

Supplementary Data

Arabidopsis Response to Low-Phosphate Conditions Includes Active Changes in Actin Filaments and PIN2 Polarization and Is Dependent on Strigolactone Signaling

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Figure legends

Figure S1. PIN2 plasma-membrane localization and polarity in epidermal cells of the primary-root elongation zone in PIN2::PIN2–GFP seedlings grown under high (2 mM) and low (1 μ M) phosphate (Pi) conditions (120 h postgermination).

(A–D) PIN2–GFP signal in roots of Col-0 (A, B) and *max2-1* (*max2-1-22*; C, D) seedlings grown on high-Pi plates (A, C) and low-Pi plates (B, D) (scale bars = 50 μ m).

(E) Intensity of the PIN2–GFP signal in the apical plasma membrane of Col-0 and *max2-1-22* roots grown on high-Pi plates (white columns) and low-Pi plates (black columns).

(F) Polarity of the PIN2–GFP signal in the plasma membrane of Col-0 and *max2-1-22* roots grown on high-Pi plates (white columns) and low-Pi plates (black columns). Polarity index was determined as the ratio of intensity on the polar versus lateral sides, divided by 2 (Dhonukshe *et al.*, 2008).

Cells (n = 50–60) from 10 plants were examined per each of three replicates. Different lowercase letters above the bars indicate statistically significant differences between means by one-way ANOVA pairwise Student's t-test ($P \leq 0.05$). Arrows point toward the roots.

Figure S2. AUX1 plasma-membrane localization in epidermal cells of the primary-root elongation zone in AUX1::AUX1–YFP seedlings grown under high (2 mM) and low (1 μ M) phosphate (Pi) conditions (48 h postgermination).

(A–B) AUX1–YFP signal in roots of Col-0 (A, B) seedlings grown on high-Pi plates (A) and low-Pi plates (B) (scale bars = 50 μ m).

(C) Intensity of the AUX1–YFP signal in the apical plasma membrane of Col-0 roots grown on high-Pi plates (white columns) and low-Pi plates (black columns).

Cells (n = 50–60) from 10 plants were examined per each of three replicates. Different lowercase letters above the bars indicate statistically significant differences between means by one-way ANOVA pairwise Student's t-test ($P \leq 0.05$). Arrows point toward the roots.

Figure S3. Quantification of PIN2-containing BFA bodies in the epidermal cells of the primary-root elongation zone in PIN2::PIN2–GFP seedlings grown under high (2 mM) and low (1 μ M) phosphate (Pi) conditions (120 h postgermination)

(A–D) PIN2-containing BFA bodies (green – PIN–GFP signal; red – FM4-64) in Col-0 (A, B) and *max2-1-22* (C, D) roots grown on high-Pi plates (A, C) and low-Pi plates (B, D) and treated with BFA (100 μ M, 1 h) (scale bars = 50 μ m).

(E) Number of PIN2-containing BFA bodies per cell in the apical plasma membrane of Col-0 and *max2-1-22* roots grown on high-Pi plates (white columns) and low-Pi plates (black columns) and treated with BFA (100 μ M, 1 h).

(F) Percentage of cells with PIN2-containing BFA bodies in the apical plasma membrane of Col-0 and *max2-1-22* roots grown on high-Pi plates (white columns) and low-Pi plates (black columns) and treated with BFA (100 μ M, 1 h).

Cells (n = 50–60) from 10 plants were examined per each of three replicates. Different lowercase letters above the bars indicate statistically significant differences between means by one-way ANOVA pairwise Student's t-test ($P \leq 0.05$). Arrows indicate PIN2-containing BFA bodies.

Figure S1.

