University School of Medicine, Boston, Massachusetts;
- 3. Department of Epidemiology and the Welch Center for Prevention, Epidemiology, and Clinical Research, Johns Hopkins University, Baltimore, Maryland;
- 4. Department of Epidemiology and Preventive Medicine, Regensburg University Medical Center, Regensburg, Germany;
- 5. Renal Division, University Hospital of Freiburg, Freiburg, Germany;
- 6. Division of Nephrology, University of Washington, Seattle, Washington;
- 7. Institute of Genetic Medicine, European Academy of Bolzano/Bozen (EURAC), Italy and Affiliated Institute of the University of Lübeck, Lübeck, Germany:
- 8. Division of Nephrology, Brigham and Women's Hospital and Harvard Medical School, Boston Massachusetts;
- 9. Interfaculty Institute for Genetics and Functional Genomics, University of Greifswald, Greifswald, Germany;
- 10. Department of Biostatistics, Boston University School of Public Health and NHLBI's Framingham Heart Study, Boston Massachusetts;
- 11. Cardiovascular Health Research Unit and Department of Biostatistics, University of Washington, Seattle, Washington;
- 12. University of Maryland School of Medicine, Baltimore, Maryland;

- 13. Medstar Research Institute, Baltimore, Maryland;
- 14. Clinical Research Branch, National Institute on Aging, Baltimore, Maryland;
- 15. Division of Nephrology, University of California, San Francisco Medical School and San Francisco VA Medical Center, San Francisco, California;
- 16. Department of Medical Genetics, University of Lausanne, Lausanne, Switzerland;
- 17. Swiss Institute of Bioinformatics, Lausanne, Switzerland;
- 18. MRC Epidemiology Unit, Institute of Metabolic Science, Addenbrooke's Hospital, Cambridge, United Kingdom;
- 19. NHLBI's Framingham Heart Study and the Center for Population Studies, Framingham, Massachusetts;
- 20. Jackson State University, Jackson, Mississippi;
- 21. Loyola University, Maywood, Illinois;
- 22. Department of Cardiology, University Medical Center Groningen, University of Groningen, Groningen, The Netherlands;
- 23. University of Iceland, Reykjavik, Iceland;
- 24. Icelandic Heart Association, Hjartavernd, Holtasmara, Kopavogur, Iceland;
- 25. Department of Biostatistical Sciences, Wake Forest University, Division of Public Health Sciences, Winston-Salem, North Carolina;
- 26. Division of Biomedical Statistics and Informatics, Mayo Clinic, Rochester, Minnesota;
- 27. MRC Human Genetics Unit, Institute of Genetics and Molecular Medicine, Western General Hospital, Crewe Road, Edinburgh, Scotland;
- 28. Innsbruck Medical University, Division of Genetic Epidemiology, Innsbruck, Austria;
- 29. Department of Medicine, University of Leipzig, Leipzig, Germany;
- 30. Department of Medical Epidemiology and Biostatistics, Karolinska Institutet, Stockholm, Sweden;
- 31. Laboratory of Epidemiology, Demography, and Biometry, NIA, Bethesda, Maryland;
- 32. University of Maryland School of Medicine, Geriatric Research and Education Clinical Center, Veterans Administration Medical Center, Baltimore, Maryland:
- 33. University of Maryland School of Medicine, Baltimore, Maryland;
- 34. McKusick-Nathans Institute of Genetic Medicine, Johns Hopkins Medical Institutions, Baltimore, Maryland;
- 35. University of North Carolina at Chapel Hill, Chapel Hill, North Carolina;
- 36. Human Genetics Center, University of Texas Health Science Center, Houston, Texas;
- 37. Laboratory of Clinical Investigation, National Institute on Aging, Baltimore, Maryland;
- 38. Laboratory of Neurogenetics, National Institute on Aging, Bethesda, Maryland;
- 39. University of Pennsylvania Division of Cardiology, Perelman Center for Advanced Medicine, Philadelphia, Pennsylvania;
- 40. University of Pennsylvania Renal Electrolyte and Hypertension Division, Philadelphia, Pennsylvania;
- 41. Departments of Epidemiology and Medicine, University of Washington, Seattle, Washington;
- 42. Cardiovascular Health Research Unit, Departments of Medicine, Epidemiology, and Health Services and Group Health Research Institute, Group Health Cooperative, Seattle, Washington;
- 43. Medical Genetics Institute, Cedars-Sinai Medical Center, Los Angeles, California;
- 44. Department of Internal Medicine, Centre Hospitalier Universitaire Vaudois, Lausanne, Switzerland;
- 45. Genetics Division, GlaxoSmithKline, King of Prussia, Pennsylvania;
- 46. Center for Human Genetic Research and Diabetes Research Center (Diabetes Unit), Massachusetts General Hospital, Boston, Massachusetts, Program in Medical and Population Genetics, Broad Institute, Cambridge, Massachusetts, and Department of Medicine, Harvard Medical School, Boston, Massachusetts;
- 47. Department of General Internal Medicine, Massachussetts General Hospital, Boston, Massachusetts;
- 48. Department of Internal Medicine, Division of Nephrology and Hypertension, Mayo Clinic, Rochester, Minnesota;
- 49. Department of Epidemiology, Graduate School of Public Health, University of Pittsburgh, Pittsburgh, Pennsylvania;
- 50. St Olav University Hospital, Trondheim, Norway;
- 51. Faculty of Medicine, Norwegian University of Science and Technology (NTNU), Trondheim, Norway;
- 52. Molecular Medicine, Department of Medical Sciences, Science for Life Laboratory, Uppsala University, Uppsala, Sweden;
- 53. Institute of Epidemiology, Helmholtz Zentrum München, German Research Center for Environmental Health, Neuherberg, Germany;
- 54. Zentrum für Innere Medizin, Klinik für Innere Medizin II Kardiologie, Universitätsklinikum Ulm, Ulm, Germany;
- 55. University Medical Centre Mannheim, 5th Department of Medicine, Mannheim, Germany;

- 56. NEFR Unit Université Catholique de Louvain Medical School, Brussels, Belgium;
- 57. Center for Public Health Genomics, Charlottesville, Virginia;
- 58. Department of Internal Medicine, University Medical Center, Groningen, University of Groningen, Groningen, The Netherlands;
- 59. First Department of Internal Medicine, Paracelsus Medical University, Salzburg, Austria;
- 60. Institute of Anatomy and Cell Biology, University of Greifswald, Greifswald, Germany;
- 61. Institute of Pharmacology, University of Greifswald, Greifswald, Germany;
- 62. Clinic for Prosthodontic Dentistry, Gerostomatology and Material Science, University of Greifswald, Greifswald, Germany;
- 63. Nephrology Clinic for Internal Medicine A, University of Greifswald, Greifswald, Germany;
- 64. Institute for Community Medicine, University of Greifswald, Greifswald, Germany;
- 65. Wellcome Trust Centre for Human Genetics, and Oxford Centre for Diabetes, Endocrinology and Metabolism, University of Oxford, United Kingdom;
- 66. Centre for Population Health Sciences, University of Edinburgh, Edinburgh, Scotland;
- 67. University of Michigan School of Public Health, Department of Epidemiology, University of Michigan, Ann Arbor, Michigan;
- 68. Gen-Info Ltd., Zagreb, Croatia;
- 69. Brigham and Women's Hospital, Harvard Medical School, Boston, Massachusetts;
- 70. Center for Population Health Sciences, University of Edinburgh Medical School, Edinburgh, Scotland;
- 71. Department of Public Health and Caring Sciences, Uppsala University, Uppsala, Sweden;
- 72. University of Maryland School of Medicine, Baltimore, Maryland;
- 73. Welch Center for Prevention, Epidemiology & Clinical Research, Johns Hopkins University, Baltimore, Maryland;
- 74. General Internal Medicine, University of California, San Francisco, San Francisco, California;
- 75. Samuel Lunenfeld Research Institute of Mount Sinai Hospital, Prosserman Centre for Health Research, Toronto, Ontario, Canada;
- 76. The Hospital for Sick Children, Toronto, Ontario, Canada;
- 77. Institute of Medical Informatics, Biometry and Epidemiology, Ludwig-Maximilians-Universität, Munich, Germany;
- 78. Klinikum Grosshadern, Munich, Germany;
- 79. Service of Medical Genetics, Centre Hospitalier Universitaire Vaudois and University of Lausanne, Lausanne, Switzerland;
- 80. Department of Biostatistics, Boston University School of Public Health, Boston, Massachusetts;
- 81. Institute of Physiology, University of Greifswald, Greifswald, Germany;
- 82. University of Mississippi Division of Nephrology, University of Mississippi, Jackson, Mississippi;
- 83. University Institute of Social and Preventive Medicine, Centre Hospitalier Universitaire Vaudois and University of Lausanne, IUMSP, Lausanne, Switzerland; and
- 84. Division of Endocrinology, Brigham and Women's Hospital and Harvard Medical School, Boston, Massachusetts

## **The GEFOS Consortium**

Karol Estrada<sup>1,2,3</sup>, Unnur Styrkarsdottir<sup>4,139</sup>, Evangelos Evangelou<sup>5,139</sup>, Yi-Hsiang Hsu<sup>6,7,139</sup>, Emma L Duncan<sup>8,9,139</sup>, Evangelia E Ntzani<sup>5,139</sup>, Ling Oei<sup>1,2,3,139</sup>, Omar M E Albagha<sup>10</sup>, Najaf Amin<sup>2</sup>, John P Kemp<sup>11</sup>, Daniel L Koller<sup>12</sup>, Guo Li<sup>13</sup>, Ching-Ti Liu<sup>14</sup>, Ryan L Minster<sup>15</sup>, Alireza Moayyeri<sup>16,17</sup>, Liesbeth Vandenput<sup>18</sup>, Dana Willner<sup>8,19</sup>, Su-Mei Xiao<sup>20,21</sup>, Laura M Yerges-Armstrong<sup>22</sup>, Hou-Feng Zheng<sup>23</sup>, Nerea Alonso<sup>10</sup>, Joel Eriksson<sup>18</sup>, Candace M Kammerer<sup>15</sup>, Stephen K Kaptoge<sup>16</sup>, Paul J Leo<sup>8</sup>, Gudmar Thorleifsson<sup>4</sup>, Scott G Wilson<sup>17,24,25</sup>, James F Wilson<sup>26,27</sup>, Ville Aalto<sup>28,29</sup>, Markku Alen<sup>30</sup>, Aaron K Aragaki<sup>31</sup>, Thor Aspelund<sup>32,33</sup>, Jacqueline R Center<sup>34,35,36</sup>, Zoe Dailiana<sup>37</sup>, David J Duggan<sup>38</sup>, Melissa Garcia<sup>39</sup>, Natàlia Garcia-Giralt<sup>40</sup>, Sylvie Giroux<sup>41</sup>, Göran Hallmans<sup>42</sup>, Lynne J Hocking<sup>43</sup>, Lise Bjerre Husted<sup>44</sup>, Karen A Jameson<sup>45</sup>, Rita Khusainova<sup>46,47</sup>, Ghi Su Kim<sup>48</sup>, Charles Kooperberg<sup>31</sup>, Theodora Koromila<sup>49</sup>, Marcin Kruk<sup>50</sup>, Marika Laaksonen<sup>51</sup>, Andrea Z Lacroix<sup>31</sup>, Seung Hun Lee<sup>48</sup>, Ping C Leung<sup>52</sup>, Joshua R Lewis<sup>24,25</sup>, Laura Masi<sup>53</sup>, Simona Mencej-Bedrac<sup>54</sup>, Tuan V Nguyen<sup>34,35</sup>, Xavier Nogues<sup>40</sup>, Millan S Patel<sup>55</sup>, Janez Prezelj<sup>56</sup>, Lynda M Rose<sup>57</sup>, Serena Scollen<sup>58</sup>, Kristin Siggeirsdottir<sup>32</sup>, Albert V Smith<sup>32,33</sup>, Olle Svensson<sup>59</sup>, Stella Trompet<sup>60,61</sup>, Olivia Trummer<sup>62</sup>, Natasja M van Schoor<sup>63</sup>, Jean Woo<sup>64</sup>, Kun Zhu<sup>24,25</sup>, Susana Balcells<sup>65</sup>, Maria Luisa Brandi<sup>53</sup>, Brendan M Buckley<sup>66</sup>, Sulin Cheng<sup>67,68</sup>, Claus Christiansen<sup>69</sup>, Cyrus Cooper<sup>45</sup>, George Dedoussis<sup>70</sup>, Ian Ford<sup>71</sup>, Morten Frost<sup>72,73</sup>, David Goltzman<sup>74</sup>, Jesús González-Macías<sup>75,76</sup>, Mika Kähönen<sup>77,78</sup>, Magnus Karlsson<sup>79</sup>, Elza Khusnutdinova<sup>46,47</sup>, Jung-Min Koh<sup>48</sup>, Panagoula Kollia<sup>49</sup>, Bente Lomholt Langdahl<sup>44</sup>, William D Leslie<sup>80</sup>, Paul Lips<sup>81,82</sup>, Östen Ljunggren<sup>83</sup>, Roman S Lorenc<sup>50</sup>, Janja Marc<sup>54</sup>, Dan Mellström<sup>18</sup>, Barbara Obermayer-Pietsch<sup>62</sup>, José M Olmos<sup>75,76</sup>, Ulrika Pettersson-Kymmer<sup>84</sup>, David M Reid<sup>43</sup>, José A Riancho<sup>75,76</sup>, Paul M Ridker<sup>57,85</sup>, François Rousseau<sup>41,86,87</sup>, P Eline Slagboom<sup>88,3</sup>, Nelson LS Tang<sup>89,90</sup>, Roser Urreizti<sup>65</sup>, Wim Van Hul<sup>91</sup>, Jorma Viikari<sup>92,93</sup>, María T Zarrabeitia<sup>94</sup>, Yurii S Aulchenko<sup>2</sup>, Martha Castano-Betancourt<sup>1,2,3</sup>, Elin Grundberg<sup>95,96,97</sup>, Lizbeth Herrera<sup>1</sup>, Thorvaldur Ingvarsson<sup>98,99,33</sup>, Hrefna Johannsdottir<sup>4</sup>, Tony Kwan<sup>95,96</sup>, Rui Li<sup>100</sup>, Robert Luben<sup>16</sup>, Carolina Medina-Gómez<sup>1,2</sup>, Stefan Th Palsson<sup>4</sup>, Sjur Reppe<sup>101</sup>, Jerome I Rotter<sup>102</sup>, Gunnar Sigurdsson<sup>103,33</sup>, Joyce B J van Meurs<sup>1,2,3</sup>, Dominique Verlaan<sup>95,96</sup>, Frances MK Williams<sup>17</sup>, Andrew R Wood<sup>104</sup>, Yanhua Zhou<sup>14</sup>, Kaare M Gautvik<sup>101,105,106</sup>, Tomi Pastinen<sup>95,96,107</sup>, Soumya Raychaudhuri<sup>108,109</sup>, Jane A Cauley<sup>110</sup>, Daniel I Chasman<sup>57,85</sup>, Graeme R Clark<sup>8</sup>, Steven R Cummings<sup>111</sup>, Patrick Danoy<sup>8</sup>, Elaine M Dennison<sup>45</sup>, Richard Eastell<sup>112</sup>, John A Eisman<sup>34,35,36</sup>, Vilmundur Gudnason<sup>32,33</sup>, Albert Hofman<sup>2,3</sup>, Rebecca D Jackson<sup>113,114</sup>, Graeme Jones<sup>115</sup>, J Wouter Jukema<sup>60,116,117</sup>, Kay-Tee Khaw<sup>16</sup>, Terho Lehtimäki<sup>118,119</sup>, Yongmei Liu<sup>120</sup>, Mattias Lorentzon<sup>18</sup>, Eugene McCloskey<sup>112,121</sup>, Braxton D Mitchell<sup>22</sup>, Kannabiran Nandakumar<sup>6,7</sup>, Geoffrey C Nicholson<sup>122</sup>, Ben A Oostra<sup>123</sup>, Munro Peacock<sup>124</sup>, Huibert A P Pols<sup>1,2</sup>, Richard L Prince<sup>24,25</sup>, Olli Raitakari<sup>28,29</sup>, lan R Reid<sup>125</sup>, John Robbins<sup>126</sup>, Philip N Sambrook<sup>127</sup>, Pak Chung Sham<sup>128,129</sup>, Alan R Shuldiner <sup>22,130</sup>, Frances A Tylavsky<sup>131</sup>, Cornelia M van Duijn<sup>2</sup>, Nick J Wareham<sup>132</sup>, L Adrienne Cupples<sup>14,133</sup>, Michael J Econs<sup>124,12</sup>, David M Evans<sup>11</sup>, Tamara B Harris<sup>39</sup>, Annie Wai Chee Kung<sup>20,21</sup>, Bruce M Psaty<sup>134,135</sup>, Jonathan Reeve<sup>136</sup>, Timothy D Spector<sup>17</sup>, Elizabeth A Streeten<sup>22,130</sup>, M Carola Zillikens<sup>1</sup>, Unnur Thorsteinsdottir<sup>4,33,140</sup>, Claes

Ohlsson<sup>18,140</sup>, David Karasik<sup>6,7,140</sup>, J Brent Richards<sup>137,17,140</sup>, Matthew A Brown<sup>8,140</sup>, Kari Stefansson<sup>4,33,140</sup>, André G Uitterlinden<sup>1,2,3,140</sup>, Stuart H Ralston<sup>10,140</sup>, John P A Ioannidis<sup>138,5,140</sup>, Douglas P Kiel<sup>6,7,140</sup>, Fernando Rivadeneira<sup>1,2,3,140</sup>

- 1. Department of Internal Medicine, Erasmus Medical Center, Rotterdam, The Netherlands.
- 2. Department of Epidemiology, Erasmus Medical Center, Rotterdam, The Netherlands.
- 3. Netherlands Genomics Initiative (NGI)-sponsored Netherlands Consortium for Healthy Aging (NCHA), Leiden, The Netherlands.
- 4. deCODE Genetics, Reykjavik, Iceland.
- 5. Department of Hygiene and Epidemiology, University of Ioannina, Ioannina, Greece.
- 6. Institute for Aging Research, Hebrew SeniorLife, Boston, USA.
- 7. Department of Medicine, Harvard Medical School, Boston, USA.
- 8. Human Genetics Group, University of Queensland Diamantina Institute, Brisbane, Australia.
- 9. Department of Endocrinology, Royal Brisbane and Women's Hospital, Brisbane, Australia.
- 10. Rheumatic Diseases Unit, Institute of Genetics and Molecular Medicine, University of Edinburgh, Edinburgh, UK.
- 11. Medical Research Council (MRC) Centre for Causal Analyses in Translational Epidemiology, University of Bristol, Bristol, UK.
- 12. Department of Medical and Molecular Genetics, Indiana University School of Medicine, Indianapolis, USA.
- 13. Cardiovascular Health Research Unit, University of Washington, Seattle, USA.
- 14. Department of Biostatistics, Boston University School of Public Health, Boston, USA.
- 15. Department of Human Genetics, University of Pittsburgh, Pittsburgh, PA, USA.
- 16. Department of Public Health and Primary Care, University of Cambridge, Cambridge, UK.
- 17. Department of Twin Research and Genetic Epidemiology, King's College London, London, UK.
- 18. Centre for Bone and Arthritis Research, Institute of Medicine, Sahlgrenska Academy, University of Gothenburg, Gothenburg, Sweden.
- 19. Australian Centre for Ecogenomics, University of Queensland, Brisbane, Australia.
- 20. Department of Medicine, The University of Hong Kong, Hong Kong, China.
- 21. Research Centre of Heart, Brain, Hormone and Healthy Aging, The University of Hong Kong, Hong Kong, China.
- 22. Department of Medicine, Division of Endocrinology, Diabetes and Nutrition, University of Maryland School of Medicine, Baltimore, MD, USA.
- 23. Department of Human Genetics, Lady Davis Institute, McGill University, Montreal, Canada.
- 24. School of Medicine and Pharmacology, University of Western Australia, Perth, Australia.
- 25. Department of Endocrinology and Diabetes, Sir Charles Gairdner Hospital, Perth, Australia.
- 26. Centre for Population Health Sciences, University of Edinburgh, Edinburgh, UK.
- 27. MRC Human Genetics Unit, MRC Institute of Genetics and Molecular Medicine at the University of Edinburgh, Edinburgh, UK.
- 28. Department of Clinical Physiology, Turku University Hospital, Turku, Finland.
- 29. Research Centre of Applied and Preventive Cardiovascular Medicine, University of Turku, Turku, Finland.
- 30. Department of Medical Rehabilitation, Oulu University Hospital and Institute of Health Sciences, Oulu, Finland.
- 31. Division of Public Health Sciences, Fred Hutchinson Cancer Research Center, Seattle, USA.
- 32. Icelandic Heart Association, Kopavogur, Iceland.
- 33. Faculty of Medicine, University of Iceland, Reykjavik, Iceland.
- 34. Osteoporosis and Bone Biology Program, Garvan Institute of Medical Research, Sydney, Australia.
- 35. Department of Medicine, University of New South Wales, Sydney, Australia.
- 36. Department of Endocrinology, St Vincents Hospital, Sydney, Australia.
- 37. Department of Orthopaedic Surgery, Medical School University of Thessalia, Larissa, Greece.
  - 38. Translational Genomics Research Institute, Phoenix, USA.
- 39. Laboratory of Epidemiology, Demography, and Biometry, National Institute on Aging, Bethesda, MD, USA.
- 40. Department of Internal Medicine, Hospital del Mar, Instituto Municipal de Investigación Médica (IMIM), Red Temática de Investigación Cooperativa en Envejecimiento y Fragilidad (RETICEF), Universitat Autònoma de Barcelona (UAB), Barcelone, Spain.

