

Defining the sequence requirements for the positioning of base J in DNA using SMRT sequencing

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SUPPLEMENTARY DATA

Supplementary Figure S1. Selection of IPD ratio cut-off for calling Js. Based on the synthetic oligonucleotides containing J, any set of peaks with an IPD ratio >1.6 at a T position or at 2 or 6 nucleotides downstream from a T, is scored as J. The 1.6 cut-off was chosen to minimize false-positive and false-negative rates (FPR, FNR). **(A)** The distribution of single molecule IPD ratios (in 0.1 bins) at all positions in the J-less 25.2S plasmid (grown in the JBP2-null strain, blue bars) was subtracted from the total distribution from all samples at all positions (red bars), to estimate the distribution of IPD ratios due to J (green bars). FPR was defined as the percentage of IPD ratios greater than the cut-off value in the JBP2-null sample; FNR was defined as the percentage of IPD ratios smaller than the cut-off value in the “Difference” population due to J. **(B)** A cut-off of 1.6 was chosen to minimize the estimated false positive and false negative rates (FPR and FNR).

Supplementary Figure S2. Partial DNA digest with restriction enzyme DdeI. Supplementary Figure S2 shows blots of plasmid digests with restriction endonuclease DdeI, hybridized with probes corresponding to the genomic *Leishmania* DNA, inserted into the plasmid. Here we present our interpretation of the main partial digest products observed. This interpretation is based on the assumption that all partial digest products are due to a partial J-modification of the DdeI sites investigated, as we do not see these products in plasmid DNA from *E. coli*. **(A)** Overview of the DdeI restriction sites in the 25.2L plasmid and the potential hybridization products when the DdeI sites are fully or partially cut because of the presence of J. **(B)** Plasmid DNA was purified from *L. tarentolae* and digested with DdeI and increasing amounts were loaded on gel. Also genomic DNA from wild type and JBP2-null *L. tarentolae* was isolated and digested with DdeI. We also isolated DNA from JBP2-null cultures grown in the presence of BrdU, which lowers J-levels further (1). The DNA was size-fractionated in an agarose gel, blotted and probed.

The main partial digest band visible with both probes is a 1358-bp band spanning the two DdeI sites at positions 4238 and 4308 in the plasmid containing the 25.2L insert. SMRT sequencing shows that these sites are indeed partially modified as shown by the sequence maps in Supplementary Figure S3 which result in an estimated modification of 30% (site 4238) and 34% (site 4308) based on the analysis of individual molecules. Note that these two sites are not part of detectable “doublet”, in contrast to the surrounding J- residues that are $>75\%$ modified and part of a doublet.

There are several other partial digest bands in Supplementary Figure S2. The weak 1054 band can be explained by partial cutting of site 4238, and the prominent 374 band, only seen by the 25.2S probe, by partial cleavage of site 4308. The larger bands, seen by both probes, cannot be unambiguously allocated, as we do not see any additional modified DdeI sites in the SMRT sequence. Notably, the 2602 band might also be located between DdeI sites 1991 and 4612. The larger bands only seen by the 25.2S probe, also require modified DdeI sites, not visibly modified in the SMRT sequence.

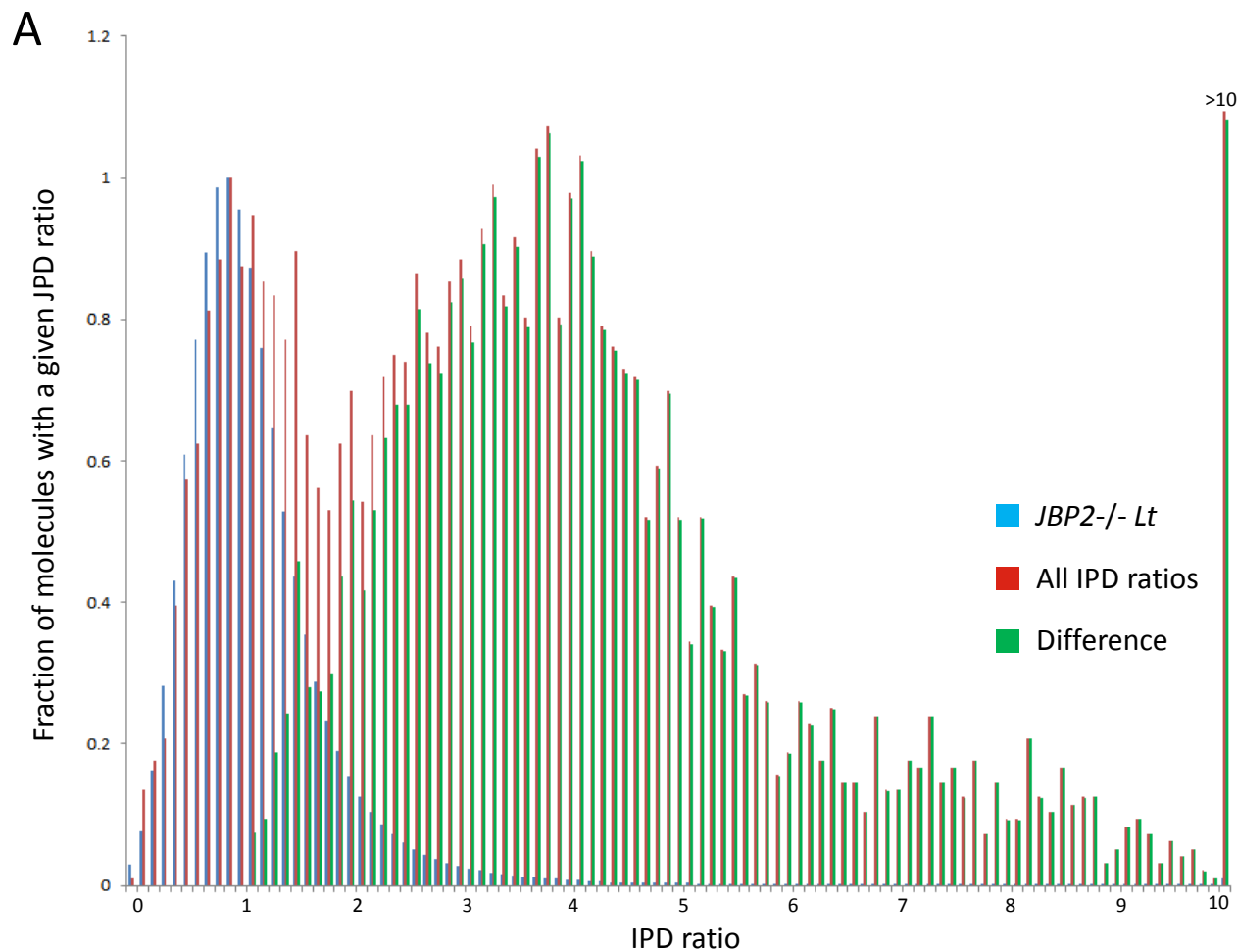
Genomic DNA was also digested with DdeI. The partially cut 1.4 kb fragment is in wild-type DNA and disappears in the JBP2-null mutant.

Supplementary Figure S3. Nucleotide-resolution kinetograms of plasmids isolated from *L. tarentolae*. Plasmids containing a cSSR (25.2S, 25.2L 28.2 or 12.1) or the telomeric repeat sequence (GGGTTA)₁₀ were grown in *L. tarentolae*, purified and SMRT sequenced. The IPD ratio plots for the complete plasmids are shown. The sequences have been aligned in the vector sequences and the relevant restriction sites, plasmids fragments, and G-quadruplexes have been indicated

Supplementary Figure S4. Average kinetic signature for base J. All cases in which there was only one T within 7 nucleotides with an IPD ratio >1.6 at relative positions 0 (T), +1, or +2 were included in the calculation; thirteen cases were found (sequences listed above). The direction of the sequencing polymerase is from left to right.

References

1. Vainio, S., Genest, P.A., Ter Riet, B., Van Luenen, H.G.A.M. and Borst, P. (2009) Evidence that J-binding protein 2 is a thymidine hydroxylase catalyzing the first step in the biosynthesis of DNA base J. *Mol. Biochem. Parasitol.*, **164**, 157-161.



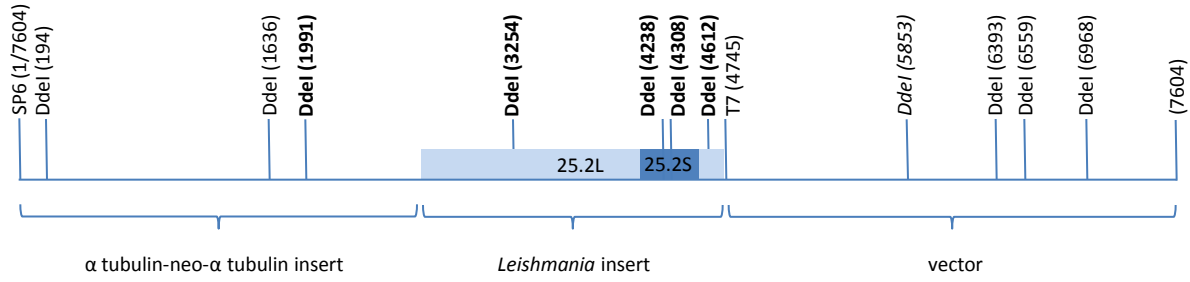
B

cut-off	FPR	FNR
1	17.2%	0.0%
1.1	15.2%	0.0%
1.2	13.2%	0.0%
1.3	11.3%	1.1%
1.4	9.6%	2.1%
1.5	8.1%	4.0%
1.6	6.9%	6.2%
1.7	5.8%	10.1%
1.8	4.9%	12.0%
1.9	4.2%	13.8%
2	3.6%	15.6%
2.1	3.1%	18.3%
2.2	2.7%	21.3%
2.3	2.4%	23.5%
2.4	2.1%	26.1%
2.5	1.9%	28.9%
2.6	1.7%	31.8%
2.7	1.5%	34.4%
2.8	1.4%	37.3%
2.9	1.3%	39.7%
3	1.2%	41.9%

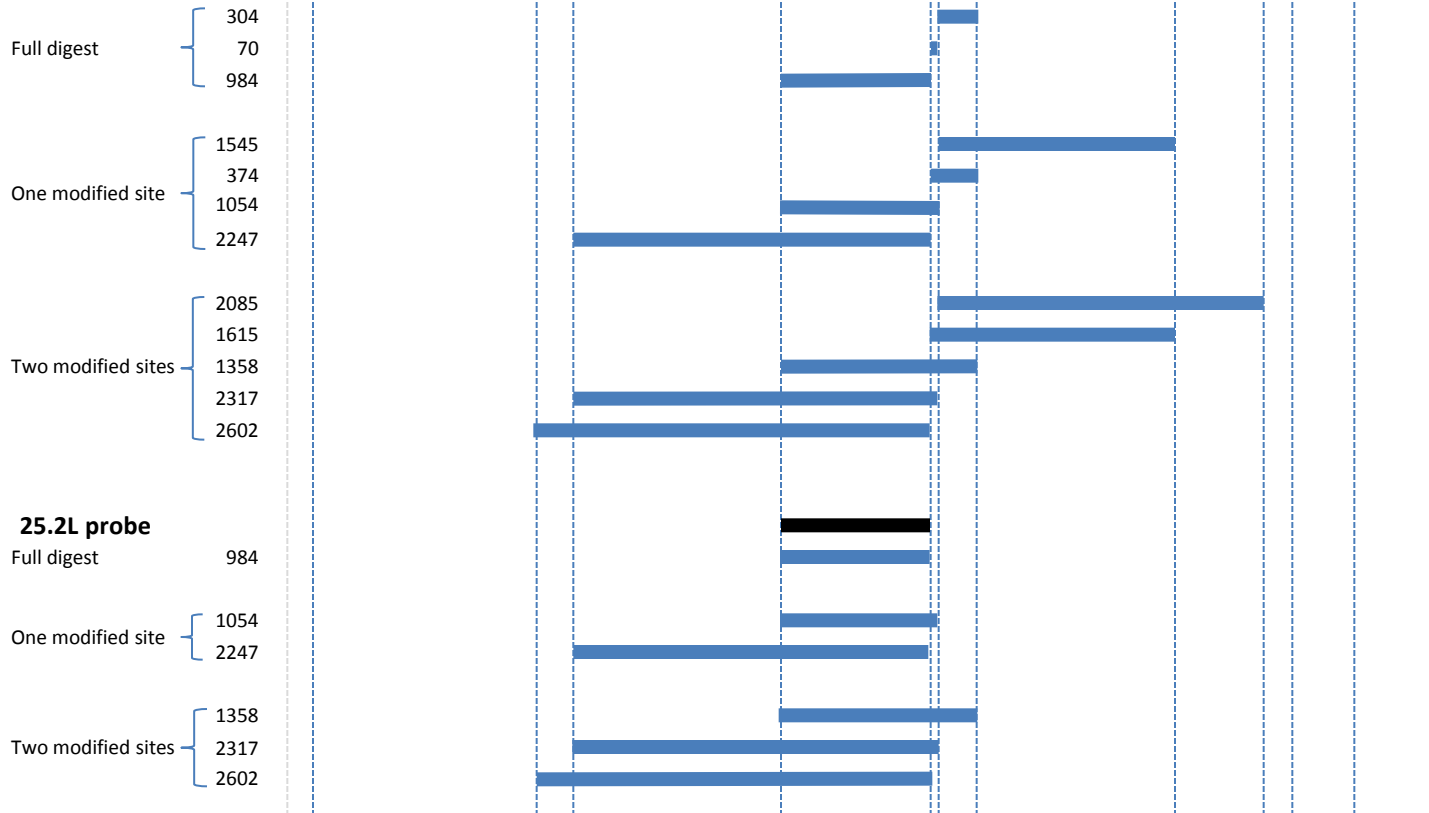
Supplementary Figure S1

A

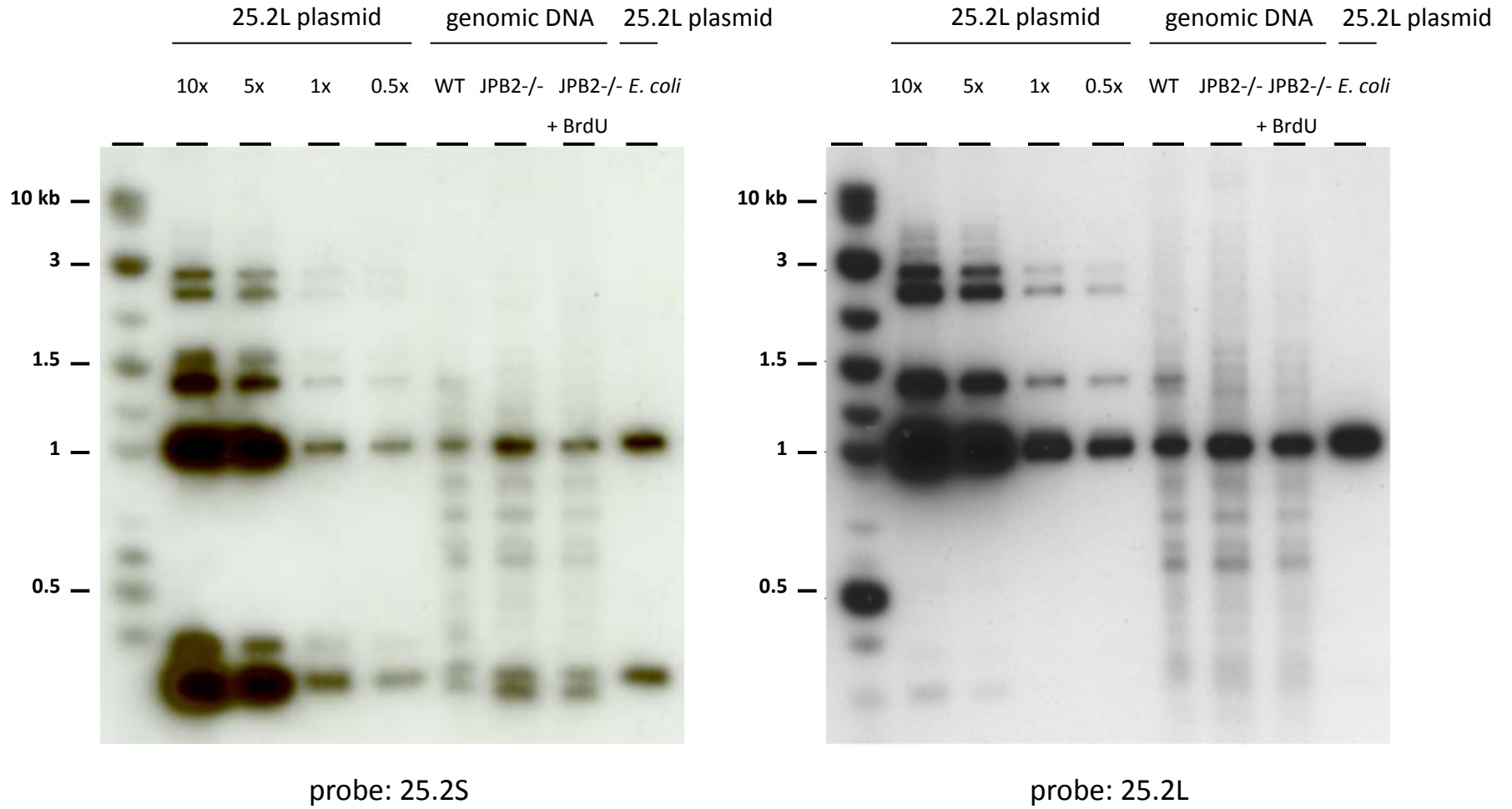
25.2L plasmid



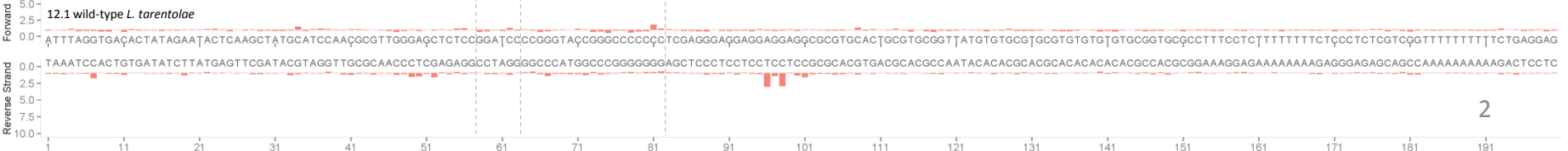
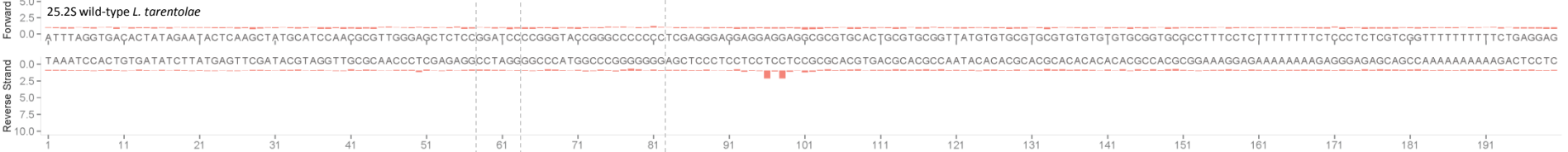
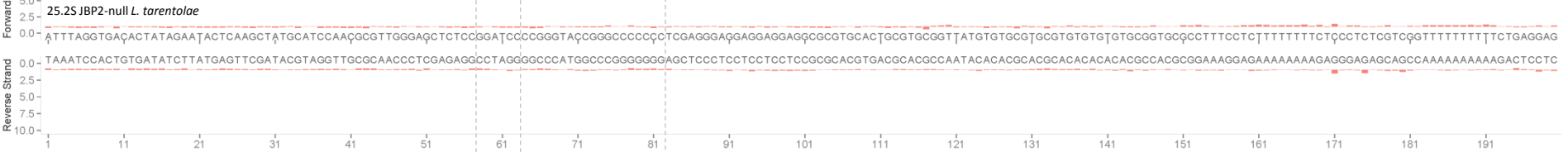
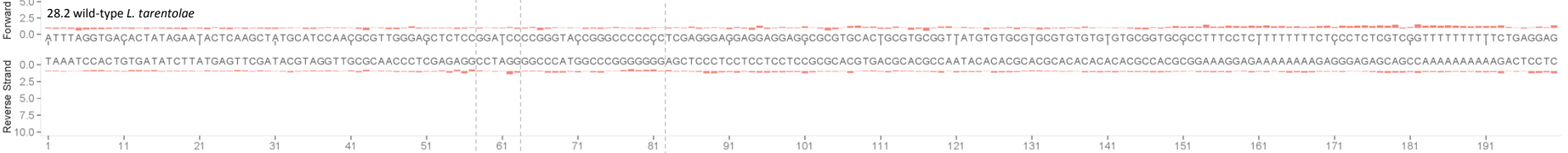
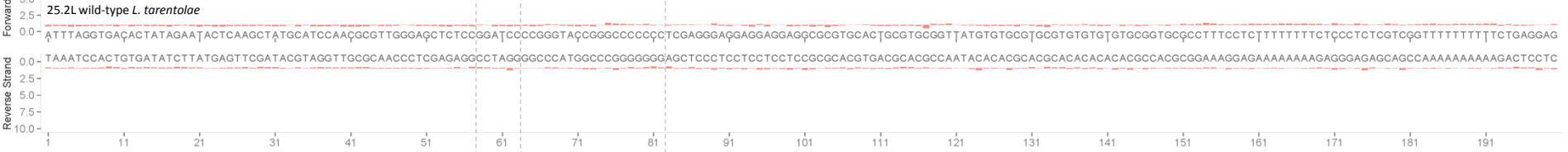
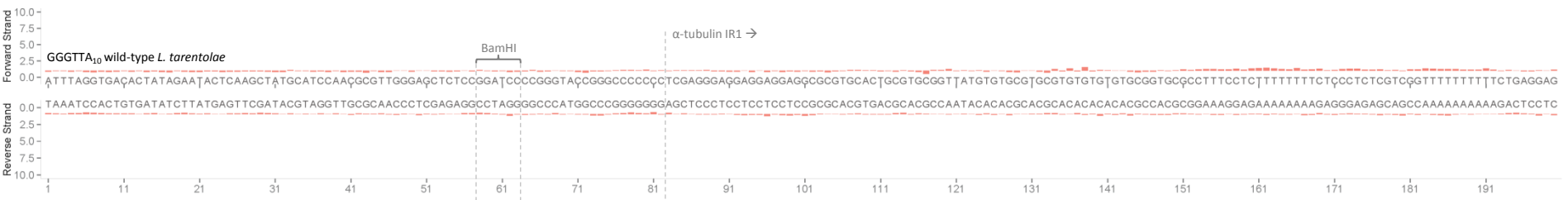
25.2S probe



