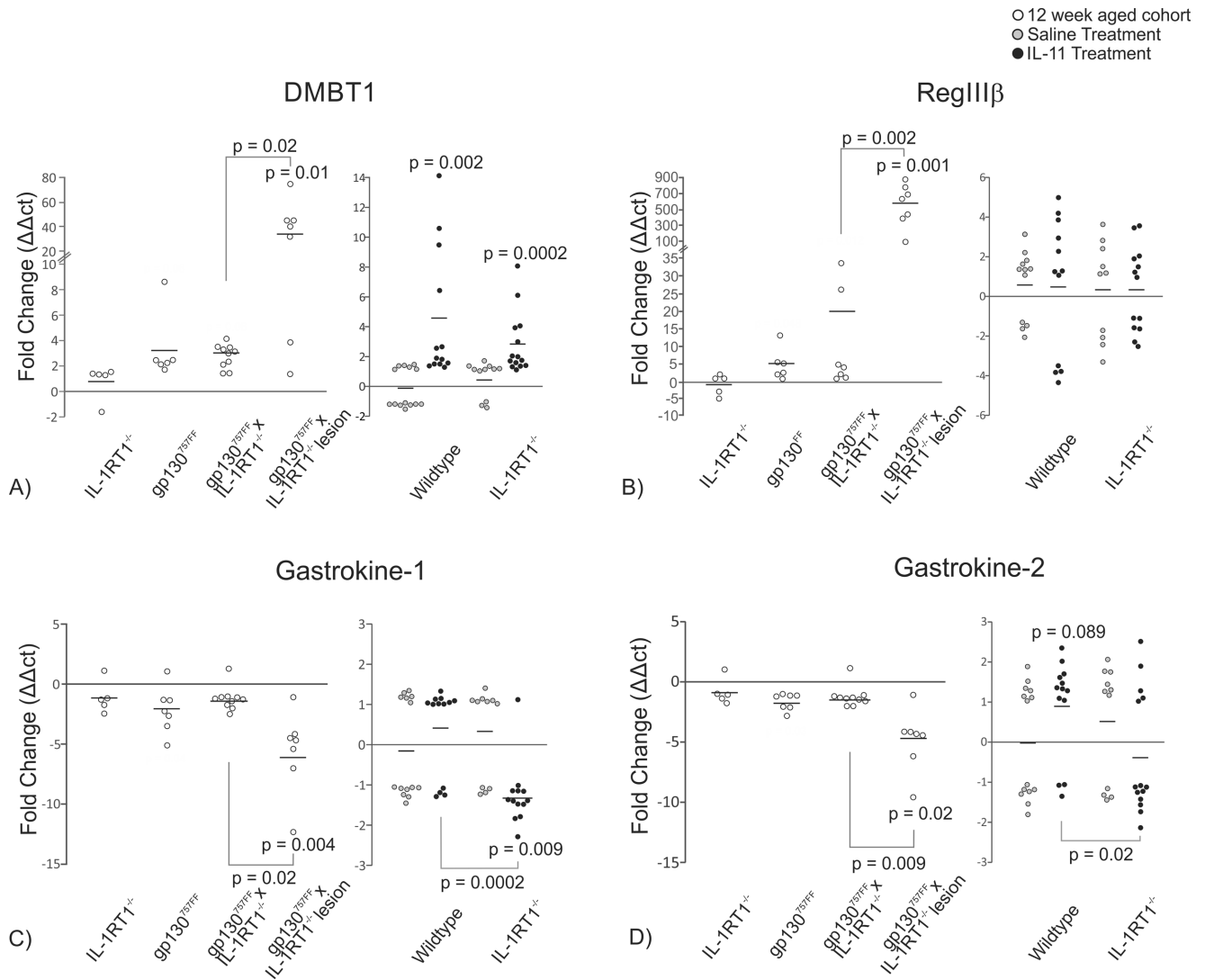