- 41. Unité de recherche en génétique humaine et moléculaire, Centre de recherche du Centre hospitalier universitaire de Québec Hôpital St-François-d'Assise (CHUQ/HSFA), Québec City, Canada.
- 42. Department of Public Health and Clinical Medicine, Umeå Unviersity, Umeå, Sweden.
- 43. Musculoskeletal Research Programme, Division of Applied Medicine, University of Aberdeen, Aberdeen, UK.
- 44. Department of Endocrinology and Internal Medicine, Aarhus University Hospital, Aarhus C, Denmark.
- 45. MRC Lifecourse Epidemiology Unit, University of Southampton, Southampton, UK.
- 46. Ufa Scientific Centre of Russian Academy of Sciences, Institute of Biochemistry and Genetics, Ufa, Russia.
- 47. Biological Department, Bashkir State University, Ufa, Russia.
- 48. Division of Endocrinology and Metabolism, Asan Medical Center, University of Ulsan College of Medicine, Seoul, South Korea.
- 49. Department of Genetics and Biotechnology, Faculty of Biology, University of Athens, Athens, Greece.
- 50. Department of Biochemistry and Experimental Medicine, The Children's Memorial Health Institute, Warsaw, Poland.
- 51. Department of Food and Environmental Sciences, University of Helsinki, Helsinki, Finland.
- 52. Jockey Club Centre for Osteoporosis Care and Control, The Chinese University of Hong Kong, Hong Kong SAR, China.
- 53. Department of Internal Medicine, University of Florence, Florence, Italy.
- 54. Department of Clinical Biochemistry, University of Ljubljana, Ljubljana, Slovenia.
- 55. Department of Medical Genetics, University of British Columbia, Vancouver, Canada.
- 56. Department of Endocrinology, University Medical Center, Ljubljana, Slovenia.
- 57. Division of Preventive Medicine, Brigham and Women's Hospital, Boston, USA.
- 58. Department of Medicine, University of Cambridge, Cambridge, UK.
- 59. Department of Surgical and Perioperative Sciences, Umeå Unviersity, Umeå, Sweden.
- 60. Department of Cardiology, Leiden University Medical Center, Leiden, The Netherlands.
- 61. Department of Gerontology and Geriatrics, Leiden University Medical Center, Leiden, The Netherlands.
- 62. Department of Internal Medicine, Division of Endocrinology and Metabolism, Medical University Graz, Graz, Austria.
- 63. Department of Epidemiology and Biostatistics, Extramuraal Geneeskundig Onderzoek (EMGO) Institute for Health and Care Research, Vrije Universiteit (VU) University Medical Center, Amsterdam, The Netherlands.
- 64. Department of Medicine and Therapeutics, The Chinese University of Hong Kong, Hong Kong SAR, China.
- 65. Department of Genetics, University of Barcelona, Centro de Investigación Biomédica en Red de Enfermedades Raras (CIBERER), Institut de Biomedicina de la Universitat de Barcelona (IBUB), Barcelone, Spain.
- 66. Department of Pharmacology and Therapeutics, University College Cork, Cork, Ireland.
- 67. Department of Health Sciences, University of Jyväskylä, Jyväskylä, Finland.
- 68. Department of Orthopaedics and Traumatology, Kuopio University Hospital, Kuopio, Finland.
- 69. Center for Clinical and Basic Research (CCBR)-Synarc, Ballerup, Denmark.
- 70. Department of Nutrition and Dietetics, Harokopio University, Athens, Greece.
- 71. Robertson Center for Biostatistics, University of Glasgow, Glasgow, United Kingdom.
- 72. Department of Endocrinology, Odense University Hospital, Odense, Denmark.
- 73. Clinical Institute, University of Southern Denmark, Odense, Denmark.
- 74. Department of Medicine, McGill University, Montreal, Canada.
- 75. Department of Medicine, University of Cantabria, Santander, Spain.
- 76. Department of Internal Medicine, Hospital Universitario Marqués de Valdecilla and Instituto de Formación e Investigación Marqués de Valdecilla (IFIMAV), Santander, Spain.
- 77. Department of Clinical Physiology, Tampere University Hospital, Tampere, Finland.
- 78. Department of Clinical Physiology, University of Tampere School of Medicine, Tampere, Finland.
- 79. Clinical and Molecular Osteoporosis Research Unit, Department of Clinical Sciences and Department of Orthopaedics, Lund University, Malmö, Sweden
- 80. Department of Internal Medicine, University of Manitoba, Winnipeg, Canada.
  - 81. Department of Endocrinology, Vrije Universiteit (VU) University Medical Center, Amsterdam, The Netherlands.

- 82. Extramuraal Geneeskundig Onderzoek (EMGO) Institute for Health and Care Research, Vrije Universiteit (VU) University Medical Center, Amsterdam, The Netherlands.
- 83. Department of Medical Sciences, University of Uppsala, Uppsala, Sweden.
- 84. Department of Pharmacology and Neuroscience, Umeå University, Umeå, Sweden.
- 85. Harvard Medical School, Boston, USA.
- 86. Department of Molecular Biology, Medical Biochemistry and Pathology, Université Laval, Québec City, Canada.
- 87. The APOGEE-Net/CanGèneTest Network on Genetic Health Services and Policy, Université Laval, Québec City, Canada.
- 88. Department of Molecular Epidemiology, Leiden University Medical Center, Leiden, The Netherlands.
- 89. Department of Chemical Pathology, The Chinese University of Hong Kong, Hong Kong SAR, China.
- 90. Li Ka Shing Institute of Health Sciences, The Chinese University of Hong Kong, Hong Kong SAR, China.
- 91. Department of Medical Genetics, University of Antwerp, Antwerp, Belgium.
- 92. Department of Medicine, Turku University Hospital, Turku, Finland.
- 93. Department of Medicine, University of Turku, Turku, Finland.
- 94. Department of Legal Medicine, University of Cantabria, Santander, Spain.
- 95. Department of Human Genetics, McGill University, Montreal, Canada.
- 96. McGill University and Genome Québec Innovation Centre, Montreal, Canada.
- 97. Wellcome Trust Sanger Institute, Hinxton, UK.
- 98. Department of Orthopedic Surgery, Akureyri Hospital, Akureyri, Iceland.
- 99. Institution of Health Science, University Of Akureyri, Akureyri, Iceland.
- 100. Department of Epidemiology and Biostatistics, Lady Davis Institute, McGill University, Montreal, Canada.
- 101. Department of Medical Biochemistry, Oslo University Hospital, Oslo, Norway.
- 102. Medical Genetics Institute, Cedars-Sinai Medical Center, Los Angeles, USA.
- 103. Department of Endocrinology and Metabolism, University Hospital, Reykjavik, Iceland.
- 104. Genetics of Complex Traits, Peninsula College of Medicine and Dentistry, University of Exeter, Exeter, England.
- 105. Department of Clinical Biochemistry, Lovisenberg Deacon Hospital, Oslo, Norway.
- 106. Institute of Basic Medical Sciences, University of Oslo, Oslo, Norway.
- 107. Department of Medical Genetics, McGill University Health Centre, Montreal, Canada.
- 108. Division of Genetics and Rheumatology, Brigham and Women's Hospital, Harvard Medical School, Boston, United States.
- 109. Program in Medical and Population Genetics, Broad Institute, Cambridge, United States.
- 110. Department of Epidemiology, University of Pittsburgh, Pittsburgh, USA.
- 111. California Pacific Medical Center, San Francisco, CA, USA.
- 112. National Institute for Health and Research (NIHR) Musculoskeletal Biomedical Research Unit, University of Sheffield, Sheffield, UK.
- 113. Department of Internal Medicine, The Ohio State University, Columbus, USA.
- 114. Center for Clinical and Translational Science, The Ohio State University, Columbus, USA.
- 115. Menzies Research Institute, University of Tasmania, Hobart, Australia.
- 116. Durrer Center for Cardiogenetic Research, Amsterdam, The Netherlands.
- 117. Interuniversity Cardiology Institute of the Netherlands, Utrecht, The Netherlands.
- 118. Department of Clinical Chemistry, Tampere University Hospital, Tampere, Finland.
- 119. Department of Clinical Chemistry, University of Tampere School of Medicine, Tampere, Finland.
- 120. Center for Human Genomics, Wake Forest University School of Medicine, Winston-Salem, NC, USA.
- 121. Academic Unit of Bone Metabolism, Metabolic Bone Centre, University of Sheffield, Sheffield, UK.
- 122. Rural Clinical School, The University of Queensland, Toowoomba, Australia.
- 123. Department of Clinical Genetics, Erasmus Medical Center, Rotterdam, The Netherlands.
- 124. Department of Medicine, Indiana University School of Medicine, Indianapolis, USA.
- 125. Department of Medicine, University of Auckland, Auckland, New Zealand.
- 126. Department of Medicine, University of Davis, Sacramento, CA, USA.
- 127. Kolling Institute, Royal North Shore Hospital, University of Sydney, Sydney, Australia.

- 128. Department of Psychiatry, The University of Hong Kong, Hong Kong, China.
- 129. Centre for Reproduction, Development and Growth, The University of Hong Kong, Hong Kong, China.
- 130. Geriatric Research and Education Clinical Center (GRECC), Veterans Administration Medical Center, Baltimore, MD, USA.
- 131. Department of Preventive Medicine, University of Tennessee College of Medicine, Memphis, TN, USA.
- 132. MRC Epidemiology Unit Box 285, Medical Research Council, Cambridge, UK.
- 133. Framingham Heart Study, Framingham, USA.
- 134. Departments of Medicine, Epidemiology and Health Services, University of Washington, Seattle, USA.
- 135. Group Health Research Institute, Group Health Cooperative, Seattle, USA.
- 136. Medicine box 157, University of Cambridge, Cambridge, UK.
- 137. Departments of Medicine, Human Genetics, Epidemiology and Biostatistics, Lady Davis Institute, McGill University, Montreal, Canada.
- 138. Stanford Prevention Research Center, Stanford University, Stanford, USA.

### **The GENIE Consortium**

Niina Sandholm<sup>1-3</sup>, Rany M Salem<sup>4-6</sup>, Amy Jayne McKnight<sup>7</sup>, Eoin P Brennan<sup>8-9</sup>, Carol Forsblom<sup>1-2</sup>, Tamara Isakova<sup>10</sup>, Gareth J McKay<sup>7</sup>, Winfred W Williams<sup>6,11</sup>, Denise M Sadlier<sup>8-9</sup>, Ville-Petteri Mäkinen<sup>1-2,12</sup>, Elizabeth J Swan<sup>7</sup>, Cameron Palmer<sup>4-5</sup>, Andrew P Boright<sup>13</sup>, Emma Ahlqvist<sup>14</sup>, Harshal A Deshmukh<sup>15</sup>, Benjamin J Keller<sup>16</sup>, Huateng Huang<sup>17</sup>, Aila Ahola<sup>1-2</sup>, Emma Fagerholm<sup>1-2</sup>, Daniel Gordin<sup>1-2</sup>, Valma Harjutsalo<sup>1-2,18</sup>, Bing He<sup>19</sup>, Outi Heikkilä<sup>1-2</sup>, Kustaa Hietala<sup>1,20</sup>, Janne Kytö<sup>1,20</sup>, Päivi Lahermo<sup>21</sup>, Markku Lehto<sup>1-2</sup>, Anne-May Österholm<sup>19</sup>, Maija Parkkonen<sup>1-2</sup>, Janne Pitkäniemi<sup>22</sup>, Milla Rosengård-Bärlund<sup>1-2</sup>, Markku Saraheimo<sup>1-2</sup>, Cinzia Sarti<sup>22</sup>, Jenny Söderlund<sup>1-2</sup>, Aino Soro-Paavonen<sup>1-2</sup>, Anna Syreeni<sup>1-2</sup>, Lena M Thorn<sup>1-2</sup>, Heikki Tikkanen<sup>23</sup>, Nina Tolonen<sup>1-2</sup>, Karl Tryggvason<sup>19</sup>, Jaakko Tuomilehto<sup>18,24-26</sup>, Johan Wadén<sup>1-2</sup>, Geoffrey V Gill<sup>27</sup>, Sarah Prior<sup>28</sup>, Candace Guiducci<sup>4</sup>, Daniel B Mirel<sup>4</sup>, Andrew Taylor<sup>4,11</sup>, Mohsen Hosseini<sup>29-30</sup>, DCCT/EDIC Research Group<sup>§31-32</sup>, Hans-Henrik Parving<sup>33-34</sup>, Peter Rossing<sup>35</sup>, Lise Tarnow<sup>35</sup>, Claes Ladenvall<sup>14</sup>, François Alhenc-Gelas<sup>36</sup>, Pierre Lefebvre<sup>37</sup>, Vincent Rigalleau<sup>38</sup>, Ronan Roussel<sup>39-40</sup>, David-Alexandre Tregouet<sup>41</sup>, Anna Maestroni<sup>42</sup>, Silvia Maestroni<sup>42</sup>, Henrik Falhammar<sup>43-44</sup>, Tianwei Gu<sup>43</sup>, Anna Möllsten<sup>45</sup>, Dan Cimponeriu<sup>46</sup>, Ioana Mihai<sup>47</sup>, Maria Mota<sup>47</sup>, Eugen Mota<sup>47</sup>, Cristian Serafinceanu<sup>48</sup>, Monica Stavarachi<sup>46</sup>, Robert L Hanson<sup>49</sup>, Robert G Nelson<sup>49</sup>, Matthias Kretzler<sup>50</sup>, Helen M Colhoun<sup>15</sup>, Nicolae Mircea Panduru<sup>48</sup>, Harvest F Gu<sup>43</sup>, Kerstin Brismar<sup>43</sup>, Gianpaolo Zerbini<sup>42</sup>, Samy Hadjadj<sup>51-52</sup>, Michel Marre<sup>39-40</sup>, Leif Groop<sup>14</sup>, Maria Lajer<sup>35</sup>, Shelley B Bull<sup>53-54</sup>, Daryl Waggott<sup>53</sup>, Andrew D Paterson<sup>30,54</sup>, David A Savage<sup>7</sup>, Stephen C Bain<sup>28</sup>, Finian Martin<sup>8-9</sup>, Joel N Hirschhorn<sup>4-6</sup>, Catherine Godson<sup>8-9</sup>, Jose C Florez<sup>4,6,11</sup>, Per-Henrik Groop<sup>1-2,55</sup> and Alexander P Maxwell<sup>7,56</sup>

- 1. Folkhälsan Institute of Genetics, Folkhälsan Research Center, Biomedicum Helsinki, Helsinki, Finland.
- 2. Division of Nephrology, Department of Medicine, Helsinki University Central Hospital, Helsinki, Finland.
- 3. Department of Biomedical Engineering and Computational Science, Aalto University, Espoo, Finland.
- 4. Program in Medical and Population Genetics, Broad Institute, Cambridge, MA, USA.
- 5. Endocrine Research Unit, Department of Endocrinology, Children's Hospital, Boston, MA, USA.
- 6. Department of Medicine, Harvard Medical School, Boston, MA, USA.
- 7. Nephrology Research, Centre for Public Health, Queen's University of Belfast, Belfast, UK.
- 8. Diabetes Research Centre, Conway Institute, School of Medicine and Medical Sciences, University College Dublin, Dublin, Ireland.
- 9. Mater Misericordiae Hospital, Dublin, Ireland.
- 10. Division of Nephrology and Hypertension, University of Miami, Miami, FL, USA.
- 11. Center for Human Genetic Research, Massachusetts General Hospital, Boston, MA, USA.
- 12. Institute of Clinical Medicine, Department of Internal Medicine, Biocenter Oulu and Clinical Research Center, University of Oulu, Oulu, Finland.
- 13. Department of Medicine, University of Toronto, Toronto, Canada.
- 14. Department of Clinical Sciences, Diabetes and Endocrinology, Skåne University Hospital, Lund University, Malmö, Sweden.
- 15. Wellcome Trust Centre for Molecular Medicine, University of Dundee, Dundee, Scotland, UK.
- 16. Computer Science, Eastern Michigan University, Ypsilanti, MI, USA.
- 17. Division of Nephrology, Internal Medicine, University of Michigan, Ann Arbor, MI, USA.
- 18. Diabetes Prevention Unit, National Institute for Health and Welfare, 00271 Helsinki, Finland.
- 19. Division of Matrix Biology, Department of Medical Biochemistry and Biophysics, Karolinska Institutet, Stockholm, Sweden.
- 20. Department of Ophthalmology, Helsinki University Central Hospital, Helsinki, Finland.

- 21. Institute for Molecular Medicine Finland, Helsinki, Finland.
- 22. Hjelt Institute, Department of Public Health, University of Helsinki, Helsinki, Finland.
- 23. Unit for Sports and Exercise Medicine, Institute of Clinical Medicine, University of Helsinski, Finland.
- 24. South Ostrobothnia Central Hospital, 60220 Seinäjoki, Finland.
- 25. Red RECAVA Grupo RD06/0014/0015, Hospital Universitario La Paz, 28046 Madrid, Spain.
- 26. Centre for Vascular Prevention, Danube-University Krems, 3500 Krems, Austria.
- 27. Diabetes Endocrine Unit, University of Liverpool, Clinical Sciences Centre, Aintree University Hospital, Liverpool, UK.
- 28. Institute of Life Sciences, Swansea University, Swansea, UK.
- 29. Institute of Medical Sciences, University of Toronto, Toronto, Canada.
- 30. Program in Genetics and Genome Biology, Hospital for Sick Children, Toronto, Canada.
- 31. NIDDK, National Institutes of Health, Bethesda, MD, USA.
- 32. Biostatics Division, The George Washington University, Washington, DC, USA.
- 33. Department of Medical Endocrinology, University Hospital of Copenhagen, Copenhagen, Denmark.
- 34. Faculty of Health Sciences, University of Aarhus, Aarhus, Denmark.
- 35. Steno Diabetes Center, Gentofte, Denmark.
- 36. INSERM U872, Paris-Descartes University, Pierre and Marie Curie University, Paris, France.
- 37. CHU Sart Tilman, Liège, Belgium.
- 38. CHU Bordeaux, Bordeaux, France.
- 39. Diabetes Department, Hôpital Bichat-Claude Bernard, Assistance Publique des Hôpitaux de Paris, Paris, France.
- 40. INSERM U 695, Université Denis Diderot Paris 7, Paris, France.
- 41. INSERM UMR\_S 937, ICAN Institute for Cardiometabolism and Nutrition, Pierre & Marie Curie University, 75013 Paris, France.
- 42. Complications of Diabetes Unit, Division of Metabolic and Cardiovascular Sciences, San Raffaele Scientific Institute, 20132 Milano, Italy.
- 43. Department of Molecular Medicine and Surgery, Karolinska Institutet, Stockholm, Sweden.
- 44. Department of Endocrinology, Metabolism and Diabetes, Karolinska University Hospital, Stockholm, Sweden.
- 45. Department of Clinical Sciences, Paediatrics, Umeå University, Umeå, Sweden.
- 46. Genetics Department of Bucharest University, Bucharest, Romania.
- 47. University of Medicine and Pharmacy of Craiova, Craiova, Romania.
- 48. "Carol Davila" University of Medicine and Pharmacy, Bucharest, Romania.
- 49. Diabetes Epidemiology and Clinical Research Section, NIDDK, Phoenix, AZ, USA.
- 50. Internal Medicine, Center for Computational Medicine and Bioinformatics, University of Michigan, Ann Arbor, MI, USA.
- 51. CHU Poitiers Endocrinology, University of Poitiers, Poitiers, France.
- 52. INSERM CIC0802, CHU Poitiers, Poitiers, France.
- 53. Prosserman Centre for Health Research, Samuel Lunenfeld Research Institute, Toronto, Canada.
- 54. Division of Biostatistics, Dalla Lana School of Public Health, University of Toronto, Toronto, Canada.
- 55. Baker IDI Heart and Diabetes Institute, Melbourne, Australia.
- 56. Regional Nephrology Unit, Level 11, Tower Block, Belfast City Hospital, Belfast, UK.

## The GLGC Consortium

Cristen J. Willer, 1,2,3,4 Ellen M. Schmidt, Sebanti Sengupta, Gina M. Peloso, 5,6,7 Stefan Gustafsson, 8,9 Stavroula Kanoni, <sup>10</sup> Andrea Ganna, <sup>8,9,11</sup> Jin Chen, <sup>4</sup> Martin L. Buchkovich, <sup>12</sup> Samia Mora, <sup>13,14</sup> Jacques S. Beckmann, <sup>15,16</sup> Jennifer L. Bragg-Gresham, <sup>4</sup> Hsing-Yi Chang, <sup>17</sup> Ayşe Demirkan, <sup>18</sup> Heleen M. Den Hertog, <sup>19</sup> Ron Do,<sup>6</sup> Louise A. Donnelly,<sup>20</sup> Georg B. Ehret,<sup>21,22</sup> Tõnu Esko,<sup>7,23,24</sup> Mary F. Feitosa,<sup>25</sup> Teresa Ferreira,<sup>26</sup> Krista Fischer, 23 Pierre Fontanillas, 7 Ross M. Fraser, 27 Daniel F. Freitag, 28 Deepti Gurdasani, 10,28 Kauko Heikkilä,<sup>29</sup> Elina Hyppönen,<sup>30</sup> Aaron Isaacs,<sup>18,31</sup> Anne U. Jackson,<sup>4</sup> Åsa Johansson,<sup>32,33</sup> Toby Johnson,<sup>34,35</sup> Marika Kaakinen, 36,37 Johannes Kettunen, Marcus E. Kleber, 40,41 Xiaohui Li,42 Jian'an Luan,43 Leo-Pekka Lyytikäinen, 44,45 Patrik K.E. Magnusson, 11 Massimo Mangino, 46 Evelin Mihailov, 23,24 May E. Montasser, 47 Martina Müller-Nurasyid, 48,49,50 Ilja M. Nolte, 51 Jeffrey R. O'Connell, 47 Cameron D. Palmer, 7,52,53 Markus Perola, <sup>23,38,39</sup> Ann-Kristin Petersen, <sup>48</sup> Serena Sanna, <sup>54</sup> Richa Saxena, <sup>55</sup> Susan K. Service, <sup>56</sup> Sonia Shah, <sup>57</sup> Dmitry Shungin. 58,59,60 Carlo Sidore, 4,54,61 Ci Song, 8,9,11 Rona J. Strawbridge, 62,63 Ida Surakka, 38,39 Toshiko Tanaka, <sup>64</sup> Tanya M. Teslovich, <sup>4</sup> Gudmar Thorleifsson, <sup>65</sup> Evita G. Van den Herik, <sup>19</sup> Benjamin F. Voight, <sup>66,67</sup> Kelly A. Volcik, 68 Lindsay L. Waite, 69 Andrew Wong, 70 Ying Wu, 12 Weihua Zhang, 71,72 Devin Absher, 69 Gershim Asiki, <sup>73</sup> Inês Barroso, <sup>10,74</sup> Latonya F. Been, <sup>75</sup> Jennifer L. Bolton, <sup>27</sup> Lori L Bonnycastle, <sup>76</sup> Paolo Brambilla, <sup>77</sup> Mary S. Burnett, <sup>78</sup> Giancarlo Cesana, <sup>79</sup> Maria Dimitriou, <sup>80</sup> Alex S.F. Doney, <sup>20</sup> Angela Döring, <sup>81,82</sup> Paul Elliott, <sup>37,83</sup> Stephen E. Epstein, <sup>78</sup> Gudmundur Ingi Eyjolfsson, <sup>84</sup> Bruna Gigante, <sup>85</sup> Mark O. Goodarzi, <sup>86</sup> Harald Grallert, <sup>87</sup> Martha L. Gravito, <sup>75</sup> Christopher J. Groves, <sup>88</sup> Göran Hallmans, <sup>89</sup> Anna-Liisa Hartikainen, <sup>90</sup> Caroline Hayward, <sup>91</sup> Dena Hernandez, <sup>92</sup> Andrew A. Hicks, <sup>93</sup> Hilma Holm, <sup>65</sup> Yi-Jen Hung, <sup>94</sup> Thomas Illig, <sup>87,95</sup> Michelle R. Jones, <sup>86</sup> Pontiano Kaleebu, 73 John J.P. Kastelein, 96 Kay-Tee Khaw, 97 Eric Kim, 42 Norman Klopp, 87,95 Pirjo Komulainen, 98 Meena Kumari,<sup>57</sup> Claudia Langenberg,<sup>43</sup> Terho Lehtimäki,<sup>44,45</sup> Shih-Yi Lin,<sup>99</sup> Jaana Lindström,<sup>100</sup> Ruth J.F. Loos, 43,101,102,103 François Mach, 21 Wendy L McArdle, 104 Christa Meisinger, 81 Braxton D. Mitchell, 47 Gabrielle Müller, 105 Ramaiah Nagaraja, 106 Narisu Narisu, 76 Tuomo V.M. Nieminen, 107,108,109 Rebecca N. Nsubuga, 73 Isleifur Olafsson, <sup>110</sup> Ken K. Ong, <sup>43,70</sup> Aarno Palotie, <sup>38,111,112</sup> Theodore Papamarkou, <sup>10,28,113</sup> Cristina Pomilla, <sup>10,28</sup> Anneli Pouta, 90,114 Daniel J. Rader, 115,116 Muredach P. Reilly, 115,116 Paul M. Ridker, 13,14 Fernando Rivadeneira, 117,118,119 Igor Rudan, 27 Aimo Ruokonen, 120 Nilesh Samani, 121,122 Hubert Scharnagl, 123 Janet Seeley, 73,124 Kaisa Silander, 38,39 Alena Stančáková, 125 Kathleen Stirrups, 10 Amy J. Swift, 76 Laurence Tiret, 126 Andre G. Uitterlinden, 117,118,119 L. Joost van Pelt, 127,128 Sailaja Vedantam, 7,52,53 Nicholas Wainwright, 10,28 Cisca Wijmenga, 128,129 Sarah H. Wild, 27 Gonneke Willemsen, 130 Tom Wilsgaard, 131 James F. Wilson, 27 Elizabeth H. Young, 10,28 Jing Hua Zhao, 43 Linda S. Adair, 132 Dominique Arveiler, 133 Themistocles L. Assimes, 134 Stefania Bandinelli, 135 Franklyn Bennett, 136 Murielle Bochud, 137 Bernhard O. Boehm, 138, 139 Dorret I. Boomsma, 130 Ingrid B. Borecki, <sup>25</sup> Stefan R. Bornstein, <sup>140</sup> Pascal Bovet, <sup>137,141</sup> Michel Burnier, <sup>142</sup> Harry Campbell, <sup>27</sup> Aravinda Chakravarti,<sup>22</sup> John C. Chambers,<sup>71,72,143</sup> Yii-Der Ida Chen,<sup>144,145</sup> Francis S. Collins,<sup>76</sup> Richard S. Cooper,<sup>146</sup> John Danesh, <sup>28</sup> George Dedoussis, <sup>80</sup> Ulf de Faire, <sup>85</sup> Alan B. Feranil, <sup>147</sup> Jean Ferrières, <sup>148</sup> Luigi Ferrucci, <sup>64</sup>

