

**Sumoylation of CCAAT/enhancer-binding protein α is implicated in
hematopoietic stem/progenitor cell development through regulating *runx1* in zebrafish**

Hao Yuan¹, Tao Zhang¹, Xiaohui Liu¹, Min Deng², Wenqing Zhang³, Zilong Wen⁴, Saijuan Chen¹,
Zhu Chen¹, Hugues de The^{1,5}, Jun Zhou^{1,*} and Jun Zhu^{1,5,*}

Supplementary Figure Legends

Figure S1. HSPCs are reduced in SUMO-deficient embryos. (A) WISH assay of *runx1* at 30 hpf. Boxed regions indicate the AGM. Blue arrows identify *runx1*-positive cells in the AGM. (B) Fluorescent images of the Tg (*cmyb*-EGFP) line at 72 hpf. Boxed regions indicate the CHT. White arrows identify *cmyb*-positive cells in the CHT. (C) Quantification of *cmyb*-EGFP positive cells in the CHT region of Tg (*cmyb*-EGFP) line at 72 hpf. Data shown are the mean \pm SD, $n \geq 3$, * $P < 0.01$ by student's *t*-test. (D) WISH assay of *cmyb* in the *p53* mutant embryos at 72 hpf.

Figure S2. Apoptosis of HSPC is not affected in SUMO-deficient embryos. (A-I) Double immunostaining of *cmyb*-EGFP (A-C) and TUNEL (D-F) in the CHT of Tg (*cmyb*-EGFP) line at 72 hpf. The bottom panel shows merged images (G-I). (J) Quantification of TUNEL and *cmyb*-EGFP double positive cells in the CHT of Tg (*cmyb*-EGFP) line at 72 hpf. Data shown are the mean \pm SD, $n \geq 5$, P values stand for student's *t*-test.

Figure S3. WISH and Luciferase activity assays. (A) WISH assay of *runx1* at 36 hpf.

Overexpression of *SUMO-cebpa* could rescue the HSPC defects of SUMOs morphant. Blue arrows identify *runx1*-positive cells in the AGM. (B) Luciferase activity assays were performed in 293T cells using various *C/ebpα* constructs indicated. The Renilla plasmid was used as an internal control. Note that *SUMO-C/ebpα ΔBR* lost the repressive effect on the transcriptional activity of WT *C/ebpα*. (C) WISH assay of *cmyb* at 72 hpf. Overexpression of *SUMO-cebpa*, *POZ-cebpa* or *SUMO-cebpa ΔBR* alone had no obvious effect on HSPC development.

Figure S4. Targeted disruption of *cebpa* gene with TALENs in zebrafish. (A) Partial structure and sequence of the *cebpa* gene, showing the target sites (underlined) of the left and right TALENs. The *Sal* I site used for detecting mutations is highlighted in red. (B) Sequencing results of *cebpa* mutation induced by TALENs in F1 fish line. The TALENs binding sites are underlined. Deletions are indicated by dashes. (C) Sudan black staining of embryos from incross of heterozygous *cebpa* mutants at 72 hpf. Red arrows indicated Sudan black positive cells in the CHT. (D) Genotyping of the corresponding embryos in (C) by genomic PCR amplification and *Sal* I digestion.

Figure S1.