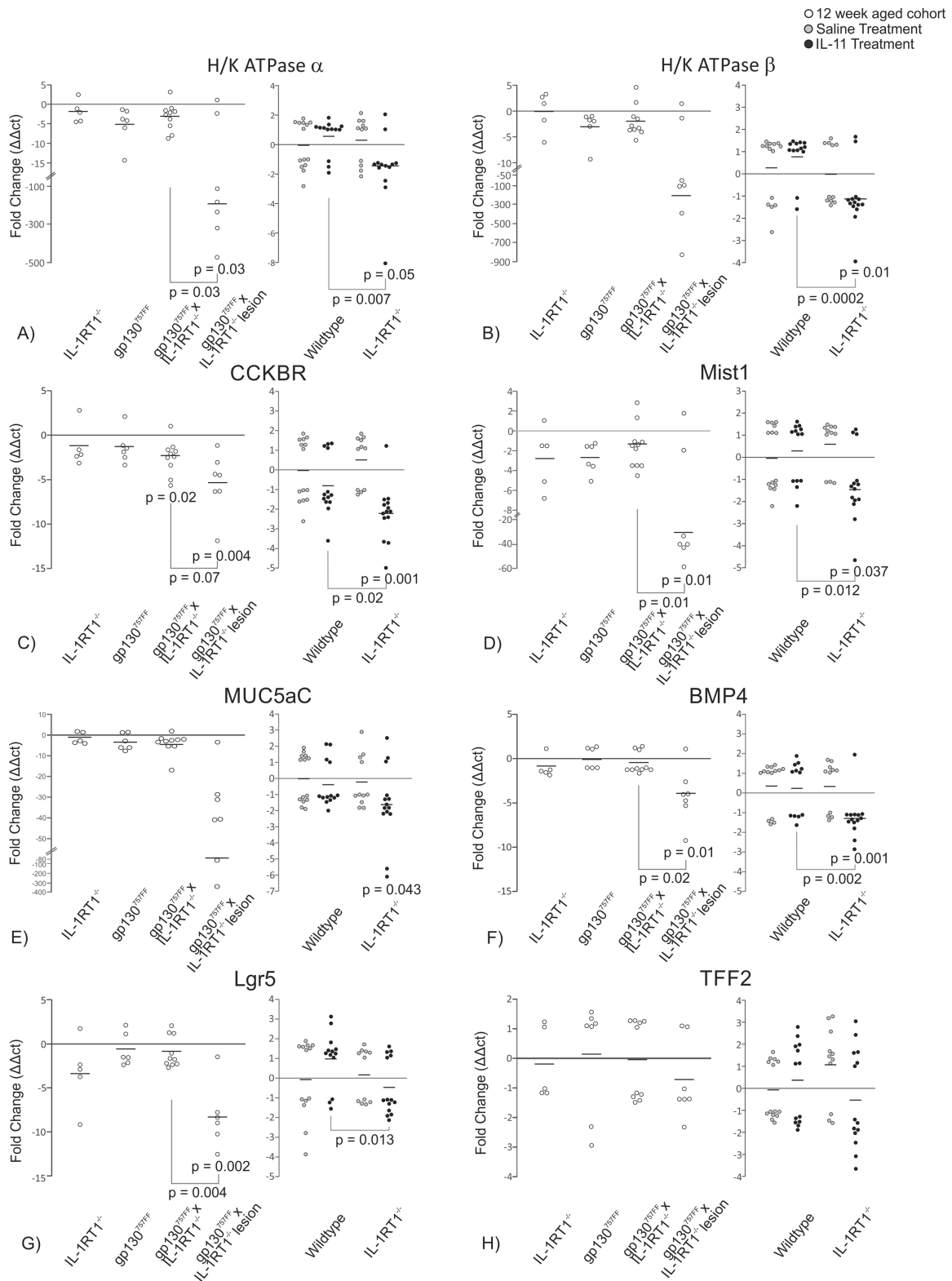