Nelson B. Freimer, 56,149 Christian Gieger, 48 Leif C. Groop, 150,151 Vilmundur Gudnason, 152 Ulf Gyllensten, 32 Anders Hamsten, 62,63,153 Tamara B. Harris, 154 Aroon Hingorani, 57 Joel N. Hirschhorn, 7,52,53 Albert Hofman, 117,119 G. Kees Hovingh, 96 Chao Agnes Hsiung, 155 Steve E. Humphries, 156 Steven C. Hunt, 157 Kristian Hveem, 158 Carlos Iribarren, <sup>159</sup> Marjo-Riitta Järvelin, <sup>36,37,83,114,160</sup> Antti Jula, <sup>161</sup> Mika Kähönen, <sup>162</sup> Jaakko Kaprio, <sup>29,38,163</sup> Antero Kesäniemi, 164 Mika Kivimaki, 57 Jaspal S. Kooner, 72,143,165 Peter J. Koudstaal, 19 Ronald M. Krauss, 166 Diana Kuh,<sup>70</sup> Johanna Kuusisto,<sup>167</sup> Kirsten O. Kyvik,<sup>168,169</sup> Markku Laakso,<sup>167</sup> Timo A. Lakka,<sup>98,170</sup> Lars Lind,<sup>171</sup> Cecilia M. Lindgren, <sup>26</sup> Nicholas G. Martin, <sup>172</sup> Winfried März, <sup>41,123,173</sup> Mark I. McCarthy, <sup>26,88</sup> Colin A. McKenzie, <sup>174</sup> Pierre Meneton, <sup>175</sup> Andres Metspalu, <sup>23,24</sup> Leena Moilanen, <sup>176</sup> Andrew D. Morris, <sup>20</sup> Patricia B. Munroe, <sup>34,35</sup> Inger Njølstad, <sup>131</sup> Nancy L. Pedersen, <sup>11</sup> Chris Power, <sup>30</sup> Peter P. Pramstaller, <sup>93,177,178</sup> Jackie F. Price, <sup>27</sup> Bruce M. Psaty, 179,180 Thomas Quertermous, 134 Rainer Rauramaa, 98,181 Danish Saleheen, 28,182,183 Veikko Salomaa, 184 Dharambir K. Sanghera, <sup>75</sup> Jouko Saramies, <sup>185</sup> Peter E.H. Schwarz, <sup>140,186</sup> Wayne H-H Sheu, <sup>187</sup> Alan R. Shuldiner, <sup>47,188</sup> Agneta Siegbahn, <sup>8,33,171</sup> Tim D. Spector, <sup>46</sup> Kari Stefansson, <sup>65,189</sup> David P. Strachan, <sup>190</sup> Bamidele O. Tayo, <sup>146</sup> Elena Tremoli, <sup>191</sup> Jaakko Tuomilehto, <sup>100,192,193,194</sup> Matti Uusitupa, <sup>195,196</sup> Cornelia M. van Duijn, <sup>18,31</sup> Peter Vollenweider, <sup>197</sup> Lars Wallentin, <sup>33,171</sup> Nicholas J. Wareham, <sup>43</sup> John B. Whitfield, <sup>172</sup> Bruce H.R. Wolffenbuttel, 128,198 Jose M. Ordovas, 199,200,201 Eric Boerwinkle, 68 Colin N.A. Palmer, 20 Unnur Thorsteinsdottir, 65,189 Daniel I. Chasman, 13,14 Jerome I. Rotter, 42 Paul W. Franks, 58,60,202 Samuli Ripatti, 10,38,39 L. Adrienne Cupples, 5,203 Manjinder S. Sandhu, 10,28 Stephen S. Rich, 204 Michael Boehnke, Panos Deloukas, 10 Sekar Kathiresan, 6,7,205,206 Karen L. Mohlke, 12 Erik Ingelsson, 8,9,26 Gonçalo R. Abecasis<sup>4</sup>

- 1. Department of Internal Medicine, Division of Cardiovascular Medicine, University of Michigan, Ann Arbor, Michigan 48109, USA
- 2. Department of Computational Medicine and Bioinformatics, University of Michigan, Ann Arbor, Michigan 48109, USA
- 3. Department of Human Genetics, University of Michigan, Ann Arbor, Michigan 48109, USA
- 4. Center for Statistical Genetics, Department of Biostatistics, University of Michigan, Ann Arbor, Michigan 48109, USA
- 5. Department of Biostatistics, Boston University School of Public Health, Boston, Massachusetts 02118, USA
- 6. Center for Human Genetic Research, Massachusetts General Hospital, Boston, Massachusetts 02114, USA
- 7. Broad Institute, Program in Medical and Population Genetics, Cambridge, Massachusetts 02142, USA
- 8. Department of Medical Sciences, Molecular Epidemiology, Uppsala University, Uppsala, Sweden
- 9. Science for Life Laboratory, Uppsala University, Uppsala, Sweden
- 10. Wellcome Trust Sanger Institute, Wellcome Trust Genome Campus, CB10 1SA, Hinxton, United Kingdom
- 11. Department of Medical Epidemiology and Biostatistics, Karolinska Institutet, Stockholm, Sweden
- 12. Department of Genetics, University of North Carolina, Chapel Hill, NC 27599 USA
- 13. Division of Preventive Medicine, Brigham and Women's Hospital, 900 Commonwealth Ave., Boston MA 02215, USA
- 14. Harvard Medical School, Boston MA 02115, USA
- 15. Service of Medical Genetics, Lausanne University Hospital, Lausanne, Switzerland
- 16. Department of Medical Genetics, University of Lausanne, Lausanne, Switzerland
- 17. Division of Preventive Medicine and Health Services Research, Institute of Population Health Sciences, National Health Research Institutes, Zhunan, Taiwan
- 18. Genetic Epidemiology Unit, Department of Epidemiology, Erasmus University Medical Center, Rotterdam, The Netherlands
- 19. Department of Neurology, Erasmus Medical Center, Rotterdam, The Netherlands
- 20. Medical Research Institute, University of Dundee, Ninewells Hospital and Medical School. Dundee, DD1 9SY, United Kingdom

- 21. Cardiology, Department of Specialities of Medicine, Geneva University Hospital, Rue Gabrielle-Perret-Gentil 4, 1211 Geneva 14, Switzerland
- 22. Center for Complex Disease Genomics, McKusick-Nathans Institute of Genetic Medicine, Johns Hopkins University School of Medicine, Baltimore, MD 21205. USA
- 23. Estonian Genome Center of the University of Tartu, Tartu, Estonia
- 24. Institute of Molecular and Cell Biology, University of Tartu, Tartu, Estonia
- 25. Department of Genetics, Washington University School of Medicine, USA
- 26. Wellcome Trust Centre for Human Genetics, University of Oxford, Oxford, OX3 7BN, United Kingdom
- 27. Centre for Population Health Sciences, University of Edinburgh, Teviot Place, Edinburgh, EH8 9AG, Scotland, United Kingdom
- 28. Department of Public Health and Primary Care, University of Cambridge, Cambridge, United Kingdom
- 29. Hjelt Institute, Department of Public Health, University of Helsinki, Finland
- 30. Centre For Paediatric Epidemiology and Biostatistics/MRC Centre of Epidemiology for Child Health, University College of London Institute of Child Health, London, United Kingdom
- 31. Centre for Medical Systems Biology, Leiden, the Netherlands
- 32. Department of Immunology, Genetics and Pathology, Uppsala University, Uppsala, Sweden
- 33. Uppsala Clinical Research Center, Uppsala University, Uppsala, Sweden
- 34. Genome Centre, Barts and The London School of Medicine and Dentistry, Queen Mary University of London, London, UK
- 35. Clinical Pharmacology, NIHR Cardiovascular Biomedical Research Unit, William Harvey Research Institute, Barts and The London School of Medicine and Dentistry Queen Mary University of London, London, UK
- 36. Biocenter Oulu, University of Oulu, Oulu, Finland
- 37. Institute of Health Sciences, University of Oulu, Finland
- 38. Institute for Molecular Medicine Finland FIMM, University of Helsinki, Finland
- 39. Public Health Genomics Unit, National Institute for Health and Welfare, Helsinki, Finland
- 40. Department of Internal Medicine II Cardiology, University of Ulm Medical Centre, Ulm, Germany
- 41. Mannheim Institute of Public Health, Social and Preventive Medicine, Medical Faculty of Mannheim, University of Heidelberg, Ludolf-Krehl-Strasse 7-11, 68167 Mannheim, Germany
- 42. Medical Genetics Institute, Cedars-Sinai Medical Center, Los Angeles, CA 90048, USA
- 43. MRC Epidemiology Unit, Institute of Metabolic Science, Box 285, Addenbrooke's Hospital, Hills Road, Cambridge, CB2 0QQ, United Kingdom
- 44. Department of Clinical Chemistry, Fimlab Laboratories, Tampere 33520, Finland
- 45. Department of Clinical Chemistry, University of Tampere School of Medicine, Tampere 33014, Finland
- 46. Department of Twin Research and Genetic Epidemiology, King's College London, London, United Kingdom
- 47. Division of Endocrinology, Diabetes, and Nutrition, Department of Medicine, University of Maryland, School of Medicine, Baltimore, Maryland
- 48. Institute of Genetic Epidemiology, Helmholtz Zentrum München, Neuherberg 85764, Germany
- 49. Department of Medicine I, University Hospital Grosshadern, Ludwig-Maximilians University, Munich, Germany
- 50. Institute of Medical Informatics, Biometry and Epidemiology, Ludwig-Maximilians-University of Munich, Munich, Germany
- 51. Department of Epidemiology, University of Groningen, University Medical Center Groningen, The Netherlands
- 52. Division of Endocrinology, Children's Hospital Boston, Massachusetts 02115, USA
- 53. Division of Genetics, Program in Genomics, Children's Hospital, Boston, Massachusetts 02115, USA
- 54. Istituto di Ricerca Genetica e Biomedica, CNR, Monserrato, 09042, Italy
- 55. Massachusetts General Hospital/Broad Institute, Harvard University, Cambridge, MA, USA
- 56. Center for Neurobehavioral Genetics, The Semel Institute for Neuroscience and Human Behavior, University of California, Los Angeles, USA
- 57. Genetic Epidemiology Group, Department of Epidemiology and Public Health, UCL, London WC1E 6BT, United Kingdom
- 58. Department of Clinical Sciences, Genetic & Molecular Epidemiology Unit, Lund University Diabetes Center, Scania University Hosptial, Malmö, Sweden
- 59. Department of Odontology, Umeå University, Umeå, Sweden
- 60. Department of Public Health and Primary Care, Unit of Medicine, Umeå University, Umeå, Sweden
- 61. Dipartimento di Scienze Biomediche, Universita di Sassari, 07100 SS, Italy
- 62. Atherosclerosis Research Unit, Department of Medicine Solna, Karolinska University Hospital, Karolinska Institutet, Stockholm, Sweden

- 63. Center for Molecular Medicine, Karolinska University Hospital, Stockholm, Sweden
- 64. Clinical Research Branch, National Institute Health, Baltimore, MD, USA
- 65. deCODE Genetics/Amgen, 101 Reykjavik, Iceland
- 66. Department of Genetics, University of Pennsylvania School of Medicine, Philadelphia PA, 19104, USA
- 67. Department of Systems Pharmacology and Translational Therapeutics, University of Pennsylvania School of Medicine, Philadelphia PA, 19104, USA
- 68. Human Genetics Center, University of Texas Health Science Center School of Public Health, Houston, TX 77030, USA
- 69. HudsonAlpha Institute for Biotechnology, Huntsville, AL, USA
- 70. MRC Unit for Lifelong Health and Ageing, 33 Bedford Place, London, WC1B 5JU, United Kingdom
- 71. Department of Epidemiology and Biostatistics, School of Public Health, Imperial College London, London, United Kingdom
- 72. Ealing Hospital, Southall, Middlesex UB1 3HW, United Kingdom
- 73. MRC/UVRI Uganda Research Unit on AIDS, Entebbe, Uganda
- 74. University of Cambridge Metabolic Research Laboratories and NIHR Cambridge Biomedical Research Centre, Level 4, Institute of Metabolic Science Box 289 Addenbrooke's Hospital Cambridge CB2 OQQ, UK
- 75. Department of Pediatrics, University of Oklahoma Health Sciences Center, Oklahoma City, OK, USA
- 76. Genome Technology Branch, National Human Genome Research Institute, NIH, Bethesda, MD 20892, USA
- 77. Department of Experimental Medicine, University of Milano Bicocca, Italy
- 78. MedStar Health Research Institute, 6525 Belcrest Road, Suite 700, Hyattsville, MD 20782, USA
- 79. Research Centre on Public Health, University of Milano Bicocca, Italy
- 80. Department of Dietetics-Nutrition, Harokopio University, 70 El. Venizelou Str, Athens, Greece
- 81. Institute of Epidemiology I, Helmholtz Zentrum München, Neuherberg 85764, Germany
- 82. Institute of Epidemiology II, Helmholtz Zentrum München, Neuherberg 85764, Germany
- 83. Department of Epidemiology and Biostatistics, MRC Health Protection Agency (HPA) Centre for Environment and Health, School of Public Health, Imperial College London, UK
- 84. The Laboratory in Mjodd, 108 Reykjavik, Iceland
- 85. Division of Cardiovascular Epidemiology, Institute of Environmental Medicine, Karolinska Institutet, Stockholm, Sweden
- 86. Division of Endocrinology, Diabetes and Metabolism, Department of Medicine, Cedars-Sinai Medical Center, Los Angeles, CA 90048, USA
- 87. Research Unit of Molecular Epidemiology, Helmholtz Zentrum München, Neuherberg 85764, Germany
- 88. Oxford Centre for Diabetes, Endocrinology and Metabolism, University of Oxford, OX3 7LJ, United Kingdom
- 89. Department of Public Health and Clinical Medicine, Nutritional research, Umeå University, Umeå, Sweden
- 90. Department of Clinical Sciences/Obstetrics and Gynecology, Oulu University Hospital, Oulu, Finland
- 91. MRC Human Genetics Unit, Institute of Genetics and Molecular Medicine, Western General Hospital, Edinburgh, Scotland, United Kingdom
- 92. Laboratory of Neurogenetics, National Institute on Aging, Bethesda, MD 20892, USA
- 93. Center for Biomedicine, European Academy Bozen/Bolzano (EURAC), Bolzano, Italy Affiliated Institute of the University of Lübeck, Lübeck, Germany
- 94. Division of Endocrinology & Metabolism, Tri-Service General Hospital, National Defense Medical Center, Taipei, Taiwan
- 95. Hannover Unified Biobank, Hannover Medical School, Hannover 30625, Germany
- 96. Department of Vascular Medicine, Academic Medical Center, Amsterdam, The Netherlands
- 97. Clinical Gerontology Unit, University of Cambridge, Cambridge, United Kingdom
- 98. Kuopio Research Institute of Exercise Medicine, Kuopio, Finland
- 99. Division of Endocrine and Metabolism, Department of Internal Medicine, Taichung Veterans General Hospital, School of Medicine, National Yang-Ming University, Taipei, Taiwan
- 100. Diabetes Prevention Unit, National Institute for Health and Welfare, 00271 Helsinki, Finland
- 101. The Genetics of Obesity and Related Metabolic Traits Program, The Icahn School of Medicine at Mount Sinai, New York, USA
- 102. The Charles Bronfman Institute for Personalized Medicine, The Icahn School of Medicine at ount Sinai, New York, USA
- 103. The Mindich Child Health and Development Institute, The Icahn School of Medicine at Mount Sinai, New York
- 104. School of Social and Community Medicine, University of Bristol, Oakfield House, Oakfield Grove, Bristol BS8 2BN, United Kingdom

- 105. Institute for Medical Informatics and Biometrics, University of Dresden, Medical Faculty Carl Gustav Carus, Fetscherstrasse 74, 01307 Dresden, Germany
- 106. Laboratory of Genetics, National Institute on Aging, Baltimore, MD21224, USA
- 107. Department of Clinical Pharmacology, University of Tampere School of Medicine, Tamperew 33014, Finland
- 108. Department of Internal Medicine, Päijät-Häme Central Hospital, Lahti, Finland
- 109. Division of Cardiology, Helsinki University Central Hospital, Helsinki, Finland
- 110. Department of Clinical Biochemistry, Landspitali University Hospital, 101 Reykjavik, Iceland
- 111. Department of Medical Genetics, Haartman Institute, University of Helsinki and Helsinki University Central Hospital, Helsinki, Finland
- 112. Genetic Epidemiology Group, Wellcome Trust Sanger Institute, Hinxton, Cambridge, United ingdom
- 113. Department of Statistical Sciences, University College of London, London, United Kingdom
- 114. National Institute for Health and Welfare, Oulu, Finland
- 115. Cardiovascular Institute, Perelman School of Medicine at the University of Pennsylvania, 3400 Civic Center Blvd, Building 421, Translational Research Center, Philadelphia, PA 19104-5158, USA
- 116. Division of Translational Medicine and Human Genetics, Perelman School of Medicine at the University of Pennsylvania, 3400 Civic Center Blvd, Building 421, Translational Research Center, Philadelphia, PA 19104-5158, USA
- 117. Department of Epidemiology, Erasmus University Medical Center, Rotterdam, the Netherlands
- 118. Department of Internal Medicine, Erasmus University Medical Center, Rotterdam, the Netherlands
- 119. Netherlands Genomics Initiative (NGI)-sponsored Netherlands Consortium for Healthy Aging NCHA), Leiden, The Netherlands
- 120. Department of Clinical Sciences/Clinical Chemistry, University of Oulu, Oulu, Finland
- 121. National Institute for Health Research Leicester Cardiovascular Biomedical Research Unit, Glenfield Hospital, Leicester LE3 9QP, UK
- 122. Department of Cardiovascular Sciences, University of Leicester, Glenfield Hospital, Leicester, LE3 9QP, UK
- 123. Clinical Institute of Medical and Chemical Laboratory Diagnostics, Medical University of Graz, Austria
- 124. School of International Development, University of East Anglia, Norwich NR4 7TJ, United Kingdom
- 125. University of Eastern Finland and Kuopio University Hospital, 70210 Kuopio, Finland
- 126. INSERM UMRS 937, Pierre and Marie Curie University, Paris, France
- 127. Department of Laboratory Medicine, University of Groningen, University Medical Center Groningen, The Netherlands
- 128. LifeLines Cohort Study, University of Groningen, University Medical Center Groningen, The Netherlands
- 129. Department of Genetics, University of Groningen, University Medical Center Groningen, The Netherlands
- 130. Department of Biological Psychology, VU Univ, Amsterdam, The Netherlands
- 131. Department of Community Medicine, Faculty of Health Sciences, University of Tromsø, Tromsø, Norway
- 132. Department of Nutrition, University of North Carolina, Chapel Hill, NC, USA
- 133. Department of Epidemiology and Public Health, EA 3430, University of Strasbourg, Faculty of Medicine, Strasbourg, France
- 134. Department of Medicine, Stanford University School of Medicine, Stanford, CA, USA
- 135. Geriatric Unit, Azienda Sanitaria Firenze (ASF), Florence, Italy
- 136. Chemical Pathology, Department of Pathology, University of the West Indies, Mona, Kingston 7, Jamaica
- 137. Institute of Social and Preventive Medicine (IUMSP), Lausanne University Hospital, Route de la Corniche 10, 1010 Lausanne, Switzerland
- 138. Division of Endocrinology and Diabetes, Department of Internal Medicine, Ulm University Medical Centre, Ulm, Germany
- 139. Lee Kong Chian School of Medicine, Nanyang Technological University, Singapore
- 140. Department of Medicine III, University of Dresden, Medical Faculty Carl Gustav Carus, Fetscherstrasse 74, 01307 Dresden, Germany
- 141. Ministry of Health, Victoria, Republic of Seychelles
- 142. Service of Nephrology, Lausanne University Hospital, Lausanne, Switzerland
- 143. Imperial College Healthcare NHS Trust, London, United Kingdom
- 144. Division of Reproductive Endocrinology, Department of Obstetrics and Gynecology, Cedars-Sinai Medical Center, Los Angeles, California, USA
- 145. Department of Medicine, University of California Los Angeles, Los Angeles, California, USA
- 146. Department of Preventive Medicine and Epidemiology, Loyola University Medical School, Maywood, Illinois 60153, USA
- 147. Office of Population Studies Foundation, University of San Carlos, Talamban, Cebu City, Philippines
- 148. Department of Cardiology, Toulouse University School of Medicine, Rangueil Hospital, Toulouse, France

