

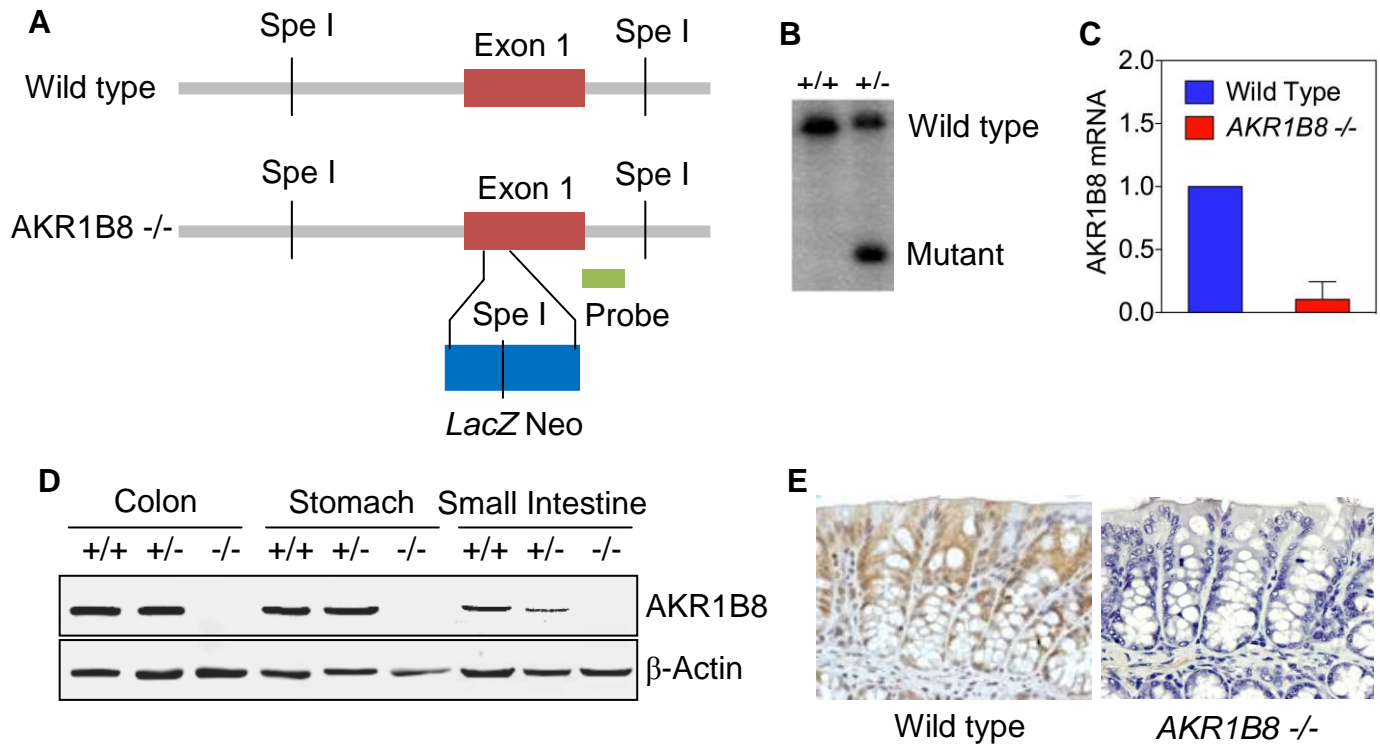
## **Supplemental Data**

Yi Shen, Jun Ma, Ruilan Yan, Hongyan Ling, Xiaoning Li, Wancai Yang, John Gao, Chenfei Huang, Yiwen Bu, Yingchun He, Laxiang Wan, Mei Chris Huang, William F Stenson, Duanfang Liao, and Deliang Cao