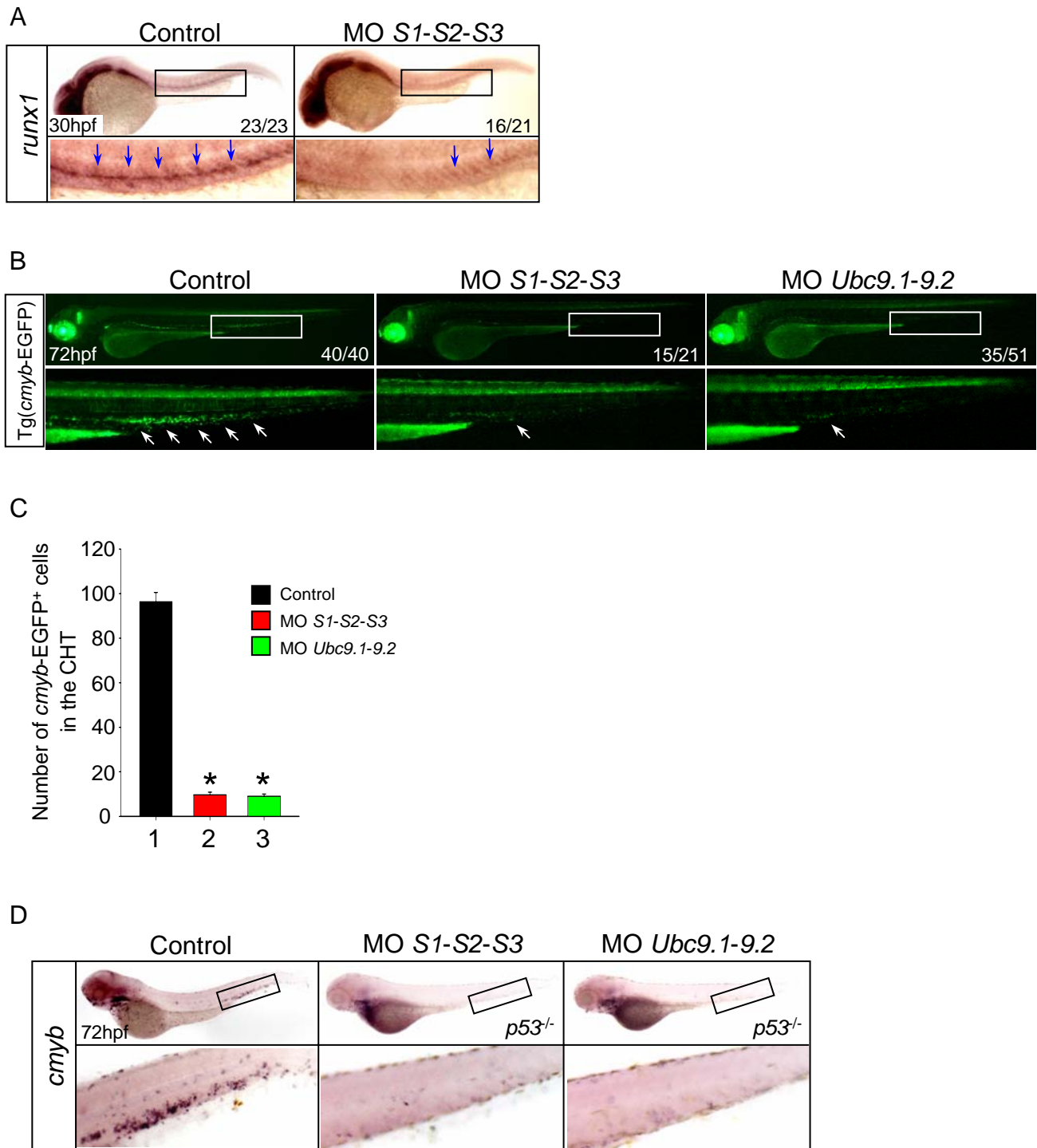


Figure S2.

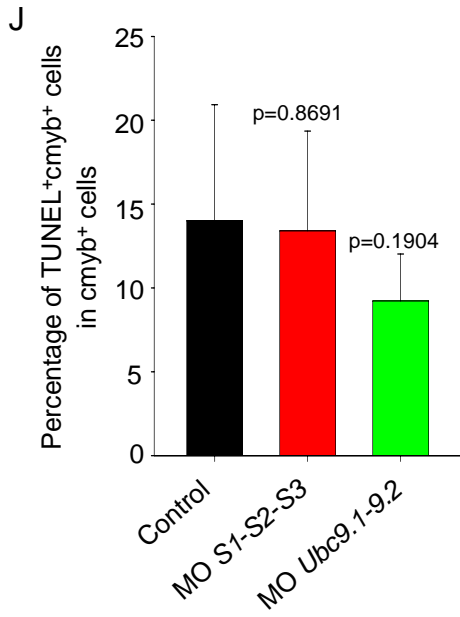
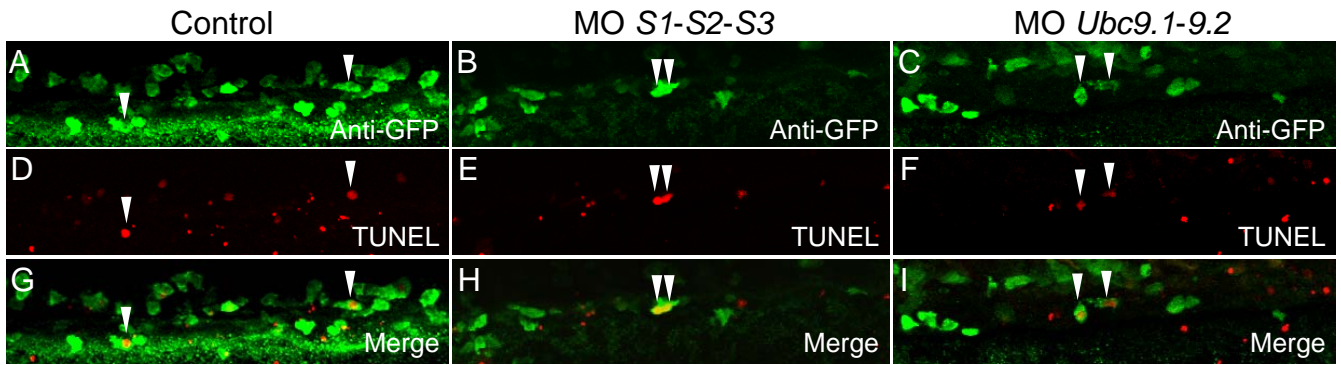


Figure S3.