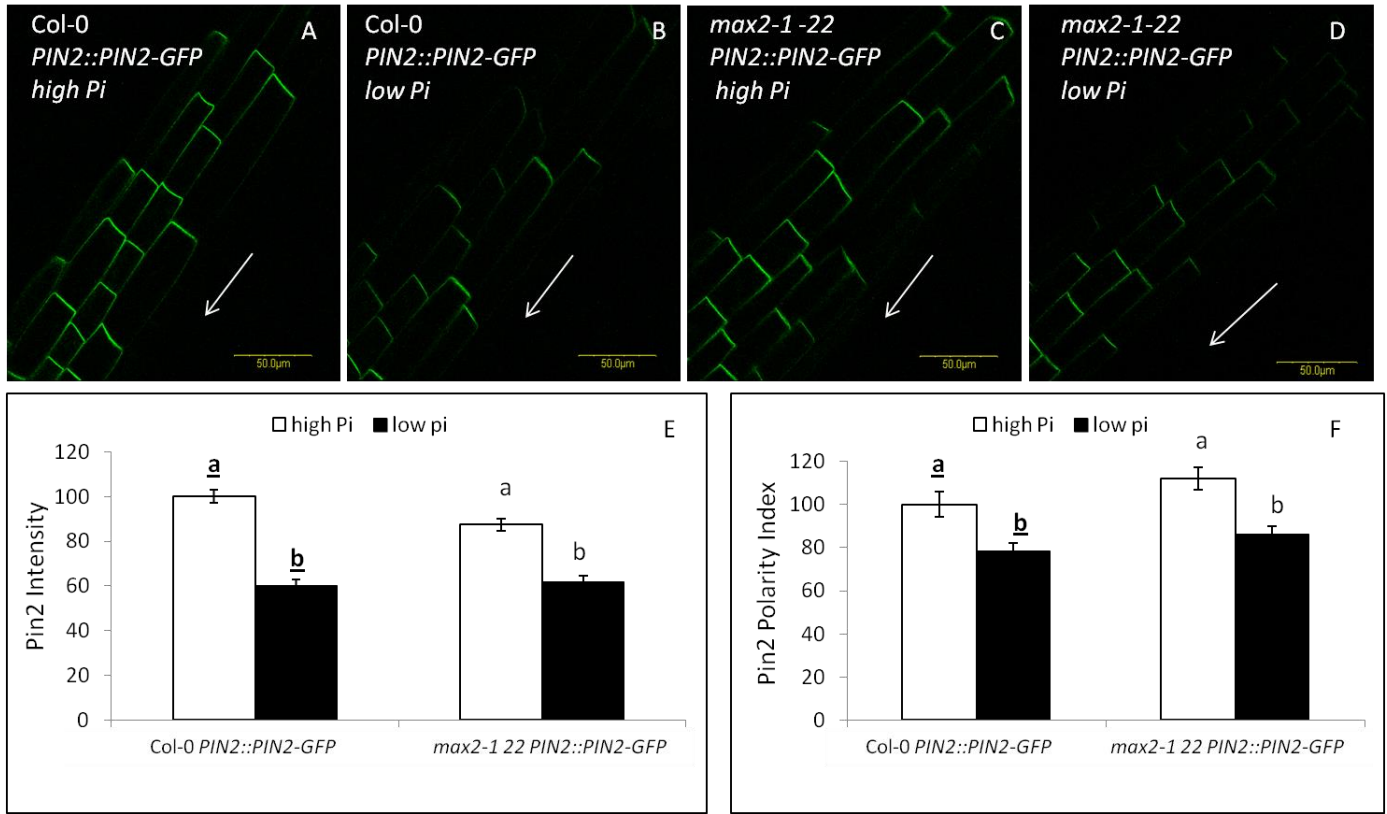


Figure S2.

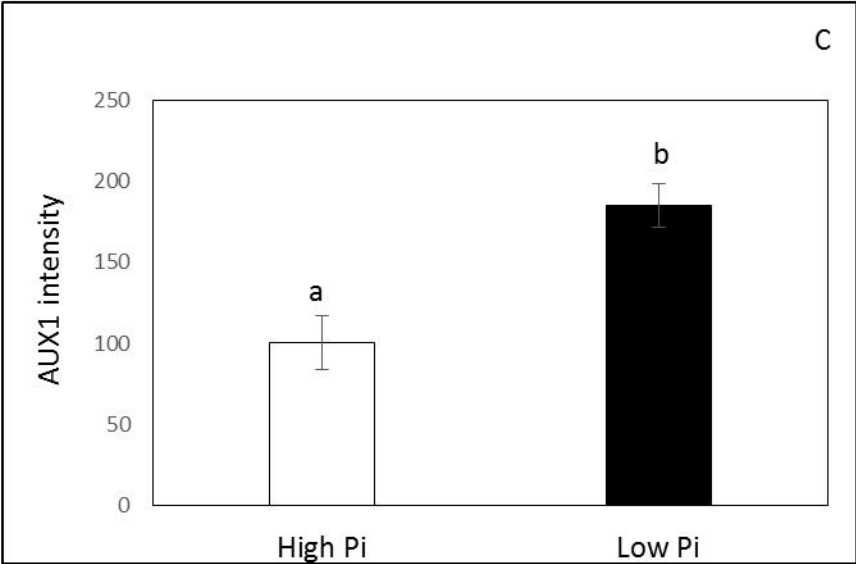
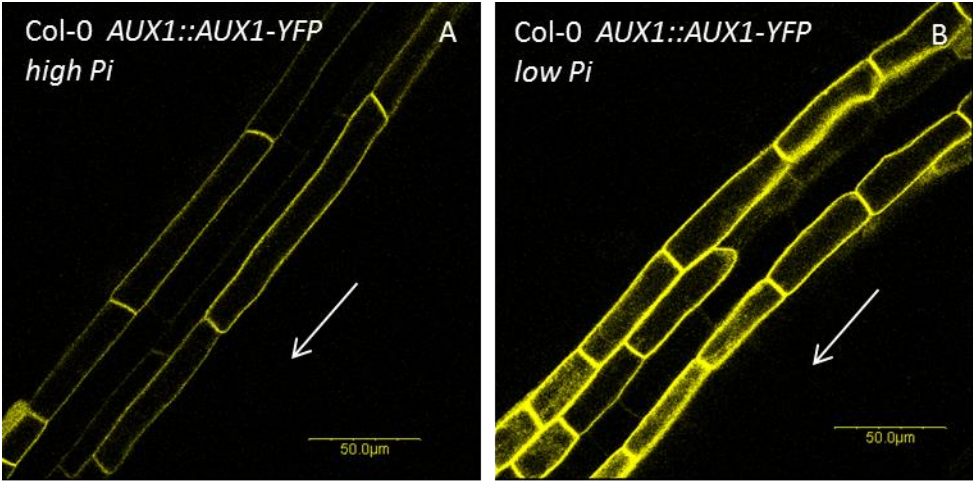


Figure S3.