- 149. Department of Psychiatry, University of California, Los Angeles, USA
- 150. Department of Clinical Sciences, Lund University, SE-20502, Malmö, Sweden
- 151. Department of Medicine, Helsinki University Hospital, FI-00029 Helsinki, Finland
- 152. Icelandic Heart Association, Kopavogur, Iceland
- 153. Department of Cardiology, Karolinska University Hospital, Stockholm, Sweden
- 154. Laboratory of Epidemiology, Demography, and Biometry, National Institute on Ageing, Bethesda, MD, USA
- 155. Institute of Population Health Sciences, National Health Research Institutes, Zhunan, Taiwan
- 156. Cardiovascular Genetics, BHF Laboratories, Institute Cardiovascular Science, University College London, London, United Kingdom
- 157. Cardiovascular Genetics, University of Utah School of Medicine, Salt Lake City, UT, USA
- 158. HUNT Research Centre, Department of Public Health and General Practice, Norwegian University of Science and Technology, Levanger, Norway
- 159. Kaiser Permanente, Division of Research, Oakland, CA, USA
- 160. Unit of Primary Care, Oulu University Hospital, Oulu, Finland
- 161. Department of Chronic Disease Prevention, National Institute for Health and Welfare, Turku, Finland
- 162. Department of Clinical Physiology, University of Tampere School of Medicine, Tampere 33014, Finland
- 163. Department of Mental Health and Substance Abuse Services, National Institute for Health and Welfare, Helsinki, Finland
- 164. Institute of Clinical Medicine, Department of Medicine, University of Oulu and Clinical Research Center, Oulu University Hospital, Oulu, Finland
- 165. National Heart & Lung Institute, Imperial College London, Hammersmith Hospital, London, United Kingdom
- 166. Children's Hospital Oakland Research Institute, 5700 Martin Luther King Junior Way, Oakland, CA 94609, USA
- 167. Department of Medicine, University of Eastern Finland and Kuopio University Hospital, 70210 Kuopio, Finland
- 168. Institute of Regional Health Services Research, University of Southern Denmark, Odense, Denmark
- 169. Odense Patient data Explorative Network (OPEN), Odense University Hospital, Odense, Denmark
- 170. Institute of Biomedicine/Physiology, University of Eastern Finland, Kuopio Campus, Finland
- 171. Department of Medical Sciences, Uppsala University, Uppsala, Sweden
- 172. Queensland Institute of Medical Research, Locked Bag 2000, Royal Brisbane Hospital, Queensland 4029, Australia
- 173. Synlab Academy, Synlab Services GmbH, Gottlieb-Daimler-Straße 25, 68165 Mannheim, Germany
- 174. Tropical Metabolism Research Unit, Tropical Medicine Research Institute, University of the West Indies, Mona, Kingston 7, Jamaica
- 175. U872 Institut National de la Santé et de la Recherche Médicale, Centre de Recherche des Cordeliers, 75006 Paris, France
- 176. Department of Medicine, Kuopio University Hospital, Kuopio, Finland
- 177. Department of Neurology, General Central Hospital, Bolzano, Italy
- 178. Department of Neurology, University of Lübeck, Lübeck, Germany
- 179. Cardiovascular Health Research Unit, Departments of Medicine, Epidemiology, and Health Services, University of Washington, Seattle, WA, USA
- 180. Group Health Research Institute, Group Health Cooperative, Seattle, WA, USA
- 181. Department of Clinical Physiology and Nuclear Medicine, Kuopio University Hospital, Kuopio, Finland
- 182. Center for Non-Communicable Diseases, Karachi, Pakistan
- 183. Department of Medicine, University of Pennsylvania, USA
- 184. Unit of Chronic Disease Epidemiology and Prevention, National Institute for Health and Welfare, Helsinki, Finland
- 185. South Karelia Central Hospital, Lappeenranta, Finland
- 186. Paul Langerhans Institute Dresden, German Center for Diabetes Research (DZD), Dresden, Germany
- 187. Division of Endocrine and Metabolism, Department of Internal Medicine, Taichung Veterans General Hospital, Taichung, Taiwan
- 188. Geriatric Research and Education Clinical Center, Veterans Administration Medical Center, Baltimore, Maryland
- 189. Faculty of Medicine, University of Iceland, 101 Reykjavík, Iceland
- 190. Division of Population Health Sciences and Education, St George's, University of London, Cranmer Terrace, London SW17 0RE, United Kingdom
- 191. Department of Pharmacological Sciences, University of Milan, Monzino Cardiology Center, IRCCS, Milan, Italy
- 192. Centre for Vascular Prevention, Danube-University Krems, 3500 Krems, Austria
- 193. King Abdulaziz University, Faculty of Medicine, Jeddah 21589, Saudi Arabia
- 194. Red RECAVA Grupo RD06/0014/0015, Hospital Universitario La Paz, 28046
- 195. Institute of Public Health and Clinical Nutrition, University of Eastern Finland, Finland

- 196. Research Unit, Kuopio University Hospital, Kuopio, Finland
- 197. Department of Medicine, Lausanne University Hospital, Switzerland
- 198. Department of Endocrinology, University of Groningen, University Medical Center Groningen, The Netherlands
- 199. Department of Cardiovascular Epidemiology and Population Genetics, National Center for rdiovascular Investigation, Madrid, Spain
- 200. IMDEA-Alimentacion, Madrid, Spain
- 201. Nutrition and Genomics Laboratory, Jean Mayer-USDA Human Nutrition Research Center on Aging at Tufts University, Boston, MA, USA
- 202. Department of Nutrition, Harvard School of Public Health, Boston, MA, USA
- 203. Framingham Heart Study, Framingham, MA, USA
- 204. Center for Public Health Genomics, University of Virginia, Charlottesville, VA 22908, USA
- 205. Cardiovascular Research Center, Massachusetts General Hospital, Boston, Massachusetts 02114, USA
- 206. Department of Medicine, Harvard Medical School, Boston, Massachusetts 02115, USA

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### **Authors**

Georg B. Ehret<sup>1,2,3</sup>, Patricia B. Munroe<sup>4</sup>, Kenneth M. Rice<sup>5</sup>, Murielle Bochud<sup>2</sup>, Andrew D. Johnson<sup>6,7</sup>, Daniel I. Chasman<sup>8,9</sup>, Albert V. Smith<sup>10,11</sup>, Martin D. Tobin<sup>12</sup>, Germaine C. Verwoert<sup>13,14,15</sup>, Shih-Jen Hwang<sup>6,16,7</sup>, Vasyl Pihur<sup>1</sup>, Peter Vollenweider<sup>17</sup>, Paul F. O'Reilly<sup>18</sup>, Najaf Amin<sup>13</sup>, Jennifer L Bragg-Gresham<sup>19</sup>, Alexander Teumer<sup>20</sup>, Nicole L. Glazer<sup>21</sup>, Lenore Launer<sup>22</sup>, Jing Hua Zhao<sup>23</sup>, Yurii Aulchenko<sup>13</sup>, Simon Heath<sup>24</sup>, Siim Sõber<sup>25</sup>, Afshin Parsa<sup>26</sup>, Jian'an Luan<sup>23</sup>, Pankaj Arora<sup>27</sup>, Abbas Dehghan<sup>13,14,15</sup>, Feng Zhang<sup>28</sup>, Gavin Lucas<sup>29</sup>, Andrew A. Hicks<sup>30</sup>, Anne U. Jackson<sup>31</sup>, John F Peden<sup>32</sup>, Toshiko Tanaka<sup>33</sup>, Sarah H. Wild<sup>34</sup>, Igor Rudan<sup>35,36</sup>, Wilmar Igl<sup>37</sup>, Yuri Milaneschi<sup>33</sup>, Alex N. Parker<sup>38</sup>, Cristiano Fava<sup>39,40</sup>, John C. Chambers<sup>18,41</sup>, Ervin R. Fox<sup>42</sup>, Meena Kumari<sup>43</sup>, Min Jin Go<sup>44</sup>, Pim van der Harst<sup>45</sup>, Wen Hong Linda Kao<sup>46</sup>, Marketa Sjögren<sup>39</sup>, D. G. Vinay<sup>47</sup>, Myriam Alexander<sup>48</sup>, Yasuharu Tabara<sup>49</sup>, Sue Shaw-Hawkins<sup>4</sup>, Peter H. Whincup<sup>50</sup>, Yongmei Liu<sup>51</sup>, Gang Shi<sup>52</sup>, Johanna Kuusisto<sup>53</sup>, Bamidele Tayo<sup>54</sup>, Mark Seielstad<sup>55,56</sup>, Xueling Sim<sup>57</sup>, Khanh-Dung Hoang Nguyen<sup>1</sup>, Terho Lehtimäki<sup>58</sup>, Giuseppe Matullo<sup>59,60</sup>, Ying Wu<sup>61</sup>, Tom R. Gaunt<sup>62</sup>, N. Charlotte Onland-Moret<sup>63,64</sup>, Matthew N. Cooper<sup>65</sup>, Carl G.P. Platou<sup>66</sup>, Elin Org<sup>25</sup>, Rebecca Hardy<sup>67</sup>, Santosh Dahgam<sup>68</sup>, Jutta Palmen<sup>69</sup>, Veronique Vitart<sup>70</sup>, Peter S. Braund<sup>71,72</sup>, Tatiana Kuznetsova<sup>73</sup>, Cuno S.P.M. Uiterwaal<sup>63</sup>, Adebowale Adeyemo<sup>74</sup>, Walter Palmas<sup>75</sup>, Harry Campbell<sup>35</sup>, Barbara Ludwig<sup>76</sup>, Maciej Tomaszewski<sup>71,72</sup>, Ioanna Tzoulaki<sup>77,78</sup>, Nicholette D. Palmer<sup>79</sup>, CARDIoGRAM consortium<sup>80</sup>, CKDGen Consortium<sup>80</sup>, KidneyGen Consortium<sup>80</sup>, EchoGen consortium<sup>80</sup>, CHARGE-HF consortium<sup>80</sup>, Thor Aspelund<sup>10,11</sup>, Melissa Garcia<sup>22</sup>, Yen-Pei C. Chang<sup>26</sup>, Jeffrey R. O'Connell<sup>26</sup>, Nanette I. Steinle<sup>26</sup>, Diederick E. Grobbee<sup>63</sup>, Dan E. Arking<sup>1</sup>, Sharon L. Kardia<sup>81</sup>, Alanna C. Morrison<sup>82</sup>, Dena Hernandez<sup>83</sup>, Samer Najjar<sup>84,85</sup>, Wendy L. McArdle<sup>86</sup>, David Hadley<sup>50,87</sup>, Morris J. Brown<sup>88</sup>, John M. Connell<sup>89</sup>, Aroon D. Hingorani<sup>90</sup>, Ian N.M. Day<sup>62</sup>, Debbie A. Lawlor<sup>62</sup>, John P. Beilby<sup>91,92</sup>, Robert W. Lawrence<sup>65</sup>, Robert Clarke<sup>93</sup>, Rory Collins<sup>93</sup>, Jemma C Hopewell<sup>93</sup>, Halit Ongen<sup>32</sup>, Albert W. Dreisbach<sup>42</sup>, Yali Li<sup>94</sup>, J H. Young<sup>95</sup>, Joshua C. Bis<sup>21</sup>, Mika Kähönen<sup>96</sup>, Jorma Viikari<sup>97</sup>, Linda S. Adair<sup>98</sup>, Nanette R. Lee<sup>99</sup>, Ming-Huei Chen<sup>100</sup>, Matthias Olden<sup>101,102</sup>, Cristian Pattaro<sup>30</sup>, Judith A. Hoffman Bolton<sup>103</sup>, Anna Köttgen<sup>104,103</sup>, Sven Bergmann<sup>105,106</sup>, Vincent Mooser<sup>107</sup>, Nish Chaturvedi<sup>108</sup>, Timothy M. Frayling<sup>109</sup>, Muhammad Islam<sup>110</sup>, Tazeen H. Jafar<sup>110</sup>, Jeanette Erdmann<sup>111</sup>, Smita R. Kulkarni<sup>112</sup>, Stefan R. Bornstein<sup>76</sup>, Jürgen Grässler<sup>76</sup>, Leif Groop 113,114, Benjamin F. Voight 115, Johannes Kettunen 116,126, Philip Howard 117, Andrew Taylor 43, Simonetta Guarrera<sup>60</sup>, Fulvio Ricceri<sup>59,60</sup>, Valur Emilsson<sup>118</sup>, Andrew Plump<sup>118</sup>, Inês Barroso<sup>119,120</sup>, Kay-Tee Khaw<sup>48</sup>, Alan B. Weder<sup>121</sup>, Steven C. Hunt<sup>122</sup>, Yan V. Sun<sup>81</sup>, Richard N. Bergman<sup>123</sup>, Francis S. Collins<sup>124</sup>, Lori L.

Bonnycastle<sup>124</sup>, Laura J. Scott<sup>31</sup>, Heather M. Stringham<sup>31</sup>, Leena Peltonen<sup>119,125,126,127</sup>, Markus Perola<sup>125</sup>, Erkki Vartiainen<sup>125</sup>, Stefan-Martin Brand<sup>128,129</sup>, Jan A. Staessen<sup>73</sup>, Thomas J. Wang<sup>6,130</sup>, Paul R. Burton<sup>12,72</sup>, Maria Soler Artigas<sup>12</sup>, Yanbin Dong<sup>131</sup>, Harold Snieder<sup>132,131</sup>, Xiaoling Wang<sup>131</sup>, Haidong Zhu<sup>131</sup>, Kurt K. Lohman<sup>133</sup>, Megan E. Rudock<sup>51</sup>, Susan R Heckbert<sup>134,135</sup>, Nicholas L Smith<sup>134,136,135</sup>, Kerri L Wiggins<sup>137</sup>, Ayo Doumatey<sup>74</sup>, Daniel Shriner<sup>74</sup>, Gudrun Veldre<sup>25,138</sup>, Margus Viigimaa<sup>139,140</sup>, Sanjay Kinra<sup>141</sup>, Dorairajan Prabhakaran<sup>142</sup>, Vikal Tripathy<sup>142</sup>, Carl D. Langefeld<sup>79</sup>, Annika Rosengren<sup>143</sup>, Dag S. Thelle<sup>144</sup>, Anna Maria Corsi<sup>145</sup>, Andrew Singleton<sup>83</sup>, Terrence Forrester<sup>146</sup>, Gina Hilton<sup>1</sup>, Colin A. McKenzie<sup>146</sup>, Tunde Salako<sup>147</sup>, Naoharu Iwai<sup>148</sup>, Yoshikuni Kita<sup>149</sup>, Toshio Ogihara<sup>150</sup>, Takayoshi Ohkubo<sup>149,151</sup>, Tomonori Okamura<sup>148</sup>, Hirotsugu Ueshima<sup>152</sup>, Satoshi Umemura<sup>153</sup>, Susana Eyheramendy<sup>154</sup>, Thomas Meitinger<sup>155,156</sup>, H.-Erich Wichmann<sup>157,158,159</sup>, Yoon Shin Cho<sup>44</sup>, Hyung-Lae Kim<sup>44</sup>, Jong-Young Lee<sup>44</sup>, James Scott<sup>160</sup>, Joban S. Sehmi<sup>160,41</sup>, Weihua Zhang<sup>18</sup>, Bo Hedblad<sup>39</sup>, Peter Nilsson<sup>39</sup>, George Davey Smith<sup>62</sup>, Andrew Wong<sup>67</sup>, Narisu Narisu<sup>124</sup>, Alena Stančáková<sup>53</sup>, Leslie J. Raffel<sup>161</sup>, Jie Yao<sup>161</sup>, Sekar Kathiresan<sup>162,27</sup>, Chris O'Donnell<sup>163,27,9</sup>, Stephen M. Schwartz<sup>134</sup>, M. Arfan Ikram<sup>13,15</sup>, W. T. Longstreth Jr.<sup>164</sup>, Thomas H. Mosley<sup>165</sup>, Sudha Seshadri<sup>166</sup>, Nick R.G. Shrine<sup>12</sup>, Louise V. Wain<sup>12</sup>, Mario A. Morken<sup>124</sup>, Amy J. Swift<sup>124</sup>, Jaana Laitinen<sup>167</sup>, Inga Prokopenko<sup>51,168</sup>, Paavo Zitting<sup>169</sup>, Jackie A. Cooper<sup>69</sup>, Steve E. Humphries<sup>69</sup>, John Danesh<sup>48</sup>, Asif Rasheed<sup>170</sup>, Anuj Goel<sup>32</sup>, Anders Hamsten<sup>171</sup>, Hugh Watkins<sup>32</sup>, Stephan J.L. Bakker<sup>172</sup>, Wiek H. van Gilst<sup>45</sup>, Charles S. Janipalli<sup>47</sup>, K. Radha Mani<sup>47</sup>, Chittaranjan S. Yajnik<sup>112</sup>, Albert Hofman<sup>13</sup>, Francesco U.S. Mattace-Raso<sup>13,14</sup>, Ben A. Oostra<sup>173</sup>, Ayse Demirkan<sup>13</sup>, Aaron Isaacs<sup>13</sup>, Fernando Rivadeneira<sup>13,14</sup>, Edward G Lakatta<sup>174</sup>, Marco Orru<sup>175,176</sup>, Angelo Scuteri<sup>174</sup>, Mika Ala-Korpela<sup>177,178,179</sup>, Antti J Kangas<sup>177</sup>, Leo-Pekka Lyytikäinen<sup>58</sup>, Pasi Soininen<sup>177,178</sup>, Taru Tukiainen<sup>180,181,177</sup>, Peter Würtz<sup>177,18,180</sup>, Rick Twee-Hee Ong<sup>56,57,182</sup>, Marcus Dörr<sup>183</sup>, Heyo K. Kroemer<sup>184</sup>, Uwe Völker<sup>20</sup>, Henry Völzke<sup>185</sup>, Pilar Galan<sup>186</sup>, Serge Hercberg<sup>186</sup>, Mark Lathrop<sup>24</sup>, Diana Zelenika<sup>24</sup>, Panos Deloukas<sup>119</sup>, Massimo Mangino<sup>28</sup>, Tim D. Spector<sup>28</sup>, Guangju Zhai<sup>28</sup>, James F. Meschia<sup>187</sup>, Michael A. Nalls<sup>83</sup>, Pankaj Sharma<sup>188</sup>, Janos Terzic<sup>189</sup>, M. J. Kranthi Kumar<sup>47</sup>, Matthew Denniff<sup>71</sup>, Ewa Zukowska-Szczechowska<sup>190</sup>, Lynne E. Wagenknecht<sup>79</sup>, F. Gerald R. Fowkes<sup>191</sup>, Fadi J. Charchar<sup>192</sup>, Peter E.H. Schwarz<sup>193</sup>, Caroline Hayward<sup>70</sup>, Xiuqing Guo<sup>161</sup>, Charles Rotimi<sup>74</sup>, Michiel L. Bots<sup>63</sup>, Eva Brand<sup>194</sup>, Nilesh J. Samani<sup>71,72</sup>, Ozren Polasek<sup>195</sup>, Philippa J. Talmud<sup>69</sup>, Fredrik Nyberg<sup>68,196</sup>, Diana Kuh<sup>67</sup>, Maris Laan<sup>25</sup>, Kristian Hveem<sup>66</sup>, Lyle J. Palmer<sup>197,198</sup>, Yvonne T. van der Schouw<sup>63</sup>, Juan P. Casas<sup>199</sup>, Karen L. Mohlke<sup>61</sup>, Paolo Vineis<sup>200,60</sup>, Olli Raitakari<sup>201</sup>, Santhi K. Ganesh<sup>202</sup>, Tien Y. Wong<sup>203,204</sup>, E Shyong Tai<sup>205,57,206</sup>, Richard S. Cooper<sup>54</sup>, Markku Laakso<sup>53</sup>, Dabeeru C. Rao<sup>207</sup>, Tamara B. Harris<sup>22</sup>, Richard W. Morris<sup>208</sup>, Anna F. Dominiczak<sup>209</sup>, Mika Kivimaki<sup>210</sup>, Michael G. Marmot<sup>210</sup>, Tetsuro Miki<sup>49</sup>, Danish Saleheen<sup>170,48</sup>, Giriraj R. Chandak<sup>47</sup>, Josef Coresh<sup>211</sup>, Gerjan Navis<sup>212</sup>, Veikko Salomaa<sup>125</sup>, Bok-Ghee Han<sup>44</sup>, Xiaofeng Zhu<sup>94</sup>, Jaspal S. Kooner<sup>160,41</sup>, Olle Melander<sup>39</sup>, Paul M Ridker<sup>8,213,9</sup>, Stefania Bandinelli<sup>214</sup>, Ulf B. Gyllensten<sup>37</sup>, Alan F. Wright<sup>70</sup>, James F. Wilson<sup>34</sup>, Luigi Ferrucci<sup>33</sup>, Martin Farrall<sup>32</sup>, Jaakko Tuomilehto<sup>215,216,217,218</sup>, Peter P. Pramstaller<sup>30,219</sup>, Roberto Elosua<sup>29,220</sup>, Nicole Soranzo<sup>119,28</sup>, Eric J.G. Sijbrands<sup>13,14</sup>, David Altshuler<sup>221,115</sup>, Ruth J.F. Loos<sup>23</sup>, Alan R. Shuldiner<sup>26,222</sup>, Christian Gieger<sup>157</sup>, Pierre Meneton<sup>223</sup>, Andre G. Uitterlinden<sup>13,14,15</sup>, Nicholas J. Wareham<sup>23</sup>, Vilmundur Gudnason<sup>10,11</sup>, Jerome I. Rotter<sup>161</sup>, Rainer Rettig<sup>224</sup>, Manuela Uda<sup>175</sup>, David P. Strachan<sup>50</sup>, Jacqueline C.M. Witteman<sup>13,15</sup>, Anna-Liisa Hartikainen<sup>225</sup>, Jacques S. Beckmann<sup>105,226</sup>, Eric Boerwinkle<sup>227</sup>, Ramachandran S. Vasan<sup>6,228</sup>, Michael Boehnke<sup>31</sup>, Martin G. Larson<sup>6,229</sup>, Marjo-Riitta Järvelin<sup>18,230,231,232,233</sup>, Bruce M. Psaty<sup>21,135\*</sup>, Gonçalo R Abecasis<sup>19\*</sup>, Aravinda Chakravarti<sup>1</sup>, Paul Elliott<sup>18,233\*</sup>, Cornelia M. van Duijn<sup>13,234\*</sup>, Christopher Newton-Cheh<sup>27,115</sup>, Daniel Levy<sup>6,16,7</sup>, Mark J. Caulfield<sup>4</sup>, Toby Johnson<sup>4</sup>

- Center for Complex Disease Genomics, McKusick-Nathans Institute of Genetic Medicine, Johns Hopkins University School of Medicine, Baltimore, MD 21205, USA
- 2. Institute of Social and Preventive Medicine (IUMSP), Centre Hospitalier Universitaire Vaudois and University of Lausanne, Bugnon 17, 1005 Lausanne, Switzerland
- 3. Cardiology, Department of Specialties of Internal Medicine, Geneva University Hospital, Rue Gabrielle-Perret-Gentil 4, 1211 Geneva 14, Switzerland
- 4. Clinical Pharmacology and The Genome Centre, William Harvey Research Institute, Barts and The London School of Medicine and Dentistry, Queen Mary University of London, London EC1M 6BQ, UK
- 5. Department of Biostatistics, University of Washington, Seattle, WA, USA
- 6. Framingham Heart Study, Framingham, MA, USA
- 7. National Heart Lung, and Blood Institute, Bethesda, MD, USA
- 8. Division of Preventive Medicine, Brigham and Women's Hospital, 900 Commonwealth Avenue East, Boston MA 02215, USA
- 9. Harvard Medical School, Boston, MA, USA
- 10. Icelandic Heart Association, Kopavogur, Iceland
- 11. University of Iceland, Reykajvik, Iceland
- 12. Department of Health Sciences, University of Leicester, University Rd, Leicester LE1 7RH, UK
- 13. Department of Epidemiology, Erasmus Medical Center, PO Box 2040, 3000 CA, Rotterdam, The Netherlands
- 14. Department of Internal Medicine, Erasmus Medical Center, Rotterdam, The Netherlands
- 15. Netherlands Consortium for Healthy Aging (NCHA), Netherland Genome Initiative (NGI), The Netherlands
- 16. Center for Population Studies, National Heart Lung, and Blood Institute, Bethesda, MD, USA
- 17. Department of Internal Medicine, Centre Hospitalier Universitaire Vaudois, 1011 Lausanne, Switzerland
- 18. Department of Epidemiology and Biostatistics, School of Public Health, Imperial College London, Norfolk Place, London W2 1PG, UK
- 19. Center for Statistical Genetics, Department of Biostatistics, University of Michigan School of Public Health, Ann Arbor, MI 48103, USA
- 20. Interfaculty Institute for Genetics and Functional Genomics, Ernst-Moritz-Arndt-University Greifswald, 17487 Greifswald, Germany
- 21. Cardiovascular Health Research Unit, Departments of Medicine, Epidemiology and Health Services, University of Washington, Seattle, WA, USA
- 22. Laboratory of Epidemiology, Demography, Biometry, National Institute on Aging, National Institutes of Health, Bethesda, Maryland 20892, USA
- 23. MRC Epidemiology Unit, Institute of Metabolic Science, Cambridge CB2 0QQ, UK
- 24. Centre National de Génotypage, Commissariat à L'Energie Atomique, Institut de Génomique, Evry, France
- 25. Institute of Molecular and Cell Biology, University of Tartu, Riia 23, Tartu 51010, Estonia
- 26. University of Maryland School of Medicine, Baltimore, MD, USA, 21201, USA
- 27. Center for Human Genetic Research, Cardiovascular Research Center, Massachusetts General Hospital, Boston, Massachusetts, 02114, USA
- 28. Department of Twin Research & Genetic Epidemiology, King's College London, UK
- 29. Cardiovascular Epidemiology and Genetics, Institut Municipal d'Investigacio Medica, Barcelona Biomedical Research Park, 88 Doctor Aiguader, 08003 Barcelona, Spain
- 30. Institute of Genetic Medicine, European Academy Bozen/Bolzano (EURAC), Viale Druso 1, 39100 Bolzano, Italy Affiliated Institute of the University of Lübeck, Germany
- 31. Department of Biostatistics, Center for Statistical Genetics, University of Michigan, Ann Arbor, Michigan, 48109, USA
- 32. Department of Cardiovascular Medicine, The Wellcome Trust Centre for Human Genetics, University of Oxford, OX3 7BN, UK