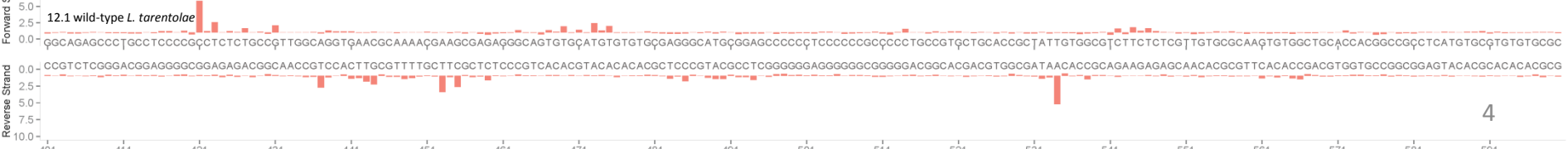
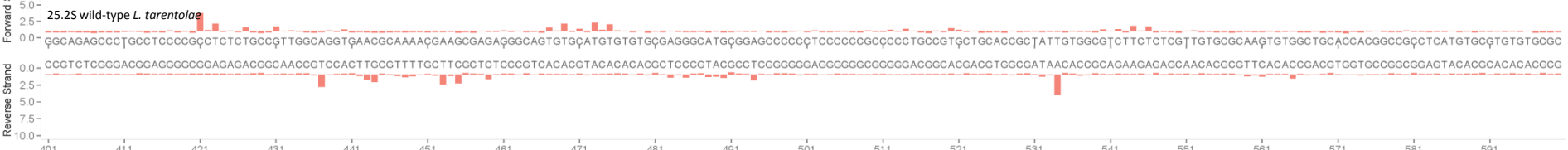
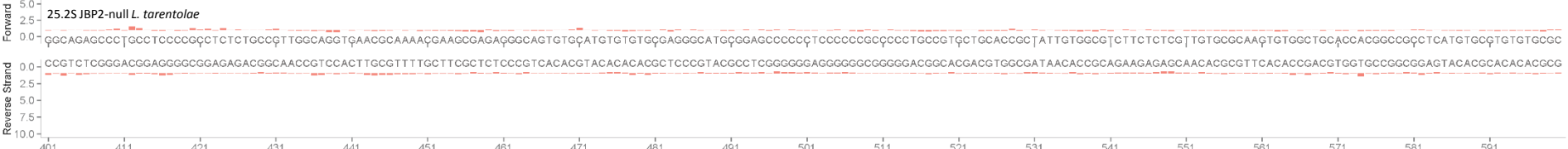
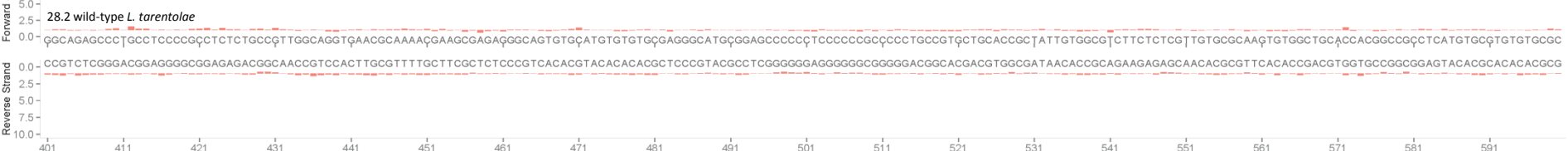
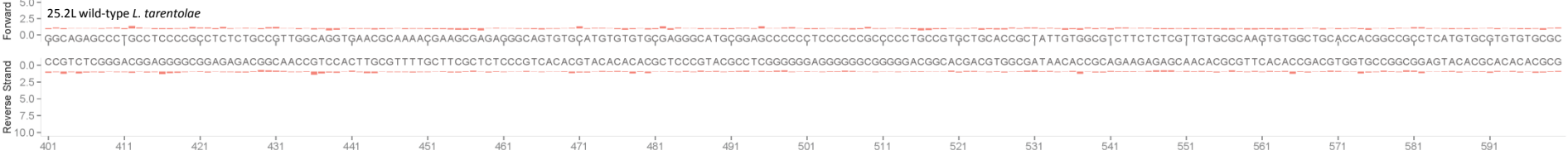
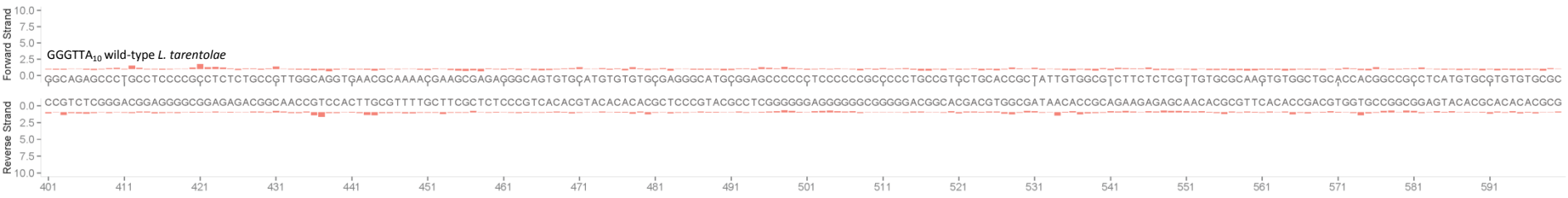
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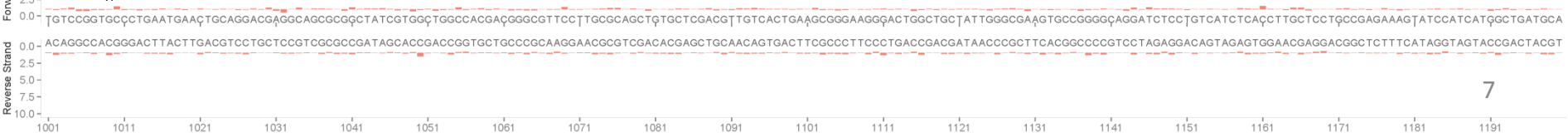
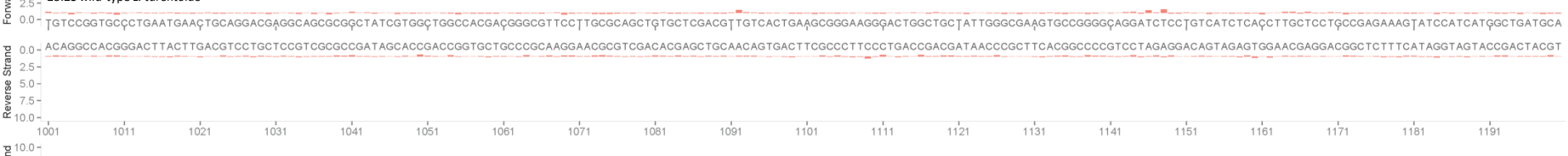
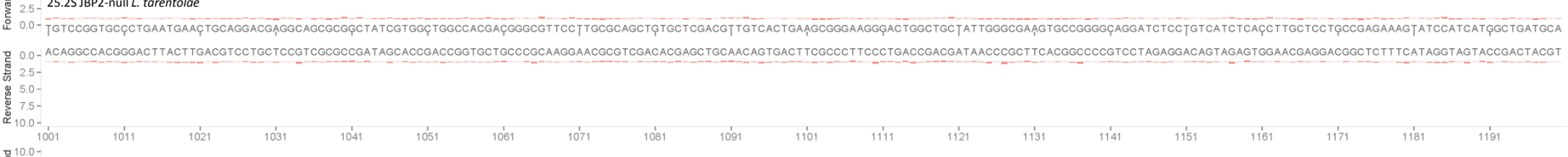
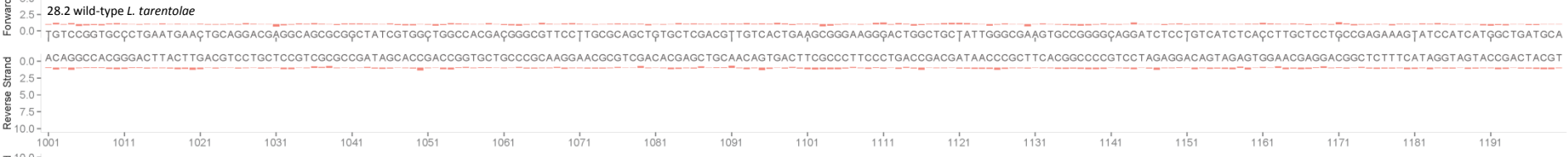
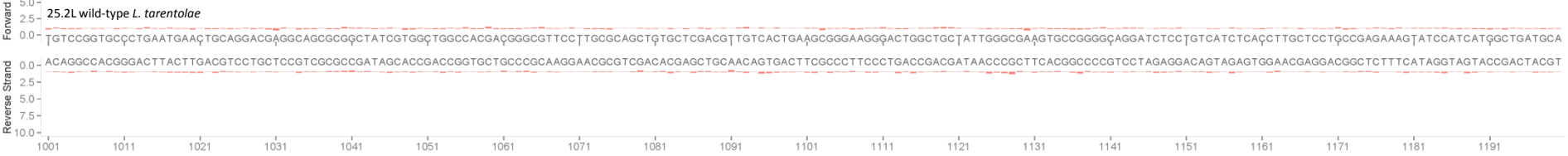
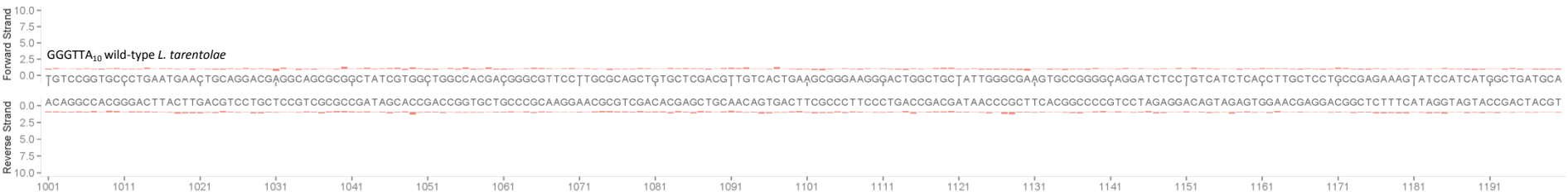
Supplementary Figure S3. Nucleotide-resolution kintograms.

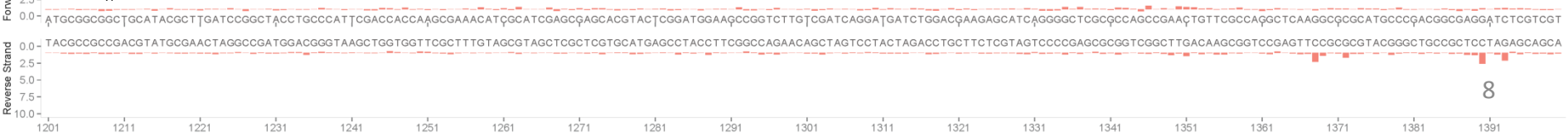
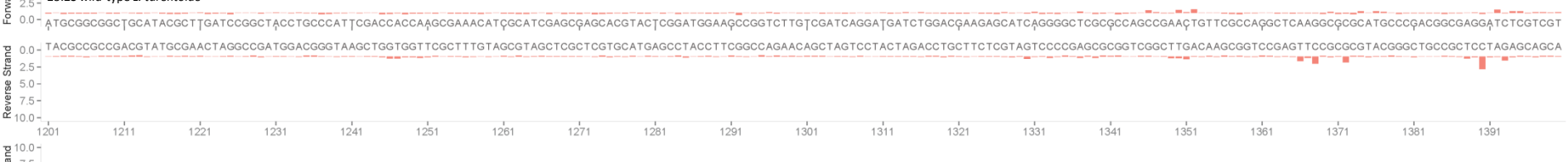
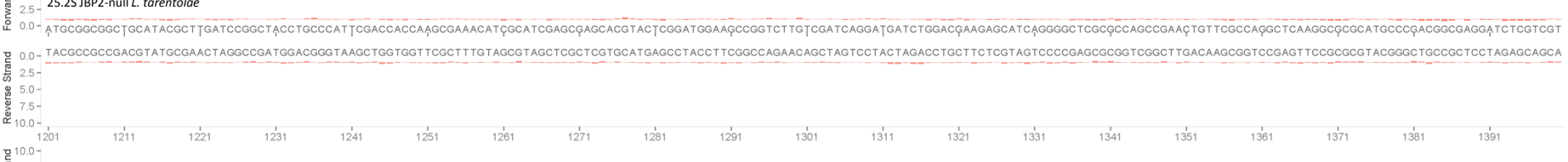
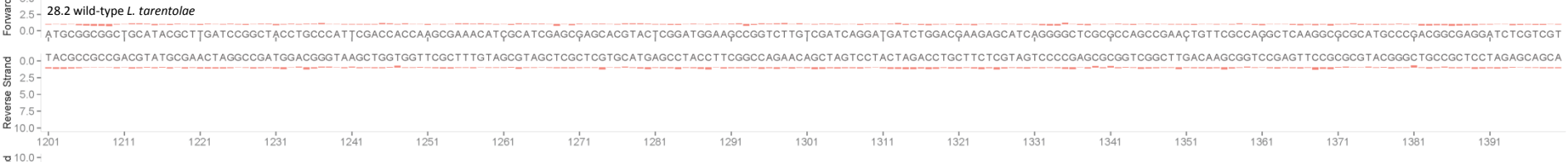
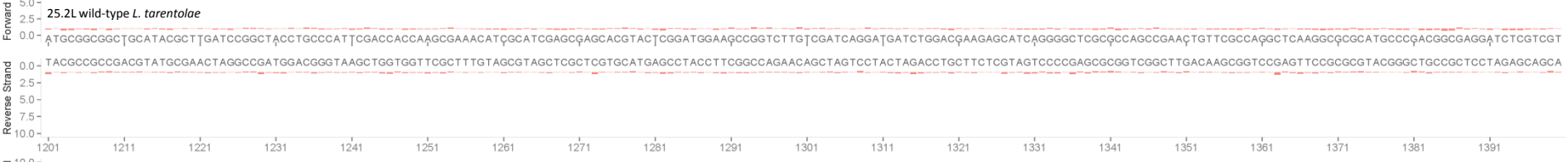
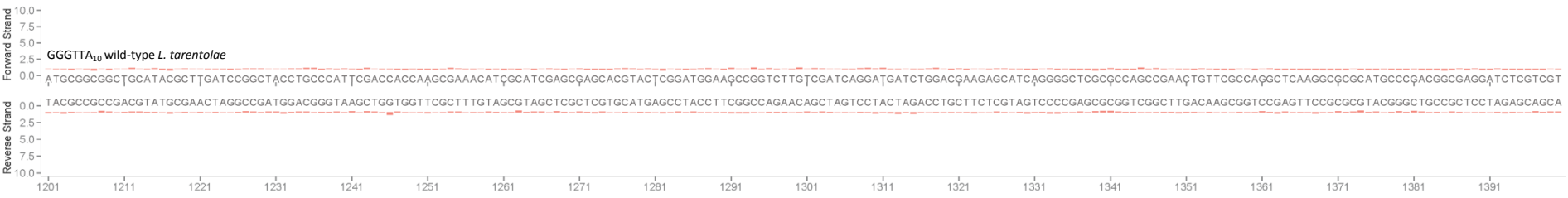


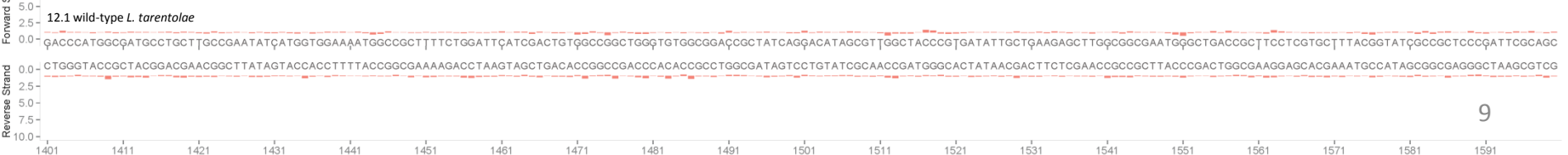
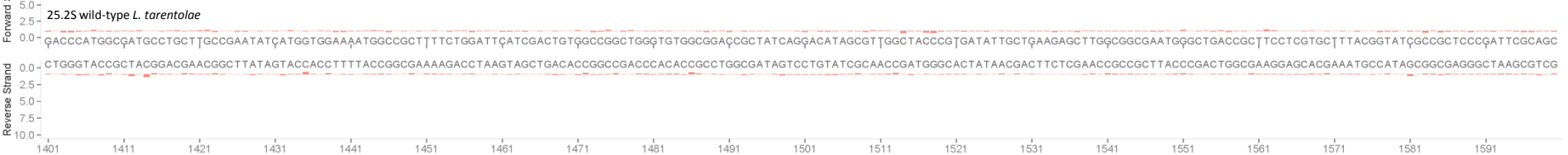
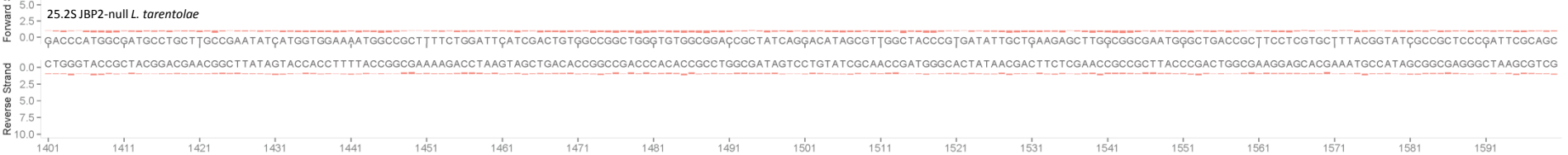
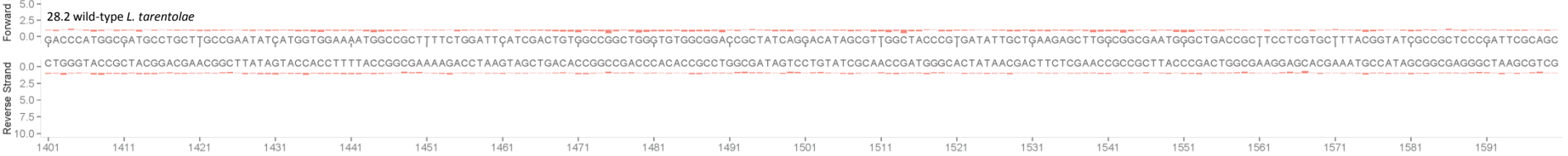
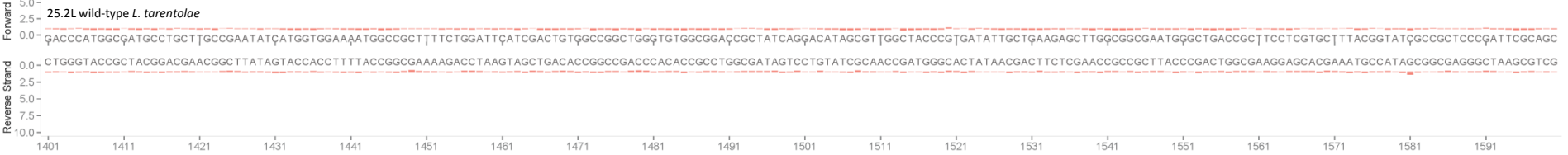
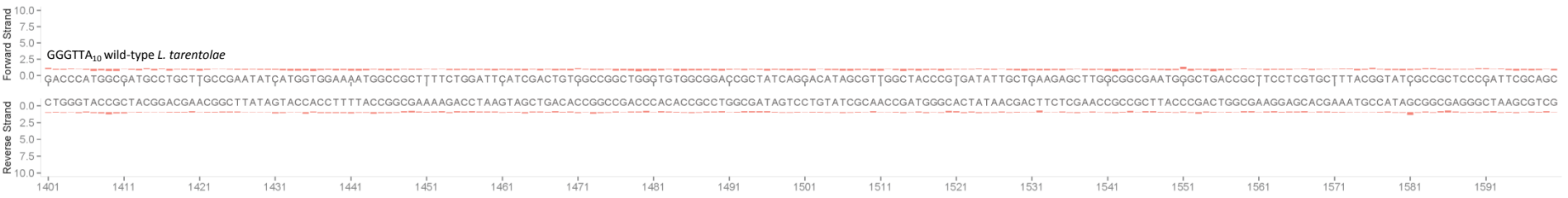


