

File S1

Strain construction

C. albicans strain AF1006 producing C-terminally HA-tagged Sch9 was constructed by transformation of heterozygous strain CAS2 by a tagging cassette generated by oligonucleotides Sch9-HA for/rev, as described (SCHAEKEL *et al.* 2013). Correct chromosomal integration was verified by colony PCR using primers Sch9ver and 3' test HA-tag. Both alleles of *SCH9* were deleted in *C. albicans* strain RM1000AH (SANYAL *et al.* 2004) and 8675 (JOGLEKAR *et al.* 2008) using the URA blaster method. The construction of the URA blaster deletion cassette for *SCH9* was described previously (STICHTERNOOTH *et al.* 2011). After the deletion of the first copy, the heterozygous strains were grown on 5-FOA plate to make the cells auxotroph for *URA3* to obtain RMKS1A, RMKS1B and 8675t. Then the same cassette was again used to disrupt the second allele of the gene, to get strains RMKS2A, RMKS2B and 8675T, respectively. To obtain the re-integrant of Sch9 in heterozygous and homozygous mutant background, the entire ORF along with its promoter and terminator was cloned in *KpnI* and *SalI* sites in Clp10 integration vector (MURAD *et al.* 2000). Sch9 orf was re-integrated at *RPS10* locus in the *Candida* genome using *StuI* to obtain RMKS1AR, RMKS1BR, RMKS2AR and RMKS2BR. The correct chromosomal integration of Clp10 was verified by PCR using primers UP-RPS10 and NV207. To check the binding pattern of CENP-A across the centromere in *sch9* mutant by ChIP, one copy of CENP-A was tagged with Prot A using plasmid construct pCaCse4TAPNAT (THAKUR and SANYAL 2013). pCaCse4-TAP-NAT was partially digested with *XhoI* and transformed into RMKS2A to get Prot A tagged CENP-A strain.