**Figures S1-6**

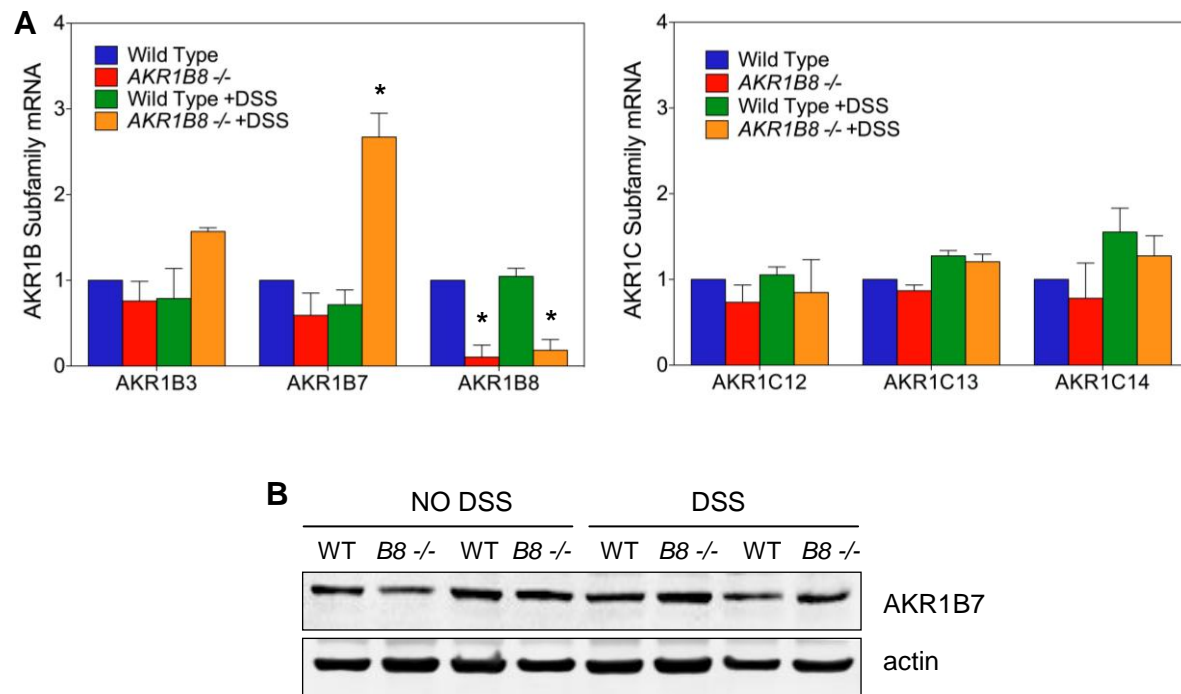
**Tables S1-7**

**Figure S1**



**Figure S1. Targeted disruption of *AKR1B8* gene.** (A) Schematic strategy for disruption of *AKR1B8* gene. Red box denotes Exon 1 of *AKR1B8* gene and blue box indicates LacZ/Neo expression cassette, a selection marker. (B) Southern blot. Mouse genomic DNA was extracted from tails, digested by SpeI, and probed by the DNA probe indicated in (A). (C) *AKR1B8* mRNA level. Data represent mean  $\pm$  SD from three mice. (D) Western blot and (E) Immunohistochemistry, indicating *AKR1B8* protein levels.

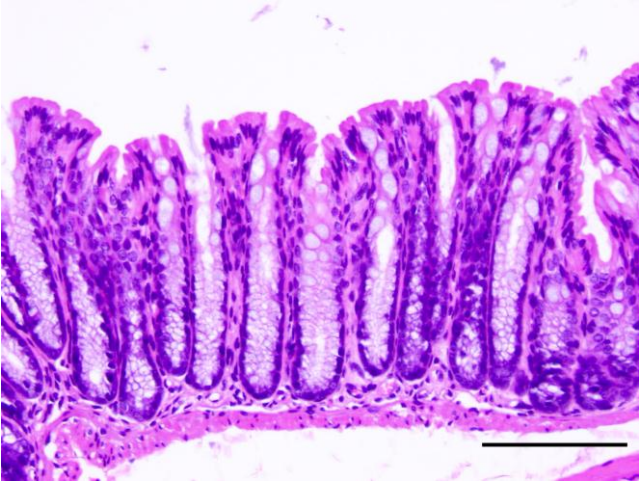
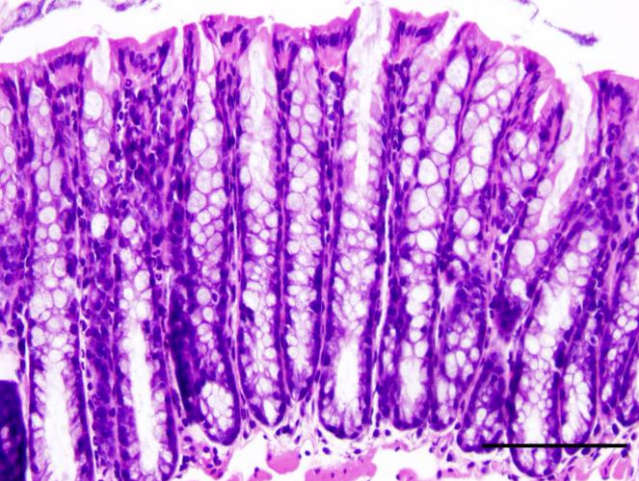
**Figure S2**