- 33. Clinical Research Branch, National Institute on Aging, Baltimore MD 21250, USA
- 34. Centre for Population Health Sciences, University of Edinburgh, EH89AG, UK
- 35. Centre for Population Health Sciences and Institute of Genetics and Molecular Medicine, College of Medicine and Vet Medicine, University of Edinburgh, EH8 9AG, UK
- 36. Croatian Centre for Global Health, University of Split, Croatia
- 37. Department of Genetics and Pathology, Rudbeck Laboratory, Uppsala University, SE-751 85 Uppsala, Sweden
- 38. Amgen, 1 Kendall Square, Building 100, Cambridge, MA 02139, USA
- 39. Department of Clinical Sciences, Lund University, Malmö, Sweden
- 40. Department of Medicine, University of Verona, Italy
- 41. Ealing Hospital, London, UB1 3HJ, UK
- 42. Department of Medicine, University of Mississippi Medical Center, USA
- 43. Genetic Epidemiology Group, Epidemiology and Public Health, UCL, London, WC1E 6BT, UK
- 44. Center for Genome Science, National Institute of Health, Seoul, Korea
- 45. Department of Cardiology, University Medical Center Groningen, University of Groningen, The Netherlands
- 46. Departments of Epidemiology and Medicine, Johns Hopkins University, Baltimore MD, USA
- 47. Centre for Cellular and Molecular Biology (CCMB), Council of Scientific and Industrial Research (CSIR), Uppal Road, Hyderabad 500 007, India
- 48. Department of Public Health and Primary Care, University of Cambridge, CB1 8RN, UK
- 49. Department of Basic Medical Research and Education, and Department of Geriatric Medicine, Ehime University Graduate School of Medicine, Toon, 791-0295, Japan
- 50. Division of Community Health Sciences, St George's University of London, London, SW17 0RE, UK
- 51. Epidemiology & Prevention, Division of Public Health Sciences, Wake Forest University School of Medicine, Winston-Salem, NC 27157, USA
- 52. Division of Biostatistics and Department of Genetics, School of Medicine, Washington University in St. Louis, Saint Louis, Missouri 63110, USA
- 53. Department of Medicine, University of Eastern Finland and Kuopio University Hospital, 70210 Kuopio, Finland
- 54. Department of Preventive Medicine and Epidemiology, Loyola University Medical School, Maywood, IL, USA
- 55. Department of Laboratory Medicine & Institute of Human Genetics, University of California San Francisco, 513 Parnassus Ave. San Francisco CA 94143, USA
- 56. Genome Institute of Singapore, Agency for Science, Technology and Research, Singapore, 138672, Singapore
- 57. Centre for Molecular Epidemiology, Yong Loo Lin School of Medicine, National University of Singapore, Singapore, 117597, Singapore
- 58. Department of Clinical Chemistry, University of Tampere and Tampere University Hospital, Tampere, 33521, Finland
- 59. Department of Genetics, Biology and Biochemistry, University of Torino, Via Santena 19, 10126, Torino, Italy
- 60. Human Genetics Foundation (HUGEF), Via Nizza 52, 10126, Torino, Italy
- 61. Department of Genetics, University of North Carolina, Chapel Hill, NC, 27599, USA
- 62. MRC Centre for Causal Analyses in Translational Epidemiology, School of Social & Community Medicine, University of Bristol, Bristol BS8 2BN, UK
- 63. Julius Center for Health Sciences and Primary Care, University Medical Center Utrecht, Heidelberglaan 100, 3508 GA Utrecht, The Netherlands
- 64. Complex Genetics Section, Department of Medical Genetics DBG, University Medical Center Utrecht, 3508 GA Utrecht, The Netherlands
- 65. Centre for Genetic Epidemiology and Biostatistics, University of Western Australia, Crawley, WA, Australia
- 66. HUNT Research Centre, Department of Public Health and General Practice, Norwegian University of Science and Technology, 7600 Levanger, Norway
- 67. MRC Unit for Lifelong Health & Ageing, London, WC1B 5JU, UK
- 68. Occupational and Environmental Medicine, Department of Public Health and Community Medicine, Institute of Medicine, Sahlgrenska Academy, University of Gothenburg, 40530 Gothenburg, Sweden
- 69. Centre for Cardiovascular Genetics, University College London, London WC1E 6JF, UK
- 70. MRC Human Genetics Unit and Institute of Genetics and Molecular Medicine, Edinburgh, EH2, UK
- 71. Department of Cardiovascular Sciences, University of Leicester, Glenfield Hospital, Leicester, LE3 9QP, UK
- 72. Leicester NIHR Biomedical Research Unit in Cardiovascular Disease, Glenfield Hospital, Leicester, LE3 9QP, UK
- 73. Studies Coordinating Centre, Division of Hypertension and Cardiac Rehabilitation, Department of Cardiovascular Diseases, University of Leuven, Campus Sint Rafaël, Kapucijnenvoer 35, Block D, Box 7001, 3000 Leuven, Belgium

- 74. Center for Research on Genomics and Global Health, National Human Genome Research Institute, Bethesda, MD 20892, USA
- 75. Columbia University, NY, USA
- 76. Department of Medicine III, Medical Faculty Carl Gustav Carus at the Technical University of Dresden, 01307 Dresden, Germany
- 77. Epidemiology and Biostatistics, School of Public Health, Imperial College, London, W2 1PG, UK
- 78. Clinical and Molecular Epidemiology Unit, Department of Hygiene and Epidemiology, University of Ioannina School of Medicine, Ioannina, Greece
- 79. Wake Forest University Health Sciences, Winston-Salem, NC 27157, USA
- 80. A list of consortium members is supplied in the Supplementary Materials
- 81. Department of Epidemiology, School of Public Health, University of Michigan, Ann Arbor, MI 48109, USA
- 82. Division of Epidemiology, Human Genetics and Environmental Sciences, School of Public Health, University of Texas at Houston Health Science Center, 12 Herman Pressler, Suite 453E, Houston, TX 77030, USA
- 83. Laboratory of Neurogenetics, National Institute on Aging, Bethesda, MD 20892, USA
- 84. Laboratory of Cardiovascular Science, Intramural Research Program, National Institute on Aging, NIH, Baltimore, Maryland, USA
- 85. Washington Hospital Center, Division of Cardiology, Washington DC, USA
- 86. ALSPAC Laboratory, University of Bristol, Bristol, BS8 2BN, UK
- 87. Pediatric Epidemiology Center, University of South Florida, Tampa, FL, USA
- 88. Clinical Pharmacology Unit, University of Cambridge, Addenbrookes Hospital, Hills Road, Cambridge CB2 2QQ, UK
- 89. University of Dundee, Ninewells Hospital & Medical School, Dundee, DD1 9SY, UK
- 90. Genetic Epidemiology Group, Department of Epidemiology and Public Health, UCL, London WC1E 6BT, UK
- 91. Pathology and Laboratory Medicine, University of Western Australia, Crawley, WA, Australia
- 92. Molecular Genetics, PathWest Laboratory Medicine, Nedlands, WA, Australia
- 93. Clinical Trial Service Unit and Epidemiological Studies Unit, University of Oxford, Oxford, OX3 7LF, UK
- 94. Department of Epidemiology and Biostatistics, Case Western Reserve University, 2103 Cornell Road, Cleveland, OH 44106, USA
- 95. Department of Medicine, Johns Hopkins University, Baltimore, USA
- 96. Department of Clinical Physiology, University of Tampere and Tampere University Hospital, Tampere, 33521, Finland
- 97. Department of Medicine, University of Turku and Turku University Hospital, Turku, 20521, Finland
- 98. Department of Nutrition, University of North Carolina, Chapel Hill, NC, 27599, USA
- 99. Office of Population Studies Foundation, University of San Carlos, Talamban, Cebu City 6000, Philippines
- 100. Department of Neurology and Framingham Heart Study, Boston University School of Medicine, Boston, MA, 02118, USA
- 101. Department of Internal Medicine II, University Medical Center Regensburg, 93053 Regensburg, Germany
- 102. Department of Epidemiology and Preventive Medicine, University Medical Center Regensburg, 93053 Regensburg, Germany
- 103. Department of Epidemiology, Johns Hopkins University, Baltimore MD, USA
- 104. Renal Division, University Hospital Freiburg, Germany
- 105. Département de Génétique Médicale, Université de Lausanne, 1015 Lausanne, Switzerland
- 106. Swiss Institute of Bioinformatics, 1015 Lausanne, Switzerland
- 107. Division of Genetics, GlaxoSmithKline, Philadelphia, Pennsylvania 19101, USA
- 108. International Centre for Circulatory Health, National Heart & Lung Institute, Imperial College, London, UK
- 109. Genetics of Complex Traits, Peninsula Medical School, University of Exeter, UK
- 110. Department of Community Health Sciences & Department of Medicine, Aga Khan University, Karachi, Pakistan
- 111. Medizinische Klinik II, Universität zu Lübeck, Lübeck, Germany
- 112. Diabetes Unit, KEM Hospital and Research Centre, Rasta Peth, Pune-411011, Maharashtra, India
- 113. Department of Clinical Sciences, Diabetes and Endocrinology Research Unit, University Hospital, Malmö, Sweden
- 114. Lund University, Malmö 20502, Sweden
- 115. Program in Medical and Population Genetics, Broad Institute of Harvard and MIT, Cambridge, Massachusetts, 02139, USA
- 116. Department of Chronic Disease Prevention, National Institute for Health and Welfare, FIN-00251 Helsinki, Finland
- 117. William Harvey Research Institute, Barts and The London School of Medicine and Dentistry, Queen Mary University of London, London EC1M 6BQ. UK
- 118. Merck Research Laboratory, 126 East Lincoln Avenue, Rahway, NJ 07065, USA

- 119. Wellcome Trust Sanger Institute, Hinxton, CB10 1SA, UK
- 120. University of Cambridge Metabolic Research Labs, Institute of Metabolic Science Addenbrooke's Hospital, CB2 OQQ, Cambridge, UK
- 121. Division of Cardiovascular Medicine, Department of Internal Medicine, University of Michigan Medical School, Ann Arbor, MI, USA
- 122. Cardiovascular Genetics, University of Utah School of Medicine, Salt Lake City, UT, USA
- 123. Department of Physiology and Biophysics, Keck School of Medicine, University of Southern California, Los Angeles, California 90033, USA
- 124. National Human Genome Research Institute, National Institutes of Health, Bethesda, Maryland 20892, USA
- 125. National Institute for Health and Welfare, 00271 Helsinki, Finland
- 126. FIMM, Institute for Molecular Medicine, Finland, Biomedicum, P.O. Box 104, 00251 Helsinki, Finland
- 127. Broad Institute, Cambridge, Massachusetts 02142, USA
- 128. Leibniz-Institute for Arteriosclerosis Research, Department of Molecular Genetics of Cardiovascular Disease, University of Münster, Münster, Germany
- 129. Medical Faculty of the Westfalian Wilhelms University Muenster, Department of Molecular Genetics of Cardiovascular Disease, University of Muenster, Muenster, Germany
- 130. Division of Cardiology, Massachusetts General Hospital, Boston, MA, USA
- 131. Georgia Prevention Institute, Department of Pediatrics, Medical College of Georgia, Augusta, GA, USA
- 132. Unit of Genetic Epidemiology and Bioinformatics, Department of Epidemiology, University Medical Center Groningen, University of Groningen, Groningen, The Netherlands
- 133. Department of Biostatical Sciences, Division of Public Health Sciences, Wake Forest University School of Medicine, Winston-Salem, NC 27157, USA
- 134. Department of Epidemiology, University of Washington, Seattle, WA, 98195, USA
- 135. Group Health Research Institute, Group Health Cooperative, Seattle, WA, USA
- 136. Seattle Epidemiologic Research and Information Center, Veterans Health Administration Office of Research & Development, Seattle, WA 98108, USA
- 137. Department of Medicine, University of Washington, 98195, USA
- 138. Department of Cardiology, University of Tartu, L. Puusepa 8, 51014 Tartu, Estonia
- 139. Tallinn University of Technology, Institute of Biomedical Engineering, Ehitajate tee 5, 19086 Tallinn, Estonia
- 140. Centre of Cardiology, North Estonia Medical Centre, Sütiste tee 19, 13419 Tallinn, Estonia
- 141. Division of Non-communicable disease Epidemiology, The London School of Hygiene and Tropical Medicine London, Keppel Street, London WC1E 7HT. UK
- 142. South Asia Network for Chronic Disease, Public Health Foundation of India, C-1/52, SDA, New Delhi 100016, India
- 143. Department of Emergency and Cardiovascular Medicine, Institute of Medicine, Sahlgrenska Academy, University of Gothenburg, 41685 Gothenburg, Sweden
- 144. Department of Biostatistics, Institute of Basic Medical Sciences, University of Oslo, 0317 Oslo, Norway
- 145. Tuscany Regional Health Agency, Florence, Italy
- 146. Tropical Medicine Research Institute, University of the West Indies, Mona, Kingston, Jamaica
- 147. University of Ibadan, Ibadan, Nigeria
- 148. Department of Genomic Medicine, and Department of Preventive Cardiology, National Cerebral and Cardiovascular Research Center, Suita, 565-8565, Japan
- 149. Department of Health Science, Shiga University of Medical Science, Otsu, 520-2192, Japan
- 150. Department of Geriatric Medicine, Osaka University Graduate School of Medicine, Suita, 565-0871, Japan
- 151. Tohoku University Graduate School of Pharmaceutical Sciences and Medicine, Sendai, 980-8578, Japan
- 152. Lifestyle-related Disease Prevention Center, Shiga University of Medical Science, Otsu, 520-2192, Japan
- 153. Department of Medical Science and Cardiorenal Medicine, Yokohama City University School of Medicine, Yokohama, 236-0004, Japan
- 154. Department of Statistics, Pontificia Universidad Catolica de Chile, Vicuña Mackena 4860, Santiago, Chile
- 155. Institute of Human Genetics, Helmholtz Zentrum Munich, German Research Centre for Environmental Health, 85764 Neuherberg, Germany
- 156. Institute of Human Genetics, Klinikum rechts der Isar, Technical University of Munich, 81675 Munich, Germany
- 157. Institute of Epidemiology, Helmholtz Zentrum Munich, German Research Centre for Environmental Health, 85764 Neuherberg, Germany

- 158. Chair of Epidemiology, Institute of Medical Informatics, Biometry and Epidemiology, Ludwig-Maximilians-Universität, 81377 Munich, Germany
- 159. Klinikum Grosshadern, 81377 Munich, Germany
- 160. National Heart and Lung Institute, Imperial College London, London, UK, W12 0HS, UK
- 161. Medical Genetics Institute, Cedars-Sinai Medical Center, Los Angeles, CA, USA
- 162. Medical Population Genetics, Broad Institute of Harvard and MIT, 5 Cambridge Center, Cambridge MA 02142, USA
- 163. National Heart, Lung and Blood Institute and its Framingham Heart Study, 73 Mount Wayte Ave., Suite #2, Framingham, MA 01702, USA
- 164. Department of Neurology and Medicine, University of Washington, Seattle, USA
- 165. Department of Medicine (Geriatrics), University of Mississippi Medical Center, Jackson, MS, USA
- 166. Department of Neurology, Boston University School of Medicine, USA
- 167. Finnish Institute of Occupational Health, Finnish Institute of Occupational Health, Appistie 1, 90220 Oulu, Finland
- 168. Wellcome Trust Centre for Human Genetics, University of Oxford, UK
- 169. Lapland Central Hospital, Department of Physiatrics, Box 8041, 96101 Rovaniemi, Finland
- 170. Center for Non-Communicable Diseases Karachi, Pakistan
- 171. Atherosclerosis Research Unit, Department of Medicine, Karolinska Institute, Stockholm, Sweden
- 172. Department of Internal Medicine, University Medical Center Groningen, University of Groningen, The Netherlands
- 173. Department of Medical Genetics, Erasmus Medical Center, Rotterdam, The Netherlands
- 174. Gerontology Research Center, National Institute on Aging, Baltimore, MD 21224, USA
- 175. Istituto di Neurogenetica e Neurofarmacologia, Consiglio Nazionale delle Ricerche, Cittadella Universitaria di Monserrato, Monserrato, Cagliari, Italy
- 176. Unita` Operativa Semplice Cardiologia, Divisione di Medicina, Presidio Ospedaliero Santa Barbara, Iglesias, Italy
- 177. Computational Medicine Research Group, Institute of Clinical Medicine, University of Oulu and Biocenter Oulu, 90014 University of Oulu, Oulu, Finland
- 178. NMR Metabonomics Laboratory, Department of Biosciences, University of Eastern Finland, 70211 Kuopio, Finland
- 179. Department of Internal Medicine and Biocenter Oulu, Clinical Research Center, 90014 University of Oulu, Oulu, Finland
- 180. Institute for Molecular Medicine Finland FIMM, 00014 University of Helsinki, Helsinki, Finland
- 181. Department of Biomedical Engineering and Computational Science, School of Science and Technology, Aalto University, 00076 Aalto, Espoo, Finland
- 182. NUS Graduate School for Integrative Sciences & Engineering (NGS) Centre for Life Sciences (CeLS), Singapore, 117456, Singapore
- 183. Department of Internal Medicine B, Ernst-Moritz-Arndt-University Greifswald, 17487 Greifswald, Germany
- 184. Institute of Pharmacology, Ernst-Moritz-Arndt-University Greifswald, 17487 Greifswald, Germany
- 185. Institute for Community Medicine, Ernst-Moritz-Arndt-University Greifswald, 17487 Greifswald, Germany
- 186. U557 Institut National de la Santé et de la Recherche Médicale, U1125 Institut National de la Recherche Agronomique, Université Paris 13, Bobigny, France
- 187. Department of Neurology, Mayo Clinic, Jacksonville, FL, USA
- 188. Imperial College Cerebrovascular Unit (ICCRU), Imperial College, London, W6 8RF, UK
- 189. Faculty of Medicine, University of Split, Croatia
- 190. Department of Internal Medicine, Diabetology, and Nephrology, Medical University of Silesia, 41-800, Zabrze, Poland
- 191. Public Health Sciences section, Division of Community Health Sciences, University of Edinburgh, Medical School, Teviot Place, Edinburgh, EH8 9AG, UK
- 192. School of Science and Engineering, University of Ballarat, 3353 Ballarat, Australia
- 193. Prevention and Care of Diabetes, Department of Medicine III, Medical Faculty Carl Gustav Carus at the Technical University of Dresden, 01307 Dresden, Germany
- 194. University Hospital Münster, Internal Medicine D, Münster, Germany
- 195. Department of Medical Statistics, Epidemiology and Medical Informatics, Andrija Stampar School of Public Health, University of Zagreb, Croatia
- 196. AstraZeneca R&D, 431 83 Mölndal, Sweden
- 197. Genetic Epidemiology & Biostatistics Platform, Ontario Institute for Cancer Research, Toronto
- 198. Samuel Lunenfeld Institute for Medical Research, University of Toronto, Canada
- 199. Faculty of Epidemiology and Population Health, London School of Hygiene and Tropical Medicine, UK

- 200. Department of Epidemiology and Public Health, Imperial College, Norfolk Place London W2 1PG, UK
- 201. Research Centre of Applied and Preventive Cardiovascular Medicine, University of Turku and the Department of Clinical Physiology, Turku University Hospital, Turku, 20521, Finland
- 202. Department of Internal Medicine, Division of Cardiovascular Medicine, University of Michigan Medical Center, Ann Arbor, Michigan, USA
- 203. Singapore Eye Research Institute, Singapore, 168751, Singapore
- 204. Department of Ophthalmology, National University of Singapore, Singapore, 119074, Singapore
- 205. Department of Medicine, Yong Loo Lin School of Medicine, National University of Singapore, Singapore, 119074, Singapore
- 206. Duke-National University of Singapore Graduate Medical School, Singapore, 169857, Singapore
- 207. Division of Biostatistics, Washington University School of Medicine, Saint Louis, MO, 63110, USA
- 208. Department of Primary Care & Population Health, UCL, London, UK, NW3 2PF, UK
- 209. BHF Glasgow Cardiovascular Research Centre, University of Glasgow, 126 University Place, Glasgow, G12 8TA, UK
- 210. Epidemiology Public Health, UCL, London, UK, WC1E 6BT, UK
- 211. Departments of Epidemiology, Biostatistics, and Medicine, Johns Hopkins University, Baltimore MD, USA
- 212. Division of Nephrology, Department of Internal Medicine, University Medical Center Groningen, University of Groningen, The Netherlands
- 213. Division of Cardiology, Brigham and Women's Hospital, 900 Commonwealth Avenue East, Boston MA 02215, USA
- 214. Geriatric Rehabilitation Unit, Azienda Sanitaria Firenze (ASF), Florence, Italy
- 215. National Institute for Health and Welfare, Diabetes Prevention Unit, 00271 Helsinki, Finland
- 216. Hjelt Institute, Department of Public Health, University of Helsinki, 00014 Helsinki, Finland
- 217. South Ostrobothnia Central Hospital, 60220 Seinäjoki, Finland
- 218. Red RECAVA Grupo RD06/0014/0015, Hospital Universitario La Paz, 28046 Madrid, Spain
- 219. Department of Neurology, General Central Hospital, 39100 Bolzano, Italy
- 220. CIBER Epidemiología y Salud Pública, 08003 Barcelona
- 221. Department of Medicine and Department of Genetics, Harvard Medical School, Boston, Massachusetts 02115, USA
- 222. Geriatric Research and Education Clinical Center, Veterans Administration Medical Center, Baltimore, MD, USA
- 223. U872 Institut National de la Santé et de la Recherche Médicale, Centre de Recherche des Cordeliers, Paris, France
- 224. Institute of Physiology, Ernst-Moritz-Arndt-University Greifswald, 17487 Greifswald, Germany
- 225. Institute of Clinical Medicine/Obstetrics and Gynecology, University of Oulu, Finland
- 226. Service of Medical Genetics, Centre Hospitalier Universitaire Vaudois, 1011 Lausanne, Switzerland
- 227. Human Genetics Center, 1200 Hermann Pressler, Suite E447 Houston, TX 77030, USA
- 228. Division of Epidemiology and Prevention, Boston University School of Medicine, Boston, MA, USA
- 229. Department of Mathematics, Boston University, Boston, MA, USA
- 230. Institute of Health Sciences, University of Oulu, BOX 5000, 90014 University of Oulu, Finland
- 231. Biocenter Oulu, University of Oulu, BOX 5000, 90014 University of Oulu, Finland
- 232. National Institute for Health and Welfare, Box 310, 90101 Oulu, Finland
- 233. MRC-HPA Centre for Environment and Health, School of Public Health, Imperial College London, Norfolk Place, London W2 1PG, UK
- 234. Centre of Medical Systems Biology (CMSB 1-2), NGI Erasmus Medical Center, Rotterdam, The Netherlands

### The International ENDOGENE Consortium

Carl A Anderson<sup>1,2</sup>, Scott D Gordon<sup>3</sup>, Qun Guo<sup>4</sup>, Anjali K Henders<sup>3</sup>, Ann Lambert<sup>5</sup>, Sang Hong Lee<sup>6</sup>, Peter Kraft<sup>7</sup>, Stephen H Kennedy<sup>5</sup>, Stuart Macgregor<sup>3</sup>, Nicholas G Martin<sup>3</sup>, Stacey A Missmer<sup>4</sup>, Grant W Montgomery<sup>3</sup>, Andrew P Morris<sup>1</sup>, Dale R Nyholt<sup>3</sup>, Jodie N Painter<sup>3</sup>, Fenella Roseman<sup>5</sup>, Susan A Treloar<sup>8</sup>, Peter M Visscher<sup>9</sup>, Leanne Wallace<sup>3</sup>, Krina T Zondervan<sup>1,5</sup>.