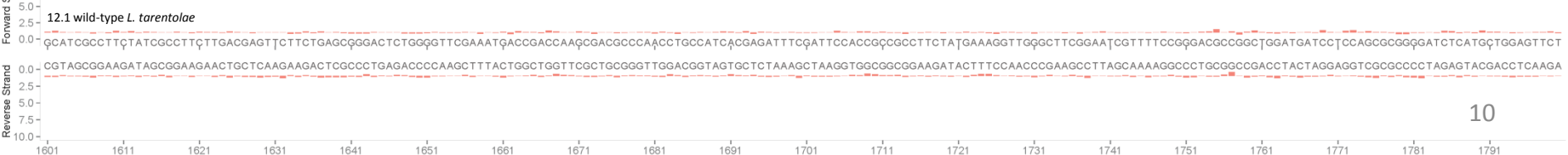
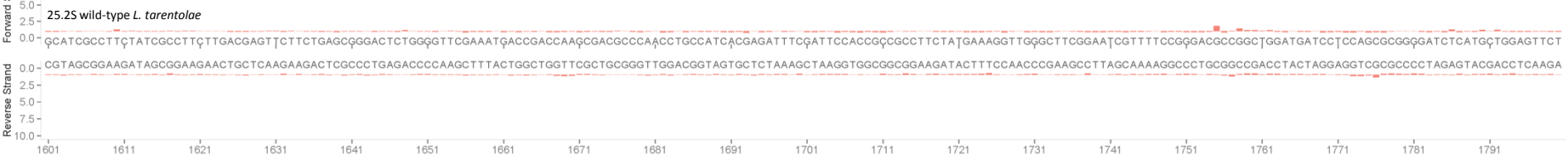
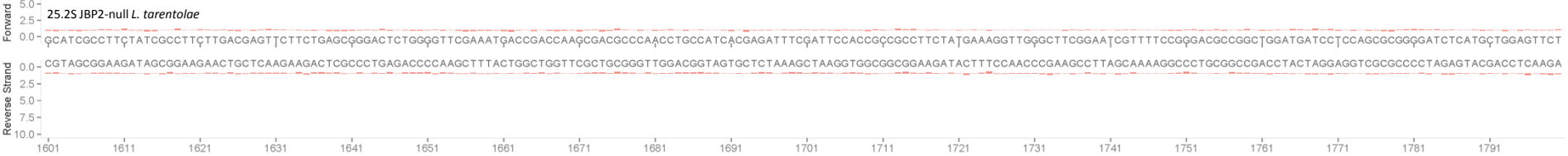
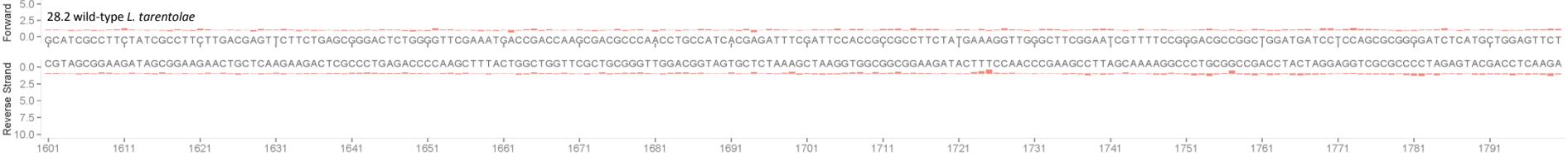
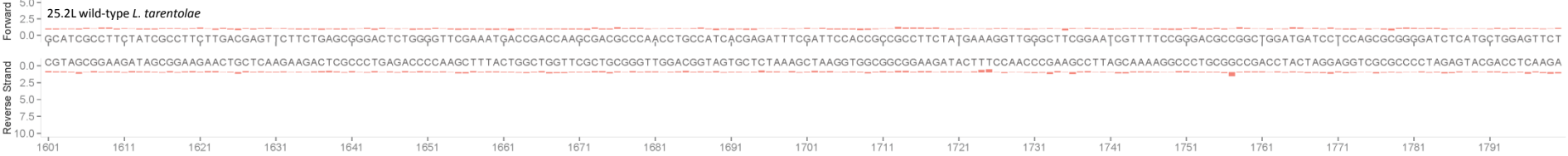
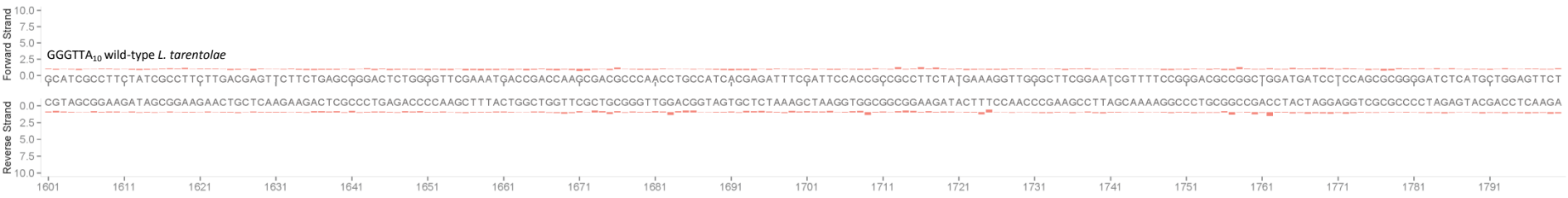


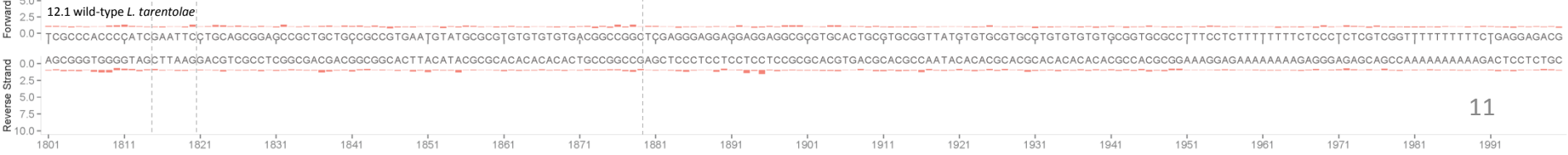
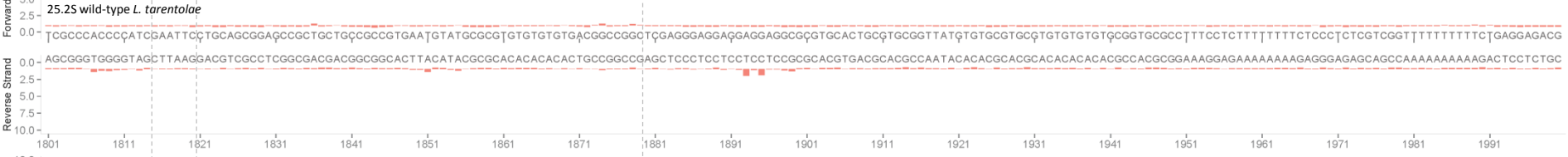
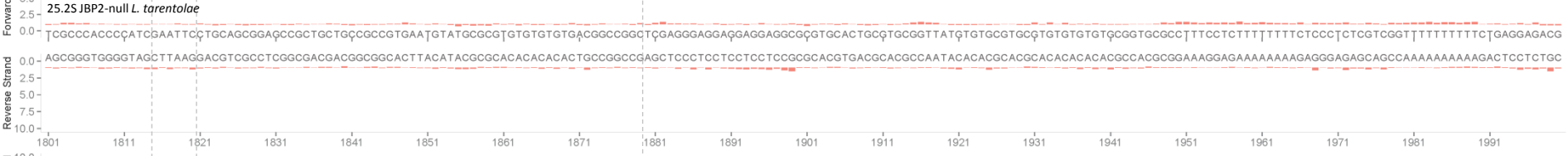
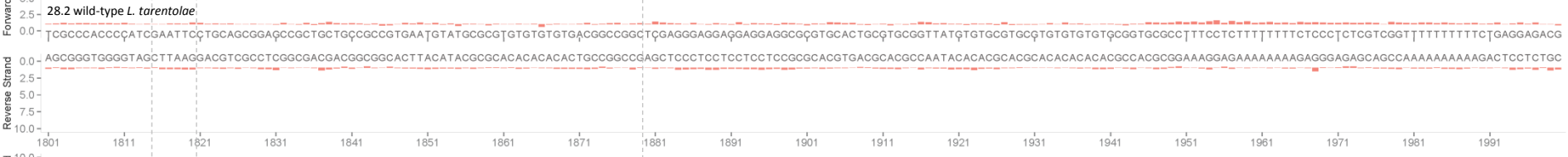
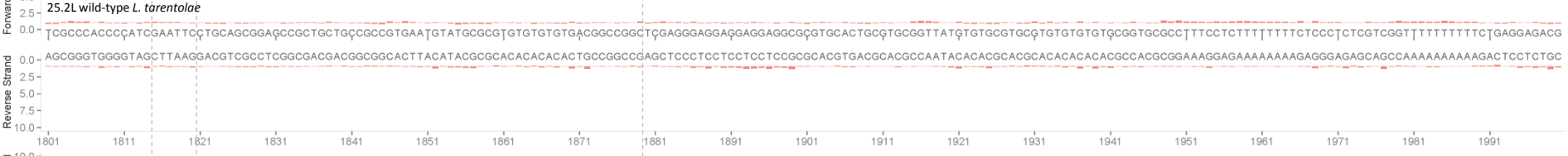
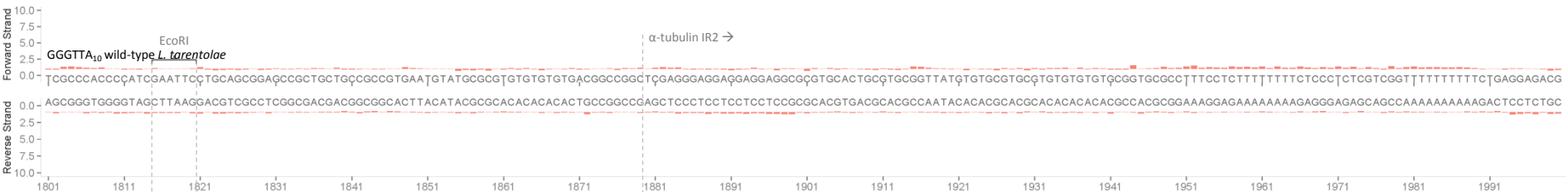


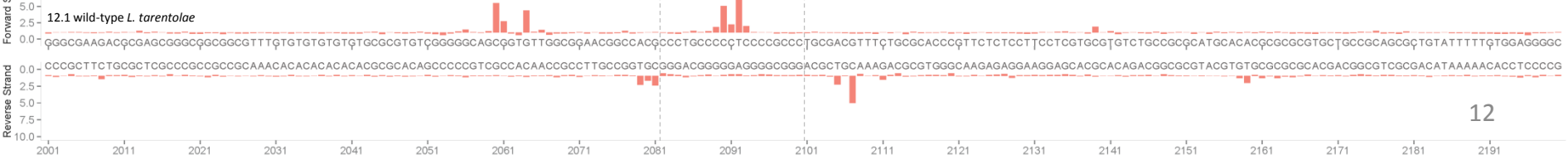
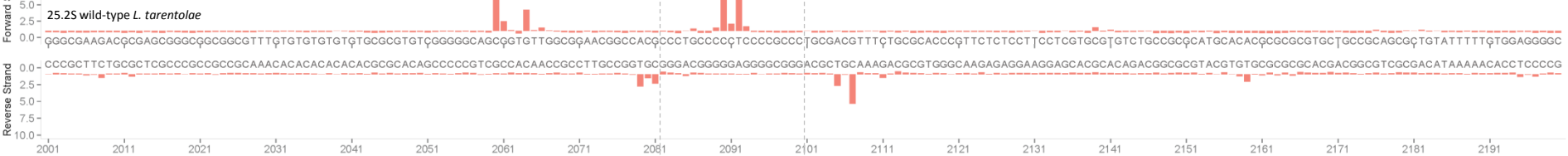
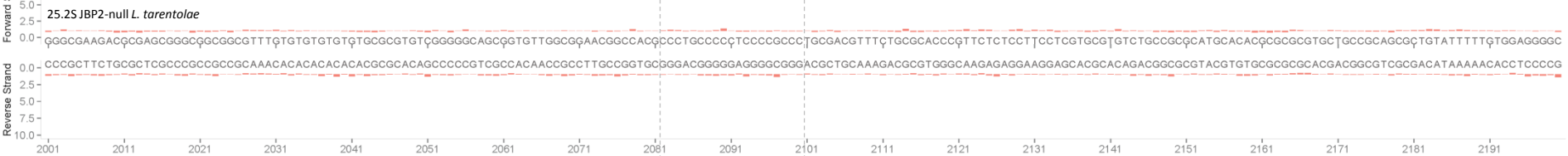
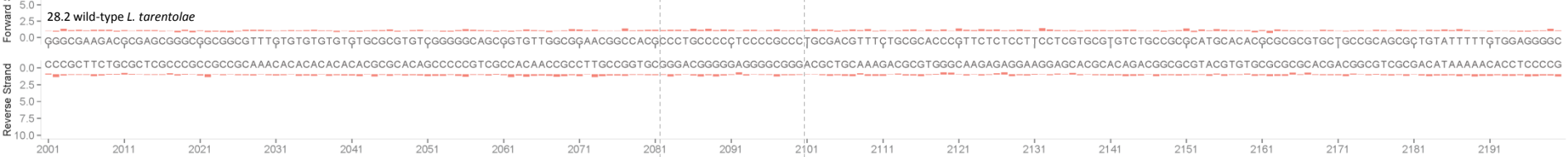
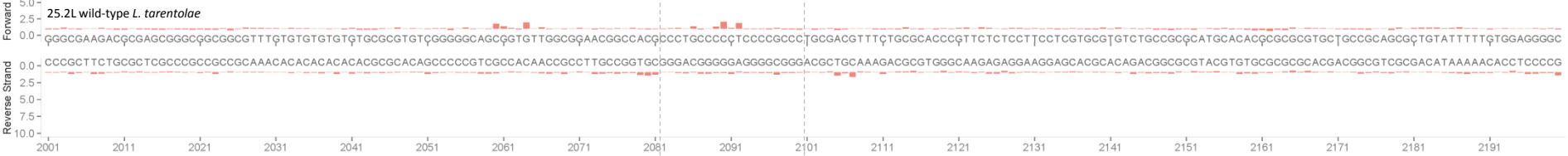
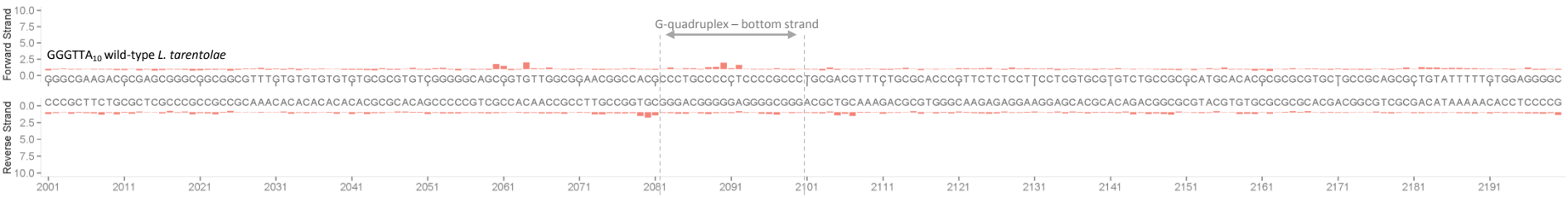


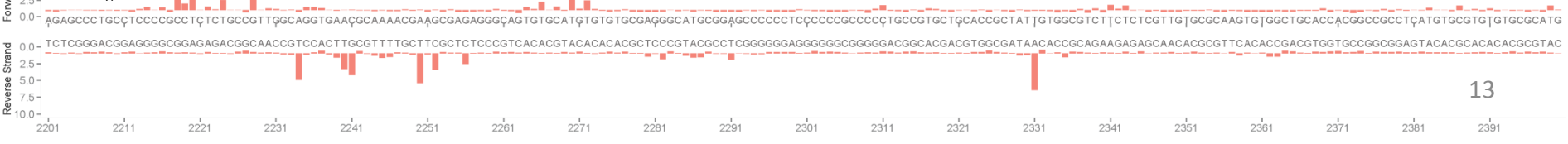
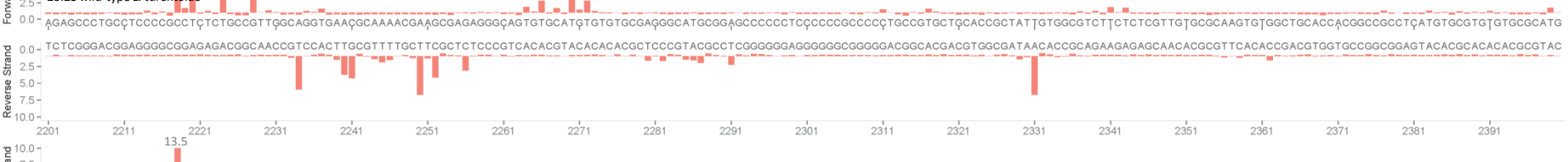
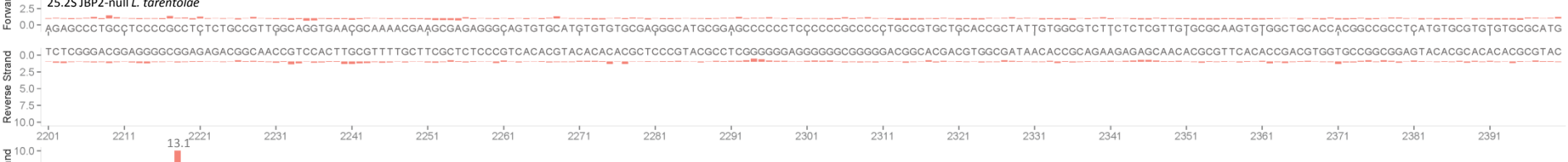
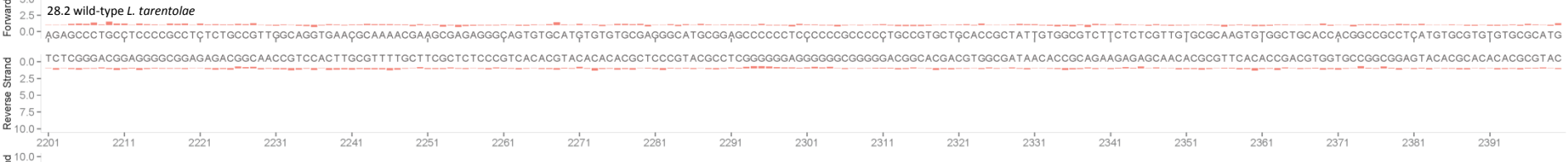
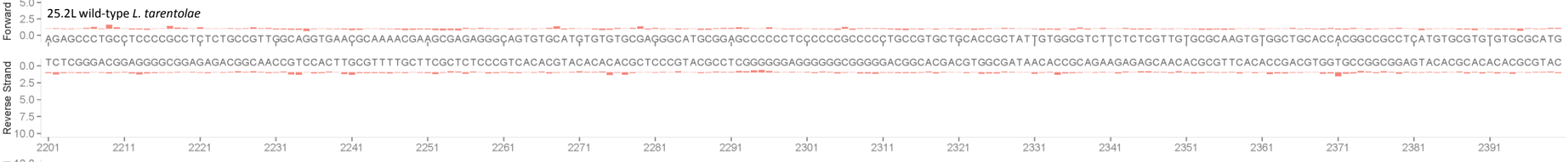
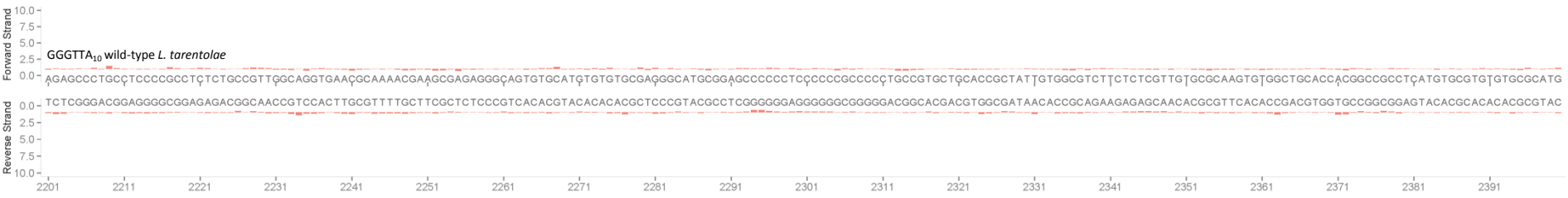