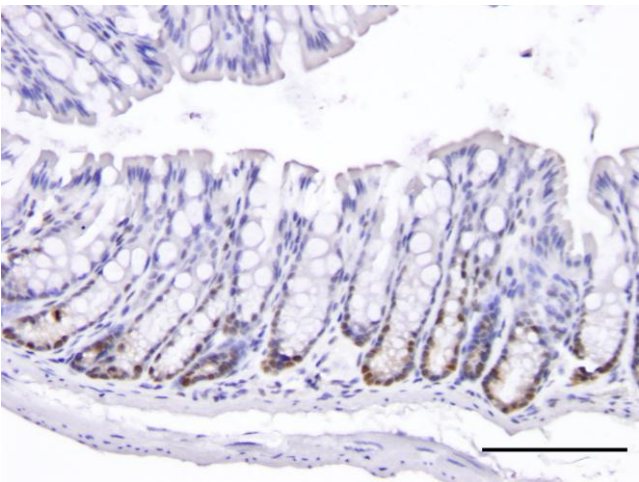
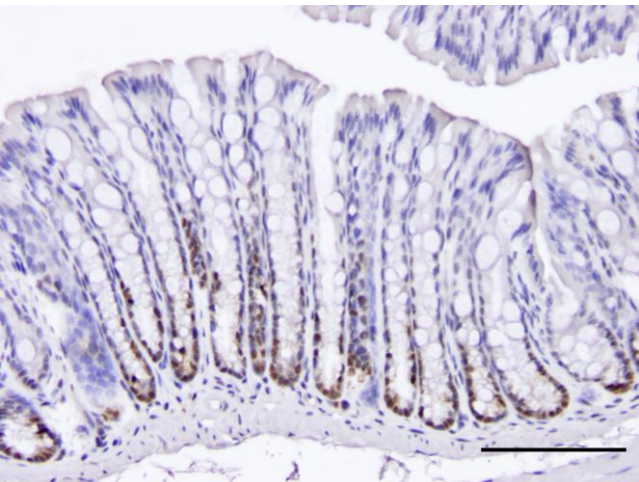
**Figure S2.** *AKR1B* and *AKR1C* subfamily expression. A) *AKR1B* and *AKR1C* subfamily mRNA levels detected by real-time RT-PCR. Data represent mean  $\pm$  SD from three mice. \*  $P < 0.01$ , compared to wild type control. *AKR1B7* was induced by DSS- induced colitis in the *AKR1B8*<sup>-/-</sup> mice. B) Western blot of *AKR1B7*. WT, wild type; B8, *AKR1B8*.