- 1. Wellcome Trust Centre for Human Genetics, University of Oxford, Oxford, UK.
- 2. Wellcome Trust Sanger Institute, Wellcome Trust Genome Campus, Hinxton, UK.
- 3. Queensland Institute of Medical Research, Herston, Queensland, Australia.
- 4. Brigham and Women's Hospital and Harvard Medical School, Boston, Massachusetts, USA.
- 5. Nuffield Department of Obstetrics and Gynaecology, University of Oxford, John Radcliffe Hospital, Oxford, UK.
- 6. Queensland Brain Institute, The University of Queensland, Brisbane, QLD 4072, Australia.
- 7. Harvard School of Public Health, Boston, Massachusetts, USA.
- 8. Centre for Military and Veterans' Health, The University of Queensland, Mayne Medical School, Queensland, Australia.
- 9. The University of Queensland Diamantina Institute, Princess Alexandra Hospital, Brisbane, QLD 4102, Australia.

### The LifeLines Cohort Study

Behrooz Z Alizadeh <sup>1</sup>, Rudolf A de Boer <sup>2</sup>, H Marike Boezen<sup>1</sup>, Marcel Bruinenberg <sup>3</sup>, Lude Franke<sup>4</sup>, Pim van der Harst <sup>2</sup>, Hans L Hillege <sup>1,2</sup>, Melanie M van der Klauw <sup>5</sup>, Gerjan Navis<sup>6</sup>, Johan Ormel <sup>7</sup>, Dirkje S Postma<sup>8</sup>, Judith GM Rosmalen<sup>7</sup>, Joris P Slaets<sup>9</sup>, Harold Snieder<sup>1</sup>, Ronald P Stolk<sup>1</sup>, Bruce HR Wolffenbuttel<sup>5</sup>, Cisca Wijmenga<sup>4</sup>

- 1. Department of Epidemiology, University of Groningen, University Medical Center Groningen, The Netherlands
- 2. Department of Cardiology, University of Groningen, University Medical Center Groningen, The Netherlands
- 3. LifeLines Cohort Study, University of Groningen, University Medical Center Groningen, The Netherlands
- 4. Department of Genetics, University of Groningen, University Medical Center Groningen, The Netherlands
- 5. Department of Endocrinology, University of Groningen, University Medical Center Groningen, The Netherlands
- 6. Department of Internal Medicine, Division of Nephrology, University of Groningen, University Medical Center Groningen, The Netherlands
- 7. Interdisciplinary Center of Psychopathology and Emotion Regulation ICPE . , Department of Psychiatry, University of Groningen, University Medical Center Groningen, The Netherlands
- 8. Department of Pulmonology, University of Groningen, University Medical Center Groningen, The Netherlands
- 9. University Center for Geriatric Medicine, University of Groningen, University Medical Center Groningen, The Netherlands

# The MAGIC Investigators

Robert A Scott, Vasiliki Lagou, 2,3 Ryan P Welch, 4,6 Eleanor Wheeler, May E Montasser, Jian'an Luan, 1 Reedik Mägi, 2,9 Rona J Strawbridge, 10,11 Emil Rehnberg, 12 Stefan Gustafsson, 12 Stavroula Kanoni, Laura J Rasmussen-Torvik, <sup>13</sup> Loïc Yengo, <sup>14,15</sup> Cecile Lecoeur, <sup>14,15</sup> Dmitry Shungin, <sup>16,18</sup> Serena Sanna, <sup>19</sup> Carlo Sidore, 5,6,19,20 Paul C D Johnson, 1 J Wouter Jukema, 22,23 Toby Johnson, 44,25 Anubha Mahajan, Niek Verweij, 16 Gudmar Thorleifsson,<sup>27</sup> Jouke-Jan Hottenga,<sup>28</sup> Sonia Shah,<sup>29</sup> Albert V Smith,<sup>30,31</sup> Bengt Sennblad,<sup>10</sup> Christian Gieger, <sup>32</sup> Perttu Salo, <sup>33</sup> Markus Perola, <sup>9,33,34</sup> Nicholas J Timpson, <sup>35</sup> David M Evans, <sup>35</sup> Beate St Pourcain, <sup>36</sup> Ying Wu,<sup>37</sup> Jeanette S Andrews,<sup>38</sup> Jennie Hui,<sup>39,40,41,42</sup> Lawrence F Bielak,<sup>43</sup> Wei Zhao,<sup>43</sup> Momoko Horikoshi,<sup>2,3</sup> Pau Navarro, 44 Aaron Isaacs, 45,46 Jeffrey R O'Connell, Kathleen Stirrups, Veronique Vitart, 44 Caroline Hayward, 44 Tönu Esko, 9,47 Evelin Mihailov, 47 Ross M Fraser, 48 Tove Fall, 12 Benjamin F Voight, 49,50 Soumya Raychaudhuri,<sup>51</sup> Han Chen,<sup>52</sup> Cecilia M Lindgren,<sup>2</sup> Andrew P Morris,<sup>2</sup> Nigel W Rayner,<sup>2,3</sup> Neil Robertson,<sup>2,3</sup> Denis Rybin,<sup>53</sup> Ching-Ti Liu,<sup>52</sup> Jacques S Beckmann,<sup>54,55</sup> Sara M Willems,<sup>46</sup> Peter S Chines,<sup>56</sup> Anne U Jackson,<sup>5,6</sup> Hyun Min Kang,<sup>5,6</sup> Heather M Stringham,<sup>5,6</sup> Kijoung Song,<sup>57</sup> Toshiko Tanaka,<sup>58</sup> John F Peden,<sup>2,59</sup> Anuj Goel, 2,60 Andrew A Hicks, 61 Ping An, 62 Martina Müller-Nurasyid, 32,63,64 Anders Franco-Cereceda, 65 Lasse Folkersen, 10,11 Letizia Marullo, 2,66 Hanneke Jansen, 67 Albertine J Oldehinkel, 68 Marcel Bruinenberg, 69 James S Pankow, <sup>70</sup> Kari E North, <sup>71,72</sup> Nita G Forouhi, <sup>1</sup> Ruth J F Loos, <sup>1</sup> Sarah Edkins, <sup>7</sup> Tibor V Varga, <sup>16</sup> Göran Hallmans, 73 Heikki Oksa, 74 Mulas Antonella, 19 Ramaiah Nagaraja, 75 Stella Trompet, 22,23 Ian Ford, 21 Stephan J L Bakker, <sup>76</sup> Augustine Kong, <sup>27</sup> Meena Kumari, <sup>77</sup> Bruna Gigante, <sup>78</sup> Christian Herder, <sup>79</sup> Patricia B Munroe, <sup>24,25</sup> Mark Caulfield, 24,25 Jula Antti, 33 Massimo Mangino, 80 Kerrin Small, 80 Iva Miljkovic, 81 Yongmei Liu, 82 Mustafa Atalay, 83 Wieland Kiess, 84,85 Alan L James, 39,86,87 Fernando Rivadeneira, 45,88,90 Andre G Uitterlinden, 45,88,89,90 Colin N A Palmer, 91 Alex S F Doney, 91 Gonneke Willemsen, 28 Johannes H Smit, 92 Susan Campbell, 44 Ozren Polasek, 93 Lori L Bonnycastle, <sup>56</sup> Serge Hercberg, <sup>94</sup> Maria Dimitriou, <sup>95</sup> Jennifer L Bolton, <sup>96</sup> Gerard R Fowkes, <sup>96</sup> Peter Kovacs, <sup>97</sup> Jaana Lindström, <sup>98</sup> Tatijana Zemunik, <sup>93</sup> Stefania Bandinelli, <sup>99</sup> Sarah H Wild, <sup>48</sup> Hanneke V Basart, <sup>100</sup> Wolfgang Rathmann, 101 Harald Grallert, 102 DIAGRAM consortium, 103 Winfried Maerz, 104,105 Marcus E Kleber, 105,106 Bernhard O Boehm, 107 Annette Peters, 108 Peter P Pramstaller, 61,109,110 Michael A Province, 62 Ingrid B Borecki, 62 Nicholas D Hastie, 44 Igor Rudan, 48 Harry Campbell, 48 Hugh Watkins, 2,60 Martin Farrall, 2,60 Michael Stumvoll, 84,111 Luigi Ferrucci, 58 Dawn M Waterworth, 57 Richard N Bergman, 112 Francis S Collins, 56 Jaakko Tuomilehto, 113,114,115,116 Richard M Watanabe, 117,118 Eco J C de Geus, 28 Brenda W Penninx, 92 Albert Hofman, 90 Ben A Oostra, 45,46,89 Bruce M Psaty, 119,120,121,122 Peter Vollenweider, 123 James F Wilson, 48 Alan F Wright, 44 G Kees Hovingh, 100 Andres Metspalu, 9,47 Matti Uusitupa, 124,125 Patrik K E Magnusson, 12 Kirsten O Kyvik, 126,127 Jaakko Kaprio, 34,128,129 Jackie F Price, 96 George V Dedoussis, 95 Panos Deloukas, 7 Pierre Meneton, 130 Lars Lind, 131 Michael Boehnke, 5,6 Alan R Shuldiner, 8,132 Cornelia M van Duijn, 45,46,89,90 Andrew D Morris, 91 Anke Toenjes, 84,111 Patricia A Peyser, 43 John P Beilby, 39,41,42 Antje Körner, 84,85 Johanna Kuusisto, 133 Markku Laakso, 133 Stefan R Bornstein, 134 Peter E H Schwarz, 134 Timo A Lakka, 83,135 Rainer Rauramaa, 135,136 Linda S

Adair, <sup>137</sup> George Davey Smith, <sup>35</sup> Tim D Spector, <sup>80</sup> Thomas Illig, <sup>102,138</sup> Ulf de Faire, <sup>78</sup> Anders Hamsten, <sup>10,11,139</sup> Vilmundur Gudnason, <sup>30,31</sup> Mika Kivimaki, <sup>77</sup> Aroon Hingorani, <sup>77</sup> Sirkka M Keinanen-Kiukaanniemi, <sup>140,141</sup> Timo E Saaristo, <sup>74,142</sup> Dorret I Boomsma, <sup>28</sup> Kari Stefansson, <sup>27,31</sup> Pim van der Harst, <sup>26</sup> Josée Dupuis, <sup>52,143</sup> Nancy L Pedersen, <sup>12</sup> Naveed Sattar, <sup>144</sup> Tamara B Harris, <sup>145</sup> Francesco Cucca, <sup>19,20</sup> Samuli Ripatti, <sup>146,147,148</sup> Veikko Salomaa, <sup>149</sup> Karen L Mohlke, <sup>37</sup> Beverley Balkau, <sup>150,151</sup> Philippe Froguel, <sup>14,15,152</sup> Anneli Pouta, <sup>153,154</sup> Marjo-Riitta Jarvelin, <sup>154,155,156,157</sup> Nicholas J Wareham, <sup>1</sup> Nabila Bouatia-Naji, <sup>14,15,158</sup> Mark I McCarthy, <sup>2,3,159</sup> Paul W Franks, <sup>16,17,160</sup> James B Meigs, <sup>161,162</sup> Tanya M Teslovich, <sup>5,6</sup> Jose C Florez, <sup>162,165</sup> Claudia Langenberg, <sup>1,77</sup> Erik Ingelsson, <sup>12</sup> Inga Prokopenko, <sup>2,3</sup> and Inês Barroso <sup>7,166,167</sup>

- 1. Medical Research Council (MRC) Epidemiology Unit, Institute of Metabolic Science, Addenbrooke's Hospital, Cambridge, UK.
- 2. Wellcome Trust Center for Human Genetics, University of Oxford, Oxford, UK.
- 3. Oxford Centre for Diabetes, Endocrinology and Metabolism, University of Oxford, Oxford, UK.
- 4. Bioinformatics Graduate Program, University of Michigan Medical School, Ann Arbor, Michigan, USA.
- 5. Center for Statistical Genetics, University of Michigan, Ann Arbor, Michigan, USA.
- 6. Department of Biostatistics, University of Michigan, Ann Arbor, Michigan, USA.
- 7. Wellcome Trust Sanger Institute, Wellcome Trust Genome Campus, Hinxton, UK.
- 8. Division of Endocrinology, Diabetes and Nutrition, University of Maryland, School of Medicine, Baltimore, Maryland, USA.
- 9. Estonian Genome Center, University of Tartu, Tartu, Estonia.
- 10. Atherosclerosis Research Unit, Department of Medicine Solna, Karolinska Institutet, Stockholm, Sweden.
- 11. Center for Molecular Medicine, Karolinska University Hospital, Stockholm, Sweden.
- 12. Department of Medical Epidemiology and Biostatistics, Karolinska Institutet, Stockholm, Sweden.
- 13. Department of Preventive Medicine, Northwestern University Feinberg School of Medicine, Chicago, Illinois, USA.
- 14. Universite Lille Nord de France, Lille, France.
- 15. Le Centre national de la recherche scientifique (CNRS) UMR8199, Institut Pasteur de Lille, France.
- 16. Department of Clinical Sciences, Genetic and Molecular Epidemiology Unit, Lund University, Skåne University Hospital Malmö, Malmö, Sweden.
- 17. Department of Public Health & Clinical Medicine, Umeå University, Umeå, Sweden.
- 18. Department of Odontology, Umeå University, Umeå, Sweden.
- 19. Istituto di Ricerca Genetica e Biomedica, CNR, Monserrato, Italy.
- 20. Dipartimento di Scienze Biomediche, Università di Sassari, Sassari, Italy.
- 21. Robertson Centre for Biostatistics, University of Glasgow, Glasgow, UK.
- 22. Interuniversity Cardiology Institute of the Netherlands (ICIN), Durrer Center for Cardiogenetic Research, Utrecht, The Netherlands.
- 23. Departmentt of Cardiology, Leiden University Medical Center, Leiden, The Netherlands.
- 24. Department of Clinical Pharmacology, William Harvey Research Institute, Barts and The London School of Medicine and Dentistry, Queen Mary University of London, Charterhouse Square, London, UK.
- 25. The Genome Centre, Barts and The London School of Medicine and Dentistry, Queen Mary University of London, Charterhouse Square, London,
- 26. Department of Cardiology, University of Groningen, University Medical Center Groningen, The Netherlands.
- 27. deCODE genetics, Rekjavik, Iceland.
- 28. Department of Biological Psychology, VU University & EMGO+ Institute, Amsterdam, The Netherlands.
- 29. University College London Genetics Institute (UGI), University College London, London, UK.
- 30. Icelandic Heart Association, Kopavogur, Iceland.
- 31. Faculty of Medicine, University of Iceland, Reykjavìk, Iceland.
- 32. Institute of Genetic Epidemiology, Helmholtz Zentrum München German Research Center for Environmental Health, Neuherberg, Germany.

- 33. Department of Chronic Disease Prevention, National Institute for Health and Welfare, Helsinki, Finland.
- 34. University of Helsinki, Institute of Molecular Medicine, Finland (FIMM), Helsinki, Finland.
- 35. MRC Council Centre for Causal Analyses in Translational Epidemiology (CAiTE) Centre, School of Social and Community Medicine, University of Bristol, UK.
- 36. School of Social and Community Medicine, University of Bristol, UK.
- 37. Department of Genetics, University of North Carolina, Chapel Hill, North Carolina, USA.
- 38. Department of Biostatistical Sciences, Division of Public Health Sciences, Wake Forest School of Medicine, Winston-Salem, North Carolina, USA.
- 39. Busselton Population Medical Research Institute, Sir Charles Gairdner Hospital, Nedlands, Western Australia, Australia.
- 40. School of Population Health, The University of Western Australia, Nedlands, Western Australia, Australia.
- 41. School of Pathology and Laboratory Medicine, The University of Western Australia, Nedlands, Western Australia, Australia.
- 42. PathWest Laboratory Medicine WA, Nedlands, Western Australia, Australia.
- 43. Department of Epidemiology, School of Public Health, University of Michigan, Ann Arbor, Michigan, USA.
- 44. MRC Human Genetics Unit at the Medical Research Council Institute of Genetics and Molecular Medicine, University of Edinburgh, Western General Hospital, Edinburgh, UK.
- 45. Centre for Medical Systems Biology (CMSB), Leiden, The Netherlands.
- 46. Genetic Epidemiology Unit, Department of Epidemiology, Erasmus University Medical Center, Rotterdam, The Netherlands.
- 47. Institute of Molecular and Cell Biology, University of Tartu, Tartu, Estonia.
- 48. Centre for Population Health Sciences, University of Edinburgh, Teviot Place, Edinburgh, UK.
- 49. The Broad Institute of Harvard and MIT, Boston, Massachusetts, USA.
- 50. Department of Pharmacology, University of Pennsylvania Perelman School of Medicine, Philadelphia, Pennsylvania, USA.
- 51. Divisions of Genetics & Rheumatology, Brigham and Women's Hospital, Boston, Massachusetts, USA.
- 52. Department of Biostatistics, Boston University School of Public Health, Boston, Massachusetts, USA.
- 53. Boston University Data Coordinating Center, Boston, Massachusetts, USA.
- 54. Department of Medical Genetics, University of Lausanne, Lausanne, Switzerland.
- 55. The Service of Medical Genetics, CHUV, University Hospital, Lausanne Switzerland.
- 56. Genome Technology Branch, National Human Genome Research Institute, National Institutes of Health (NIH), Bethesda, Maryland, USA.
- 57. Genetics, GlaxoSmithKline, Upper Merion, Pennsylvania, USA.
- 58. Clinical Research Branch, National Institute on Aging, Baltimore, Maryland, USA.
- 59. Illumina Inc., Chesterford Research Park, Essex, UK.
- 60. Department of Cardiovascular Medicine, University of Oxford, Oxford, UK.
- 61. Centre for Biomedicine, European Academy Bozen/Bolzano (EURAC), Bolzano, Italy Affiliated Institute of the University of Lübeck, Lübeck, Germany.
- 62. Division of Statistical Genomics, Washington University School of Medicine, St. Louis, Missouri, USA.
- 63. Institute of Medical Informatics, Biometry and Epidemiology, Ludwig-Maximilians-Universität, Munich, Germany.
- 64. Department of Medicine I, University Hospital Grosshadern, Ludwig-Maximilians-Universität, Munich, Germany.
- 65. Cardiothoracic Surgery Unit, Department of Molecular Medicine and Surgery, Karolinska Institutet, Stockholm, Sweden.
- 66. Department of Evolutionary Biology, Genetic Section, University of Ferrara, Ferrara, Italy.
- 67. Department of Epidemiology, University Medical Center Groningen, University of Groningen, Groningen, The Netherlands.
- 68. Interdisciplinary Center for Pathology of Emotions, University of Groningen, University Medical Center Groningen, Groningen, The Netherlands.
- 69. University Medical Center Groningen, University of Groningen, Groningen, The Netherlands.
- 70. Division of Epidemiology and Community Health, University of Minnesota, Minneapolis, Minnesota, USA.
- 71. Carolina Center for Genome Sciences, University of North Carolina-Chapel Hill, Chapel Hill, North Carolina, USA.
- 72. Department of Epidemiology, University of North Carolina-Chapel Hill, Chapel Hill, North Carolina, USA.
- 73. Department of Public Health and Clinical Medicine, Section for Nutritional Research, Umeå University Hospital, Umeå, Sweden.
- 74. Pirkanmaa Hospital District, Tampere, Finland.
- 75. Laboratory of Genetics, National Institute on Aging, NIH, Baltimore, Maryland, USA.
- 76. Department of Internal Medicine, University of Groningen, University Medical Center Groningen, Groningen, The Netherlands.

- 77. Department of Epidemiology and Public Health, University College London, London UK.
- 78. Division of Cardiovascular Epidemiology, Institute of Environmental Medicine, Karolinska Institutet, Stockholm, Sweden.
- 79. Institute for Clinical Diabetology, German Diabetes Center, Leibniz Center for Diabetes Research at Heinrich Heine University Düsseldorf, Düsseldorf, Germany.
- 80. Department of Twin Research and Genetic Epidemiology, King's College London, London, UK.
- 81. Department of Epidemiology, Center for Aging and Population Health, University of Pittsburgh, Pittsburgh, Pennsylvania, USA.
- 82. Department of Epidemiology and Prevention, Division of Public Health Sciences, Wake Forest School of Medicine, Winston-Salem, North Carolina, USA
- 83. Institute of Biomedicine, Physiology, University of Eastern Finland, Kuopio Campus, Kuopio, Finland.
- 84. University of Leipzig, IFB Adiposity Diseases, Leipzig, Germany.
- 85. Pediatric Research Center, Department of Women's & Child Health, University of Leipzig, Leipzig, Germany.
- 86. School of Medicine and Pharmacology, The University of Western Australia, Nedlands, Western Australia, Australia.
- 87. Pulmonary Physiology, Sir Charles Gairdner Hospital, Nedlands, Western Australia, Australia.
- 88. Department of Internal Medicine, Erasmus University Medical Center, Rotterdam, The Netherlands.
- 89. Netherlands Consortium for Healthy Ageing of the Netherlands (NCHAH) of the Genomics Initiative (NGI), Leiden, The Netherlands.
- 90. Department of Epidemiology, Erasmus University Medical Center, Rotterdam, The Netherlands.
- 91. Medical Research Institute, University of Dundee, Dundee, UK.
- 92. Department of Psychiatry, VU University Medical Centre, Amsterdam, The Netherlands.
- 93. Faculty of Medicine, University of Split, Split, Croatia.
- 94. U557 Institut National de la Santé et de la Recherche Médicale, U1125 Institut National de la Recherche Agronomique, Université Paris 13, Bobigny, France
- 95. Department of Dietetics-Nutrition, Harokopio University, Athens, Greece.
- 96. Centre for Population Health Sciences, University of Edinburgh, Edinburgh, UK.
- 97. University of Leipzig, Interdisciplinary Center for Clinical Research, Leipzig, Germany.
- 98. National Institute for Health and Welfare, Diabetes Prevention Unit, Helsinki, Finland.
- 99. Geriatric Department Azienda Sanitaria Firenze, Florence Italy.
- 100. Department Vascular Medicine, Academic Medical Center, Amsterdam, The Netherlands.
- 101. Institute of Biometrics and Epidemiology, German Diabetes Center, Leibniz Center for Diabetes Research at Heinrich Heine University Düsseldorf, Düsseldorf, Germany.
- 102. Research Unit of Molecular Epidemiology, Helmholtz Zentrum München German Research Center for Environmental Health, Neuherberg, Germany.
- 103. The members of this consortium are listed in the Supplementary Note.
- 104. Synlab Academy, Mannheim, Germany.
- 105. Mannheim Institute of Public Health, Social and Preventive Medicine, Medical Faculty of Mannheim, University of Heidelberg, Mannheim, Germany.
- 106. Ludwigshafen Risk and Cardiovascular Health (LURIC) Study nonprofit LLC, Freiburg, Germany.
- 107. Division of Endocrinology and Diabetes, Department of Medicine, University Hospital, Ulm, Germany.
- 108. Institute of Epidemiology II, Helmholtz Zentrum München German Research Center for Environmental Health, Neuherberg, Germany.
- 109. Department of Neurology, General Central Hospital, Bolzano, Italy.
- 110. Department of Neurology, University of Lübeck, Lübeck, Germany.
- 111. Department of Medicine, University of Leipzig, Leipzig, Germany.
- 112. Diabetes and Obesity Research Institute, Cedars-Sinai Medical Center, Los Angeles, California, USA.
- 113. Diabetes Prevention Unit, National Institute for Health and Welfare, Helsinki, Finland.
- 114. South Ostrobothnia Central Hospital, Seinäjoki, Finland.
- 115. Red RECAVA Grupo RD06/0014/0015, Hospital Universitario La Paz, Madrid, Spain.
- 116. Centre for Vascular Prevention, Danube-University Krems, Krems, Austria.
- 117. Department of Preventive Medicine, Keck School of Medicine of USC, Los Angeles, California, USA.
- 118. Department of Physiology & Biophysics, Keck School of Medicine of USC, Los Angeles, California, USA.

