



Supplementary Figure 1 | Parameters of TAP activity in monocytes are reminiscent of those in Raji cells.

(a) Flow cytometry-based translocation assay was carried out in monocytes in the absence or presence of 2 μM of the high-affinity epitope R9 and a fluorescently labeled epitope lacking the N-core glycosylation site (C4-F). The detected mean fluorescence intensities (530-nm channel) were normalized to the NST-F/ATP sample. The error bars indicate the standard error of mean ($n \geq 4$). (b) The TPT assay was carried out using 10 nM of reporter peptide NST-F and increasing concentrations of ATP. Translocation rates relative to the ADP control values are plotted versus ATP concentrations ($n = 2$). (c) The TPT assay was carried out using 10 mM ATP and increasing concentrations of NST-F peptide. Translocation rates relative to the ADP control values are plotted versus NST-F concentrations ($n = 2$). (d) Simultaneous dual flow cytometric detection of peptide translocation in the 530-nm channel (right panel) and ICP47^{AT565} binding in the 488/575-nm channel (left panel) was carried out for increasing concentrations of ICP47^{AT565} in monocytes. Numbers indicate the concentration of ICP47^{AT565} in μM (left panel) and the mean percent of transport (right panel). As a control, peptide translocation was carried out in presence of DMSO. (e) The mean fluorescence intensities detected in the 530-nm channel were ADP background subtracted and plotted against the increasing ICP47^{AT565} concentrations in order to determine the IC₅₀ value (13 nM with a 95% confidence interval from 9 nM to 18 nM) of ICP47^{AT565} binding (dotted line) in monocytes. The error bars indicate the standard error of mean ($n = 2$).