

Supplementary Material

Comparative sequence analysis of Wzb, Wzd and Wze

Among four strains of *Lb. rhamnosus* (ATCC 9595, RW-9595M, R, RW-6541M), Wzb is 100 % identical for three strains while a one amino acid change (T53 to A) occurs for the Wzb of strain RW-6541M [GenBank: AY659977] [1]. Phosphoesterase activity was predicted for Wzb, based on alignment of deduced amino acid sequences with PHP superfamily members (PHP domain, PF02811). Four conserved motifs of the PHP (polymerase and histidinol phosphatase) domain identified by Aravind & Koonin [2] were located in the amino acid sequence of Wzb. These motifs consist of conserved histidine and aspartic acid residues. Previous work [3] has demonstrated the phosphatase activity of Wzb.

Wzd is a 34 kDa protein coded by the 915 bp *wzd* gene located at the beginning of the gene clusters coding for EPS biosynthesis in *Lb. rhamnosus* ATCC 9595, just downstream of promoter P1 [1]. Hydrophobicity analysis predicts two transmembrane domains (amino acids 18 to 36 and 219 to 238, TMpredTM; [4]; http://www.ch.embnet.org/software/TMPRED_form.html), which were confirmed by SAPS (Statistical Analysis of Protein Sequences; [5]: http://www.isrec.isb-sib.ch/software/SAPS_form.html). In addition, this protein contains a coiled-coil region (not a leucine zipper) in the C-terminal intracellular region (amino acids 263 to 290). Comparison of the hydrophobicity profile of Wzd with that of CpsC from *S. pneumoniae* Rx1-19F (Fig. S1) suggests the function of co-polymerase. Comparison with similar proteins from other bacterial genera showed that Wzd corresponds to the N-terminal region of Wzc et ExoP, respectively from *E. coli* [6] et *S. meliloti* [7] and is an ortholog of EpsC from *S. thermophilus* as well as EpsA from *L. lactis*. There are three classes of polysaccharide co-polymerases (PCPs). The PCP1 are

associated with LPS chain length regulation [8, 9]. The PCP2 is involved in the synthesis of high molecular weight polysaccharides, such as CPS and EPS. Both PCP1 and PCP2 type proteins are involved in the Wzy dependent mechanism of polysaccharide biosynthesis. The last group, PCP3, participates in the biosynthesis of CPS via a mechanism employing an ABC (ATP Binding Cassette) transporter. The PCP2 proteins are divided into two subgroups: PCP2a (Gram negative bacteria) and PCP2b (Gram positive bacteria). The PCP2b subgroup includes CpsC (co-polymerase) from *S. pneumoniae* [9], the PCPs from *Staphylococcus aureus*, *S. thermophilus* and *L. lactis* [10], along with PCP from *Lb. rhamnosus* ATCC 9595 and RW-9595M (Fig S2). The alignment shows that PCP2b sequences may be further divided into three subgroups (1-lactobacilli; 2-lactococci; 3-staphylococci/streptococci). PCP2b from lactococci and lactobacilli have one tyrosine residue after the trans-membrane domain in the C-terminal region and a potential coiled-coil region, which are absent from staphylococci and streptococci. Furthermore, the PCP2b sequences from lactobacilli can be differentiated from those of lactococci, staphylococci and streptococci by the presence of a 46 amino acid sequence containing five tyrosine residues that are highly conserved within four of the five *Lactobacillus* putative PCP2b proteins examined. The only tyrosine that is present in almost all of the PCP2b sequences examined (12 out of 13 sequences; Fig. S2) is the one located in the N-terminal region just following the first transmembrane sequence.

Wze is coded by the second gene of the locus responsible for EPS biosynthesis and is cotranscribed with *wzd*. The 753 bp *wze* gene codes for a 27 kDa protein which is predicted by TMpredTM to be cytoplasmic due to the absence of any transmembrane sequences. Wze has an ATP-binding site consisting of a Walker A sequence ([AG]X₄GK[ST] where X is any amino acid and two Walker B sequences (hhhD, where h represents a hydrophobic amino acid) in the N-

terminus and a C-terminus containing a tyrosine-rich region. The Walker A site is considered a common structural motif for prokaryotic protein kinases [11]. Finally, Wze has a tyrosine residue similar to the tyrosine 569 which was described as being implicated in autophosphorylation of Wzc in *E. coli* [12]. Alignment of this sequence with proteins proposed to have a role in polysaccharide production suggests that Wze would function as a protein tyrosine kinase, orthologous to CpsD from *S. pneumoniae* Rx1-19F, to EpsD from *S. thermophilus* Sfi6 and EpsB from *L. lactis* subsp. *cremoris* [13]. In addition, Wze corresponds to the C-terminus of the Wzc protein from *E. coli* [6] as well as ExoP from *S. meliloti* [7].