Figure S3

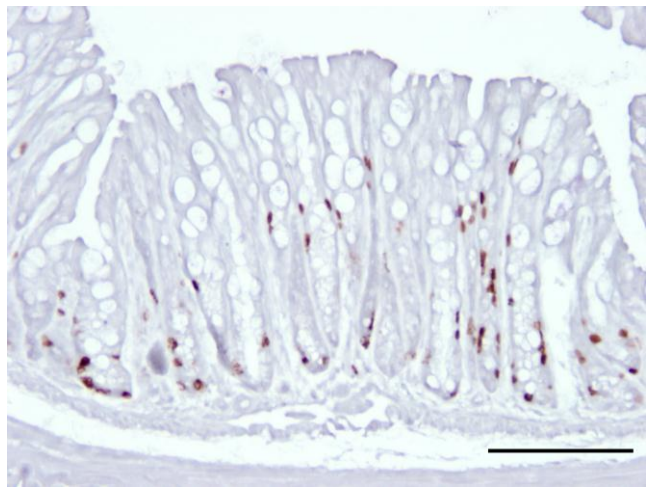
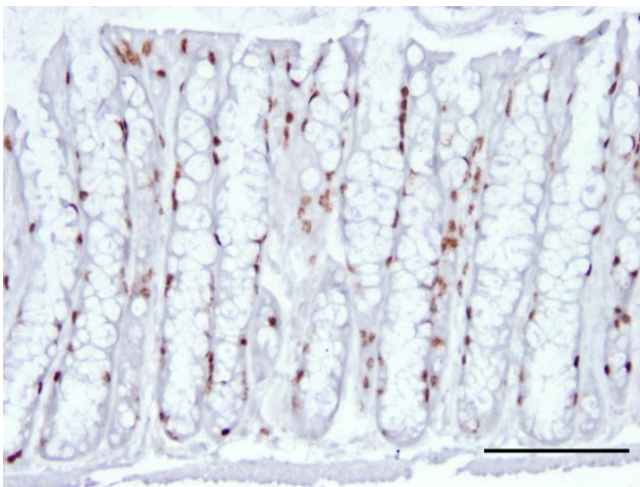
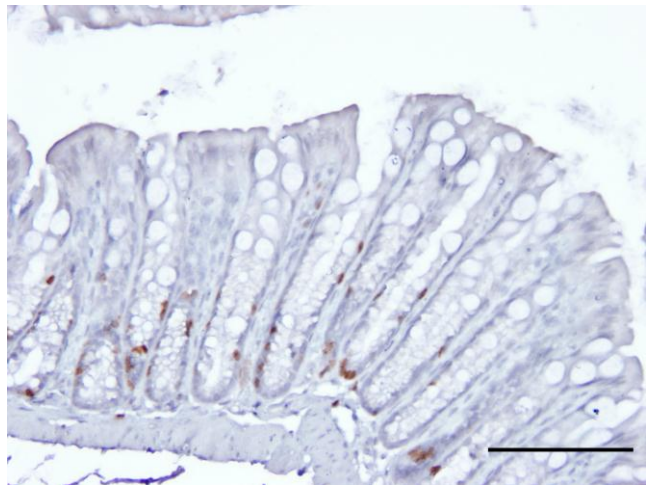
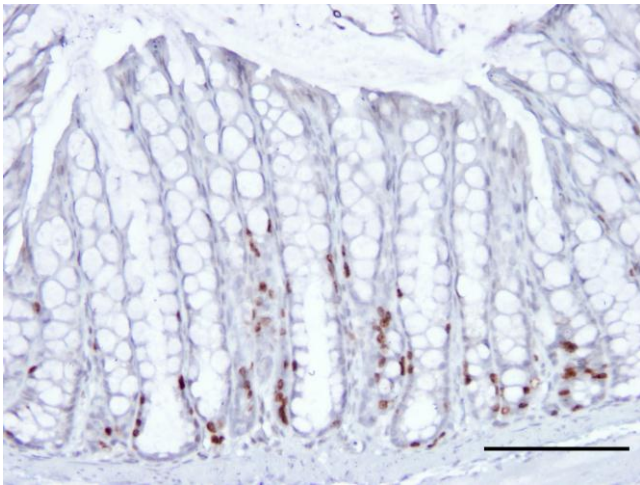
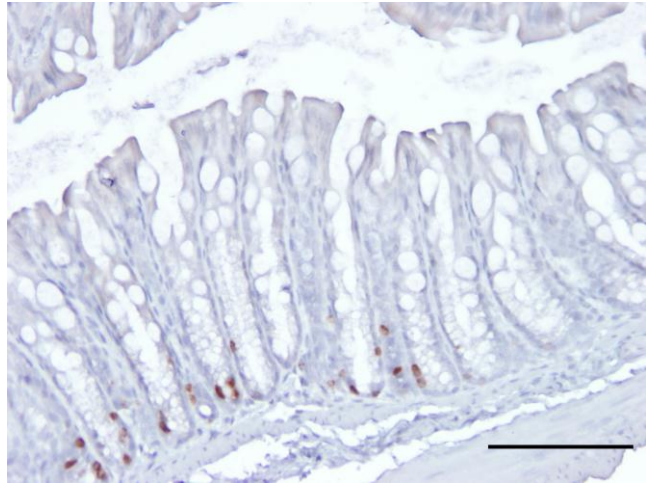
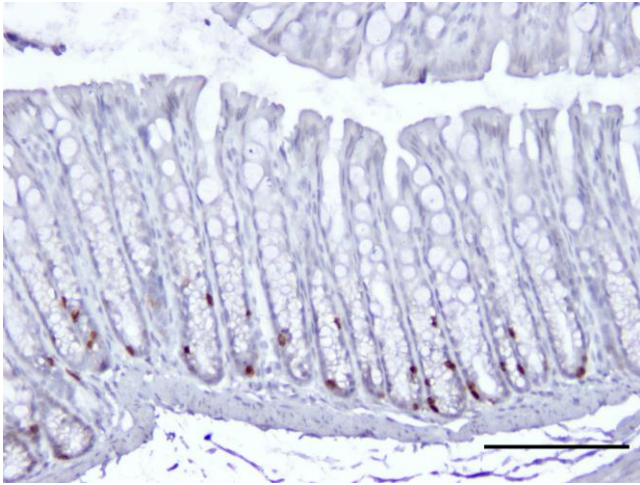
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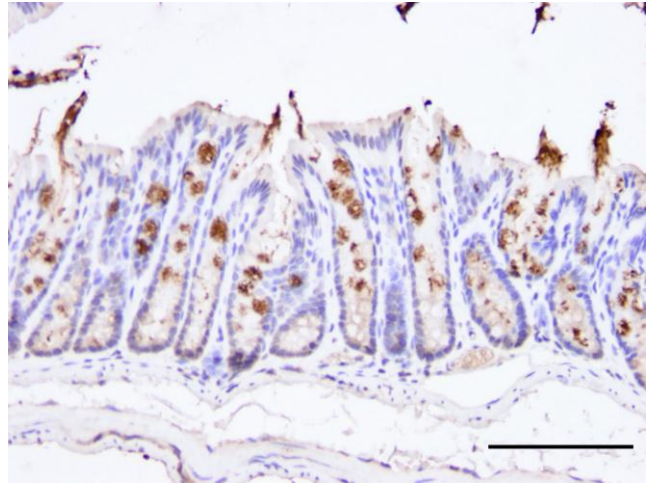
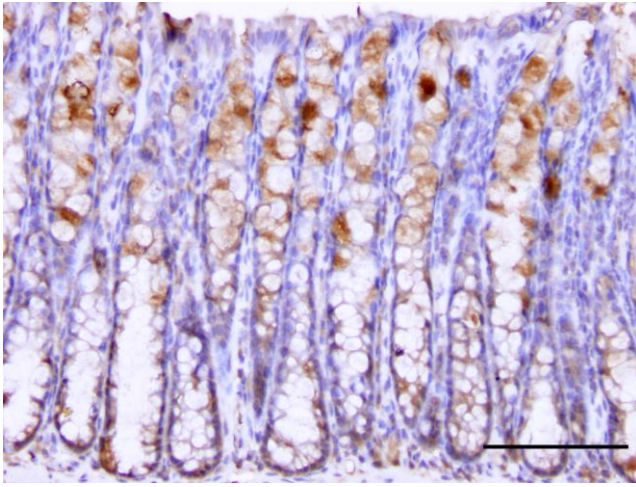
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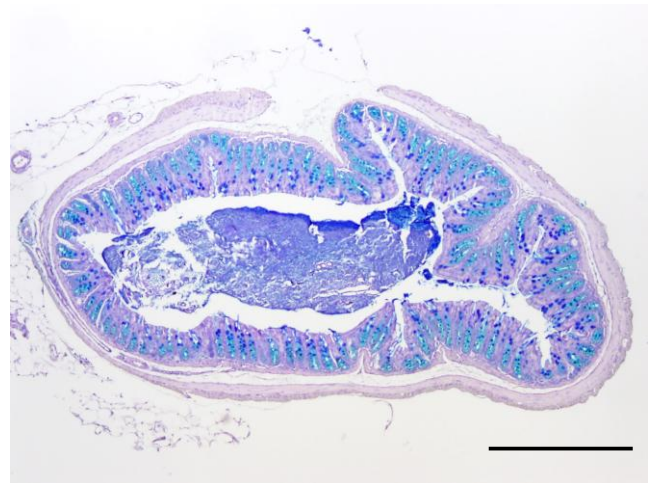
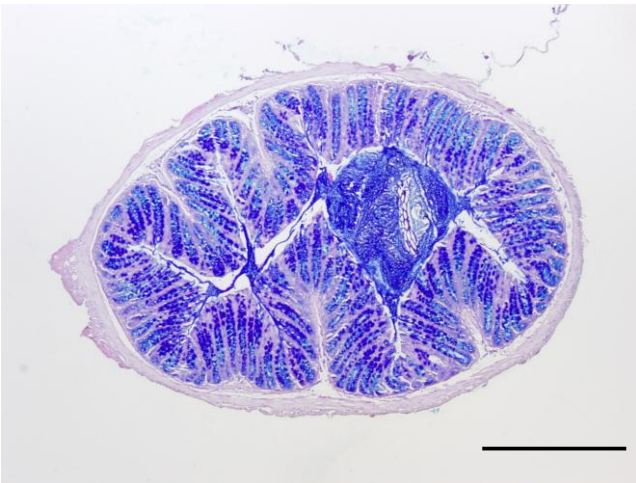
C