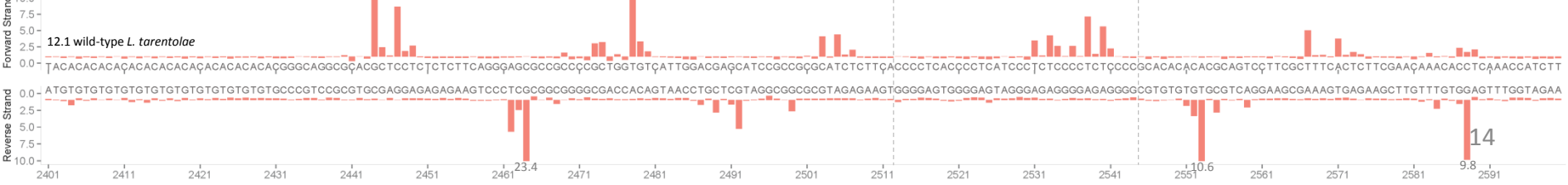
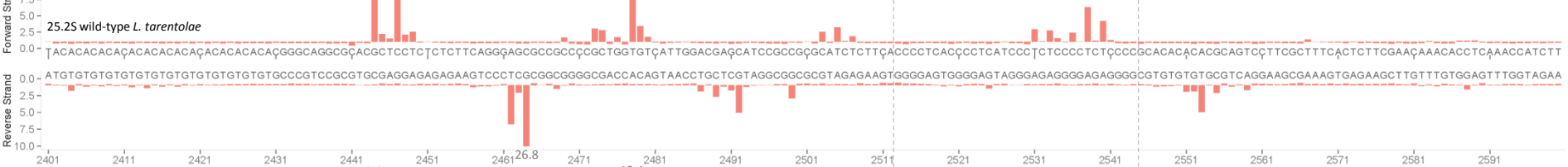
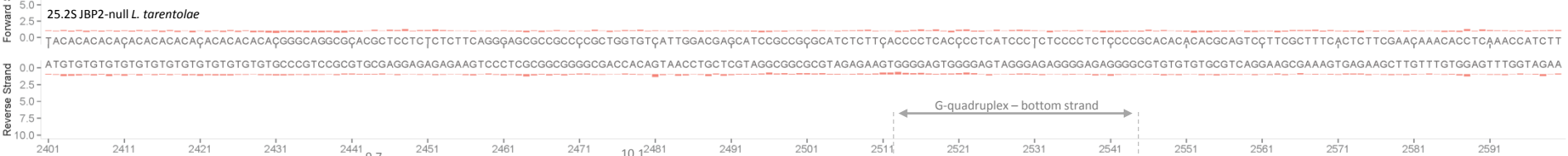
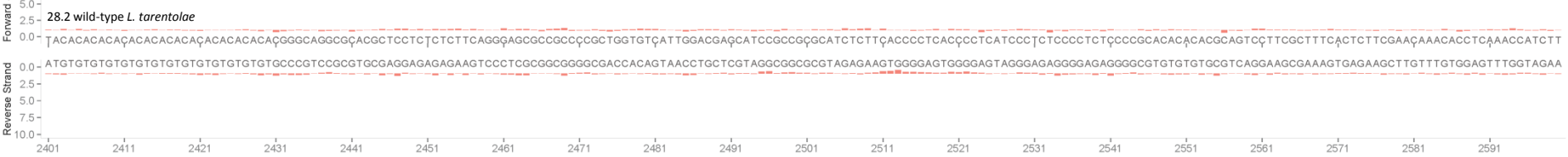
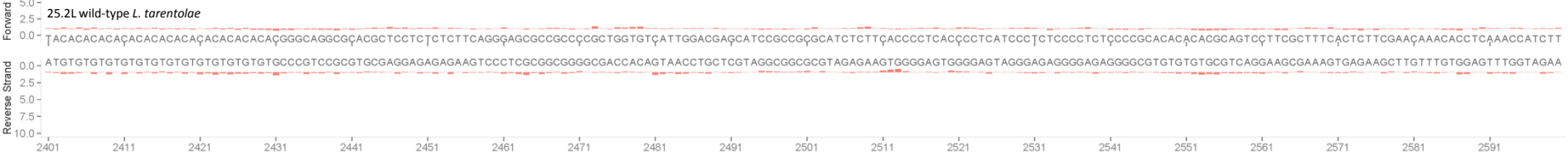
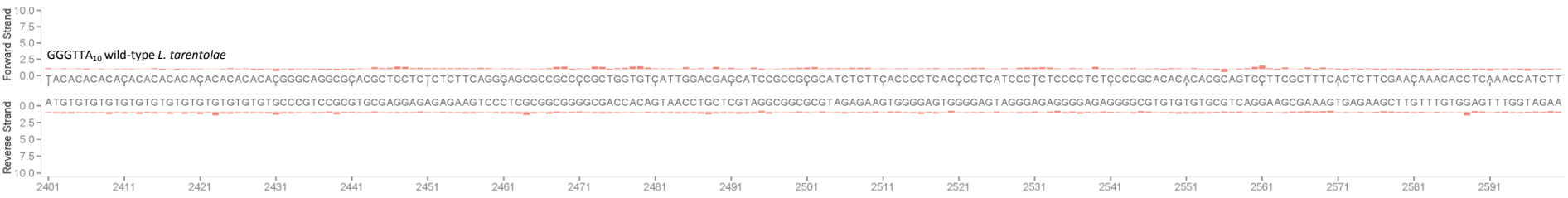


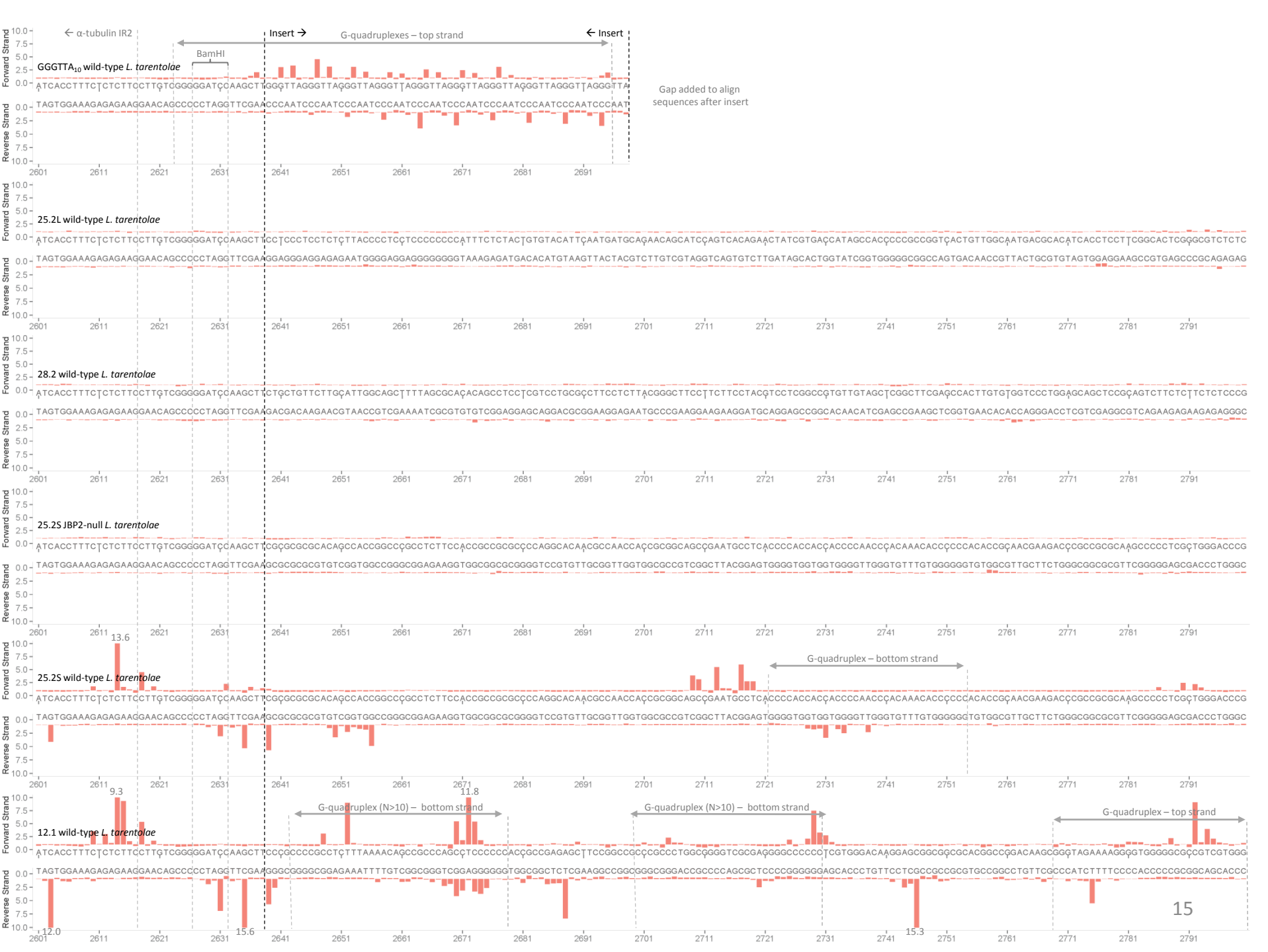


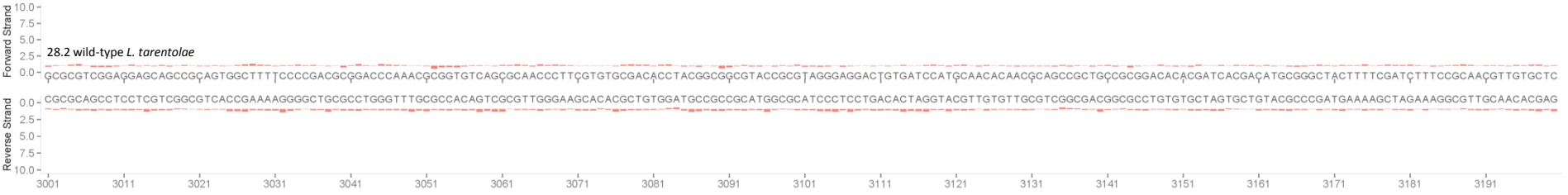
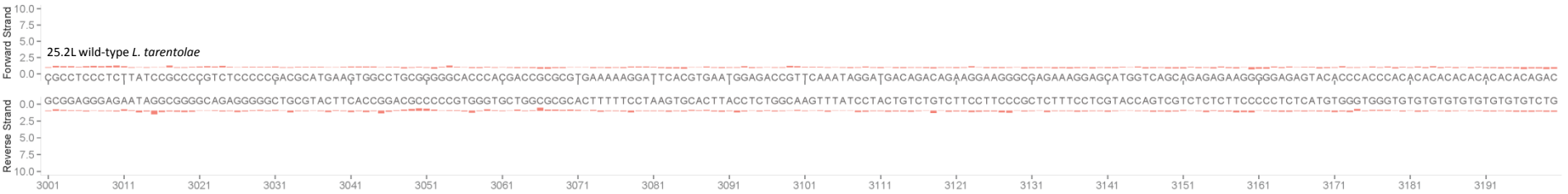


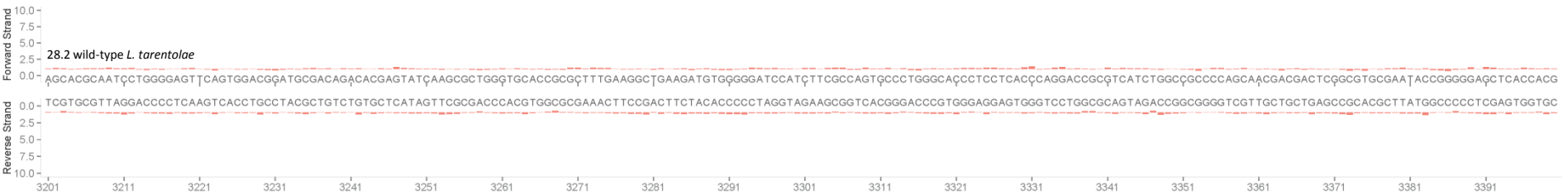
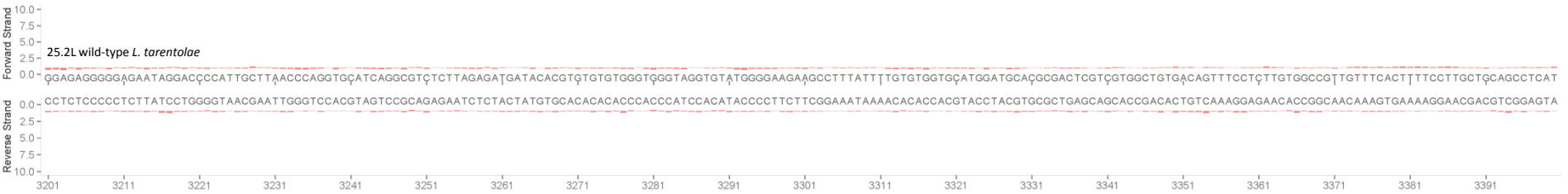


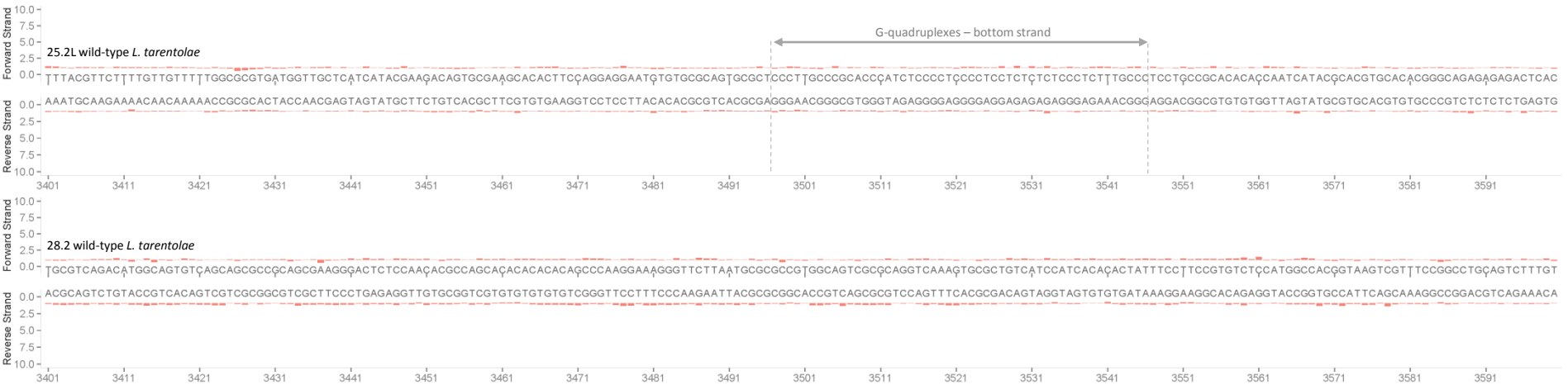


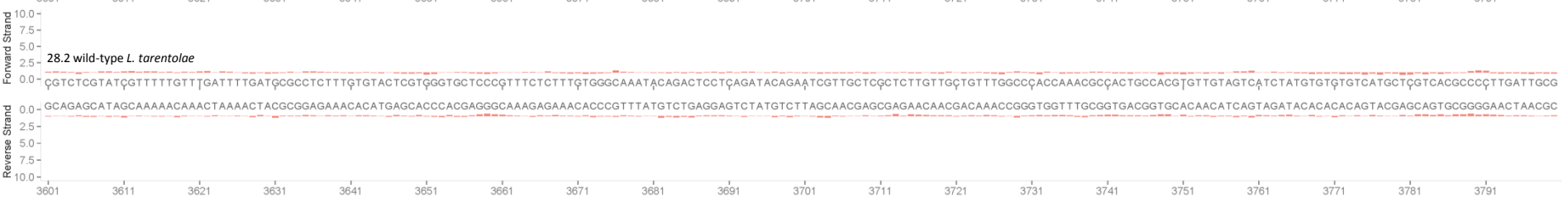
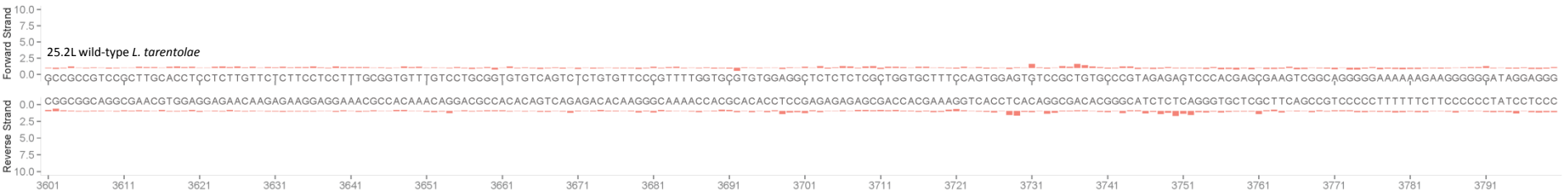


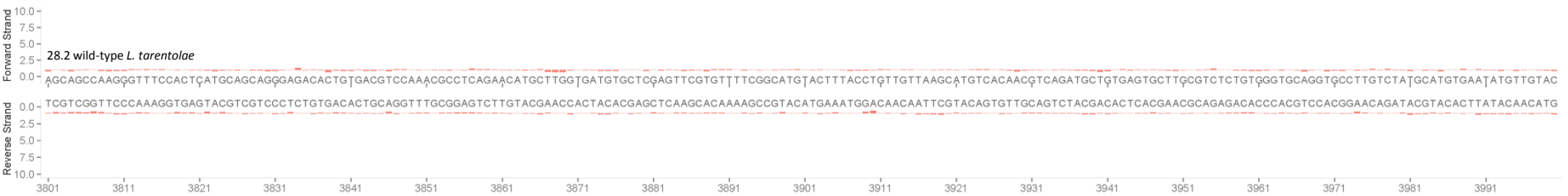
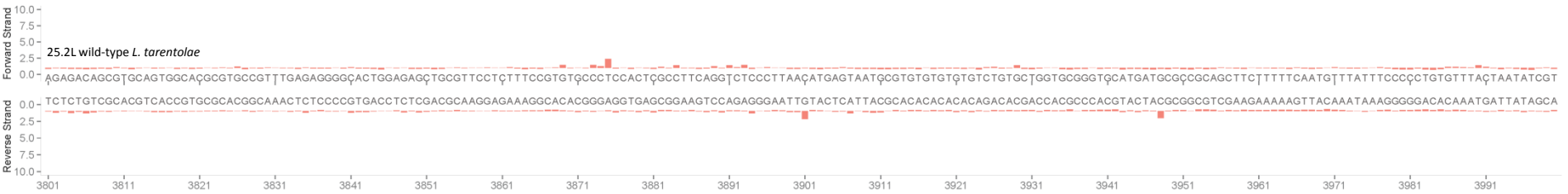


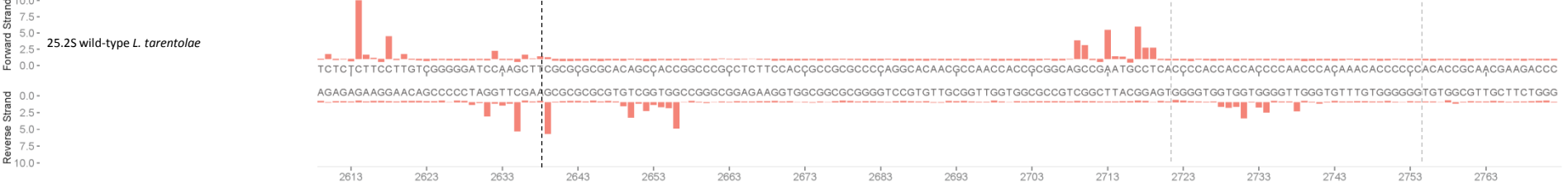
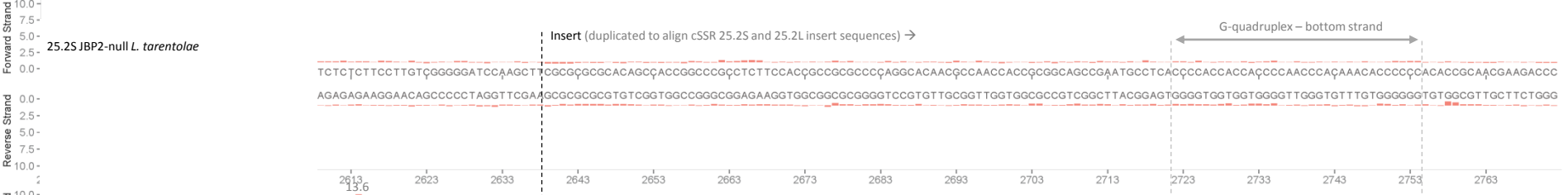
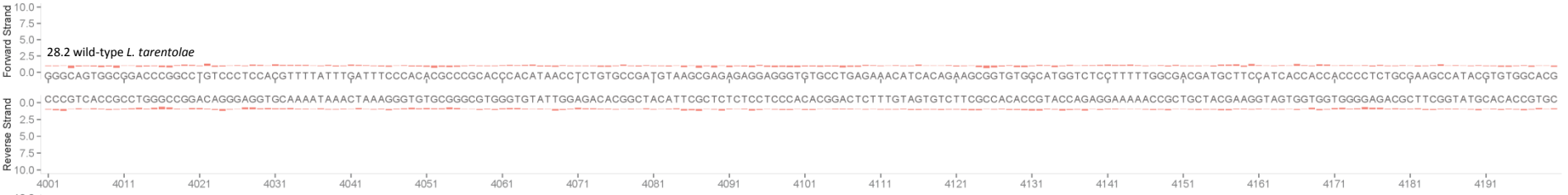
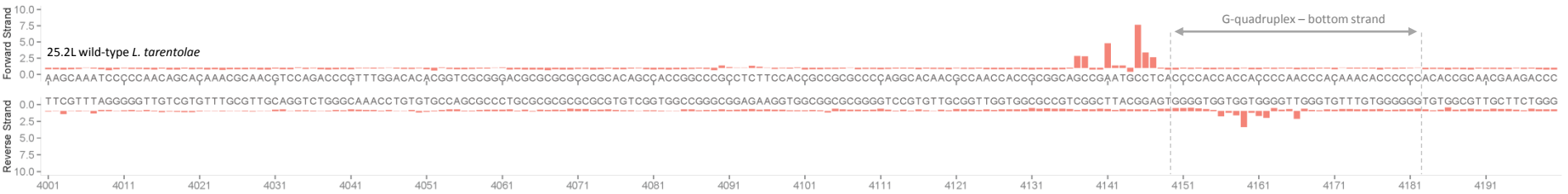