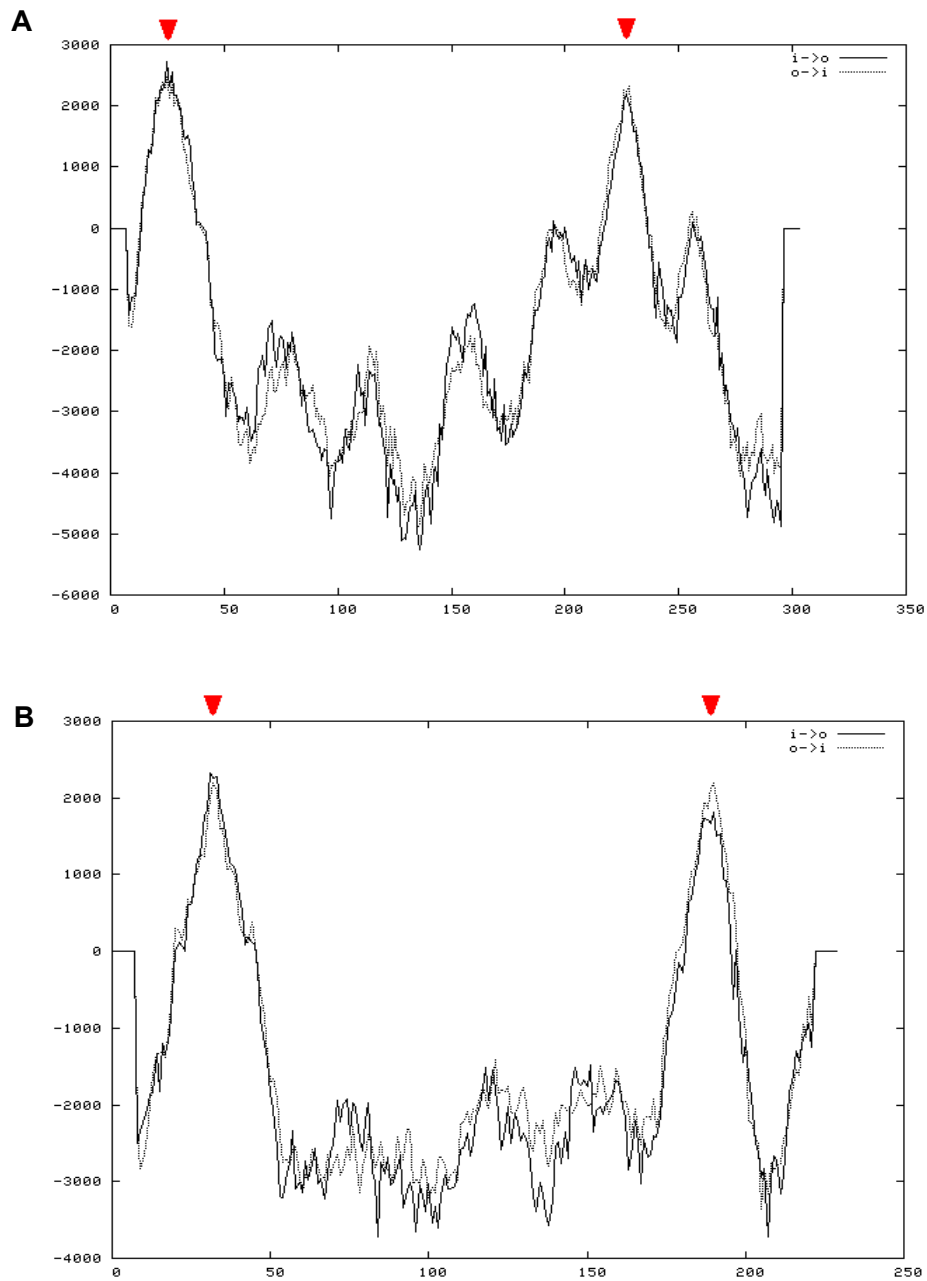


Figure S1. Comparison of transmembrane helix score plot of Wzd (A) from *L. rhamnosus* and CpsC19f (B) [GenBank: U09239].