D



E

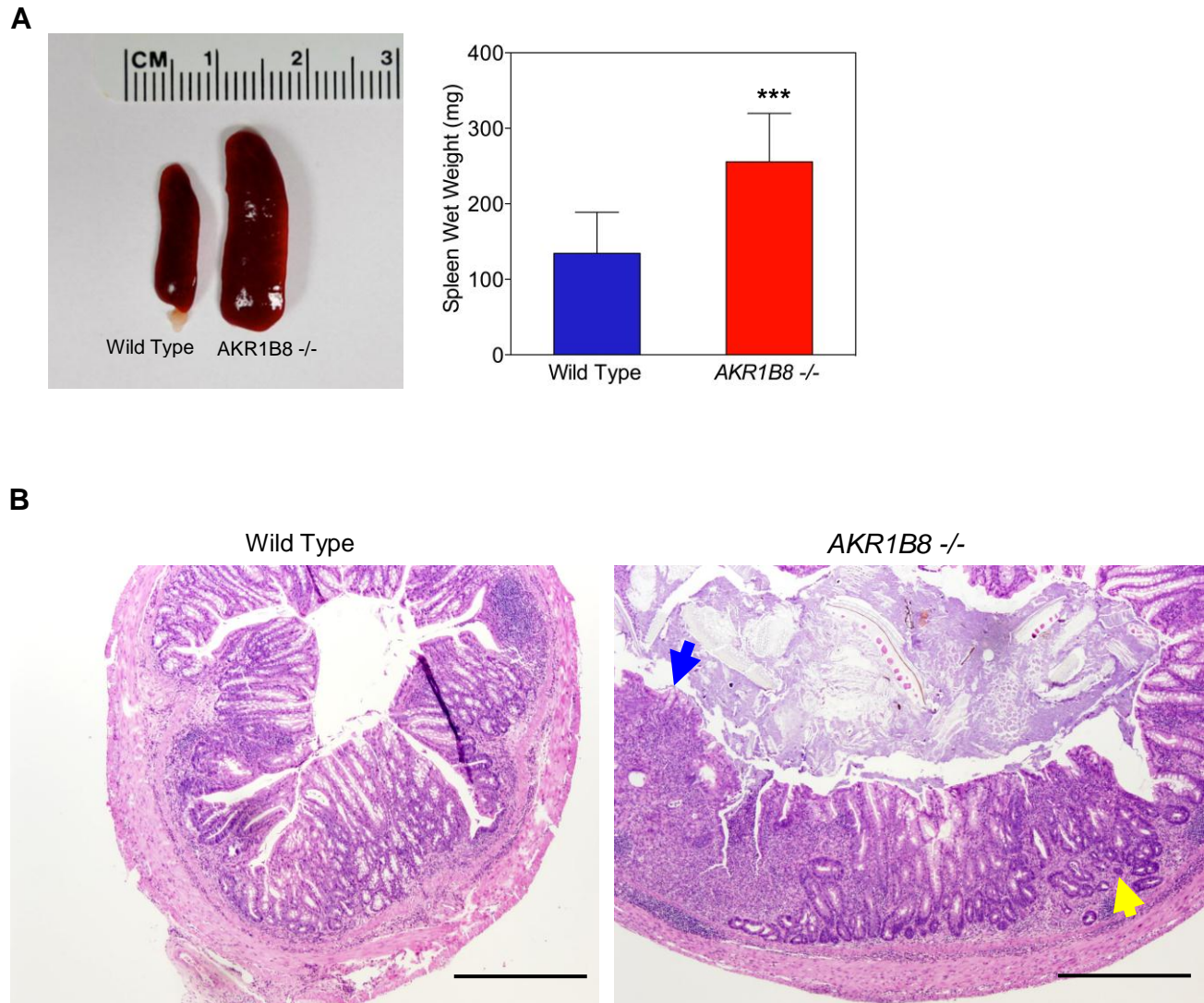


**Figure S3.** Deficiency of proliferation and self-renewal of colonic crypt cells in *AKR1B8*<sup>-/-</sup> mice. A) H & E histology; B) PCNA expression; C) BrdU labeling at 1, 24, and 48 hours; D) ITF expression; and E) Alcian blue and Periodic acid Schiff staining (PAS/AB). Scale bars indicate 50 $\mu$ m in PAS/AB, and 100 $\mu$ m in others.



