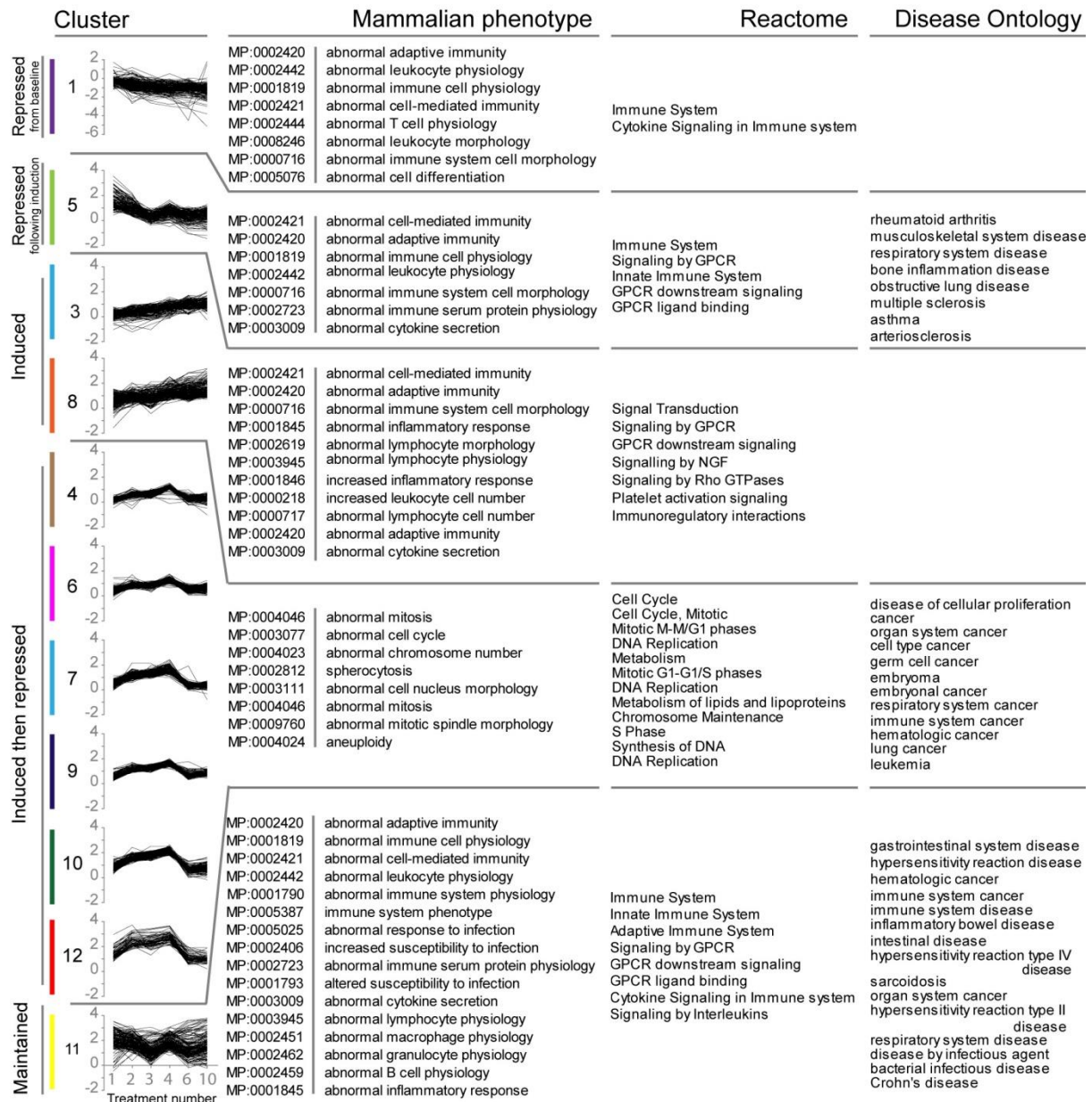
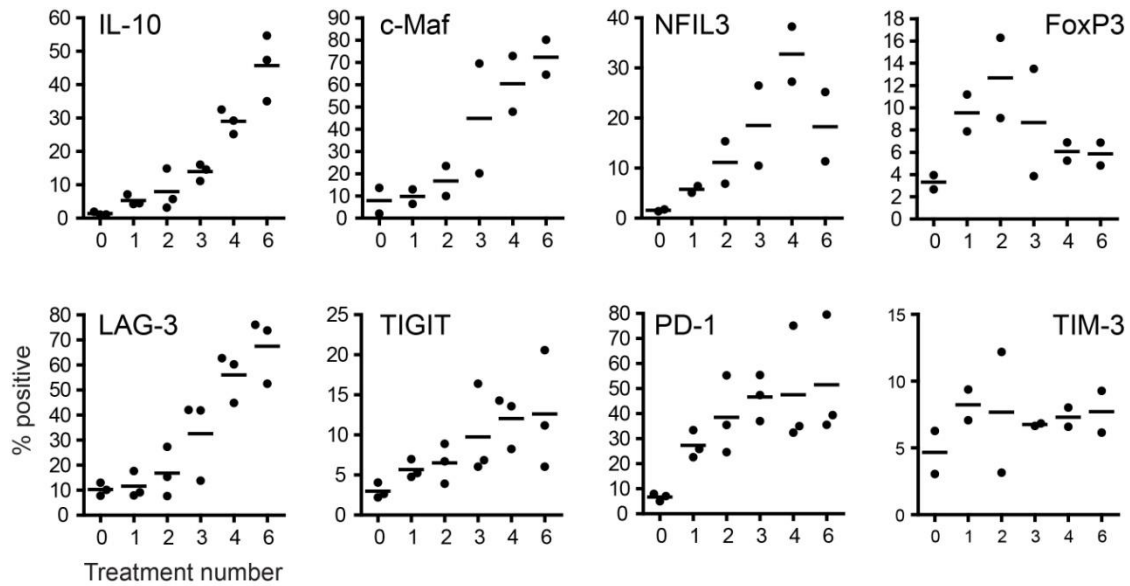