Arrows indicate transmembrane domains (TMS1, 2). Transmembrane prediction was carried out with TMpred (http://www.ch.embnet.org/software/TMPRED_form.html). The preferred orientation is in-out for TMS1 and out-in for TMS2.

EpsA B40 --MQETQEQTIDLRGIFKIIIRKRLGLILFSAIIVTILGSIYTFFFIASPVYTASTQLVVVKL 58
EpsA --MQETQEQTIDLRGIFKIIIRKRLGLILFSAIIVTILGSIYTFFFIASPVYTASTQLVVVKL 58
EpsA KF --MQETQEQTIDLRGIFKIIIRKRLGLILFSAIIVTILGGIYTFFFIASPVYTASTQLVVVKL 58
CapA -----MKEKFDLVKLLNLLKKNIKLLLILPAICLVVSAALTFVMPDKYTASTQILVNM 54
Wzd GG -----MNKQIDLSQLWNVFKRSFLAMVIFGILGLAAAYFGAKAFIAPKYESDTSLLVNR 54
EpsC P -----MNKQIDLSQLWNVFKRSFLAMVIFGILGLAAAYFGAKTFIAPKYESDTSLLVNR 54
EpsC -----MNKQIDLSQLWNVFKRSFVAMIILGILGMAAAAYFGAKTFIAPKYESDTSLLVNR 54
Wzd -----MNEQIDLARLWNVFKHSFIVMILLGLLGMFIAYFGAKTFIAPKYSASTSMLVNR 54
EpsB MENSTKTENTIDLRRLWMLLR~~AHIWSIILWAI~~GLGAVGFVLA~~AFVVEPKY~~TSTTQILVNQ 60
CpsC D39 MKEQNT--IEIDVFQLFKTLWKRKLMILLVALVTGAGAFAYSTFI~~Y~~VKPEY~~T~~STTRI~~Y~~VVN 58
CpsC MKEQNT--LEIDVLQLFRALWKRKLVILLVAITSSVAFAYSTFVIKPEFTSMTRI~~Y~~VVN 58
Wzd S MNQDNTKSDEIDVLALLHKLWTKKLLILFTAFYFAVFSFLGT~~Y~~FFI~~Q~~PT~~Y~~TSTTRI~~Y~~VVN 60
EpsC S MNQDNTKSDEIDVLALLHKLWTKKLLILFTAFYFAVFSFLGT~~Y~~FFI~~Q~~PT~~Y~~TSTTRI~~Y~~VVN 60
* : : : : . : . : : * : *

EpsA B40 PN-SDNSAAYAGEVTGNIQMANTINQVIVSPVILDKVRSNLN----- 99
EpsA PN-SDNSAAYAGEVTGNIQMANTINQVIVSPVILDKVQSNLN----- 99
EpsA KF PN-SDNSAAYAGQVTGNIQMANTINQVIVSPVILDKVQSNLN----- 99
CapA KK-SSSDLAFQ-NVQSSLSVNTYTEI~~I~~KSPRILDKVSREFDGG----- 96
Wzd GG KQ-DNDPNMQLNAQQADIQIINTYKDIITRPVVLQAVAEDLTSPQRVMVKKAKPAVY~~G~~TR 113
EpsC P KQ-DNDPNMQLNAQQADIQIINTYKDIITRPVVLQAVAEDLTSPQRVMVKKSKPAVY~~G~~TR 113
EpsC KQ-DNDPNMQLNAQQADIQIINTYKDIITRPVLEAVADDLTSPQRVMVKKAKKAVY~~G~~TR 113
Wzd KQ-DNNPNMQLNAQQADIQIINTYKDIITRPVILREVADDLTSPRRVKVKAQKAVY~~G~~TR 113
EpsB KRNAVDAQAYNAQQADVQVINTYKDIIVTSVILKDKASKWIKNP-TEVVKPAKKAKY~~K~~TL 120
CpsC D39 RNQGDKSLTNQDLQAGSYLVKDYREIILSQDALEK~~VATNLKLD~~----- 102
CpsC RDQGEKSLTNQDLQAGSSLVKDYREIILSQDVLEEVVSDLKLD----- 102
Wzd S QATDNKN-LSAQDLQAGTYLANDYKEIITSNDVLSEVIKDEKLN----- 103
EpsC S QATDNKN-LSAQDLQAGTYLANDYKEIIASNDVLSEVIKDEKLN----- 103
. . . : : : * .