**Figure S4.** DSS treatment schedules. *AKR1B8* <sup>-/-</sup> and wild type mice were fed with 2% dextran sulfate sodium (DSS) for 7 days, followed by 14 days DSS-free water, which is counted as one cycle. Animals experienced one to four cycles depending on experimental needs.

**Figure S5**

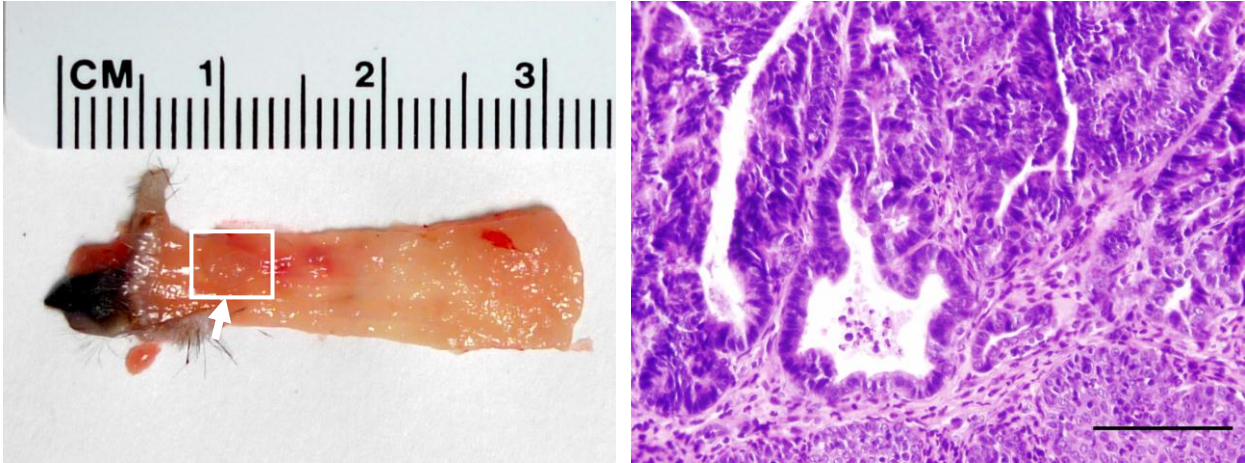


**Figure S5.** Spleen size and histological inflammation of *AKR1B8* <sup>-/-</sup> mice treated with 2% DSS. A) Spleen size. Spleen in *AKR1B8* <sup>-/-</sup> mice was significantly enlarged by >2 fold over that in wild type. Data denote mean  $\pm$  SD, n = 15, \*\*\*  $P < 0.0001$ . B) Histological inflammatory lesions in another representative mouse, showing ulcer (blue arrow). Dysplasia occurred in *AKR1B8* <sup>-/-</sup> mice (white arrow). *AKR1B8* <sup>-/-</sup> mouse colon was swelling and enlarged.