Supplementary Fig. 1 Dose escalation enhances the induction of IL-10⁺ CD4⁺ T cells in addition to minimising the risk of adverse effects during immunotherapy. Tg4 mice were treated s.c. with 3x8 μ g doses of MBP Ac1-9[4Y], with or without prior dose escalation, as illustrated (a). Splenocytes were cultured for five days with 10 μ g ml⁻¹ MBP Ac1-9[4K] and 20U ml⁻¹ IL-2. (b) Scatter plots show the percentage of viable CD4⁺ cells which are IL-10⁺, horizontal lines show means for each column ($n=3$). One of two experiments. * $P \leq 0.05$, ** $P \leq 0.01$ one-way ANOVA with Bonferroni post-test, comparing peptide-treated groups with PBS-treated control group. (c) Onset and severity of EAE in mice treated as illustrated in (a), immunised with MBP Ac1-9[4K]/CFA and pertussis toxin. Results of two independent experiments are pooled, showing mean disease score \pm SEM (PBS group and 8 μ g, with escalation, both $n=12$. 8 μ g, no escalation $n=10$).

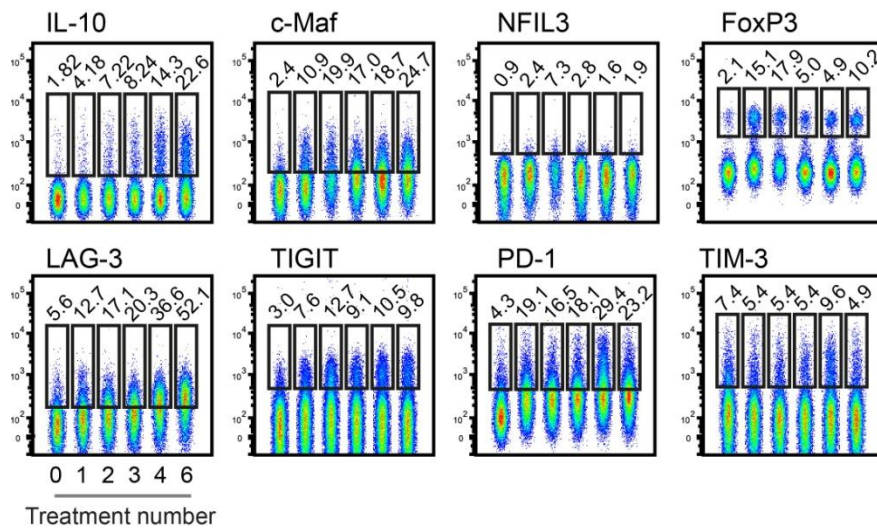


Supplementary Fig. 2 Ontology terms enriched in clusters of co-regulated CD4⁺ T cell genes during dose escalation immunotherapy. Tg4 Rag-1^{-/-} mice were EDI-treated s.c. with MBP Ac1-9[4Y]. CD4⁺ T cell transcriptome analysis was undertaken at the indicated treatment stages and transcripts were grouped by two-phase SOM clustering (see Fig. 6). Expression pattern and amplitude of expression of transcripts assigned to the twelve color-coded gene clusters described in Fig. 6b are here represented by line profiles. Significantly enriched, immunologically relevant annotations from Mammalian Phenotype, Reactome and Disease Ontology databases are shown (FDR <10⁻⁵ for MP, <5x10⁻² for Reactome and DO). Where no terms are shown, no immunologically relevant terms were enriched.