EpsA B40 -----LSDDSFQKQVTAANQTN~~S~~QVIMLT~~V~~KY~~S~~NP 129
EpsA -----LSDDSFQKQVTAANQTN~~S~~QVIMLT~~V~~KY~~S~~NP 129
EpsA KF -----LSDDSFQKQVTAANQTN~~S~~QVITL~~T~~VKY~~S~~NP 129
CapA -----YSTAELNSFLKVTNQTNSQIITVSVTTGNK 126
Wzd GG YNAVVTGTRERYVTEEAQPAKYKLPKAY~~S~~NISEEDLTKMVS~~V~~STQ~~Q~~NSQVFTVNVKDTSP 173
EpsC P YNSVTGVRERYVTEKAQPAEYKLEPAKY~~S~~NISEEDLTKMVS~~V~~STQ~~Q~~NSQVFTVNVKDTSP 173
EpsC YNAT~~T~~G~~V~~REEYVAEKAQPAKYKLRPAKY~~S~~NISADDLAKIVSVSTQ~~Q~~NSQVFTVNV~~R~~DTDP 173
Wzd YNAATGVRERYVVKEEQPAKYKLPKAY~~S~~NISEDDLDKMI~~S~~VSN~~A~~QNSQVFTVNV~~R~~DTDP 173
EpsB ADGTK-----KLVRPAEP~~AVIR~~RAGRYN-VSAKEMQKAVSVTTQ~~Q~~SQVFTI~~S~~AKSNDP 173
CpsC D39 -----MPAKTLASKVQVTVPTDTRIVSISVKDKQP 132
CpsC -----LTPKDLANKIKVTV~~P~~VDTRIVSVSVSDRVP 132
Wzd S -----LSEAELSKMVS~~V~~NIPTDTRLISISVNAKTG 133
EpsC S -----LSEAELSKMVS~~V~~NIPTDTRLISISVNAKTG 133
. : . : . : : : . . .

EpsA B40 YIAK~~K~~IADE~~T~~AKIFSSDAAKLLNVTNVN~~I~~LSKAKAQ~~T~~TPISPKPKLYLAISV~~I~~AGLV~~L~~GL 189
EpsA YIAK~~K~~IADE~~T~~AKIFSSDAAKLLNVTNVN~~I~~LSKAKAQ~~T~~TPISPKPKLYLAISV~~I~~AGLV~~L~~GL 189
EpsA KF YIAQ~~K~~IADE~~T~~AKIFSSDAAKLLNVTNVN~~I~~LSKAKAQ~~T~~TPISPKPKLYLAISV~~I~~GGLV~~L~~GL 189
CapA SESDKIVNKISKVFAHDMPKIMSVDNVTILSSAHDNAVKVSPIVSVNLV~~I~~SII~~V~~GIVLAI 186
Wzd GG VRARDIANEIAKVFEKKIAKIMSISNVSVVSRATANTIPVSPK~~L~~LSIVGLALGILIAL 233
EpsC P VRARDIANEIANVFEKKIAKIMSISNVSVVSKATADPTPVSPK~~L~~NLAGLVGLLFGILFAF 233
EpsC LRARDIANEIAKVFEKKIATIMSISNVSVVSKATATSTPVSPR~~L~~KLMTIVGLVLGVLVAF 233
Wzd VRAKDVANEIAKVFKAKIASIMS~~V~~SNVSVSRATADPTPVTPNLKIASLIGLILGMVLA 233
EpsB EKSQAIANAVAQTFFKNKIKS~~I~~MNVNNTIVSPASVG-AKTFPKT~~T~~FLTLAGVVLGLIISV 233
CpsC D39 EEASRIANSLREVAVEKIVAVTRVSDVTTLEE~~A~~R~~P~~ATTPSSPNVRRNSLFGFLGGAVTV 192
CpsC EEASRIANSLREVA~~A~~QKIIISITRVSDVTTLEE~~A~~R~~P~~ATSPSSPNIKRSTLIGFLAGVIGTS 192
Wzd S QDAQTLANKVREVASKKIKKVT~~K~~VEDVTTLEEAKLPES~~P~~SSPNIKLN~~V~~LLGAVLGGFLAV 193
EpsC S QDAQTLANKVREVASKKIKKVT~~K~~VEDVTTLEEAKLPES~~P~~SSPNIKLN~~V~~LLGAVLGGFLAV 193
: : : : . : : * . . * . * .

