# Important biological information uncovered in previously unaligned reads

# from chromatin immunoprecipitation experiments (ChIP-Seq)

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Center for Applied Plant Sciences (CAPS), The Ohio State University, Columbus, OH., USA This supplementary information document includes:

- 1. A description of Supplementary Files
- 2. Supplementary Figure S1 to S9
- 3. Supplementary Table S1 to S3
- 4. Supplementary Sections 3, 4, 5 and 6

#### **Description of Supplementary Files**

Supplementary File 1 as XLS Sheet 1 — Short Read Archive (SRA) accession numbers and dataset description for *A. thaliana* 

Supplementary File 1 as XLS Sheet 2 — Short Read Archive (SRA) accession numbers and dataset description for *H. sapiens* 

Supplementary File 1 as XLS Sheet 3 — Short Read Archive (SRA) accession numbers and dataset description for *D. melanogaster* 

Supplementary File 1 as XLS Sheet 4 — Short Read Archive (SRA) accession numbers and dataset description for *C. elegans* 

Supplementary File 2 as XLS file — Genomic locations of TAL1 recovered peaks (Sheet 1); Genomic locations and target genes of validated TAL1 peaks (Sheet 2)

Supplementary Section 3 — Commands and description of parameters for running Bowtie, SHRiMP & MACS programs; versions of reference genomes used in analysis

Supplementary Section 4 — Published peaks repeat elements

Supplementary Section 5 — Reanalyzed peaks repeat elements

Supplementary Section 6 — TAL1 recovered peaks repeat elements

#### **Supplementary Figures**



**Supplementary Figure S1: Alignment proportions of control ChIP-Seq datasets.** Each bar represents a ChIP-Seq run; several runs constitute an experiment for determining binding patterns of one transcription factor. Runs have been grouped into their respective TFs



## Supplementary Figure S2: Quality scores for maize and Arabidopsis ChIp-Seq reads. Both

aligned and unaligned reads exhibit similar quality scores.



Supplementary Figure S3: Proportions of uniquely aligned, unaligned and multiple-aligned reads for *A. thaliana* and *D. melanogaster* trimmed datasets. Each facet is a ChIP-Seq run in which each bar represents a run whose reads have been trimmed to a particular read length shown on the x-axis.



**Supplementary Figure S4: Proportions of aligned, unaligned and multiple-aligned reads for different mismatches allowed in alignment process.** Each facet is a ChIP-Seq run in which each bar represents a run whose reads have been aligned using the number of allowed mismatches showed on the x-axis.



Supplementary Figure S5: Number of reads in a ChIP-Seq dataset as a function of proportion of unaligned reads from the same dataset. Lack of correlation suggests that sequencing more reads does not guarantee increased read mappability.



Supplementary Figure S6: Taxonomic classification of previously unaligned reads from *A*. *thaliana*, *Z. mays*, *H. sapiens*, and *D. melanogaster* datasets. Taxonomic classification reveals presence of bacterial (blue), metazoan (yellow) and plant derived (green) sequences. The Arabidopsis runs 9-14 correspond to Arabidopsis genome-simulated and aligned reads reads subjected taxonomic classification as a positive control (See Supplementary File 1 Sheets 5, 6 & 7 for the accession numbers corresponding to each number in the figure).



Supplementary Figure S7: A Receiver Operating Characteristic Curve illustrating the trade-off between true positive and false positive rates of selected next generation sequencing alignment programs



Supplementary Figure S8: Motifs enriched in published (a), reanalyzed (b) and recovered (c) peaks. Statistically over-represented motifs were identified using the *de novo* motif discovery tool MEME.



**Supplementary Figure S9: ChIP-qPCR validation of selected recovered TAL1 peaks.** Each bar represents fold enrichment of peaks relative to IgG. Error bars represent standard error. Statistical significance is as a result of t-test between the validated peak enrichment and the negative controls of ACTIN, GAPDH, and negative peak; n=3. Actual p-value is indicated when > 0.05, otherwise p-value < 0.05 indicated by \*; < 0.01 by \*\*; and < 0.001 by \*\*\*;- ve: negative.