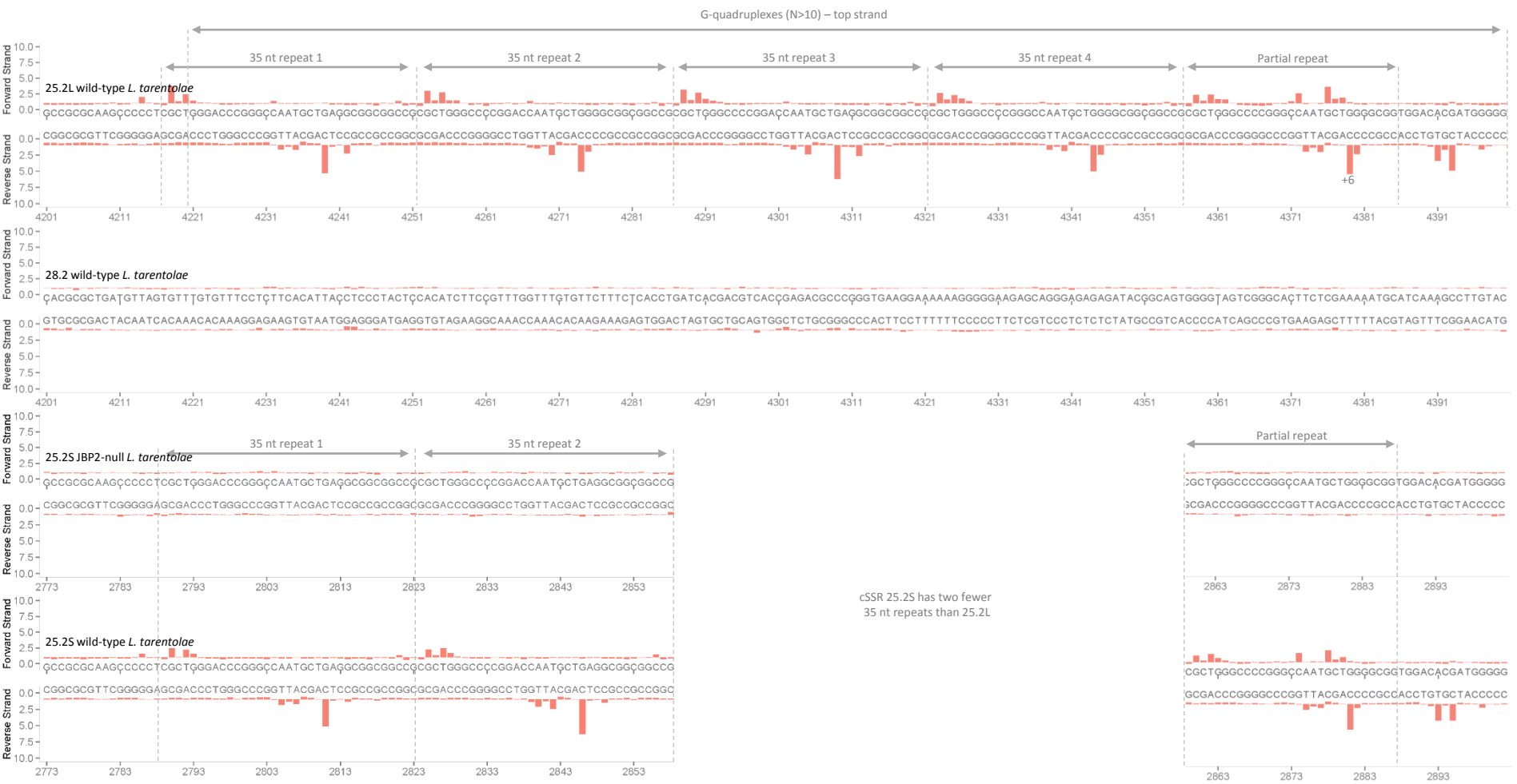


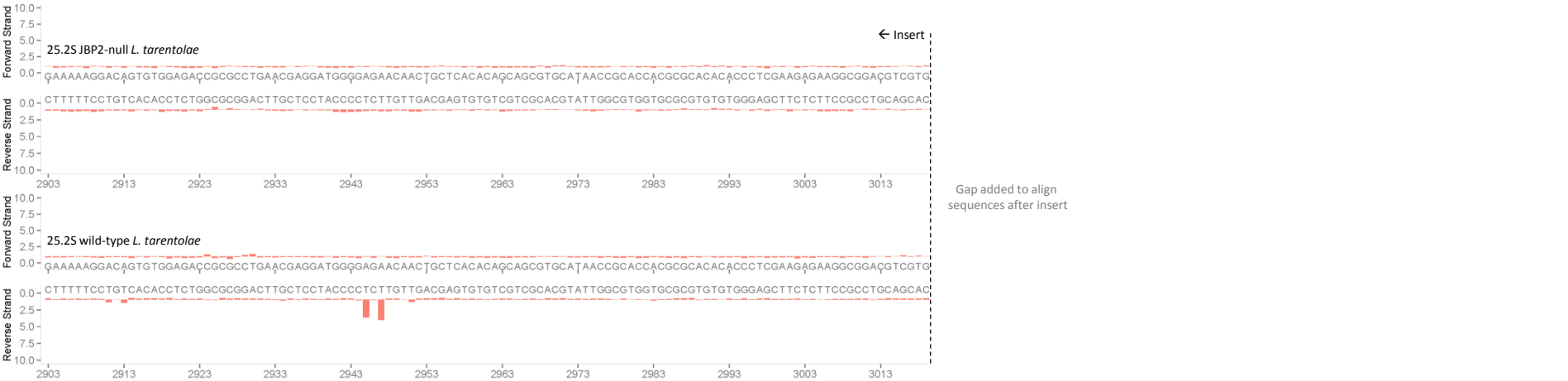
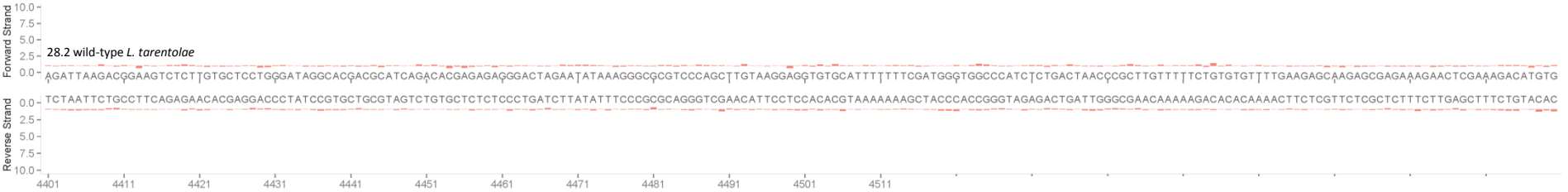
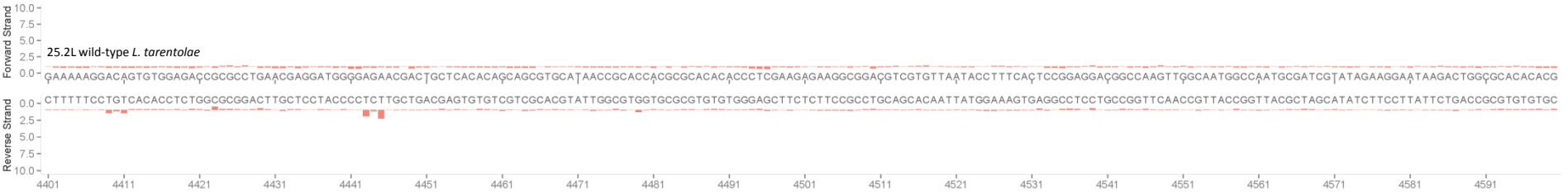


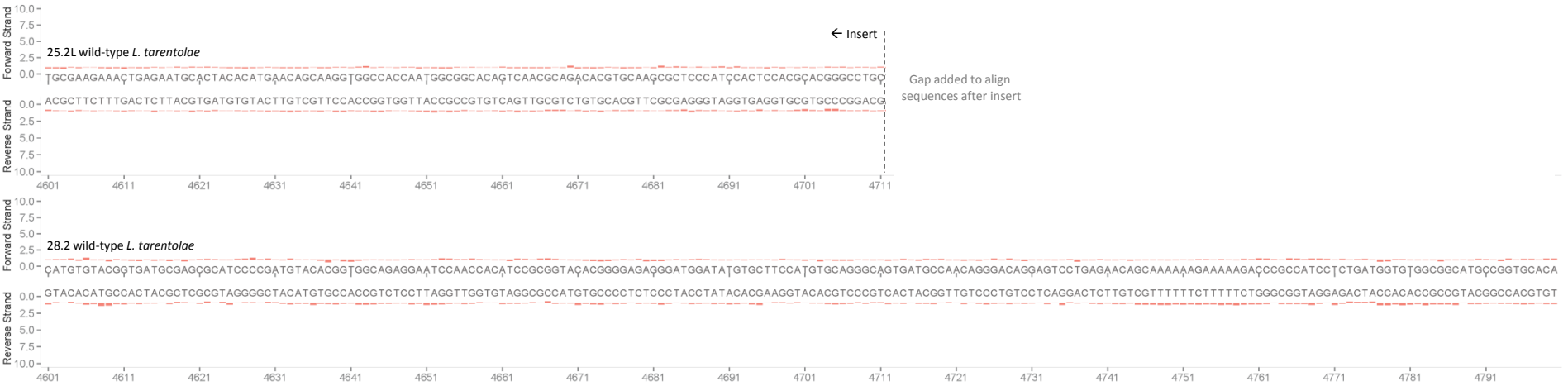


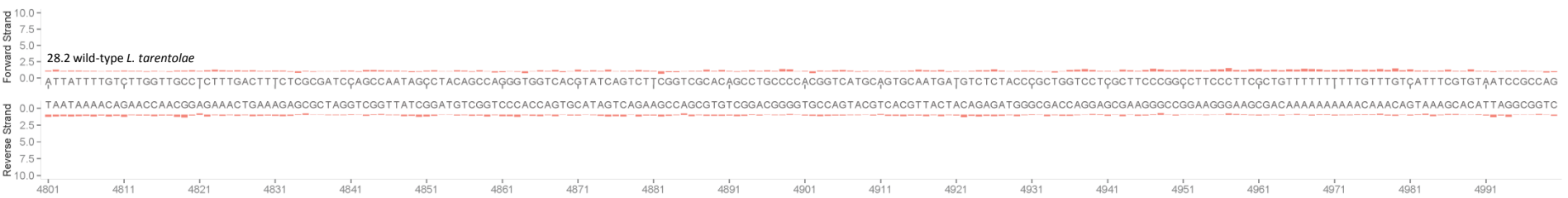


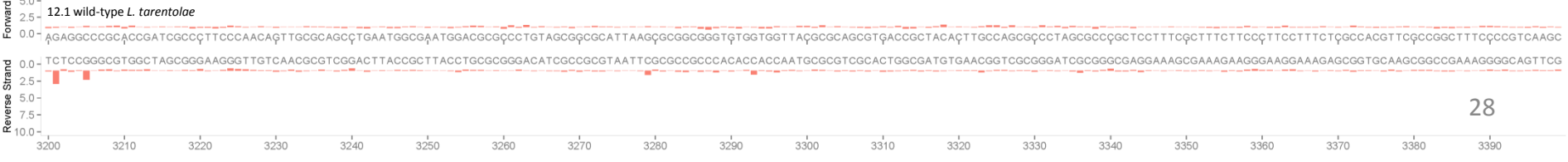
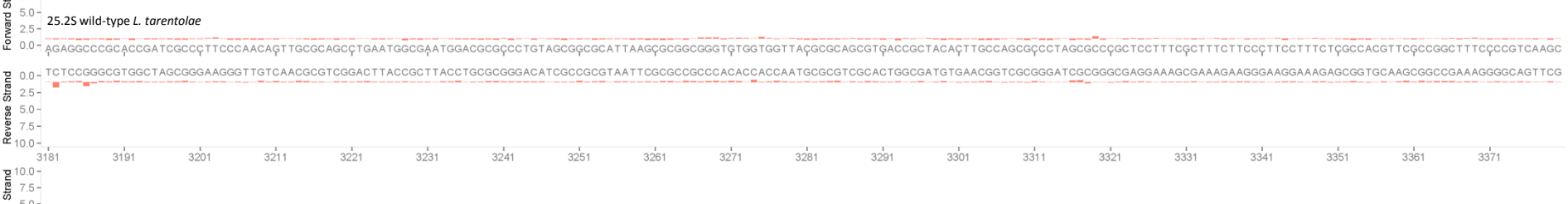
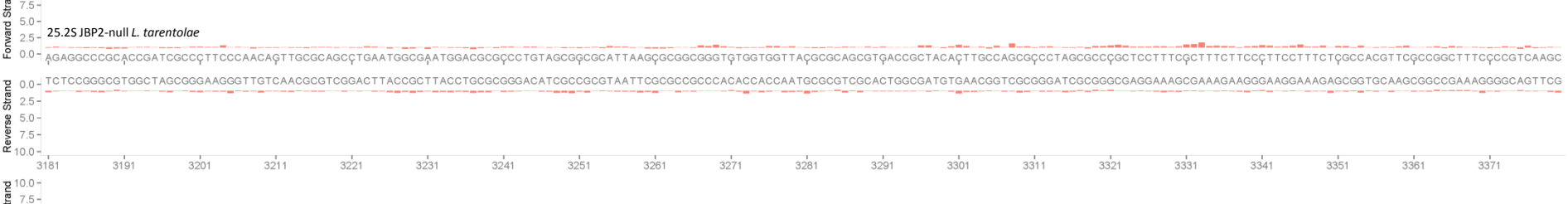
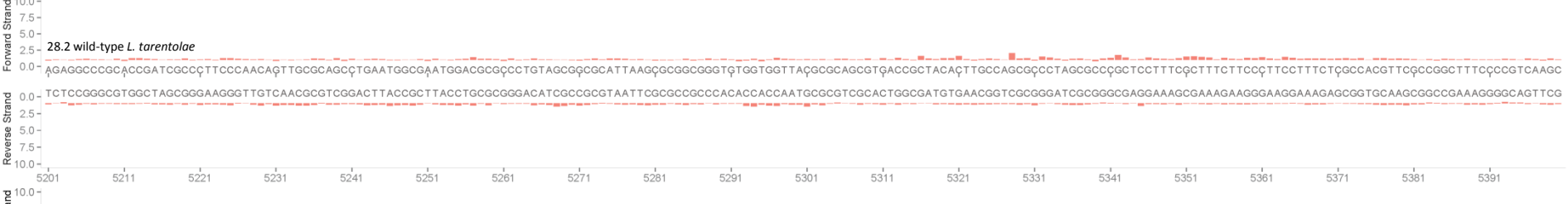
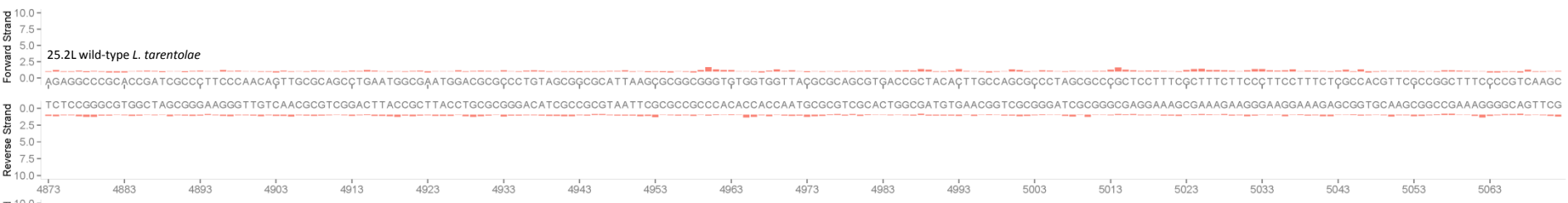
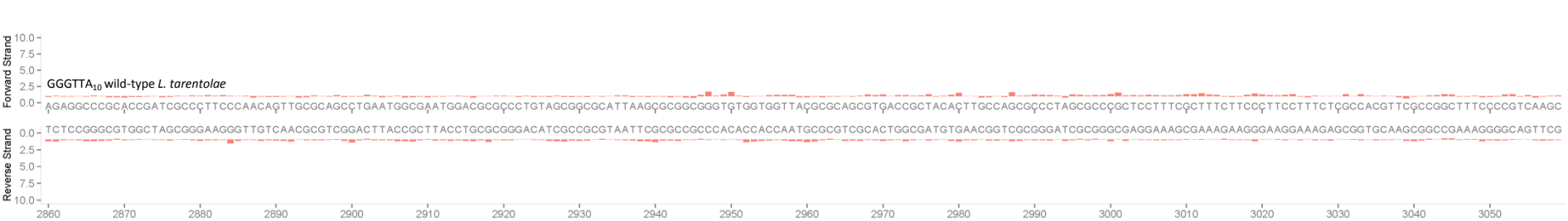


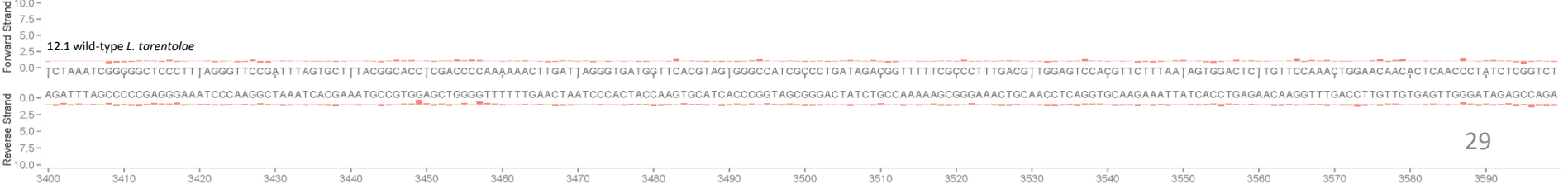
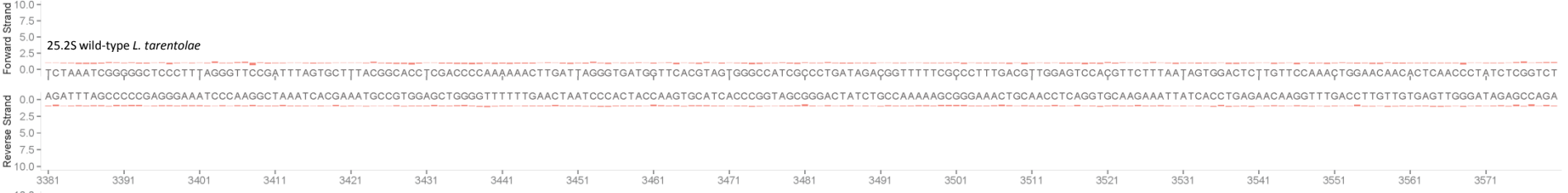
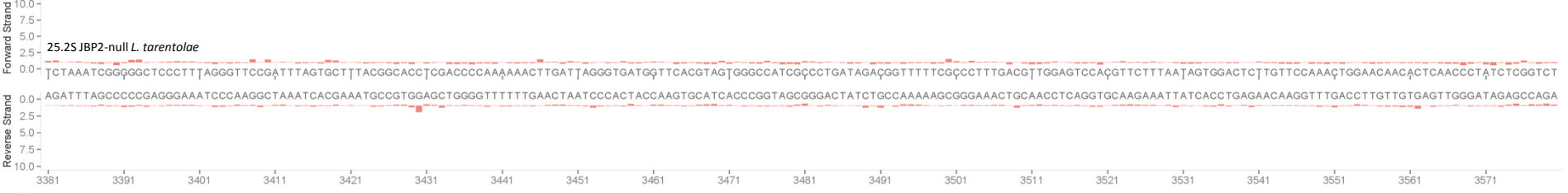
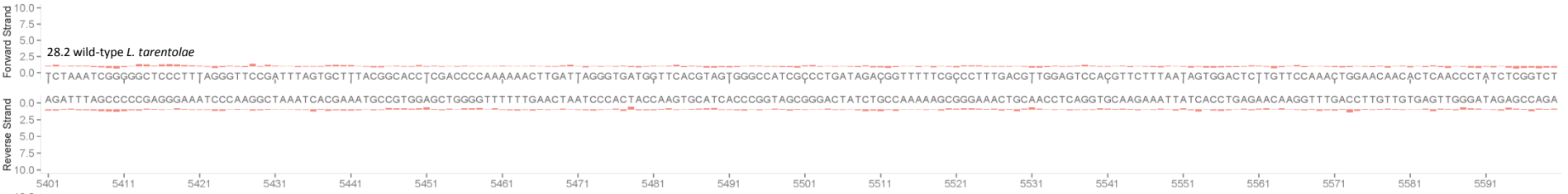
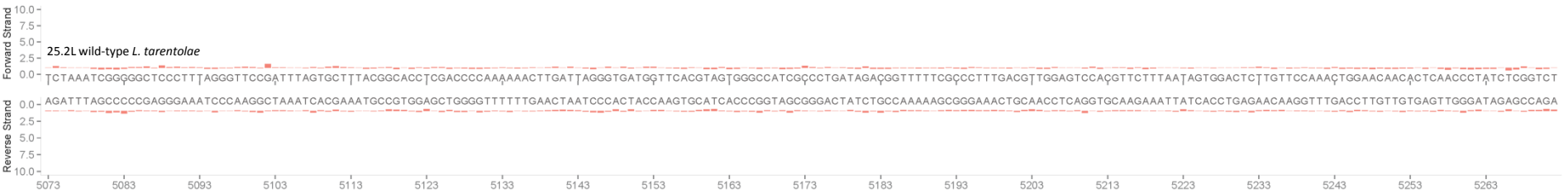
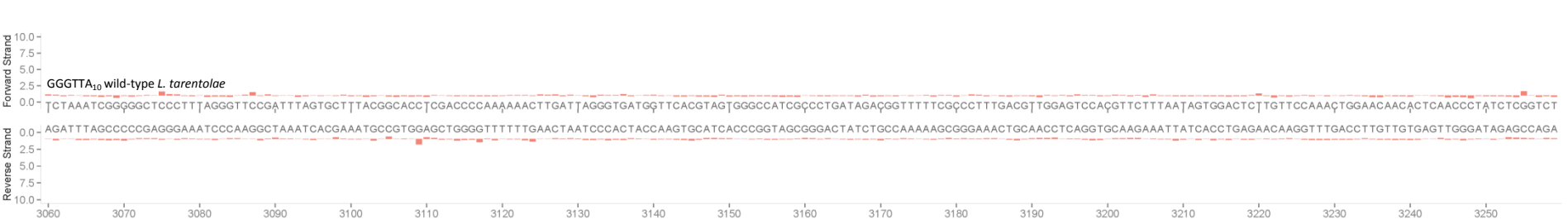