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EpsA B40 AIALLKELFDNKINKEEDIEA-LGLTVLGVTSYDQMSDFNKNTNKNGTQSGTKSSPPSDH 248
EpsA AIALLKELFDNKINKEEDIEA-LGLTVLGVTSYDQMSDFNKNTNKNGTQSGTKSSPPSDH 248
EpsA KF AIALLQELFDNKINKEEDIEA-LGLTVLGVTTYAQMSDFNKNTNKNGTQMGTKSSPPSDH 248
CapA LIIFLKELLDKRIKTEEDVESQLGLPILGS-----IQKF----- 220
Wzd GG SWGLVRELTLDQTIKIDIDFITDDLGLVNLGIVNYVVRMKDMDQAIQQSRATDSGNDVQDDL 293
EpsC P AWGLIRDLDQTIKEIDFITDGLVLDLAVNYVRRMKDMDQAIIEESKTKLQNNSDSFED 293
EpsC IWGLIRELTLDQTIKIDIDFITDDLGLVNLGIVNYVQRMKMDQAIIEAVKSNEDNN----DF 289
Wzd TVGLIRELTLDQTIKSIDFITNDLGLVNLGLVNYVQRMNDMDEAIARSKNKIIDS--EAEP 291
EpsB ALIILRDSFNTTVRDDDYLTKELGLTNLGHVSHFHLNKF SINNNNDN----- 279
CpsC D39 IAVLLIIEIFDTRVKRPEDIEDVLQIPLLGLVPDLDKMK----- 230
CpsC VIVLILELLDTRVKRPKDIEDTLQMTLLGIVPNLNKLLK----- 230
Wzd S VGVLVREILDDRVRRPEDVEDALGMTLLGIVPDTDKI----- 230
EpsC S VGVLVREILDDRVRRPEDVEDALGMTLLGIVPDTDKI----- 230
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EpsA B40 EVNRSSKRNR-- 259
EpsA EVNRSSKRNR-- 259
EpsA KF EVNRSSKRNR-- 259
CapA -----
Wzd GG DGIDFPQRSRRRI 306
EpsC P EEPDFPRRSRRRV 306
EpsC GEADFPQRSRRRI 302
Wzd ETTGFPQRSRRRV 304
EpsB ---SFGKKRRV-- 287
CpsC D39 -----
CpsC -----
Wzd S -----
EpsC S -----

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Figure S2. Alignment of polysaccharide co-polymerases of thePCP2b subclass.

Wzd from *L. rhamnosus* RW-9595M and ATCC 9595 (Wzd) [GenBank: AAW22487.1] were compared with homologous proteins including Wzd from *L. rhamnosus* GG (Wzd GG) [GenBank: CAR87948.1], EpsC from *L. paracasei* (EpsC P) [GenBank: WP_016383925], EpsC from *L. casei* LOCK919 (EpsC) [GenBank: YP_008200731], EpsB from *L. johnsonii* F19785 (EpsB) [GenBank: CAX67043.1], EpsA from *L. lactis* subsp. cremoris JFR1 (EpsA) [GenBank: AER51660.1] or NIZO B40 (EpsA B40) [GenBank: NP_053033.1], EpsA from *L. lactis* subsp. lactis KF147 (EpsA KF) [GenBank: ABX75679.1], CapA from *Staphylococcus aureus* (CapA) [GenBank: AAD52053.1], Wzd from *S. thermophilus* MR-2C (Wzd S) [GenBank: AAL32496.1], EpsC from *S. thermophilus* Sfi6 (EpsC S) [GenBank: AAC44010.1] and CpsC from *S. pneumoniae* D39 (CpsC D39) [GenBank: ABJ55335.1] or from *S. pneumoniae* Rx1-19F (CpsC)

[GenBank: AAC44960.1]. Regions corresponding to the motifs associated with transmembrane sequences are highlighted in yellow and the coiled-coil region is underlined. Tyrosines are in red. Alignments were performed using the program ClustalW2 (<http://www.ebi.ac.uk/Tools/msa/clustalw2/>) and transmembrane prediction was carried out with TMPred (http://www.ch.embnet.org/software/TMPRED_form.html).

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