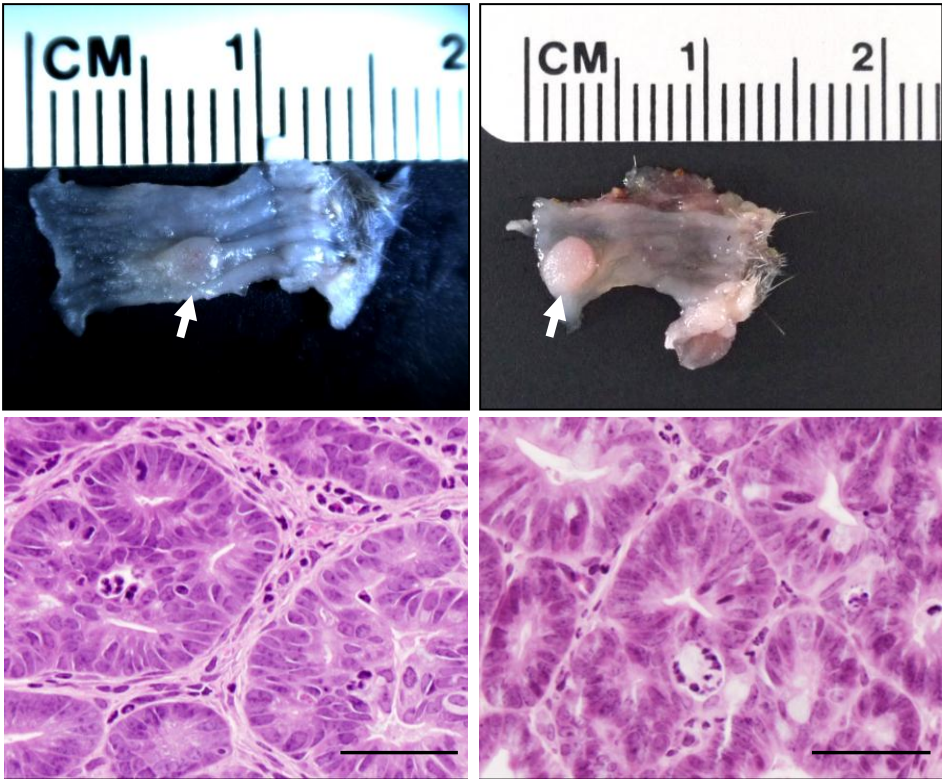


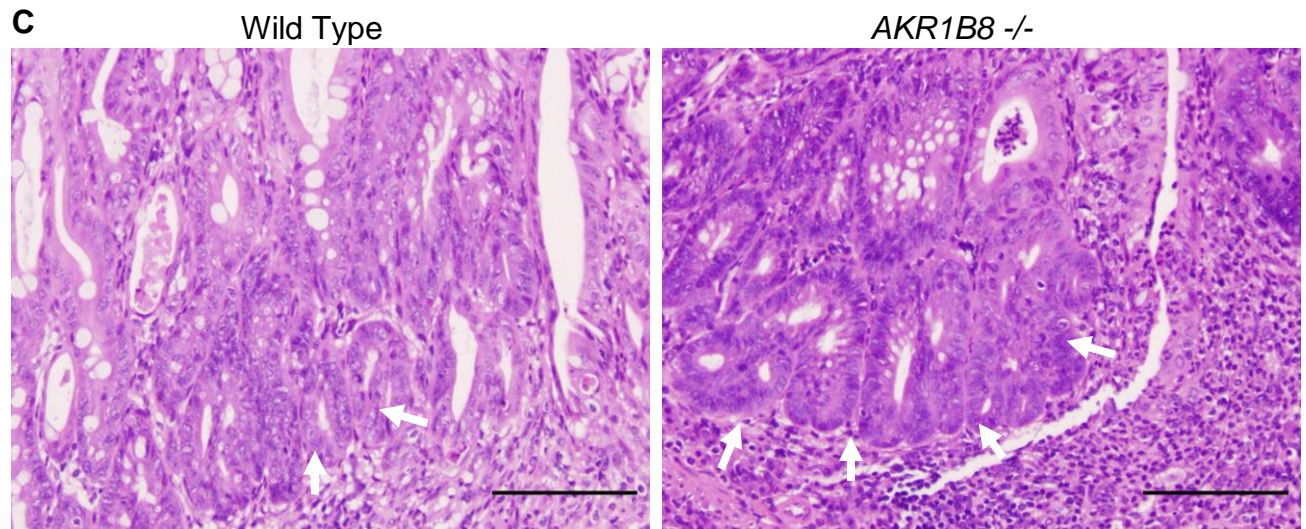
Figure S6

A



B





**Figure S6.** Colitis-associated tumorigenesis in *AKR1B8*<sup>-/-</sup> mice. A) A tumor mass (arrow) from an *AKR1B8*<sup>-/-</sup> mouse that received 2 cycles of DSS treatment. *Right:* H & E histology. B) Two more representative palpable tumors (arrows) from *AKR1B8*<sup>-/-</sup> mice received 4 cycles of DSS treatment. *Below:* H & E histology. C) Micro-dysplasias (white arrows). Dysplastic lesions in *AKR1B8*<sup>-/-</sup> mouse were at higher grade than in wild type. Scale bars indicate 200µm in (B), and 100µm in the others.

**Supplemental Table S1.** Coverage summary of Exome sequencing.

<b>Coverage Summary</b>	<b>Wild Type</b>		<b><i>AKR1B8</i> -/-</b>		<b>Tumors</b>	
<b>Reads mapped (%)</b>	34,726,317 (95.87)	30,643,916 (96.01)	37,536,791 (95.55)	34,026,043 (95.58)	37,781,378 (95.49)	32,203,249 (95.66)
Mapped confidently	32,112,619 (88.65)	28,095,115 (88.02)	34,842,474 (88.69)	31,287,103 (87.89)	34,475,395 (87.13)	29,696,048 (88.21)
Mapped repetitively	2,613,698 (7.22)	2,548,801 (7.99)	2,694,317 (6.86)	2,738,940 (7.69)	3,305,983 (8.36)	2,507,201 (7.45)
<b>Reads not mapped (%)</b>	1,496,567 (4.13)	1,274,818 (3.99)	1,750,207 (4.45)	1,572,689 (4.42)	1,785,596 (4.51)	1,460,649 (4.34)
No mapping	591,030 (1.63)	501,817 (1.57)	833,889 (2.12)	743,808 (2.09)	831,032 (2.10)	672,032 (2.00)
Low quality	842,400 (2.33)	727,953 (2.28)	865,717 (2.20)	772,845 (2.17)	854,899 (2.16)	730,565 (2.17)
Ribosomal RNA	4,797 (0.01)	4,901 (0.02)	4,540 (0.01)	4,775 (0.01)	4,195 (0.01)	3,938 (0.01)
Primer	1,721 (0.00)	299 (0.00)	856 (0.00)	548 (0.00)	783 (0.00)	611 (0.00)
Control (phiX-174)	56,529 (0.16)	39,742 (0.12)	45,127 (0.11)	50,638 (0.14)	94,560 (0.24)	53,339 (0.16)
Poly-A	46 (0.00)	47 (0.00)	34 (0.00)	40 (0.00)	65 (0.00)	79 (0.00)
Poly-C	2 (0.00)	3 (0.00)	3 (0.00)	2 (0.00)	3 (0.00)	5 (0.00)
Poly-G	0 (0.00)	2 (0.00)	1 (0.00)	0 (0.00)	0 (0.00)	0 (0.00)
Poly-T	42 (0.00)	54 (0.00)	40 (0.00)	33 (0.00)	59 (0.00)	80 (0.00)
<b>Total reads (%)</b>	36,222,884 (100.00)	31,918,734 (100.00)	39,286,998 (100.00)	35,598,732 (100.00)	39,566,974 (100.00)	33,663,898 (100.00)
<b>Total bases (%)</b>	3,622,288,400 (100.00)	3,191,873,400 (100.00)	3,928,699,800 (100.00)	3,559,873,200 (100.00)	3,956,697,400 (100.00)	3,366,389,800 (100.00)