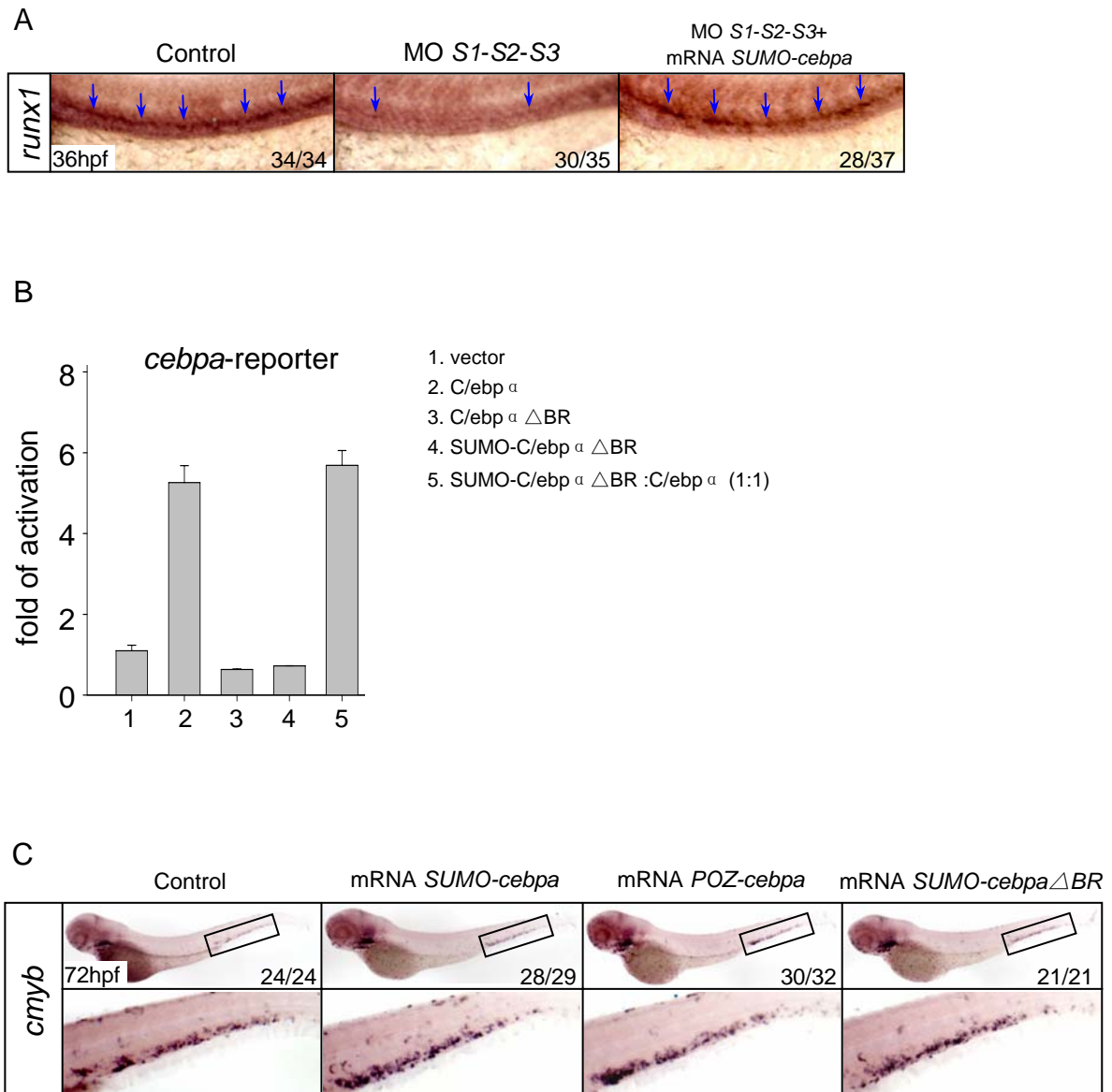


Figure S4.