- 119. Cardiovascular Health Research Unit, Departments of Medicine, University of Washington, Seattle, Washington, USA.
- 120. Group Health Research Institute, Group Health Cooperative, Seattle, Washington, USA.
- 121. Department of Epidemiology, University of Washington, Seattle, Washington, USA.
- 122. Department of Health Services, University of Washington, Seattle, Washington, USA.
- 123. Department of Internal Medicine, University Hospital and University of Lausanne, Lausanne, Switzerland.
- 124. Department of Public Health and Clinical Nutrition, University of Eastern Finland, Kuopio, Finland.
- 125. Research Unit, Kuopio University Hospital, Kuopio, Finland.
- 126. Odense Patient data Explorative Network (OPEN), Odense, Denmark.
- 127. Institute of Regional Health Services Research, Odense, Denmark.
- 128. Hjelt Institute, Department of Public Health, University of Helsinki, Helsinki, Finland.
- 129. National Institute for Health and Welfare, Department of Mental Health and Substance Abuse Services, Helsinki, Finland.
- 130. U872 Institut National de la Santé et de la Recherche Médicale, Centre de Recherche des Cordeliers, Paris, France.
- 131. Department of Medical Sciences, Uppsala University, Uppsala, Sweden.
- 132. Geriatric Research and Education Clinical Center, Veterans Administration Medical Center, Baltimore, Maryland, USA.
- 133. Department of Medicine, University of Eastern Finland and Kuopio University Hospital, Kuopio, Finland.
- 134. Department of Medicine III, University of Dresden, Medical Faculty Carl Gustav Carus, Dresden, Germany.
- 135. Kuopio Research Institute of Exercise Medicine, Kuopio, Finland.
- 136. Department of Clinical Physiology and Nuclear Medicine, Kuopio University Hospital, Kuopio, Finland.
- 137. Department of Nutrition, University of North Carolina, Chapel Hill, North Carolina, USA.
- 138. Hannover Unified Biobank, Hannover Medical School, Hannover, Germany.
- 139. Department of Cardiology, Karolinska University Hospital, Stockholm, Sweden.
- 140. Faculty of Medicine, Institute of Health Sciences, University of Oulu, Oulu, Finland.
- 141. Unit of General Practice, Oulu University Hospital, Oulu, Finland.
- 142. Finnish Diabetes Association, Tampere, Finland.
- 143. National Heart, Lung, and Blood Institute's Framingham Heart Study, Framingham, Massachusetts, USA.
- 144. British Heart Foundation (BHF) Building, Institute of Cardiovascular and Medical Sciences, University of Glasgow, Glasgow, UK.
- 145. Laboratory of Epidemiology, Demography, and Biometry, National Institute on Ageing, Bethesda, Maryland, USA.
- 146. Institute for Molecular Medicine Finland, FIMM, University of Helsinki, Helsinki, Finland.
- 147. Public Health Genomics Unit, National Institute for Health and Welfare, Helsinki, Finland.
- 148. Wellcome Trust Sanger Institute, Hinxton, UK.
- 149. Unit of Chronic Disease Epidemiology and Prevention, National Institute for Health and Welfare, Helsinki, Finland.
- 150. Inserm, Centre de recherche en Épidémiologie et Santé des Populations (CESP) Center for Research in Epidemiology and Public Health, U1018, Epidemiology of diabetes, obesity and chronic kidney disease over the lifecourse, Villejuif, France.
- 151. University Paris Sud 11, UMRS 1018, Villejuif, France.
- 152. Department of Genomics of Common Disease, School of Public Health, Imperial College London, Hammersmith Hospital, London, UK.
- 153. Department of Clinical Sciences/Obstetrics and Gynecology, University of Oulu, Oulu, Finland.
- 154. Department of Lifecourse and Services, National Institute for Health and Welfare, Oulu, Finland.
- 155. Biocenter Oulu, University of Oulu, Oulu, Finland.
- 156. Department of Epidemiology and Biostatistics, School of Public Health, MRC-HPA Centre for Environment and Health, Faculty of Medicine, Imperial College London, London, UK.
- 157. Institute of Health Sciences, University of Oulu, Oulu, Finland.
- 158. Inserm U970, Paris Cardiovascular Research Center PARCC, Paris, France.
- 159. Oxford National Institute for Health Research (NIHR) Biomedical Research Centre, Churchill Hospital, Oxford, UK.
- 160. Department of Nutrition, Harvard School of Public Health, Boston, Massachusetts, USA.
- 161. General Medicine Division, Massachusetts General Hospital, Boston, Massachusetts, USA.
- 162. Department of Medicine, Harvard Medical School, Boston, Massachusetts, USA.
- 163. Center for Human Genetic Research, Massachusetts General Hospital, Boston, Massachusetts, USA.

- 164. Diabetes Research Center, Diabetes Unit, Massachusetts General Hospital, Boston, Massachusetts, USA.
- 165. Program in Medical and Population Genetics, Broad Institute, Cambridge, Massachusetts, USA.
- 166. NIHR Cambridge Biomedical Research Centre, Institute of Metabolic Science, Addenbrooke's Hospital, Cambridge, UK.
- 167. University of Cambridge Metabolic Research Laboratories, Institute of Metabolic Science, Addenbrooke's Hospital, Cambridge, UK.

#### The MutHER Consortium

Kourosh R. Ahmadi<sup>1</sup>, Chrysanthi Ainali<sup>2</sup>, Amy Barrett<sup>3</sup>, Veronique Bataille<sup>1</sup>, Jordana T. Bell<sup>1,4</sup>, Alfonso Buil<sup>5</sup>, Panos Deloukas<sup>6</sup>, Emmanouil T. Dermitzakis<sup>5</sup>, Antigone S. Dimas<sup>4,5</sup>, Richard Durbin<sup>6</sup>, Daniel Glass<sup>1</sup>, Elin Grundberg<sup>1,6,13</sup>, Neelam Hassanali<sup>3</sup>, Åsa K. Hedman<sup>4</sup>, Catherine Ingle<sup>6</sup>, Sarah Keildson<sup>4</sup>, David Knowles<sup>7</sup>, Maria Krestyaninova<sup>8</sup>, Cecilia M. Lindgren<sup>4</sup>, Christopher E. Lowe<sup>9,10</sup>, Mark I. McCarthy<sup>3,4,11</sup>, Eshwar Meduri <sup>1,6</sup>, Paola di Meglio<sup>12</sup>, Josine L. Min<sup>4</sup>, Stephen B. Montgomery<sup>5</sup>, Frank O. Nestle<sup>12</sup>, Alexandra C. Nica<sup>5</sup>, James Nisbet<sup>6</sup>, Stephen O'Rahilly<sup>9,10</sup>, Leopold Parts<sup>6</sup>, Simon Potter<sup>6</sup>, Magdalena Sekowska<sup>6</sup>, So-Youn Shin<sup>6</sup>, Kerrin S. Small<sup>1,6</sup>, Nicole Soranzo<sup>1,6</sup>, Tim D. Spector<sup>1</sup>, Gabriela Surdulescu<sup>1</sup>, Mary E. Travers<sup>3</sup>, Loukia Tsaprouni<sup>6</sup>, Sophia Tsoka<sup>2</sup>, Alicja Wilk<sup>6</sup>, Tsun-Po Yang<sup>6</sup>, Krina T. Zondervan<sup>4</sup>

- 1. Department of Twin Research and Genetic Epidemiology, King's College London, London, UK
- 2. Department of Informatics, School of Natural and Mathematical Sciences, King's College London, Strand, London, UK
- 3. Oxford Centre for Diabetes, Endocrinology & Metabolism, University of Oxford, Churchill Hospital, Oxford, UK
- 4. Wellcome Trust Centre for Human Genetics, University of Oxford, Oxford, UK
- 5. Department of Genetic Medicine and Development, University of Geneva Medical School, Geneva, Switzerland
- 6. Wellcome Trust Sanger Institute, Wellcome Trust Genome Campus, Hinxton, UK
- 7. University of Cambridge, Cambridge, UK
- 8. European Bioinformatics Institute, Hinxton, UK
- 9. University of Cambridge Metabolic Research Labs, Institute of Metabolic Science Addenbrooke's Hospital Cambridge, UK
- 10. Cambridge NIHR Biomedical Research Centre, Addenbrooke's Hospital, Cambridge, UK
- 11. Oxford NIHR Biomedical Research Centre, Churchill Hospital, Oxford, UK
- 12. St. John's Institute of Dermatology, King's College London, London, UK
- 13. Department of Human Genetics, McGill University, McGill University and Genome Quebec Innovation Centre, H3A1A5 Montreal, Canada

#### The PAGE Consortium

Active PAGE investigators at the time of this analysis included:

**Coordinating Center**: Rutgers University, Piscataway, NJ: Tara Matise, Steve Buyske, Julia Higashio, Rasheeda Williams, Andrew Nato; University of Southern California, Los Angeles, CA: Jose Luis Ambite, Ewa Deelman.

NHGRI: Division of Genomic Medicine, NHGRI, NIH, Bethesda, MD: Teri Manolio, Lucia Hindorff.

*CALiCo*: University of North Carolina, Chapel Hill, NC: Kari E. North, Gerardo Heiss, Kira Taylor, Nora Franceschini, Christy Avery, Misa Graff, Danyu Lin, Miguel Quibrera; Baylor College of Medicine, Houston, TX: Barbara Cochran; Johns Hopkins Bloomberg School of Public Health, Baltimore, MD: Linda Kao; Penn Medical Lab, Washington DC: Jason Umans; SW Foundation for BioMedical Research, San Antonio, TX: Shelley Cole, Jean MacCluer; University of Alabama at Birmingham, Birmingham, AB: Sharina Person; University of Minnesota, Minneapolis, MN: James Pankow, Myron Gross; University of Texas Health Science Center, Houston: Eric Boerwinkle, Myriam Fornage; University of Vermont, Burlington, VT: Peter Durda, Nancy Jenny; University of Washington, Seattle, WA: Bruce Patsy, Alice Arnold, Petra Buzkova.

*EAGLE*: Vanderbilt University, Nashville, TN: Dana Crawford, Jonathan Haines, Deborah Murdock, Kim Glenn, Kristin Brown-Gentry, Tricia Thornton-Wells, Logan Dumitrescu, Janina Jeff, William S. Bush, Sabrina L. Mitchell, Robert Goodloe, Sarah Wilson, Jonathan Boston, Jennifer Malinowski, Nicole Restrepo, Matthew Oetjens, Jay Fowke, Wei Zheng; Heidelberg University, Tiffin, OH: Kylee Spencer; Pennsylvania State University, State College, PA: Marylyn Ritchie, Sarah Pendergrass.

**MEC**: University of Hawaii, Honolulu, HI: Loïc Le Marchand, Lynne Wilkens, Lani Park, Maarit Tiirikainen, Laurence Kolonel, Unhee Lim, Iona Cheng, Hansong Wang, Ralph Shohet; Keck School of Medicine, University of Southern California, Los Angeles, CA: Christopher Haiman, Daniel Stram, Brian Henderson, Kristine Monroe, Fredrick Schumacher.

**WHI**: Fred Hutchinson Cancer Research Institute (FHCRC), Seattle, WA: Charles Kooperberg, Ulrike Peters, Garnet Anderson, Chris Carlson, Ross Prentice, Andrea LaCroix, Chunyuan Wu, Cara Carty, Jian Gong, Stephanie Rosse, Alicia Young, Jeff Haessler, Jonathan Kocarnik, Yi Lin; Ohio State Medical Center, Columbus, OH: Rebecca Jackson; Translational Genomic Science Institute (TGen): David Duggan; University of Pittsburgh, PA: Lew Kuller.

# The ReproGen Consortium (age at menopause data)

Lisette Stolk, 1,2 John RB Perry, 3,4 Daniel I Chasman, 5,6 Chunyan He, 7,8 Massimo Mangino, 9 Patrick Sulem, <sup>10</sup> Maja Barbalic, <sup>11</sup> Linda Broer, <sup>12</sup> Enda M Byrne, <sup>13</sup> Florian Ernst, <sup>14</sup> Tõnu Esko, <sup>15,16,17</sup> Nora Franceschini, <sup>18</sup> Daniel F Gudbjartsson, <sup>10</sup> Jouke-Jan Hottenga, <sup>19</sup> Peter Kraft, <sup>20,21</sup> Patick F McArdle, <sup>22</sup> Eleonora Porcu,<sup>23</sup> So-Youn Shin,<sup>24</sup> Albert V Smith,<sup>25,26</sup> Sophie van Wingerden,<sup>12</sup> Guangju Zhai,<sup>9,27</sup> Wei V Zhuang,<sup>28</sup> Eva Albrecht,<sup>29</sup> Behrooz Z Alizadeh,<sup>30</sup> Thor Aspelund,<sup>25,26</sup> Stefania Bandinelli,<sup>31</sup> Lovorka Barac Lauc,<sup>32</sup> Jacques S Beckmann, 33,34 Mladen Boban, 5 Eric Boerwinkle, 1 Frank J Broekmans, 6 Andrea Burri, Harry Campbell, 7 Stephen J Chanock,<sup>38</sup> Constance Chen,<sup>20,39</sup> Marilyn C Cornelis,<sup>39</sup> Tanguy Corre,<sup>40</sup> Andrea D Coviello,<sup>41,42</sup> Pio d'Adamo, 43,44 Gail Davies, 45 Ulf de Faire, 46 Eco JC de Geus, 19,47 Ian J Deary, 45,48 George VZ Dedoussis, 49 Panagiotis Deloukas,<sup>24</sup> Shah Ebrahim,<sup>50</sup> Gudny Eiriksdottir,<sup>25</sup> Valur Emilsson,<sup>25</sup> Johan G Eriksson,<sup>51,52,53,54,55</sup> Bart CJM Fauser,<sup>36</sup> Liana Ferreli,<sup>23</sup> Luigi Ferrucci,<sup>56</sup> Krista Fischer,<sup>15</sup> Aaron R Folsom,<sup>57</sup> Melissa E Garcia,<sup>58</sup> Paolo Gasparini, 43,44 Christian Gieger, 29 Nicole Glazer, 41 Diederick E Grobbee, 59 Per Hall, 60 Toomas Haller, 15 Susan E Hankinson, 20,61 Merli Hass, 15 Caroline Hayward, 62 Andrew C Heath, 63 Albert Hofman, 2,12 Erik Ingelsson, 60 A Cecile JW Janssens, 12 Andrew D Johnson, 42 David Karasik, 42,64 Sharon LR Kardia, 65 Jules Keyzer, <sup>66</sup> Douglas P Kiel, <sup>42,64</sup> Ivana Kolcic, <sup>35</sup> Zoltán Kutalik, <sup>33,67</sup> Jari Lahti, <sup>68</sup> Sandra Lai, <sup>23</sup> Triin Laisk, <sup>69</sup> Joop SE Laven, <sup>70</sup> Debbie A Lawlor, <sup>71</sup> Jianjun Liu, <sup>72</sup> Lorna M Lopez, <sup>45,48</sup> Yvonne V Louwers, <sup>70</sup> Patrik KE Magnusson, <sup>60</sup> Mara Marongiu,<sup>23</sup> Nicholas G Martin,<sup>13</sup> Irena Martinovic Klaric,<sup>73</sup> Corrado Masciullo,<sup>40</sup> Barbara McKnight,<sup>74</sup> Sarah E Medland, <sup>13</sup> David Melzer, <sup>3</sup> Vincent Mooser, <sup>75</sup> Pau Navarro, <sup>62</sup> Anne B Newman, <sup>76</sup> Dale R Nyholt, <sup>13</sup> N. Charlotte Onland-Moret, 59 Aarno Palotie, 24,77,78 Guillaume Paré, 5,6,79 Alex N Parker, 80,81 Nancy L Pedersen, 60 Petra HM Peeters, 59,82 Giorgio Pistis, 40 Andrew S Plump, 83 Ozren Polasek, 35 Victor JM Pop, 84 Bruce M Psaty, 85,86 Katri Räikkönen, 68 Emil Rehnberg, 60 Jerome I Rotter, 87 Igor Rudan, 35,37 Cinzia Sala, 40 Andres Salumets, 15,69,88 Angelo Scuteri, 89 Andrew Singleton, 90 Jennifer A Smith, 65 Harold Snieder, 30,91 Nicole Soranzo, 9,24 Simon N Stacey, 10 John M Starr, 48,92 Maria G Stathopoulou, 49,93 Kathleen Stirrups, 24 Ronald P Stolk, 30,91 Unnur Styrkarsdottir, 10 Yan V Sun, 4 Albert Tenesa, 62,95 Barbara Thorand, 6 Daniela Toniolo, 40,97 Laufey Tryggvadottir, 26,98 Kim Tsui, 80 Sheila Ulivi, 43 Rob M van Dam, 39,99 Yvonne T van der Schouw, 59 Carla H van Gils,<sup>59</sup> Peter van Nierop,<sup>100</sup> Jacqueline M Vink,<sup>19</sup> Peter M Visscher,<sup>48,101</sup> Marlies Voorhuis,<sup>36,59</sup> Gérard Waeber, 102 Henri Wallaschofski, 103 H Erich Wichmann, 104, 105, 106 Elisabeth Widen, 77 Colette JM Wijnands-van Gent, <sup>107</sup> Gonneke Willemsen, <sup>19</sup> James F Wilson, <sup>37</sup> Bruce HR Wolffenbuttel, <sup>91,108</sup> Alan F Wright, <sup>62</sup> Laura M Yerges-Armstrong,<sup>22</sup> Tatijana Zemunik,<sup>35</sup> Lina Zgaga,<sup>37,109</sup> M. Carola Zillikens,<sup>1</sup> Marek Zygmunt,<sup>110</sup> The LifeLines Cohort Study, 91 Alice M Arnold, 74 Dorret I Boomsma, 19,47 Julie E. Buring, 5,6,111 Laura Crisponi, 23 Ellen W Demerath,<sup>57</sup> Vilmundur Gudnason,<sup>25,26</sup> Tamara B Harris,<sup>58</sup> Frank B Hu,<sup>20,39,61</sup> David J Hunter,<sup>20,39,61,21</sup> Lenore J Launer, <sup>58</sup> Andres Metspalu, <sup>15,16,17,88</sup> Grant W Montgomery, <sup>13</sup> Ben A Oostra, <sup>112</sup> Paul M Ridker, <sup>5,6,111,113</sup> Serena Sanna,<sup>23</sup> David Schlessinger,<sup>114</sup> Tim D Spector,<sup>9</sup> Kari Stefansson,<sup>10,26</sup> Elizabeth A Streeten,<sup>22</sup> Unnur

Thorsteinsdottir,<sup>10,26</sup> Manuela Uda,<sup>23</sup> André G Uitterlinden,<sup>1,2,12</sup> Cornelia M van Duijn,<sup>2</sup> Henry Völzke,<sup>115</sup> Anna Murray,<sup>3</sup> Joanne M Murabito,<sup>41,42</sup> Jenny A Visser,<sup>1</sup> and Kathryn L Lunetta<sup>28,42</sup>

- 1. Department of Internal Medicine, Erasmus MC, Rotterdam, the Netherlands
- 2. Netherlands Consortium of Healthy Aging, Rotterdam, the Netherlands
- 3. Peninsula Medical School, University of Exeter, UK
- 4. Wellcome Trust Centre for Human Genetics, University of Oxford, Oxford, UK
- 5. Division of Preventive Medicine, Brigham and Women's Hospital, Boston USA
- 6. Harvard Medical School, Boston, USA
- 7. Department of Public Health, Indiana University School of Medicine, Indianapolis, Indiana, USA
- 8. Melvin and Bren Simon Cancer Center, Indiana University, Indianapolis, Indiana, USA
- 9. Department of Twin Research and Genetic Epidemiology, King's College London, London, UK
- 10. deCODE Genetics, Reykjavik, Iceland
- 11. Human Genetics Center, University of Texas Health Science Center at Houston, Houston, Texas, USA
- 12. Department of Epidemiology, Erasmus Medical Center, Rotterdam, The Netherlands
- 13. Queensland Institute of Medical Research, Brisbane, Australia
- 14. Interfakultäres Institut für Genomforschung, Universität Greifswald, Germany
- 15. Estonian Genome Center, University of Tartu, Tartu, Estonia
- 16. Estonian Biocenter, Tartu, Estonia
- 17. Institute of Molecular and Cell Biology, University of Tartu, Tartu, Estonia
- 18. Department of Epidemiology, Gillings School of Global Public Health, University of North Carolina at Chapel Hill, Chapel Hill, North Carolina, USA
- 19. Dept Biological Psychology, VU University Amsterdam, Amsterdam, The Netherlands
- 20. Department of Epidemiology, Harvard School of Public Health, Boston, Massachusetts, USA
- 21. Broad Institute of Harvard and MIT. USA
- 22. Division of Endocrinology, Diabetes and Nutrition, University of Maryland School of Medicine, Baltimore, Maryland, USA
- 23. Istituto di Ricerca Genetica e Biomedica, Consiglio Nazionale delle Ricerche, Cagliari, Italy
- 24. Wellcome Trust Sanger Institute, Wellcome Trust Genome Campus, Hinxton, UK
- 25. Icelandic Heart Association, Kopavogur, Iceland
- 26. Faculty of Medicine, University of Iceland, Reykjavik, Iceland
- 27. Discipline of Genetics, Faculty of Medicine, Memorial University of Newfoundland, St. John's, NL, Canada
- 28. Department of Biostatistics, Boston University School of Public Health, Boston Massachusetts, USA
- 29. Institute of Genetic Epidemiology, Helmholtz Zentrum München German Research Center for Environmental Health, Neuherberg, Germany
- 30. Department of Epidemiology, University Medical Center Groningen, University of Groningen, the Netherlands
- 31. Geriatric Unit, Azienda Sanitaria di Firenze, Florence, Italy
- 32. Croation Science Foundation, Zagreb, Croatia
- 33. Department of Medical Genetics, University of Lausanne, Switzerland
- 34. Service of Medical Genetics, Centre Hospitalier Universitaire Vaudois (CHUV), University Hospital, Lausanne, Switzerland
- 35. Faculty of Medicine, University of Split, Split, Croatia
- 36. Department of Reproductive Medicine and Gynaecology, University Medical Center Utrecht, Utrecht, the Netherlands
- 37. Centre for Population Health Sciences, University of Edinburgh, Edinburgh, UK
- 38. Division of Cancer Epidemiology and Genetics, National Cancer Institute, National Institutes of Health, Bethesda, Maryland, USA
- 39. Department of Nutrition, Harvard School of Public Health, Boston, Massachusetts, USA
- 40. Division of Genetics and Cell Biology, San Raffaele Scientific Institute, Milan, Italy
- 41. Sections of General Internal Medicine, Preventive Medicine and Epidemiology, Department of Medicine, Boston University School of Medicine, Boston MA, USA
- 42. NHLBI Framingham Heart Study, Framingham, MA, USA