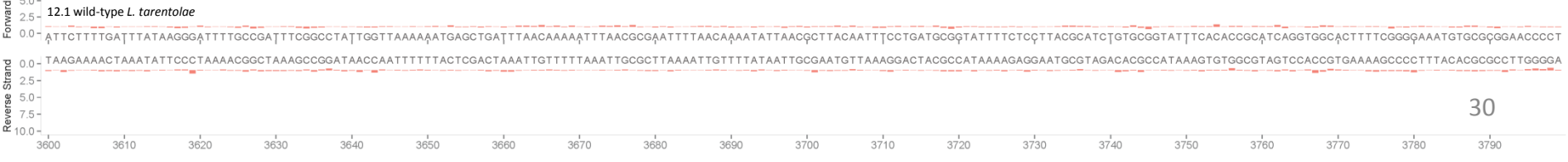
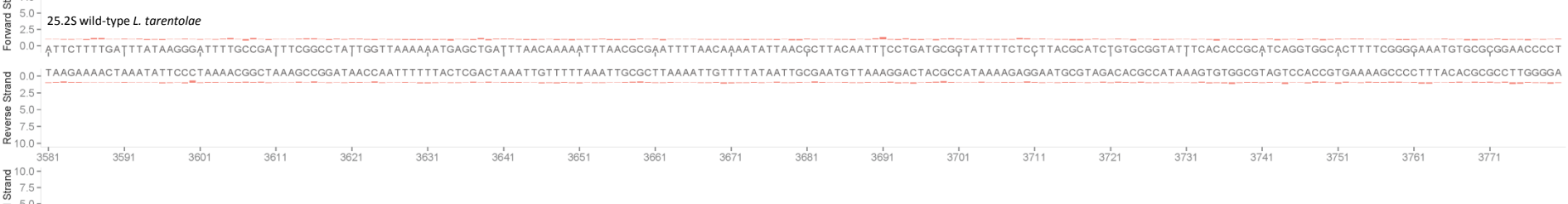
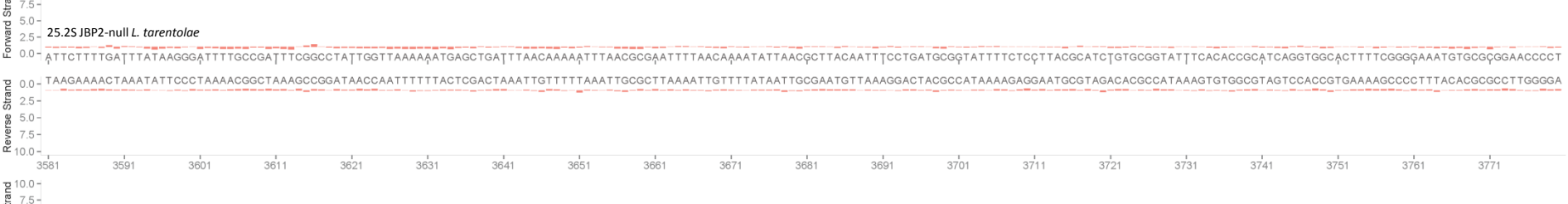
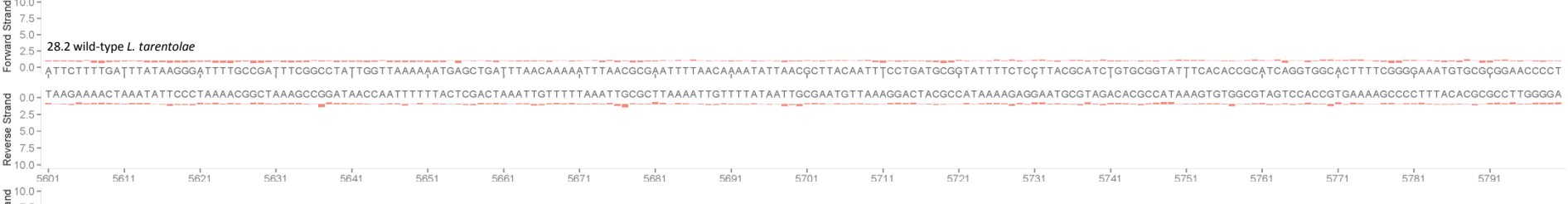
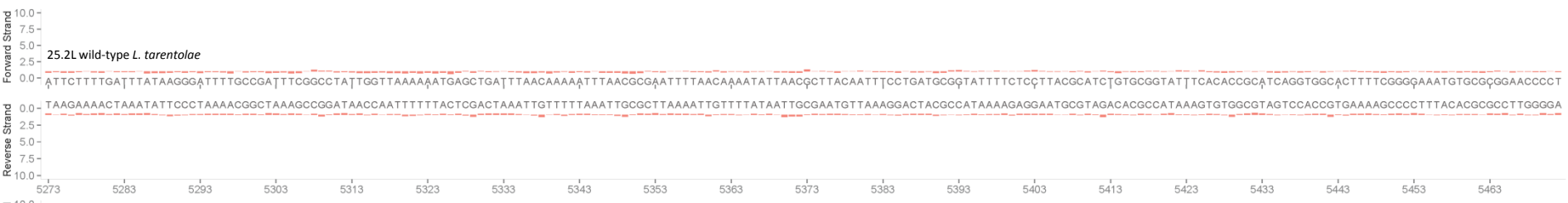
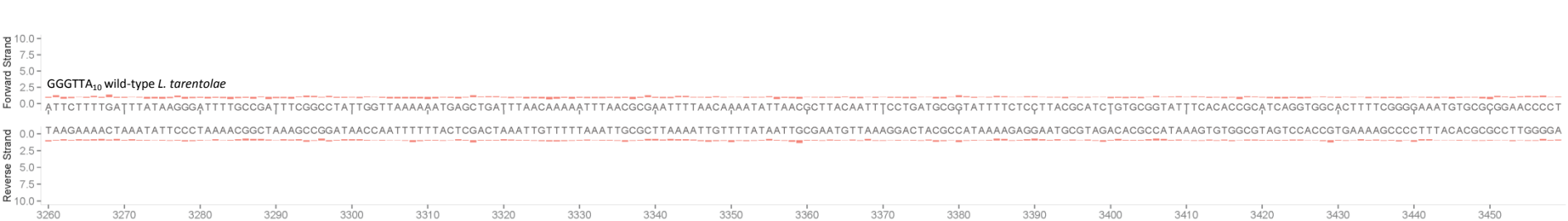


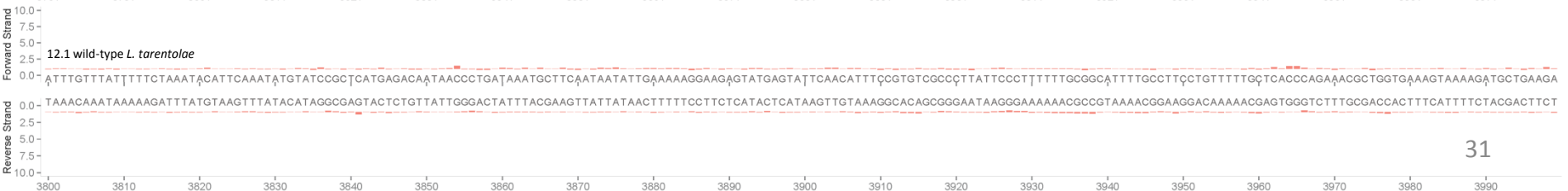
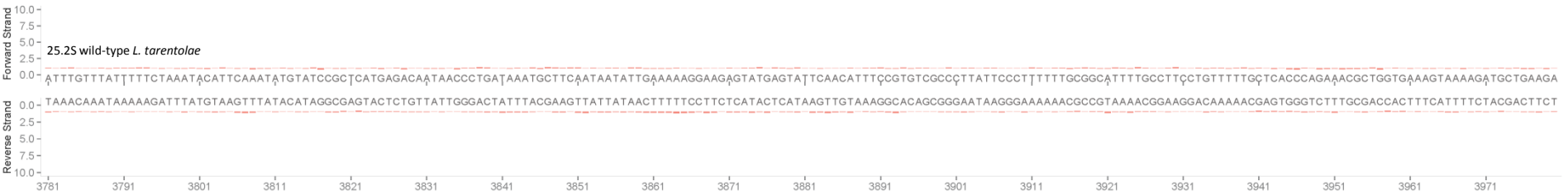
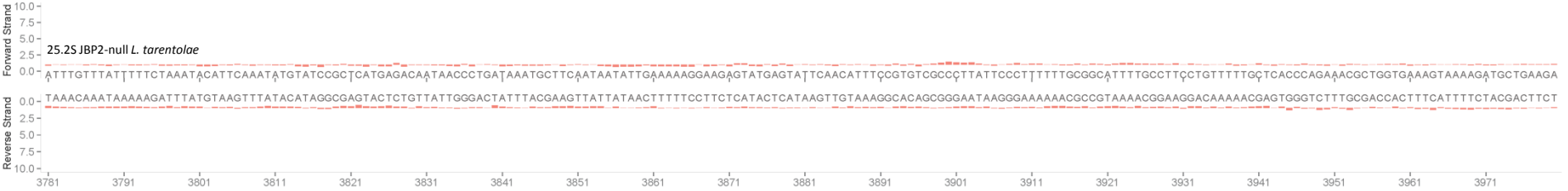
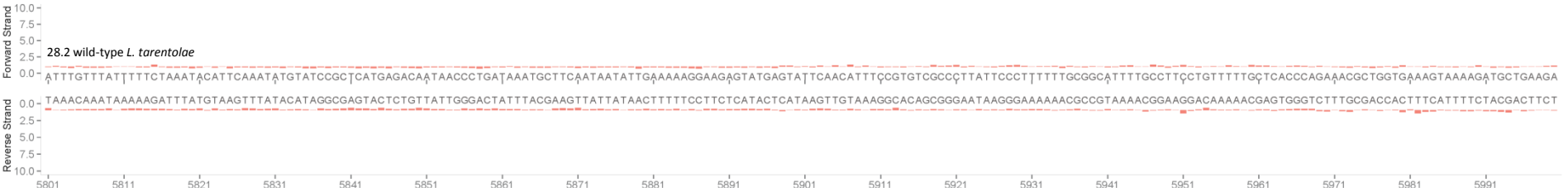
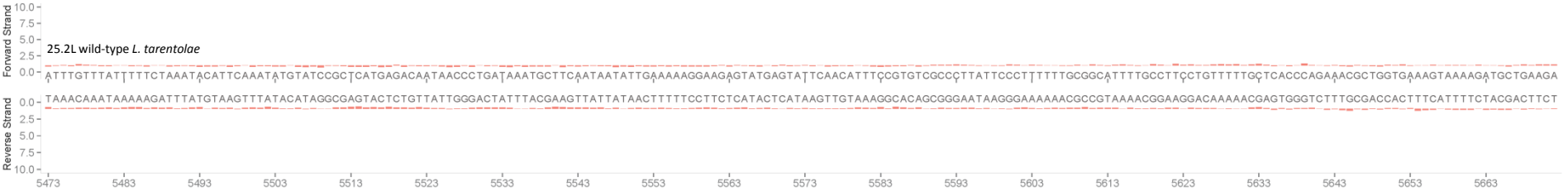
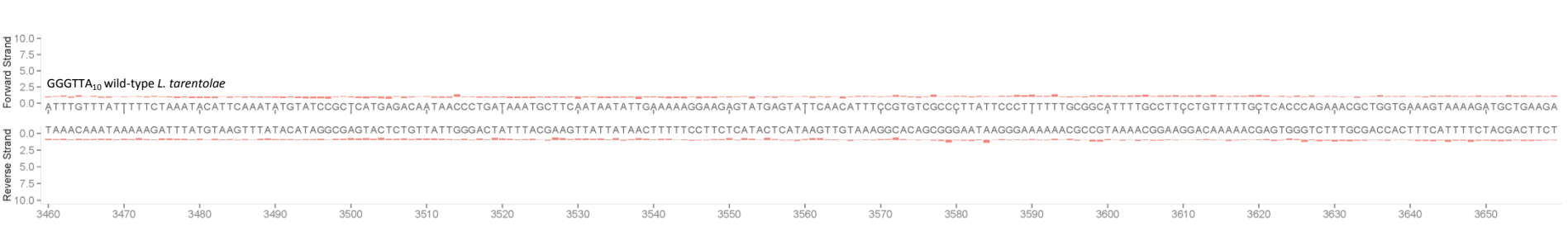






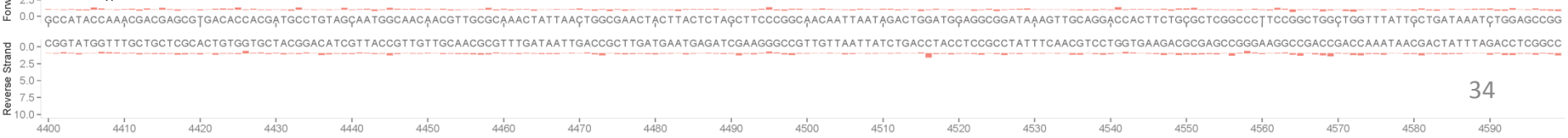
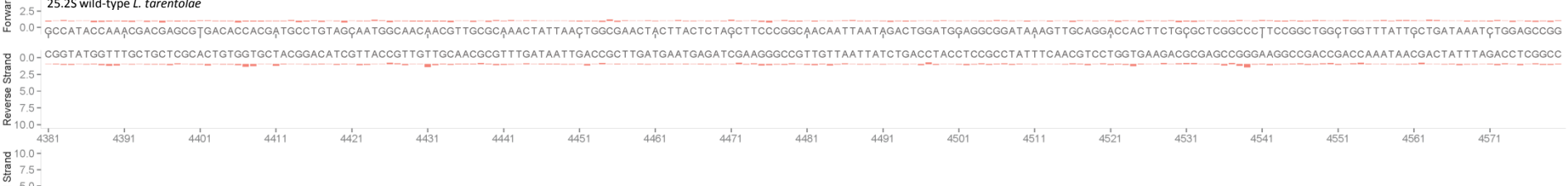
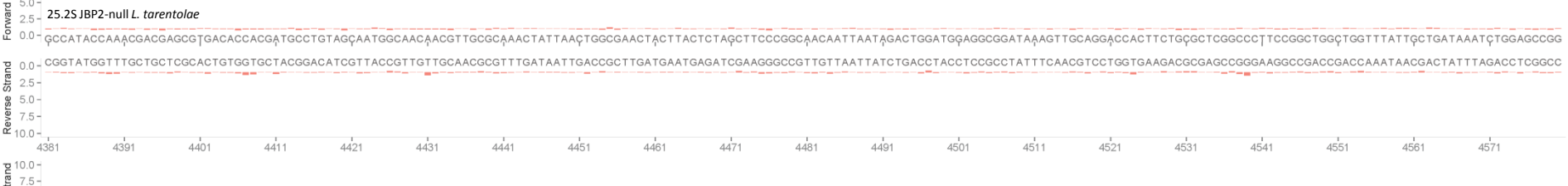
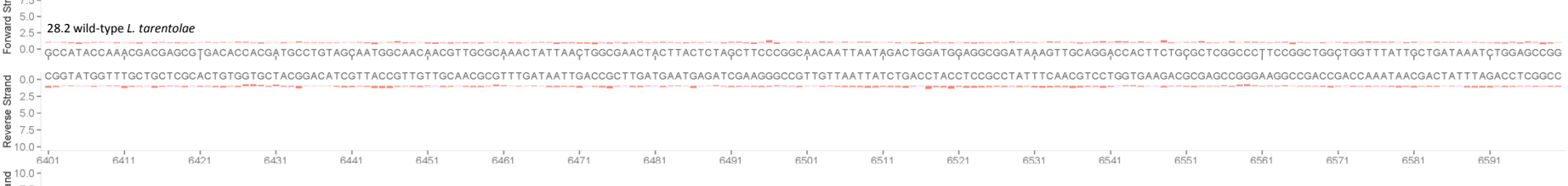
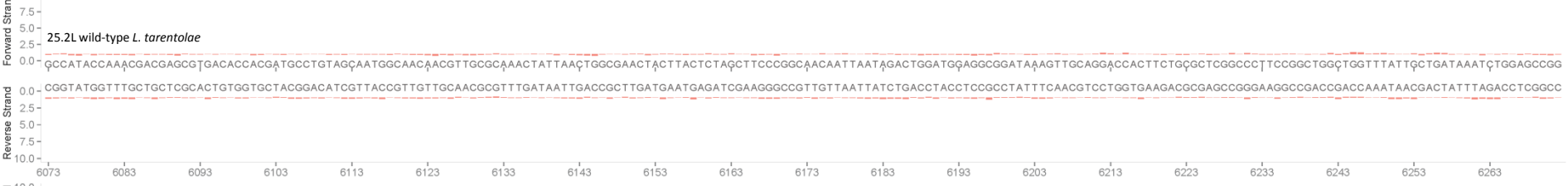
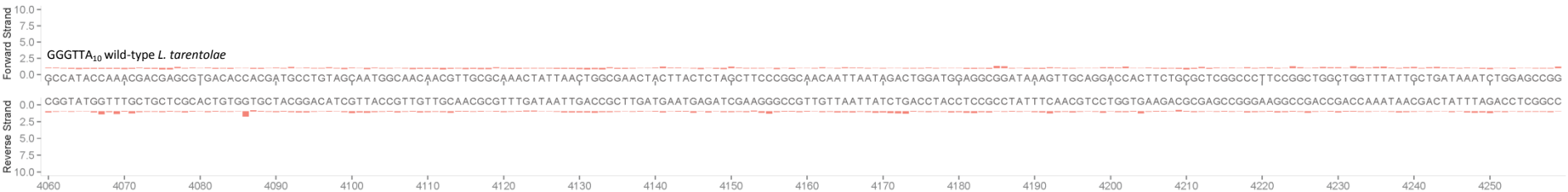


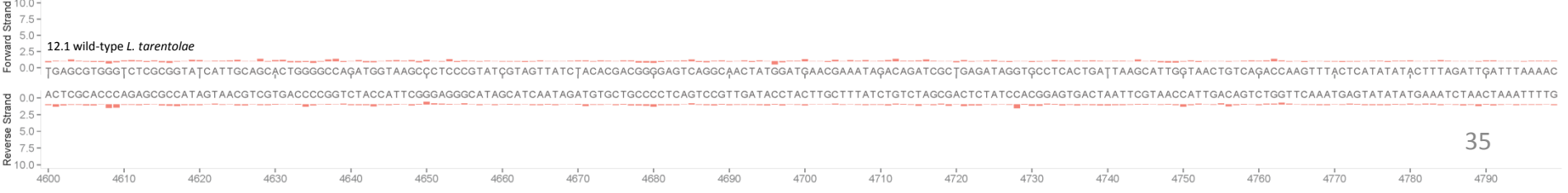
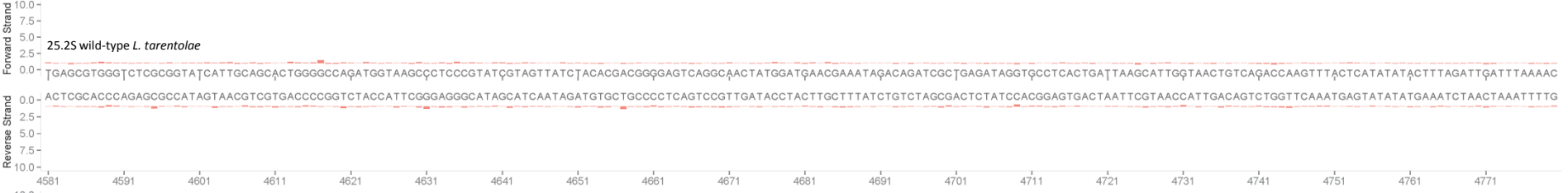
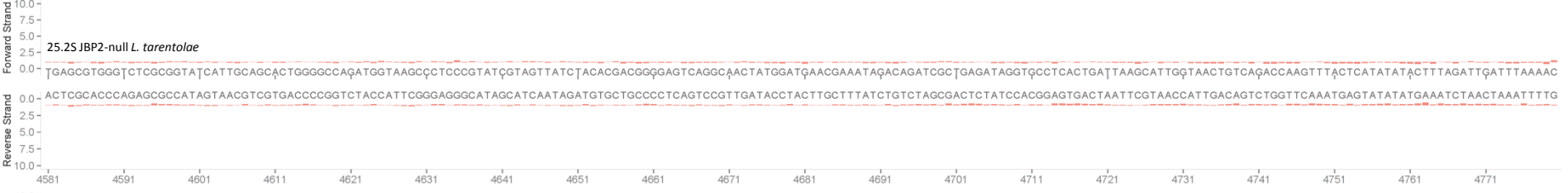
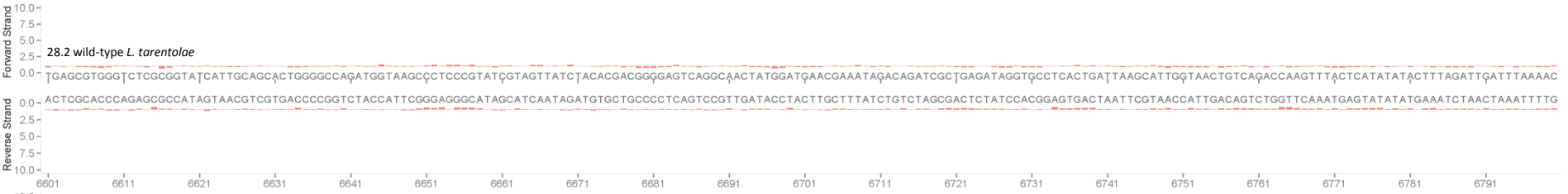
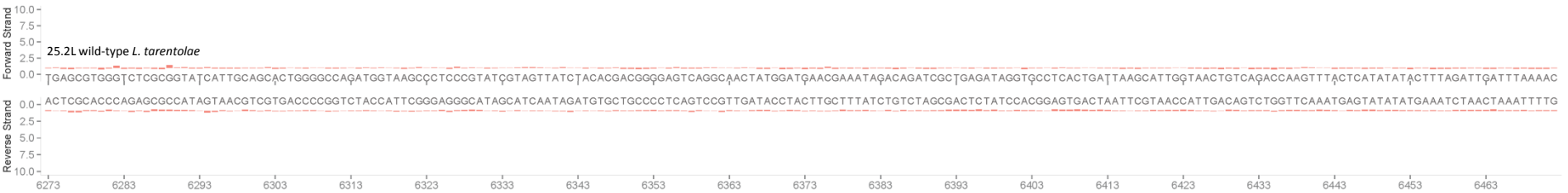
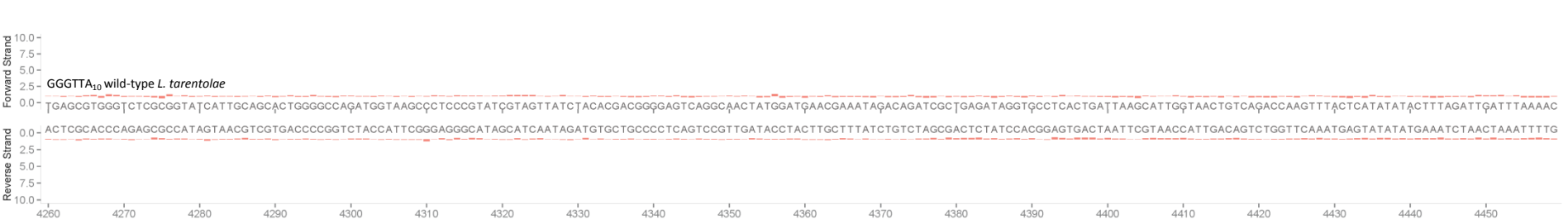