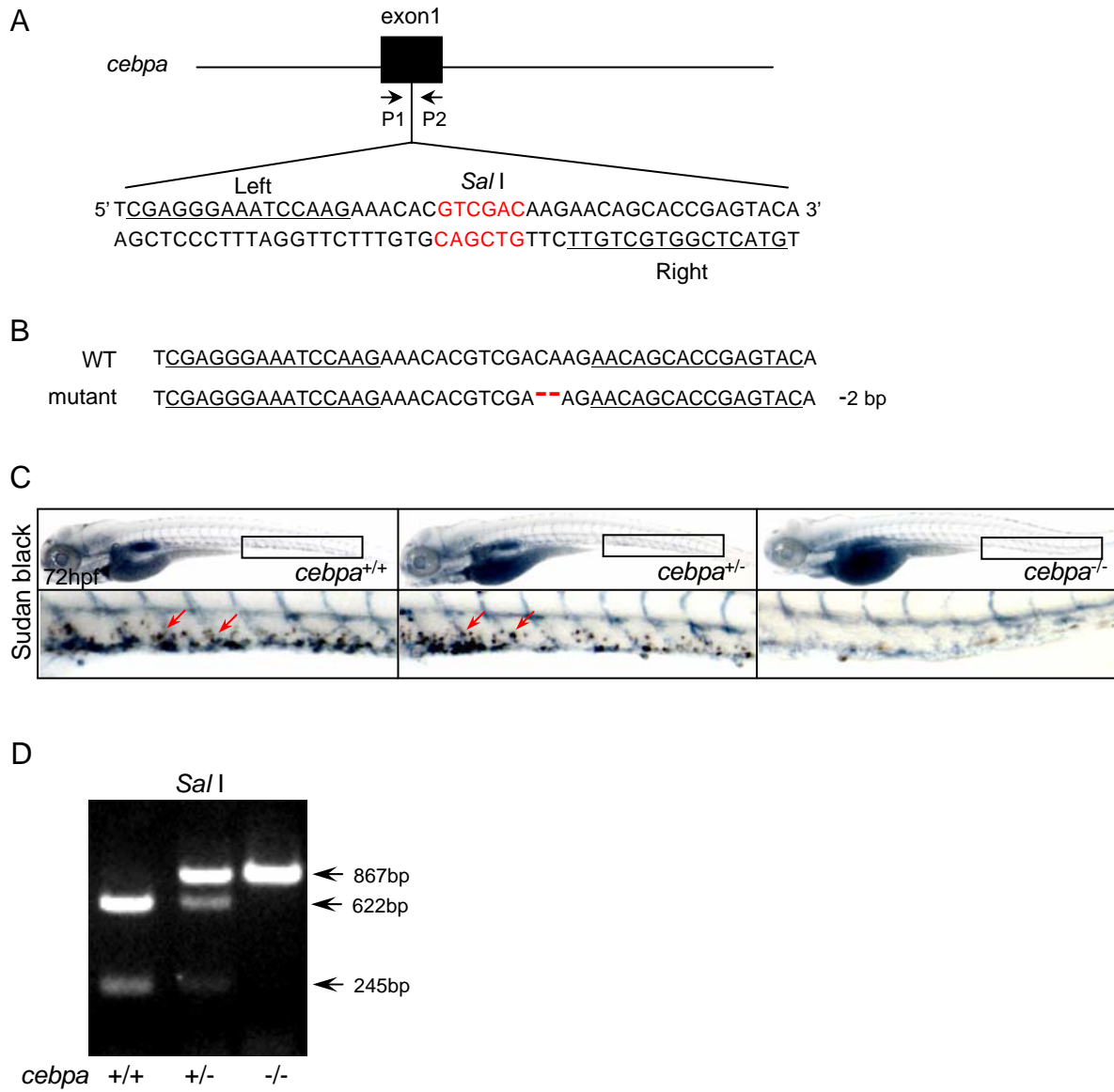


Table S1. Primer sequences

Primer name	Primer sequence	
mouse <i>Runx1</i> enhancer	Forward	GAGGATCCGGGGTGGGAGGTGTAAGTTC
	Reverse	GAGTCGACCAGGTGTCAGCAACCCAT
<i>runx1</i>	Forward	CCGGAATTCATGGACTACAAGGACGACGATGACAAAGTTTTCTTTGGGACGCCAA
	Reverse	CCGCTCGAGGCTGTCAGTATGGCCTCCAG
<i>C/ebpa</i> Δ BR	Forward	GGAGCGCAACAACATAGCCGTGAATGTGGAGACGCAACAAAAAG
	Reverse	CTTTTGTGCGTCTCCACATTCACGGCTATGTTGTTGCGCTCC
<i>SUMO2-C/ebpa</i> Δ BR	Forward	GGAGCGCAACAACATAGCCGTGAATGTGGAGACGCAACAAAAAG
	Reverse	CTTTTGTGCGTCTCCACATTCACGGCTATGTTGTTGCGCTCC
<i>cebpa</i>	Forward	ATGGAGCAAGCAAACCTCTACGAGG
	Reverse	TTAAGCGCAGTTGCCCATGGCTTTG
<i>bactin</i>	Forward	GATCTTCACTCCCCTTGTTCA
	Reverse	GGCAGCGATTCCTCATC