SUPPLEMENTARY FIGURES AND TABLES



Supplementary Figure 1: Proximal stomach mRNA expression in gp130^{757FF} xIL-1RT1^{+/-} cohort mice and wildtype and IL-1RT1^{+/-} mice constitutively administered 1ug IL-11 daily for 7 days via QRT-PCR. Analysis of gastric inflammation and pathology: (A) DMBT1, (B) RegIII β , (C) GKN-1 and (D) GKN-2. *p*-values are as indicated for statistically significant changes.



Supplementary Figure 2: Proximal stomach mRNA expression in $gp130^{757FF}$ $xIL-1RT1^{-/-}$ cohort mice and wildtype and $IL-1RT1^{-/-}$ mice constitutively administered 1ug IL-11 daily for 7 days via QRT-PCR. Analysis of gastric cell types using appropriate markers; parietal cells: (A) H/K ATPase α , (B) H/K ATPase β and (C) CCKBR; Chief cells: (D) Mist1; Mucus secreting cells (E) Muc5aC; Progenitor cells: (F) BMP4 and (G) Lrg5; and SPEM pathology: (H) TFF2. p -values are presented for statistically significant changes ($p < 0.05$).

Supplementary Table 1: Primer sequences used for mRNA expression analysis via QRT-PCR in order as presented in results

Gene	Forward Primer	Reverse Primer
IL-1 α	AACCCATGATCTGGAAGAGACC	TGGTGCTGAGATAGTGTGTTGTCC
IL-1 β	CAGGCAGTATCACTCATTGTGG	GTGCAGTTGTCTAATGGGAACG
IL-1RT1	CGGCGCATGTGCAGTTAATA	TGTAGCCGTGAGGATGATAAAGC
IL-1RAP	CCCATAAGGATGGGACTTCT	AGCGCTCCGAAGCATGACT
IL-1RT2	AAGGTTTCTGAATCCCCATTAC	TTGAACTCTCTGCCACGAACTG
IL-1RAN	TCTGACAGTGGAACGGAATGAC	TTGCATCTTGCCAGGGTCTTTT
IL-11	CTGCAAGCCCGACTGGAA	AGGCCAGGCGAGACATCA
IL-6	ACAAAGCCAGAGTCCTTCAGAGA	CTGTTAGGAGAGCATTGGAAATTG
DMBT1	GCTCCACAAGCACAGATTCC	TGCCATCTCATTGTTCTTGG
RegIII β	GAACCCATCTACTGCCTTAGACC	CTTCACATTTTGTCCCTTGTC
Gastrokine-1	CTTCAGGACCTCGATAACAATGG	TTGAGTACAAAGGCTGGTTTGG
Gastrokine-2	AATGTAGACGGAAGTGGACAGC	GCATCCTTGTTTATTCTGTGC
S100A8	GTCCTCAGTTTGTGCAGAATATAAA	GCCAGAAGCTCTGCTACTCC
S100A9	CTCTTTAGCCTTGAAGAGCAAG	TTCTTGCTCAGGGTGTGAGG
MIP2	AGTGAAGTGCCTGTCAATG	TTCAGGGTCAAGGCAAAGT
TNF α	TATGTCTCAGCCTCTTCTCATTCC	ATGATCTGAGTGTGAGGGTCTGG
IL-23	GAGCAACTTCACACCTCCCTACTAGTAG	GGCCTGGAGGCTTCGAA
Amphiregulin	CTATCTTCTCTGCCATCATC	CAACTTTTACCCTTGCAATTGCC
Arg-1	CTGCATGGGCAACCTGTGT	CCTGGTACATCTGGGAAGTTTCC
H/K ATPase α	CAAATATCATCGCCAGCTTCAA	TCCCCGTCTCGGATCACA
H/K ATPase β	CCGGTGGGTGTGGATCAG	GCAAAGAGCCCGGTCATG
CCKBR	ACAAATGTGGTCCGTGCTTCT	GCGGGAGATGAGTCCATAGG
Mist1	CCAGGGTGTCTCTTCTTTTGG	GGCGGAAGTTCACCATCCTT
Muc5aC	GCAGTTGTGTCACCATCATCTGTG	GGGGCAGTCTTGACTAACCTCTT
BMP4	GCCGTCATTCCGGATTACAT	CTCCTCACAGTGTGGCTCG
Lgr5	GGAATAAAGACGACGGCAACA	GTCCCCTCATCTTGAAGTTG
TFF2	CCCCACAACAGAAAGAAC	GGGCACTTCAAAGATCAG

Supplementary Table 2: Antibodies used for protein detection via immunoblotting

Protein	Antibody	Raised In	Company	Reference No.
β -actin	β -Actin (13E5) Rabbit mAntibody	Rabbit	Cell signaling	#4970
STAT3	STAT3 Antibody	Rabbit	Cell signaling	#9132
p-STAT3	Phospho-STAT3 (Tyr705) Antibody	Rabbit	Cell signaling	#9131
ERK1/2	p44/42 MAP Kinase Antibody	Rabbit	Cell signaling	#9102
pERK1/2	Phospho-p44/42 MAK Kinase Antibody	Rabbit	Cell signaling	#4377
AKT	AKT Antibody	Rabbit	Cell signaling	#9272
pAKT	Phospho-AKT (Ser473) Antibody	Rabbit	Cell signaling	#4058
STAT1	STAT1 Antibody	Rabbit	Cell signaling	#9172
pSTAT1	Phospho-STAT1 (Try-701) Antibody	Rabbit	Cell signaling	#9171
mIL-11	IL-11 Antibody	Mouse	WEHI	3/99-4D9-27-25

Supplementary Table 3: Fluorochromes used for Flow Cytometry analysis

Marker	Fluorochromes	Company	Reference No.
CD4	V450	BD Biosciences	560468
CD8a	APC-Cy7	BD Biosciences	561967
CD11b	PE	BD Biosciences	557397
CD11c	APC	BD Biosciences	550261
CD45R/B220	FITC	BD Biosciences	553087
Ly-6G and Ly6C (Gr-1)	PerCP-Cy5.5	BD Biosciences	552093

Immune Cell	Fluorochromes
MDSC CD11b+Gr-1+	CD45 ⁺ /CD11b ⁺ /CD11c ^{int} /Gr-1 ^{int}
Th CD4+	V450 ⁺ / APC-Cy7 ⁻
CTL CD8+	V450 ⁻ / APC-Cy7 ⁺