# Supplementary Tables

# Supplementary Table S1: Average GC content for the five genomes analyzed.

Genome	GC content, %
H. sapiens	40.38
C. elegans	35.44
D. melanogaster	41.51
Z. mays	46.81
A. thaliana	36.05

# Supplementary Table S2: GO term analysis of genes closest to TAL1 published, reanalyzed and

# recovered peaks

Published peaks GO terms:	-log <sub>10</sub> ( <i>p</i> -value)
T- cell differentiation	16.36
T-cell activation	16.09
T-cell receptor signaling pathway	13.41
Reanalyzed peaks GO terms:	
T-cell activation	18.74
T-cell differentiation	18.49
T-cell receptor signaling pathway	14.82
Recovered peaks GO terms:	
Pentose-phospahate shunt pathway	27.47
Sensory perception	17.12
Olfactory transduction	13.39
Apoptosis	2.38
T-cell activation	1.42

Supplementary Table S3: Repeat elements identified published (A), reanalyzed (B) and recovered peaks (C)

# A: Published peaks

	Number of elements	Length occupied, bp	Percentage of sequence, %
Satellites	0	0	0.00
Simple repeats	93	3888	0.50
Low complexity	22	941	0.12

# **B:** Reanalyzed peaks

	Number of elements	Length occupied, bp	Percentage of sequence, %
Satellites	2	1144	0.17
Simple repeats	65	4476	0.67
Low complexity	9	401	0.06

# C: Recovered peaks

	Number of elements	Length occupied, bp	Percentage of sequence, %
Satellites	121	61773	18.56
Simple repeats	188	31934	9.59
Low complexity	55	7685	2.31

#### Section 3

#### Bowtie alignment command and parameters:

/nfs/15/osu4902/bowtie -p 10 -q -v 3 -m 1 -S --best --strata --chunkmbs 128 -t /nfs/15/osu4902/bowtie\_db/a\_thaliana SRR016811.fastq ../sam/SRR016811.sam --un ../unaligned/SRR016811\_unaligned.fq --al ../aligned/SRR016811\_aligned.fq --max ../aligned/SRR016811\_multiple\_align.fq

Parameters:

- -p <integer>: number of processor core threads used in alignment
- -q: input files are in FASTQ format
- -v <integer>: report end-to-end hits with less than or equal v mismatches
- -m <integer>: maximum number of multiple alignments allowed
- -S: write hits in SAM format
- --best --strata: only hits in best stratum are reported
- --chunkmbs <integer>: maximum megabytes for RAM for best first search frames
- -t: print wall-clock time taken by search phases
- --un <filename>: write unaligned reads to <filename>
- --al <filename>: write aligned reads to <filename>
- --max <filename>: write multiple-aligned reads to to <filename>

#### SHRiMP alignment command and parameters:

/nfs/15/osu4902/applications/SHRiMP\_2\_2\_0/bin/gmapper-ls --qv-offset 33 -L /nfs/proj14/PAS0107/reads\_recovery/hg18-20gb-12\_12\_12\_12seeds-"\$i"of3-ls --strata --maxalignments 1 -N 12 --un SRR054882\_homo\_sapiens\_\${i}of3\_unaligned.fq -E -Q SRR054882\_homo\_sapiens\_sequence\_reads.fq >SRR054882\_homo\_sapiens\_\${i}of3.sam

Parameters: --qv-offset <integer>: interpret quality values in fastq input as PHRED+<integer> -L: reference genome idex --strata: print only best scoring hits --max-alignments <integer>: maximum alignment per read -N <integer>: number of processor core threads --un <filename>: write unaligned reads to <filename> -E: output SAM format -Q: input reads are in FASTQ format

### MACS peak calling command and parameters:

macs14 -t ChIP.bam -c Control.bam -f BAM -g h -n test -w -p 1e-5 -B

Parameters:

- -t: ChIP-seq treatment files
- -c: control files
- -f: format of alignment file
- -g: effective genome size
- -n: experiment name
- -w: Whether or not to save extended fragment pileup into a wiggle file
- -p: Pvalue cutoff for peak detection
- -B: Whether or not to save extended fragment pileup at every bp into a bedGraph file

### Genome versions:

- A. thaliana: TAIR9 genome release
- Z. mays: B73 version 2 (ZmB73\_RefGen\_v2)
- H. sapiens: version 19 (GRCh37/hg19)
- C. elegans: version WS200
- D. melanogaster: release 5.22

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	MIRS	0	0	qa	0.00 %	
LINEs	:	0	0	gd	0.00 %	
	LINE1	0	0	bp	0.00 %	
	LINE2	0	0	bp	0.00 %	
	L3/CR1	0	0	bp	0.00 %	
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Section 4

\* most repeats fragmented by insertions or deletions have been counted as one element

run with rmblastn version : 2.2.27+ The query was compared to classified sequences in ".../RepeatMasker.lib" RepBase Update 20110419-min, RM database version 20110419-min

#### 17

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file r	name: SRR070	589-origina	al_peaks.	ped.1	fasta	
sequer	nces:	3344				
total	length:	666435 bp	(666435	bp e	excl N/X·	-runs)
GC lev	vel:	48.99 %				
bases	masked:	5264 bp	( 0.79 %)	)		
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Small	RNA:	0	0	bp	0.00 \$	ō
Satell	ites:	2	1144	bp	0.17 9	2
Simple	repeats:	65	4476	bp	0.67	-
Low co	mplexity:	9	401	bp	0.06	
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The query species was assumed to be homo RepeatMasker version open-3.3.0 , default mode

run with rmblastn version : 2.2.27+ The query was compared to classified sequences in ".../RepeatMasker.lib" RepBase Update 20110419-min, RM database version 20110419-min

#### Section 5

file name: SRR070589_recovered_peaks.xls.bed.fasta sequences: 594 total length: 332838 bp (332697 bp excl N/X-runs GC level: 43.97 % bases masked: 89271 bp ( 26.82 %)
total length: 332838 bp (332697 bp excl N/X-runs   GC level: 43.97 %   bases masked: 89271 bp (26.82 %)
GC level: 43.97 % bases masked: 89271 bp (26.82 %)
bases masked: 89271 bp ( 26.82 %)
number of length percentage
elements* occupied of sequence
SINEs: 0 0 bp 0.00 %
ALUS 0 0 bp 0.00 %
MIRs 0 0 bp 0.00 %
LINEs: 0 0 bp 0.00 %
LINE1 0 0 bp 0.00 %
LINE2 0 0 bp 0.00 %
L3/CR1 0 0 bp 0.00 %
LTR elements: 0 0 bp 0.00 %
ERVL 0 0 bp 0.00 %
ERVL-MalRs 0 0 bp 0.00 %
ERV_classI 0 0 bp 0.00 %
ERV_classII 0 0 bp 0.00 %
DNA elements: 0 0 bp 0.00 %
hAT-Charlie 0 0 bp 0.00 %
TcMar-Tigger 0 0 bp 0.00 %
Unclassified: 0 0 bp 0.00 %
Total interspersed repeats: 0 bp 0.00 %
Small RNA: 0 0 bp 0.00 %
Satellites: 121 61773 bp 18.56 %
Simple repeats: 188 31934 bp 9.59 %
Low complexity: 55 7685 bp 2.31 %

\* most repeats fragmented by insertions or deletions have been counted as one element

The query species was assumed to be homo RepeatMasker version open-3.3.0 , default mode

run with rmblastn version : 2.2.27+ The query was compared to classified sequences in ".../RepeatMasker.lib" RepBase Update 20110419-min, RM database version 20110419-min

#### Section 6