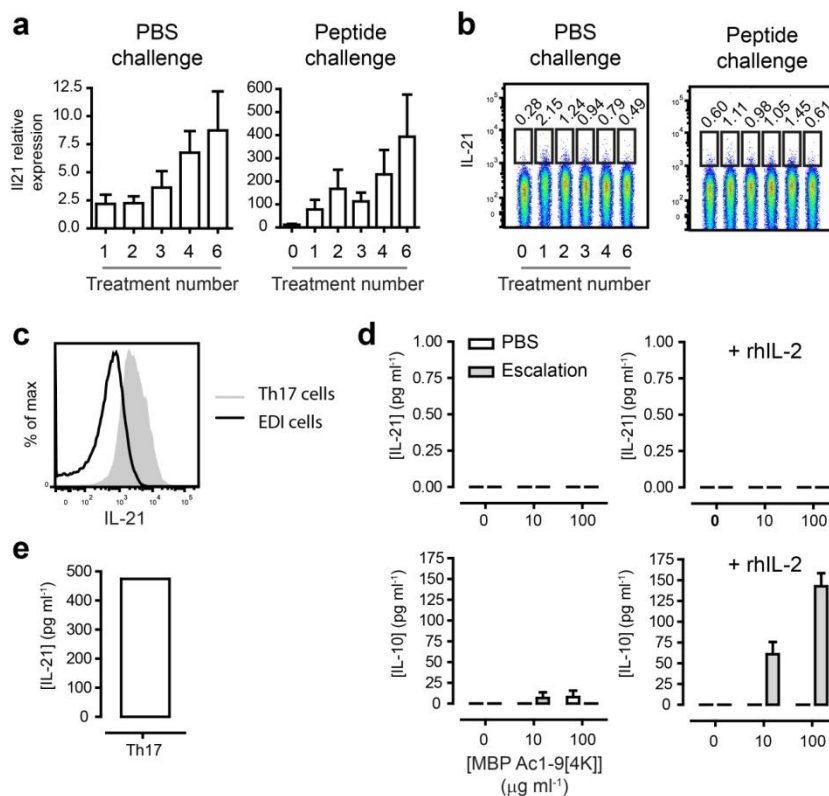
Supplementary Fig. 3 CD4⁺ T cell signature induced by dose escalation

immunotherapy. Tg4 mice were treated s.c. with an escalating dose of MBP Ac1-9[4Y] (0.08 μ g \rightarrow 0.8 μ g \rightarrow 8 μ g \rightarrow 3x80 μ g). Cells were harvested two hours after peptide challenge *in vivo*. FACS staining of IL-10, c-Maf, NFIL-3, FoxP3, LAG-3, TIGIT, PD-1 and TIM-3 in CD4⁺ T cells at the indicated stages of treatment. Scatter plots show the percentage of viable CD4⁺ cells from individual mice which express the signature proteins, horizontal lines show means for each treatment group. Pooled results from two-three independent experiments.



Supplementary Fig. 4 CD4⁺ T cell signature induced by dose escalation immunotherapy is maintained in resting cells.

Tg4 mice were treated s.c. with an escalating dose of MBP Ac1-9[4Y] (0.08µg → 0.8µg → 8µg → 3x80µg). FACS staining of IL-10, c-Maf, NFIL-3, FoxP3, LAG-3, TIGIT, PD-1 and TIM-3 in CD4⁺ T cells at the indicated stages of treatment. Resting cells were harvested two hours after PBS-challenge *in vivo*, 4 days after the indicated peptide treatment. Data are representative of two-three independent experiments.



Supplementary Fig. 5 IL-21 mRNA but not protein is produced by CD4⁺ T cells during dose escalation immunotherapy. Tg4 mice were treated s.c. with an escalating dose of MBP Ac1-9[4Y] (0.08µg → 0.8µg → 8µg → 3x80µg). (a) Quantitative RT-PCR analysis of *Il21* mRNA in CD4⁺ T cells at the indicated stages of treatment. Graphs show mean expression values + SEM for three replicate experiments, pooled ($n=3$ total). (b) IL-21 expression by CD4⁺ T cells, detected by FACS at the indicated stages of treatment. (c) FACS-based comparison of IL-21 protein expression by CD4⁺ T cells from EDI-treated mice (as above) versus Th17-polarised CD4⁺ T cells as a positive control. Representative of three similar experiments (d) CD4⁺ T cells from EDI- or PBS-treated mice were re-stimulated in the presence of irradiated APCs and a titration of MBP Ac1-9[4K]. Supernatant was collected after 72 hours to detect IL-21 and IL-10 levels by MFBI. Representative of three similar experiments, error bars show + SEM of at least two independent biological replicates, each assayed in duplicate. (e) IL-21 in the supernatant of Th17-polarised CD4⁺ T cells, detected by MFBI (positive control). Representative of two experiments, assayed in duplicate.