- 43. Institute for Maternal and Child Health, IRCCS "Burlo Garofolo" Trieste, Italy
- 44. University of Trieste, Trieste, Italy
- 45. Department of Psychology, The University of Edinburgh, Edinburgh, UK
- 46. Institute of Environmental Medicine, Karolinska Institutet, Stockholm, Sweden
- 47. EMGO+ Institute, VU Medical Centre, Amsterdam, The Netherlands
- 48. Centre for Cognitive Ageing and Cognitive Epidemiology, The University of Edinburgh, Edinburgh, UK
- 49. Department of Nutrition and Dietetics, Harokopio University, Athens, Greece
- 50. Department of Epidemiology & Population Healths, London School of Hygiene & Tropical Medicine, UK
- 51. National Institute for Health and Welfare, Finland
- 52. Department of General Practice and Primary Health Care, University of Helsinki, Finland
- 53. Helsinki University Central Hospital, Unit of General Practice, Helsinki, Finland
- 54 . Folkhalsan Research Centre, Helsinki, Finland
- 55. Vasa Central Hospital, Vasa, Finland
- 56 .Longitudinal Studies Section, Clinical Research Branch, National Institute on Aging, Baltimore, Maryland, USA
- 57. Division of Epidemiology and Community Health, School of Public Health, University of Minnesota, Minneapolis, Minnesota, USA
- 58. Laboratory of Epidemiology, Demography, and Biometry, National Institute on Aging, NIH, Bethesda, MD, USA
- 59. Julius Center for Health Sciences and Primary Care, University Medical Center Utrecht, Utrecht, the Netherlands
- 60. Department of Medical Epidemiology and Biostatistics, Karolinska Institutet, Stockholm, Sweden
- 61. Channing Laboratory, Department of Medicine, Brigham and Women.s Hospital Harvard Medical School, Boston, Massachusetts, USA
- 62. MRC Human Genetics Unit at the Medical Research Council Institute of Genetics and Molecular Medicine at the University of Edinburgh, Western General Hospital, Edinburgh, UK
- 63. Washington University St.Louis, St. Louis, MO, USA
- 64. Hebrew SeniorLife Institute for Aging Research and Harvard Medical School, Boston, Massachusetts, USA
- 65. Department of Epidemiology, University of Michigan, Ann Arbor, MI, USA
- 66. Diagnostic GP laboratory Eindhoven, Eindhoven, the Netherlands
- 67. Swiss Institute of Bioinformatics, Switzerland
- 68. Institute of Behavioural Sciences, University of Helsinki, Helsinki, Finland
- 69. Department of Obstetrics and Gynecology, University of Tartu, Tartu, Estonia
- 70. Division of Reproductive Medicine, Department of Obstetrics & Gynaecology, Erasmus MC, Rotterdam, the Netherlands
- 71. MRC Centre for Causal Analysis in Translational Epidemiology, School of Social & Community Medicine, University of Bristol, UK
- 72. Human genetic, Genome Institute of Singapore, Singapore
- 73. Institute for Migration and Ethnic Studies, Zagreb, Croatia
- 74. Department of Biostatistics, University of Washington, Seattle, WA, USA
- 75. Genetics Division, GlaxoSmithKline, King of Prussia, Pennsylvania, USA
- 76. Departments of Epidemiology and Medicine, University of Pittsburgh, Pittsburgh, Pennsylvania
- 77. Institute for Molecular Medicine Finland (FIMM), University of Helsinki, Finland
- 78. Department of Medical Genetics, University of Helsinki and University Central Hospital, Helsinki, Finland
- 79. Genetic and Molecular Epidemiology Laboratory, McMaster University, Hamilton, ON Canada
- 80. Amgen, Cambridge, MA USA
- 81. Foundation Medicine, Inc., Cambridge MA USA
- 82. Department of Epidemiology and Biostatistics, School of Public Health, Faculty of Medicine, Imperial College London, London, UK
- 83. Cardiovascular Disease, Merck Research Laboratory, Rahway, NJ, USA
- 84. Department of Clinical Health Psychology, University of Tilburg, Tilburg, the Netherlands
- 85. Departments of Medicine, Epidemiology and Health Services, University of Washington, Seattle, WA USA
- 86. Group Health Research Institute, Group Health Cooperative, Seattle, WA USA
- 87. Medical Genetics Institute, Cedars-Sinai Medical Center, Los Angeles, CA, USA
- 88. Competence Centre on Reproductive Medicine and Biology, Tartu, Estonia

- 89 .Lab Cardiovascular Sciences NIA NIH, Baltimore, USA
- 90. Laboratory of Neurogenetics, National Institute of Ageing, Bethesda, MD, USA
- 91. LifeLines Cohort Study & Biobank, University Medical Center Groningen, University of Groningen, the Netherlands
- 92. Geriatric Medicine Unit, University of Edinburgh, Edinburgh, UK
- 93. Cardiovascular Genetics Research Unit, EA4373, Université Henri Poincaré Nancy 1, Nancy, France
- 94. Department of Epidemiology, Emory University, Atlanta, GA, USA
- 95. The Roslin Institute, Royal (Dick) School of Veterinary Studies, University of Edinburgh, Roslin, UK
- 96. Institute of Epidemiology II, Helmholtz Zentrum München German Research Center for Environmental Health, Neuherberg, Germany
- 97. Institute of Molecular Genetics-CNR, Pavia, Italy
- 98. Icelandic Cancer Registry, Reykjavik, Iceland
- 99. Saw Swee Hock School of Public Health and Department of Medicine, Yong Loo Lin School of Medicine, National University of Singapore, Singapore
- 100. Municipal Health Service Brabant-Zuidoost, Helmond, the Netherlands
- 101. Genetic Epidemiology, Queensland Institute of Medical Research, Brisbane, Australia
- 102. Department of Internal Medicine, Centre Hospitalier Universitaire Vaudois (CHUV), University Hospital, Lausanne, Switzerland
- 103. Institute for Clinical Chemistry and Laboratory Medicine, University of Greifswald
- 104. Institute of Epidemiology I, Helmholtz Zentrum München German Research Center for Environmental Health, Neuherberg, Germany
- 105. Institute of Medical Informatics, Biometry and Epidemiology, Ludwig-Maximilians-Universität, Munich, Germany
- 106. Klinikum Grosshadern, Munich, Germany
- 107. POZOB Veldhoven, Veldhoven, the Netherlands
- 108. Department of Endocrinology, University Medical Center Groningen, University of Groningen, the Netherlands
- 109. Andrija Stampar School of Public Health, Medical School, University of Zagreb, Zagreb, Croatia
- 110. Klinik für Gynäkologie und Geburtshilfe, Universität Greifswald, Germany
- 111. Harvard School of Public Health, Boston, MA USA
- 112. Department of Clinical Genetics, Erasmus Medical Center, Rotterdam, The Netherlands
- 113. Division of Cardiology, Brigham and Women's Hospital, Boston, MA USA
- 114. National Institute on Aging, Intramural Research Program, Baltimore, MD, USA
- 115. Institut für Community Medicine, Universität Greifswald, Germany

# The ReproGen Consortium (age at menarche data)

Cathy E. Elks, 1 John R.B. Perry, 2 Patrick Sulem, 3 Daniel I. Chasman, 4,5 Nora Franceschini, 6 Chunyan He, 7,8 Kathryn L. Lunetta, 9,10 Jenny A. Visser, 11 Enda M. Byrne, 12,13 Diana L. Cousminer, 14 Daniel F. Gudbjartsson,<sup>3</sup> Tõnu Esko,<sup>15,16,17</sup> Bjarke Feenstra,<sup>18</sup> Jouke-Jan Hottenga,<sup>19</sup> Daniel L. Koller,<sup>20</sup> Zoltán Kutalik,<sup>21,22</sup> Peng Lin,<sup>23</sup> Massimo Mangino,<sup>24</sup> Mara Marongiu,<sup>25</sup> Patrick F. McArdle,<sup>26</sup> Albert V. Smith,<sup>27,28</sup> Lisette Stolk, 11,29 Sophie W. van Wingerden, 30 Jing Hua Zhao, Eva Albrecht, Tanguy Corre, 22 Erik Ingelsson, 33 Caroline Hayward,<sup>34</sup> Patrik K.E. Magnusson,<sup>33</sup> Erin N. Smith,<sup>35</sup> Shelia Ulivi,<sup>36</sup> Nicole M. Warrington,<sup>37</sup> Lina Zgaga,<sup>38</sup> Helen Alavere,<sup>15</sup> Najaf Amin,<sup>30</sup> Thor Aspelund,<sup>27,28</sup> Stefania Bandinelli,<sup>39</sup> Ines Barroso,<sup>40</sup> Gerald S. Berenson, 41 Sven Bergmann, 21,22 Hannah Blackburn, 40 Eric Boerwinkle, 42 Julie E. Buring, 4,43 Fabio Busonero, 25 Harry Campbell, 38 Stephen J. Chanock, 44 Wei Chen, 41 Marilyn C. Cornelis, 45 David Couper, 46 Andrea D. Coviello,<sup>47</sup> Pio d'Adamo,<sup>36</sup> Ulf de Faire,<sup>48</sup> Eco J.C. de Geus,<sup>19</sup> Panos Deloukas,<sup>40</sup> Angela Döring,<sup>31</sup> George Davey Smith, 49 Douglas F. Easton, 50 Gudny Eiriksdottir, 27 Valur Emilsson, 51 Johan Eriksson, 52,53,54,55 Luigi Ferrucci,<sup>56</sup> Aaron R. Folsom,<sup>57</sup> Tatiana Foroud,<sup>20</sup> Melissa Garcia,<sup>58</sup> Paolo Gasparini,<sup>36</sup> Frank Geller,<sup>18</sup> Christian Gieger, <sup>31</sup> The GIANT Consortium, <sup>59</sup> Vilmundur Gudnason, <sup>27,28</sup> Per Hall, <sup>33</sup> Susan E. Hankinson, <sup>43,60</sup> Liana Ferreli,<sup>25</sup> Andrew C. Heath,<sup>61</sup> Dena G. Hernandez,<sup>62</sup> Albert Hofman,<sup>63</sup> Frank B. Hu,<sup>43,45,60</sup> Thomas Illig,<sup>31</sup> Marjo-Riitta Järvelin, 64 Andrew D. Johnson, 9,65 David Karasik, 66 Kay-Tee Khaw, 67 Douglas P. Kiel, 66 Tuomas O. Kilpeläinen, 1 Ivana Kolcic, 68 Peter Kraft, 43,45,60 Lenore J. Launer, 58 Joop S.E. Laven, 69 Shengxu Li, 1 Jianjun Liu, <sup>70</sup> Daniel Levy, <sup>9,65,71</sup> Nicholas G. Martin, <sup>72</sup> Wendy L. McArdle, <sup>73</sup> Mads Melbye, <sup>18</sup> Vincent Mooser, <sup>74</sup> Jeffrey C. Murray, <sup>75</sup> Sarah S. Murray, <sup>35</sup> Michael A. Nalls, <sup>76</sup> Pau Navarro, <sup>34</sup> Mari Nelis, <sup>15,16,17</sup> Andrew R. Ness, <sup>77</sup> Kate Northstone, <sup>73</sup> Ben A. Oostra, <sup>30</sup> Munro Peacock, <sup>78</sup> Lyle J. Palmer, <sup>37</sup> Aarno Palotie, <sup>14,40,79</sup> Guillaume Paré, <sup>4,5,80</sup> Alex N. Parker, 81 Nancy L. Pedersen, 33 Leena Peltonen, 14,40,52,79,82 Craig E. Pennell, 83 Paul Pharoah, 50 Ozren Polasek, <sup>68,84</sup> Andrew S. Plump, <sup>85</sup> Anneli Pouta, <sup>52</sup> Eleonora Porcu, <sup>25</sup> Thorunn Rafnar, <sup>3</sup> John P. Rice, <sup>23</sup> Susan M. Ring, 73 Fernando Rivadeneira, 11,29,63 Igor Rudan, 38,86 Cinzia Sala, 32 Veikko Salomaa, 52 Serena Sanna, 52 David Schlessinger, 87 Nicholas J. Schork, 35 Angelo Scuteri, 25,88 Ayellet V. Segrè, 79,89 Alan R. Shuldiner, 26,90 Nicole Soranzo, 24,40 Ulla Sovio, 64 Sathanur R. Srinivasan, 41 David P. Strachan, 91 Mar-Liis Tammesoo, 15 Emmi Tikkanen, 14,52 Daniela Toniolo, 32 Kim Tsui, 81 Laufey Tryggvadottir, 92 Jonathon Tyrer, 50 Manuela Uda, 25 Rob M. van Dam, 45,93 Joyve B.J. van Meurs, 11 Peter Vollenweider, 94 Gerard Waeber, 94 Nicholas J. Wareham, 1 Dawn M. Waterworth, <sup>74</sup> Michael N. Weedon, <sup>2</sup> H. Erich Wichmann, <sup>31,95,96</sup> Gonneke Willemsen, <sup>19</sup> James F. Wilson, <sup>38</sup> Alan F. Wright,<sup>34</sup> Lauren Young,<sup>81</sup> Guangju Zhai,<sup>24</sup> Wei Vivian Zhuang,<sup>10</sup> Laura J. Bierut,<sup>23</sup> Dorret I. Boomsma, <sup>19</sup> Heather A. Boyd, <sup>18</sup> Laura Crisponi, <sup>25</sup> Ellen W. Demerath, <sup>57</sup> Cornelia M. van Duijn, <sup>30</sup> Michael J. Econs, <sup>20,78</sup> Tamara B. Harris, <sup>58</sup> David J. Hunter, <sup>43,44,45,60</sup> Ruth J.F. Loos, <sup>1</sup> Andres Metspalu, <sup>15,16,17</sup> Grant W. Montgomery, 97 Paul M. Ridker, 4,5,43,98 Tim D. Spector, 24 Elizabeth A. Streeten, 26 Kari Stefansson, 3,99 Unnur Thorsteinsdottir, 3,99 André G. Uitterlinden, 11,29,63 Elisabeth Widen, Joanne M. Murabito, 9,47 Ken K. Ong, 1,100 and Anna Murray<sup>2</sup>

- 1. Medical Research Council (MRC) Epidemiology Unit, Institute of Metabolic Science, Addenbrooke's Hospital, Cambridge, UK
- 2. Genetics of Complex Traits, Peninsula Medical School, University of Exeter, UK
- 3. deCODE Genetics, Reykjavik, Iceland
- 4. Division of Preventive Medicine, Brigham and Women's Hospital, 900 Commonwealth Avenue East, Boston MA 02215, USA
- 5. Harvard Medical School, Boston, Massachusetts, USA
- 6. Department of Epidemiology, Gillings School of Global Public Health, University of North Carolina at Chapel Hill, Chapel Hill, North Carolina, USA
- 7. Department of Public Health, Indiana University School of Medicine, Indiana, USA
- 8. Melvin and Bren Simon Cancer Center, Indiana University, Indiana, USA
- 9. The National Heart Lung and Blood Institute's Framingham Heart Study, Framingham, MA, USA
- 10. Department of Biostatistics, Boston University School of Public Health, Boston, MA, USA
- 11. Department of Internal Medicine, Erasmus MC, Rotterdam, the Netherlands
- 12. Queensland Statistical Genetics, Queensland Institute of Medical Research, Brisbane, Australia
- 13. The University of Queensland, Brisbane, Australia
- 14. Institute for Molecular Medicine Finland (FIMM), University of Helsinki, Finland
- 15. Estonian Genome Center, University of Tartu, Tartu, Estonia
- 16. Department of Biotechnology, Institute of Molecular and Cell Biology, University of Tartu, Tartu, Estonia
- 17. Genotyping Core Facility, Estonian Biocenter, Tartu, Estonia
- 18. Department of Epidemiology Research, Statens Serum Institut, Copenhagen, Denmark
- 19. Department of Biological Psychology, VU University Amsterdam, Amsterdam, The Netherlands
- 20. Department of Medical and Molecular Genetics, Indiana University School of Medicine, Indiana, USA
- 21. Department of Medical Genetics, University of Lausanne, Lausanne, Switzerland
- 22. Swiss Institute of Bioinformatics, 1015 Lausanne, Switzerland
- 23. Department of Psychiatry, Washington University School of Medicine, St. Louis, MO, USA
- 24. Department of Twin Research and Genetic Epidemiology, King's College London, London, UK
- 25. Istituto di Neurogenetica e Neurofarmacologia, Consiglio Nazionale delle Ricerche, Cagliari, Italy
- 26. Division of Endocrinology, Diabetes and Nutrition, University of Maryland School of Medicine, Baltimore, Maryland, USA
- 27. Icelandic Heart Association, Kopavogur, Iceland
- 28. University of Iceland, Reykjavik, Iceland
- 29. Netherlands Consortium of Healthy Aging, Rotterdam, the Netherlands
- 30. Genetic-Epidemiology Unit, Department of Epidemiology and Department of Clinical Genetics, Erasmus University Medical Center, Rotterdam, The Netherlands
- 31. Institute of Epidemiology, Helmholtz Zentrum München German Research Center for Environmental Health, Neuherberg, Germany
- 32. Division of Genetics and Cell Biology, San Raffaele Scientific Institute, Milan, Italy
- 33. Department of Medical Epidemiology and Biostatistics, Karolinska Institutet, Stockholm, Sweden
- 34. MRC Human Genetics Unit; Institute of Genetics and Molecular Medicine, Western General Hospital; Edinburgh, UK
- 35. Scripps Genomic Medicine, The Scripps Translational Science Institute, and The Scripps Research Institute, La Jolla, CA, USA
- 36. Medical Genetics, Department of Reproductive Sciences and Development, University of Trieste, Irieste, Italy
- 37. Centre for Genetic Epidemiology and Biostatistics University of Western Australia, Australia
- 38. Centre for Population Health Sciences, University of Edinburgh, Teviot Place, Edinburgh, EH8 9AG, Scotland
- 39. Geriatric Unit, Azienda Sanitaria di Firenze, Florence, Italy
- 40. Wellcome Trust Sanger Institute, Hinxton, Cambridge, UK
- 41. Tulane University, New Orleans, LA, USA
- 42. Human Genetics Center, University of Texas Health Science Center at Houston, Houston, Texas, USA
- 43. Department of Epidemiology, Harvard School of Public Health, Boston, MA, USA

- 44. Division of Cancer Epidemiology and Genetics, National Cancer Institute, National Institutes of Health, Department of Health and Human Services, Bethesda, MD 20892, USA
- 45. Department of Nutrition, Harvard School of Public Health, Boston, MA, USA
- 46. Collaborative Studies Coordinating Center, Department of Biostatistics, Gillings School of Global Public Health, University of North Carolina at Chapel Hill, North Carolina, USA
- 47. Sections of General Internal Medicine, Preventive Medicine and Endocrinology, Department of Medicine, Boston University School of Medicine, Boston, MA, USA
- 48. Institute of Environmental Medicine, Karolinska Institutet, Stockholm, Sweden
- 49. MRC Centre for Causal Analyses in Translational Epidemiology, Department of Social Medicine, University of Bristol, BS8 2BN, UK
- 50. Centre for Cancer Genetic Epidemiology, Departments of Oncology and Public Health and Primary Care, University of Cambridge, Cambridge, UK
- 51. MPRI, Merck & Co., Inc, 126 Lincoln Ave, Rahway, NJ 07065, USA
- 52. National Institute for Health and Welfare, Finland
- 53. Department of General Practice and Primary health Care, University of Helsinki, Finland
- 54. Helsinki University Central Hospital, Unit of General Practice, Helsinki, Finland
- 55. Folkhalsan Research Centre, Helsinki, Finland
- 56. Longitudinal Studies Section, Clinical Research Branch, National Institute on Aging, Baltimore, Maryland, USA
- 57. Division of Epidemiology and Community Health, School of Public Health, University of Minnesota, Minneapolis, Minnesota, USA
- 58. Laboratory of Epidemiology, Demography, and Biometry, Intramural Research Program, National Institute on Aging, Bethesda, Maryland, USA
- 59. A full list of members is provided in the Supplementary Note
- 60. Channing Laboratory, Department of Medicine, Brigham and Women's Hospital, and Harvard Medical School, Boston, Massachusetts, USA
- 61. Department of Psychiatry, Washington University School of Medicine, St. Louis, Missouri, USA
- 62. Laboratory of Neurogenetics, National Institute of Ageing, Bethesda, MD, USA
- 63. Department of Epidemiology, Erasmus MC, Rotterdam, the Netherlands
- 64. Department of Epidemiology and Biostatistics, School of Public Health, Imperial College London, London, UK
- 65. NHLBI Center for Population Studies, Bethesda, MD, USA
- 66. Hebrew SeniorLife Institute for Aging Research and Harvard Medical School, Boston, MA, USA
- 67. Department of Public Health and Primary Care, Institute of Public Health, University of Cambridge, Cambridge, CB2 0QQ, UK
- 68. Medical School; University of Zagreb; Zagreb, 10000; Croatia
- 69. Department of Obstetrics and Gynaecology, Erasmus MC, Rotterdam, the Netherlands
- 70. Human Genetics, Genome Institute of Singapore, Singapore
- 71. Division of Cardiology, Boston University School of Medicine, USA
- 72. Genetic Epidemiology, Queensland Institute of Medical Research, Brisbane, Australia
- 73. Avon Longitudinal Study of Parents and Children (ALSPAC), Department of Social Medicine, University of Bristol, BS8 2BN, UK
- 74. Genetics Division, GlaxoSmithKline, King of Prussia, Pennsylvania, USA
- 75. Department of Pediatrics, University of Iowa, Iowa City, IA, USA
- 76. Laboratory of Neurogenetics, Intramural Research Program, National Institute on Aging, Bethesda, Maryland, USA
- 77. Department of Oral and Dental Science, University of Bristol, BS1 2LY, UK
- 78. Department of Medicine, Indiana University School of Medicine, Indiana, USA
- 79. Broad Institute of Harvard and MIT, Cambridge, Massachusetts, USA
- 80. Genetic and Molecular Epidemiology Laboratory, McMaster University, 1200 Main St. W MDCL Rm. 3206, Hamilton, ON, L8N3Z5, Canada
- 81. Amgen, 1 Kendall Square, Building 100, Cambridge, MA 02139, USA
- 82. Deceased
- 83. School of Women's and Infants' Health, The University of Western Australia, Australia
- 84. Gen Info Ltd; Zagreb, 10000; Croatia
- 85. Cardiovascular Disease, Merck Research Laboratory, Rahway, NJ 07065, USA
- 86. Croatian Centre for Global Health; University of Split Medical School; Split, 21000; Croatia
- 87. Gerontology Research Center, National Institute on Aging, Baltimore, Maryland, USA

- 88. UOC Geriatria Istituto Nazionale Ricovero e Cura per Anziani IRCCS Rome, Italy
- 89. Department of Molecular Biology, Massachusetts General Hospital, Boston, Massachusetts, USA
- 90. Geriatric Research and Education Clinical Center, Veterans Administration Medical Center, Baltimore, Maryland, USA
- 91. Division of Community Health Sciences, St. George's, University of London, London, UK
- 92. Icelandic Cancer Registry, Reykjavik, Iceland
- 93. Departments of Epidemiology and Public Health and Medicine, Yong Loo Lin School of Medicine, National University of Singapore, Singapore
- 94. Department of Internal Medicine, BH-10 Centre Hospitalier Universitaire Vaudois (CHUV), Lausanne, Switzerland
- 95. Institute of Medical Informatics, Biometry and Epidemiology, Chair of Epidemiology, Ludwig-Maximilians-Universität, Munich, Germany
- 96. Klinikum Grosshadern, Munich, Germany
- 97. Molecular Epidemiology, Queensland Institute of Medical Research, Brisbane, Australia
- 98. Division of Cardiology, Brigham and Women's Hospital
- 99. Faculty of Medicine, University of Iceland, Reykjavik, Iceland
- 100. Department of Paediatrics, University of Cambridge, Cambridge, UK

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