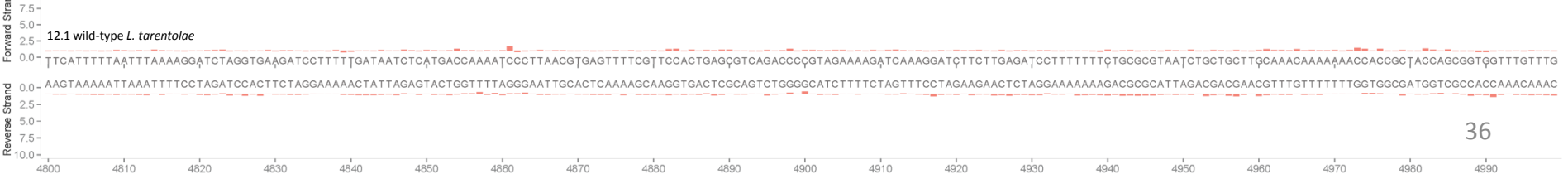
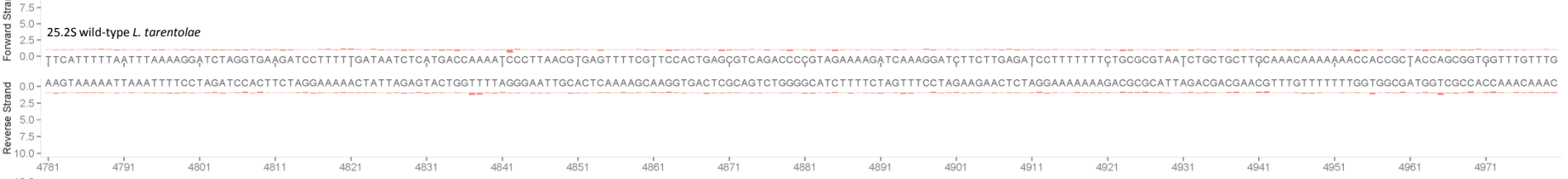
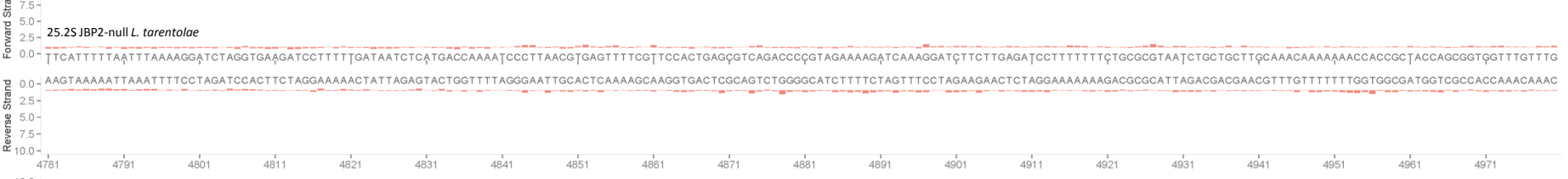
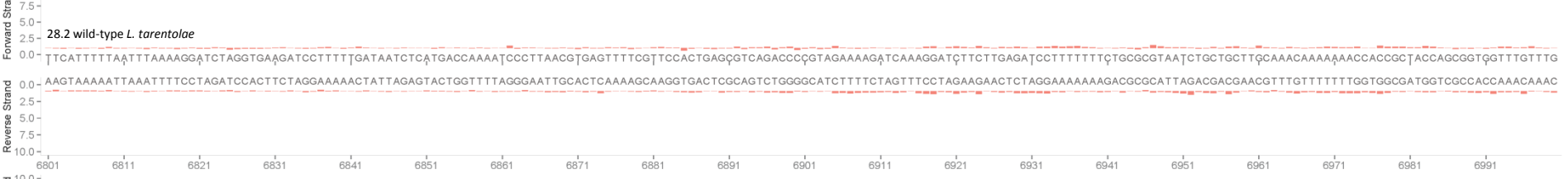
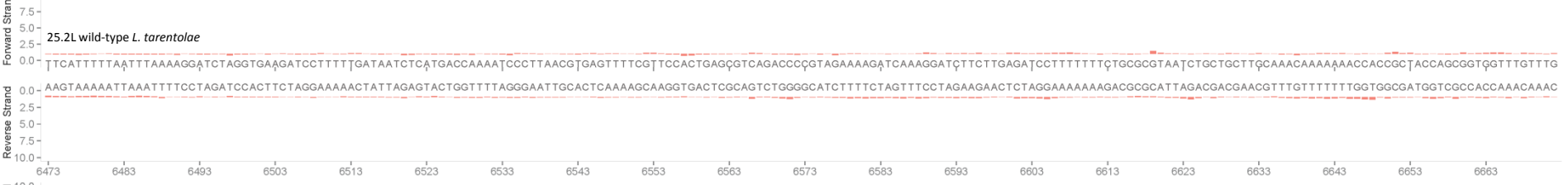
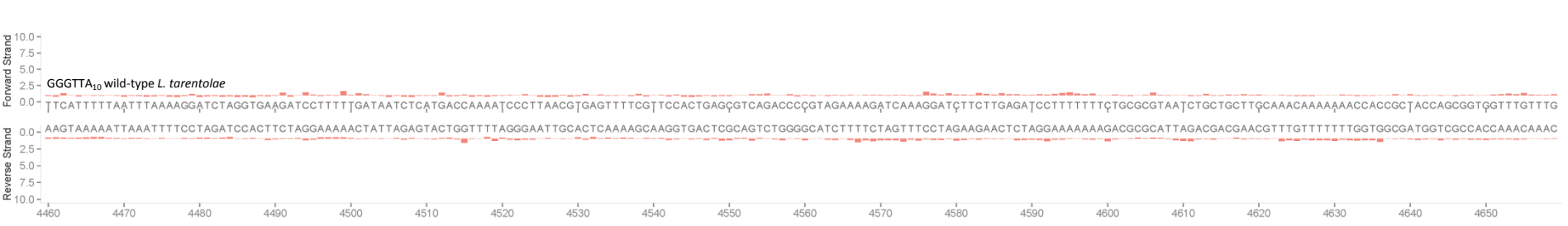


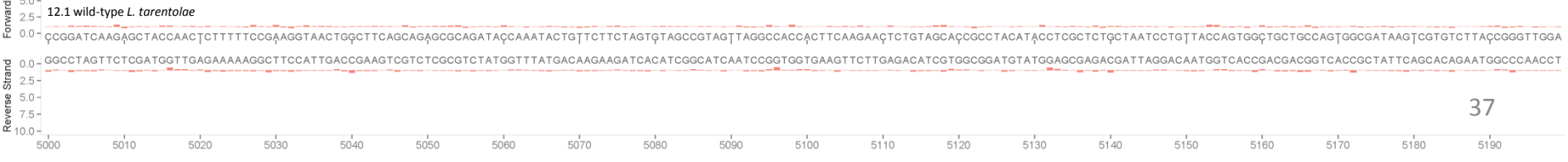
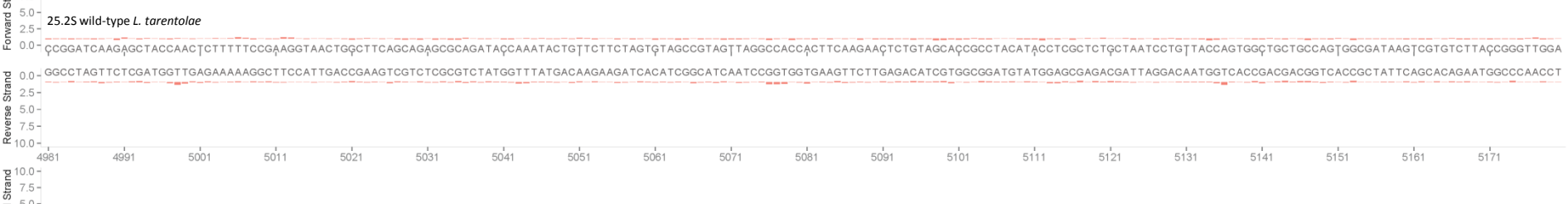
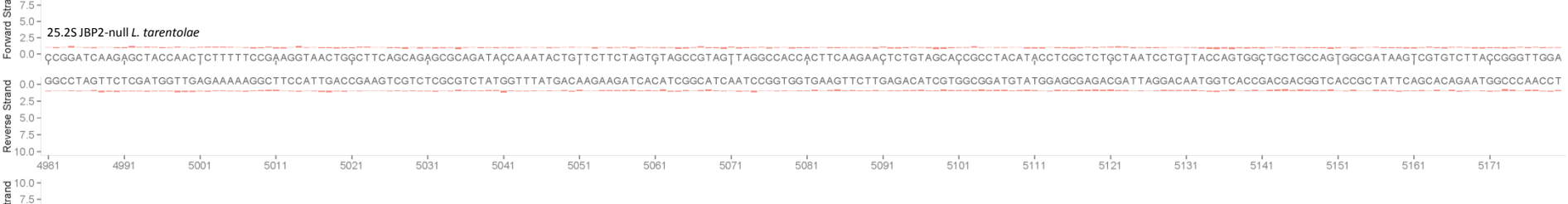
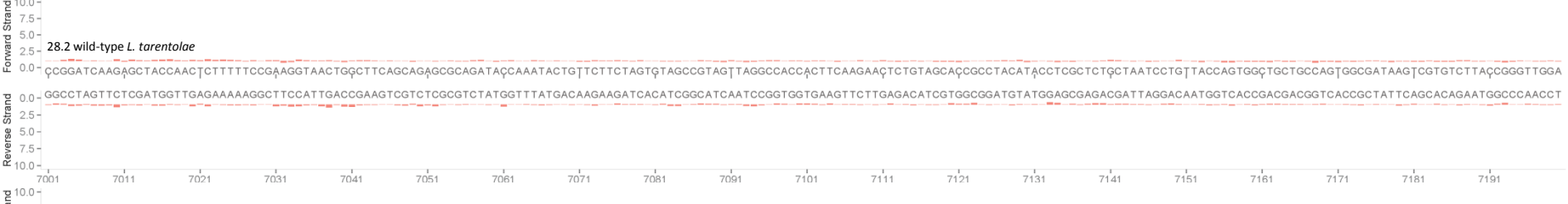
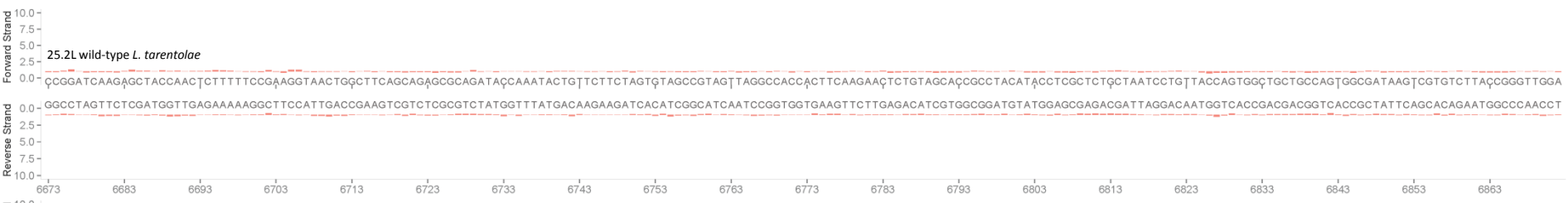
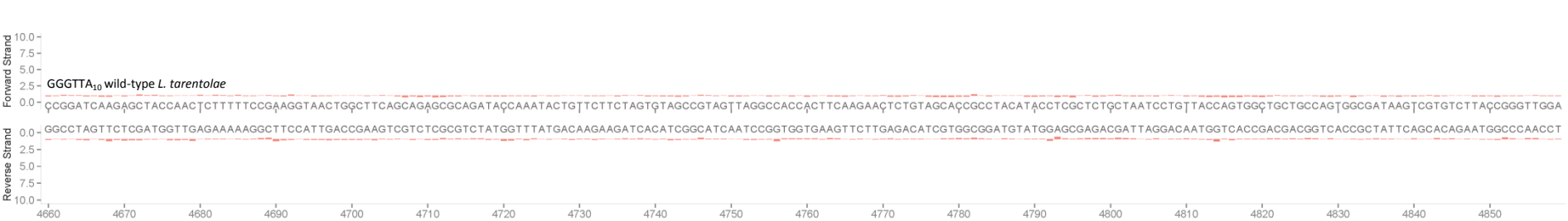


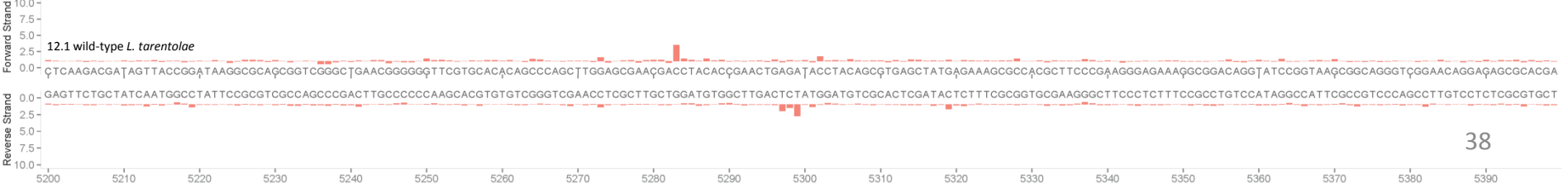
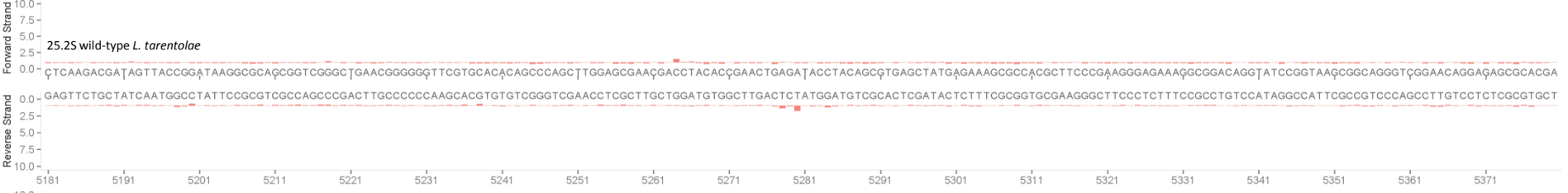
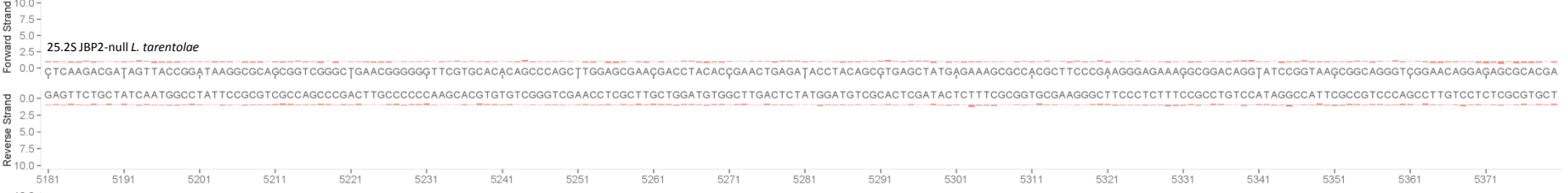
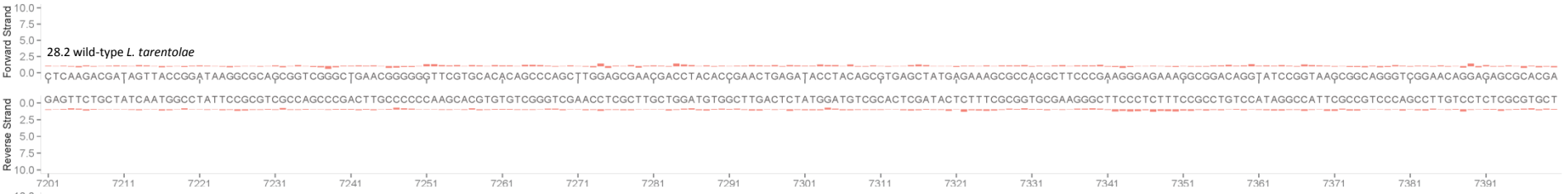
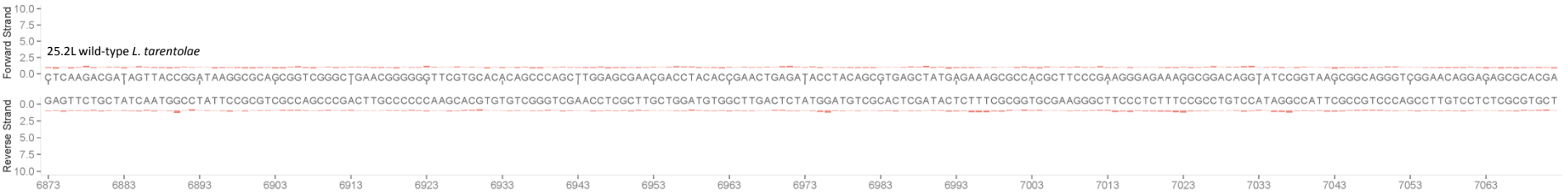
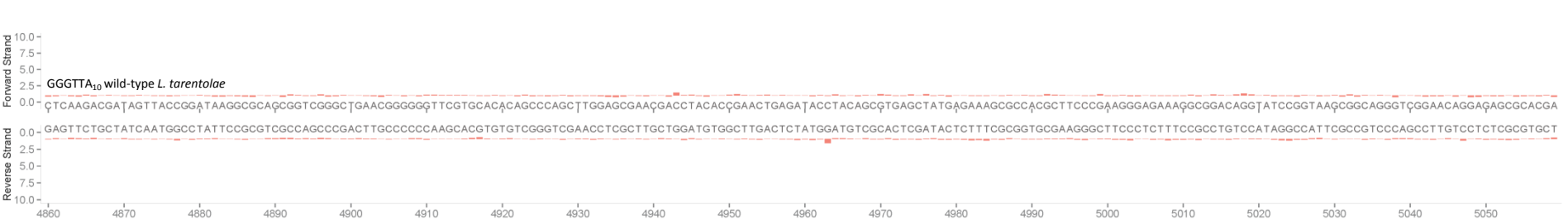


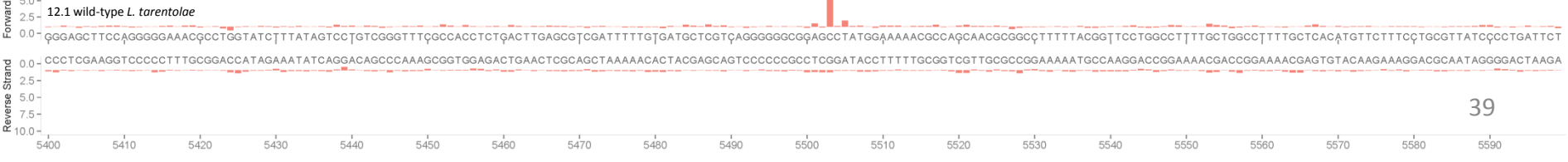
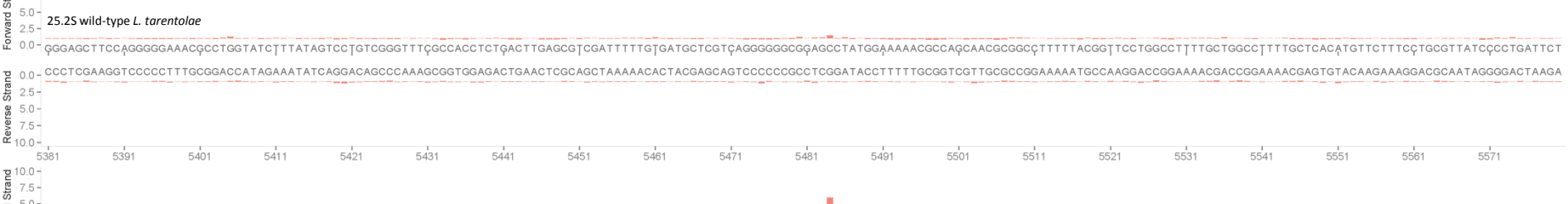
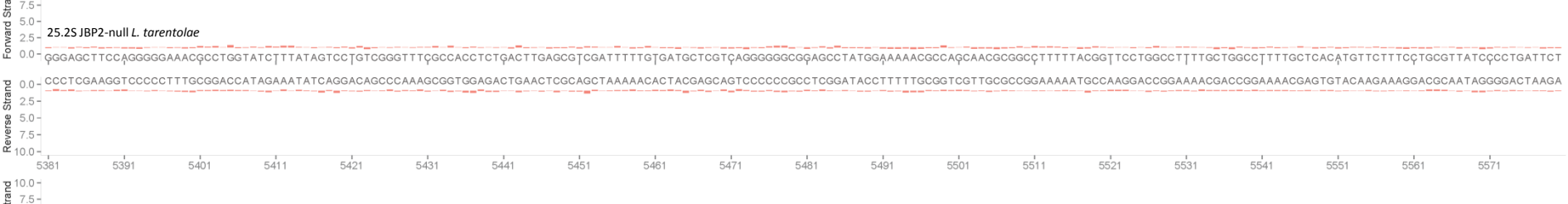
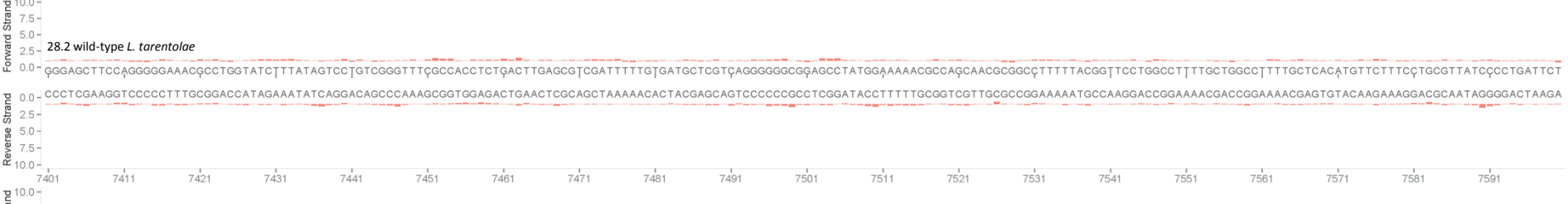
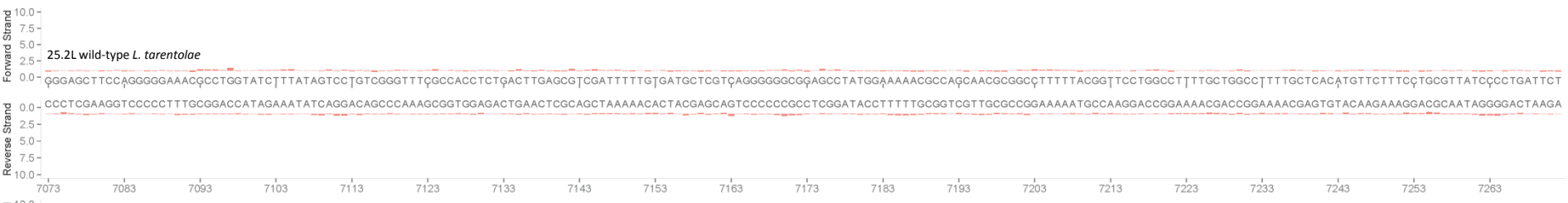
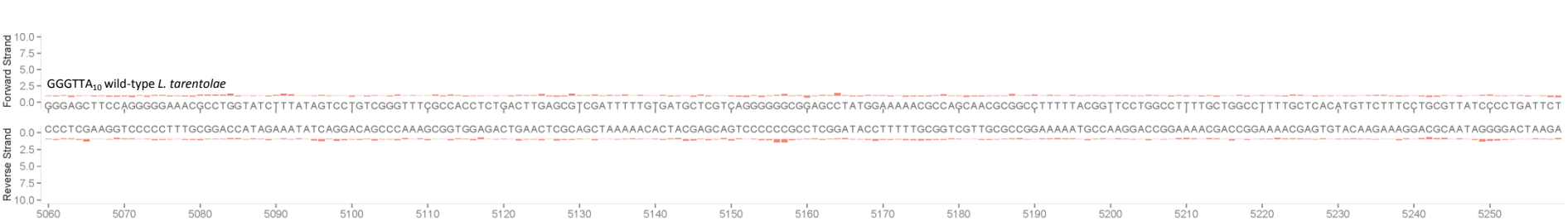


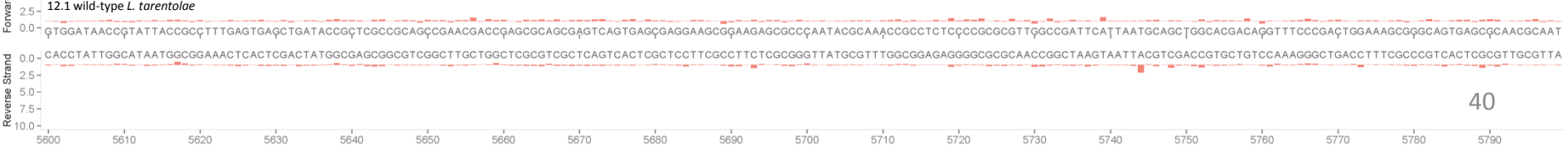
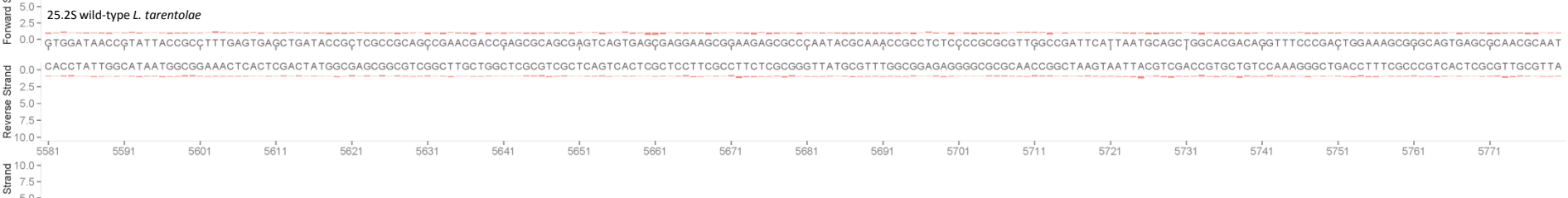
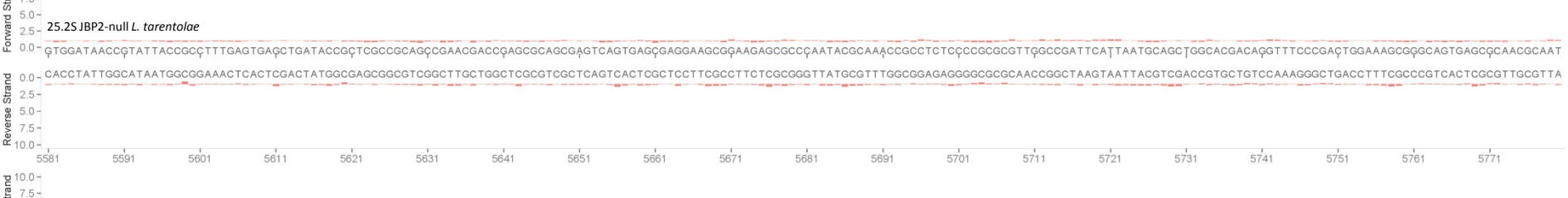
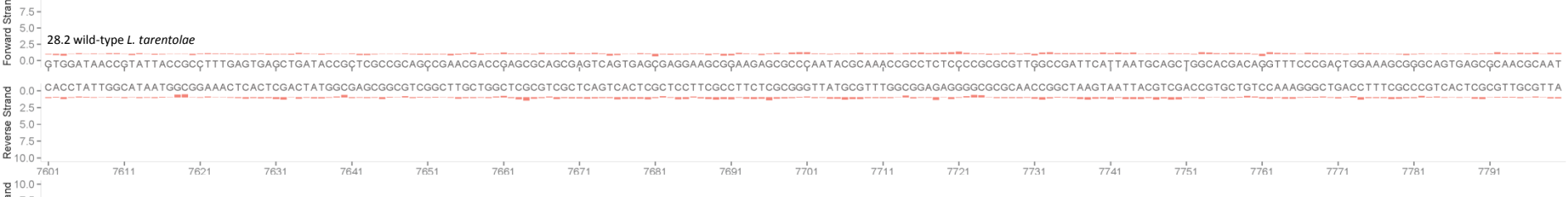
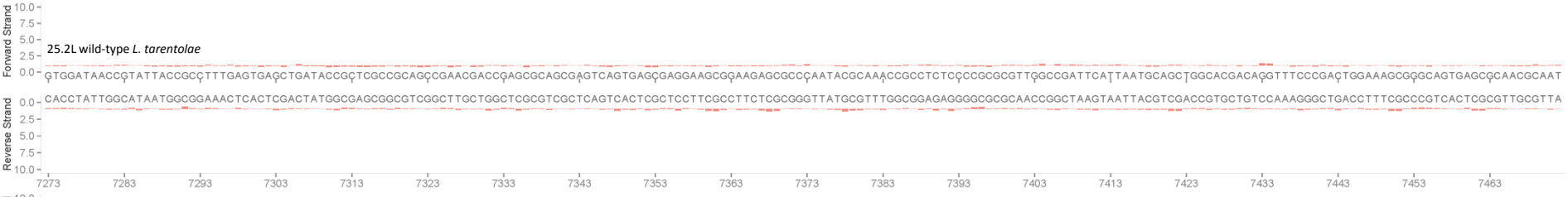
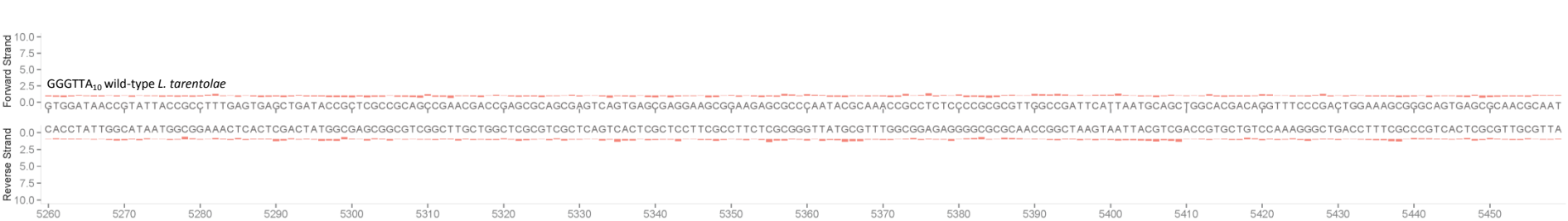


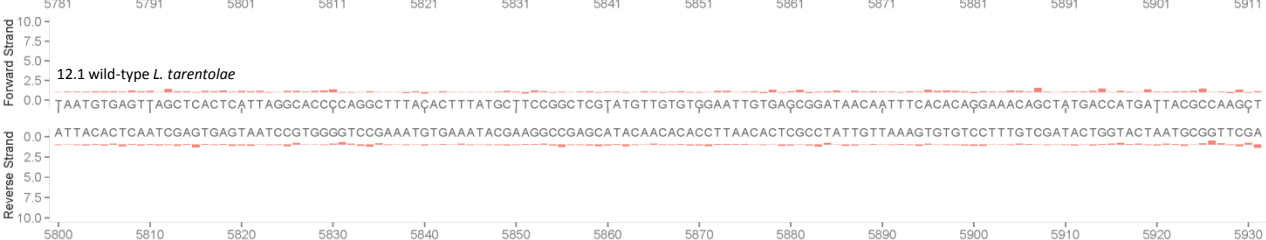
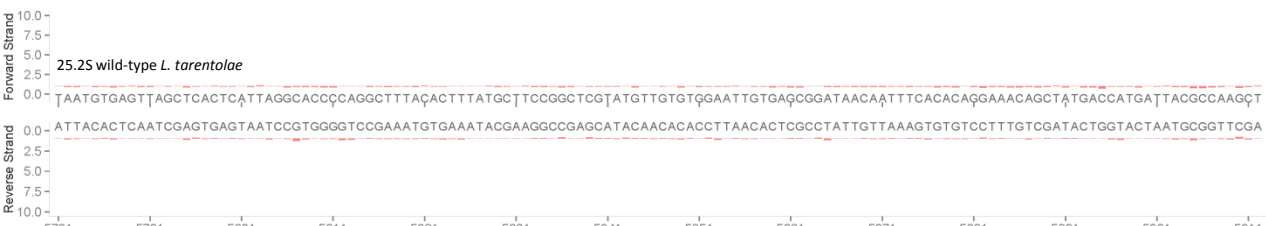
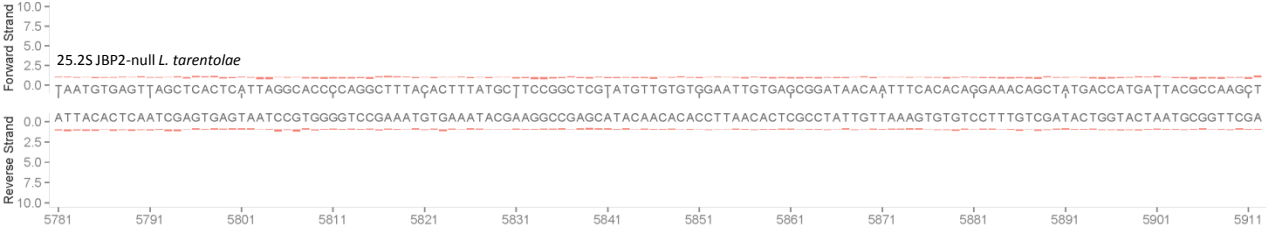
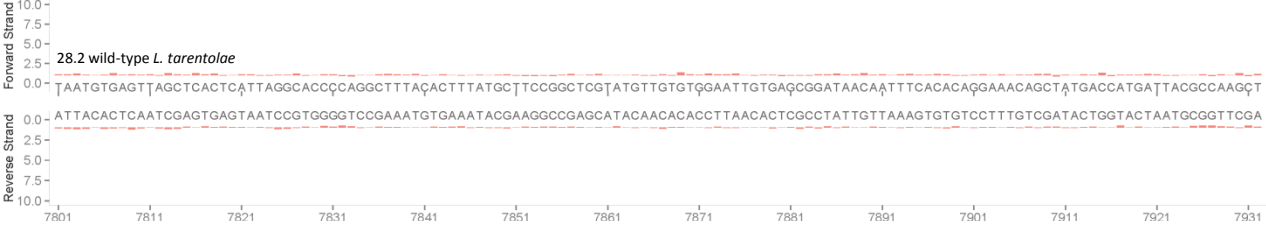
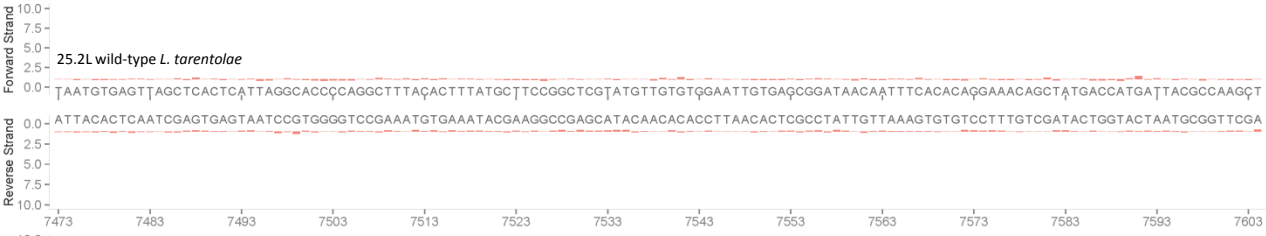
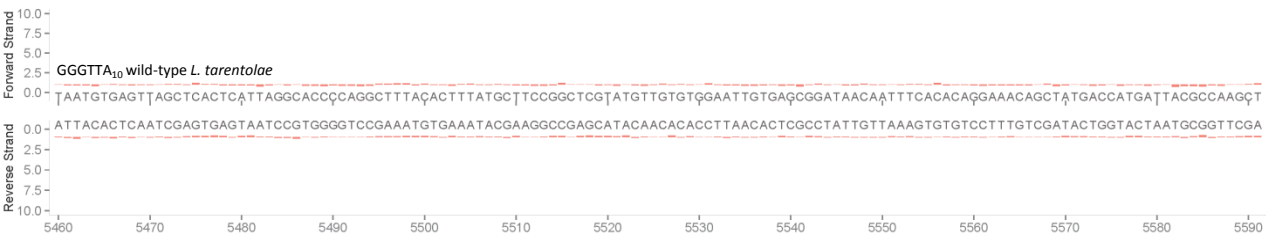


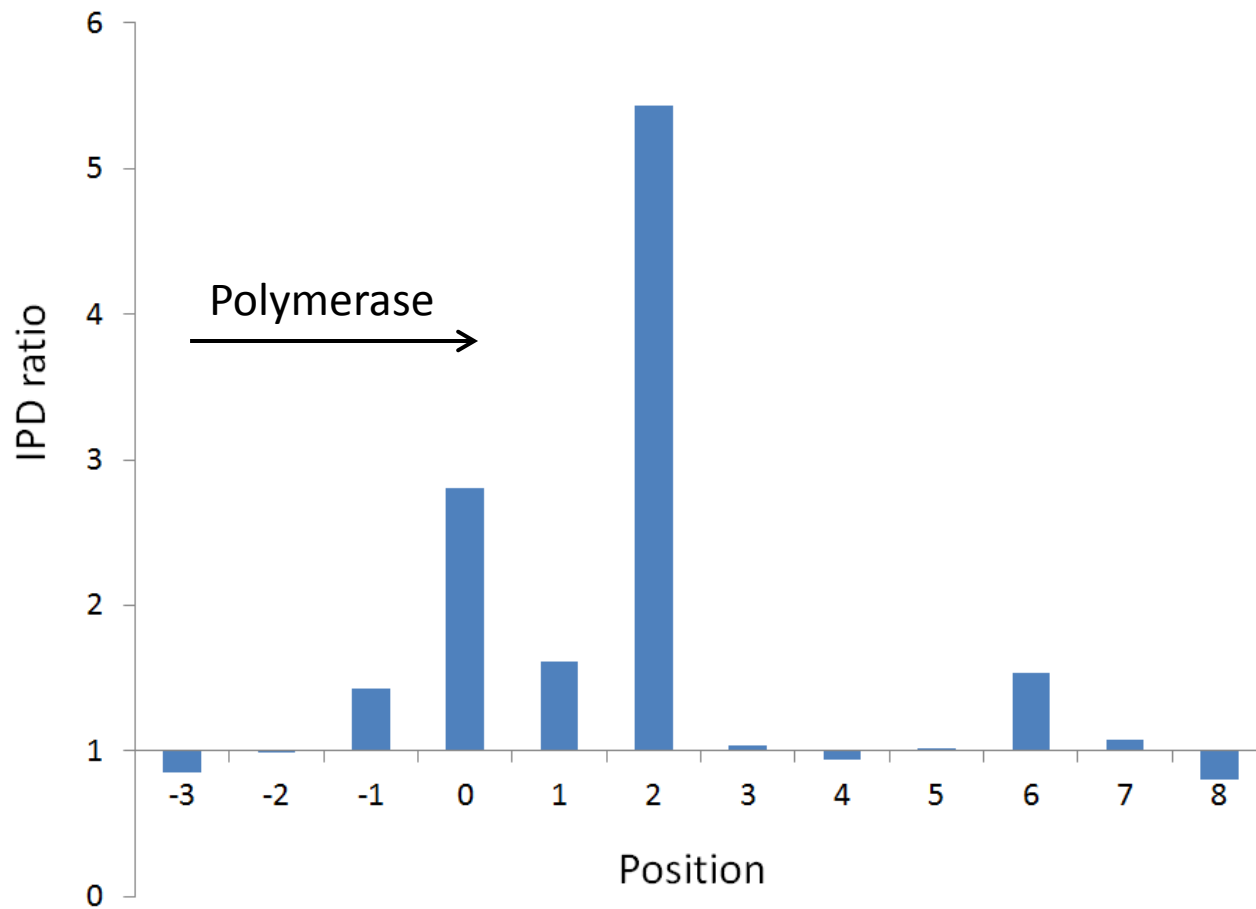












Sequences used

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 CCCTCGCGGCGG
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