**Supplemental Table S2.** Variations detected throughout the mouse Exome sequencing. Data were broken down by variant types and zygosity. \*,  $P < 0.05$  and \*\*,  $P < 0.01$  compared to wild type. SNP, single nucleotide polymorphisms; MNP, multi-nucleotide polymorphisms.

<b>Variant Types and Zygosity</b>		<b>Wild Type (n=2)</b>	<b><i>AKR1B8</i> -/- (n=2)</b>	<b>Tumors (n=2)</b>
<b>SNPs</b>	Homozygous	1470 ± 107	3480 ± 304*	4246 ± 70**
	Heterozygous	9330 ± 317	9698 ± 258	9491 ± 202
	<b>Subtotal</b>	<b>10800 ± 421</b>	<b>13178 ± 93**</b>	<b>13736 ± 474*</b>
<b>MNPs</b>	Homozygous	68 ± 1	102 ± 5**	127 ± 8**
	Heterozygous	698 ± 59	705 ± 47	760 ± 48
	<b>Subtotal</b>	<b>766 ± 57</b>	<b>806 ± 52</b>	<b>887 ± 57</b>
<b>Insertions</b>	Homozygous	94 ± 7	146 ± 19	175 ± 3**
	Heterozygous	237 ± 11	318 ± 7**	355 ± 23*
	<b>Subtotal</b>	<b>331 ± 4</b>	<b>464 ± 26**</b>	<b>530 ± 25**</b>
<b>Deletions</b>	Homozygous	142 ± 18	217 ± 28	245 ± 6**
	Heterozygous	1496 ± 30	1382 ± 25	1387 ± 99
	<b>Subtotal</b>	<b>1638 ± 12</b>	<b>1599 ± 53</b>	<b>1632 ± 52</b>
<b>Total variant loci</b>		<b>13533 ± 494</b>	<b>16046 ± 224*</b>	<b>16785 ± 660*</b>

**Supplemental Table S3.** Oncogenes and tumor suppressor genes mutated in *AKR1B8* <sup>-/-</sup> colitis and tumors only, but not in wild type colitis. Data were produced by Exome sequencing (Otogenetics, GA), n=2 mice each group.

Gene Name	Zygoty	Transcript Name	Mutant Type	Mutant Numbers	Original AA	Mutant AA	Function	Ref
Abtb1	Het	NM_030251	Non-Synonym	1	Y	H	Tumor suppressor	(50)
Chd5	Het	NM_001081376	Non-Synonym	1	R	L	Tumor suppressor	(51)
Chl1	Hom	NM_007697	Non-Synonym	3	S	A	Tumor suppressor	(52)
Chn2	Hom	NM_023543	Non-Synonym	1	M	V	Tumor suppressor	(53)
Dfna5	Hom	NM_018769	Non-Synonym	1	L	P	Tumor suppressor	(54)
Dlx5	Hom	NM_010056	Frameshift	4			Oncogene	(55)
Dok1	Hom	NM_010070	Non-Synonym	1	V	A	Tumor suppressor	(56)
Fam176a	Hom	NM_145570	Non-Synonym	1	P	S	Tumor suppressor	(57)
Fbxo5	Het	NM_025995	Non-Synonym	2	F	C	Tumor suppressor	(58)
Fgd5	Hom	NM_172731	Non-Synonym	1	Q	R	Oncogene	(59)
Fhl3	Het	NM_010213	Frameshift	1			Tumor suppressor	(60)
	Het		Non-Synonym	3	C	Y		
Herc5	Hom	NM_025992	Non-Synonym	19	S	G	E3 protein ligase; Oncogene?	(61)
	Hom		Stop Gain	1	Q	*		
Hoxa10	Hom	NM_008263	Non-Synonym	1	V	G	Transcription factor, tumor suppressor?	(62)
Hoxa13	Hom	NM_008264	Non-Synonym	1	R	A	Transcription factor, oncogene?	(63)
Hoxa5	Hom	NM_010453	Non-Synonym	1	A	V	Transcription factor, tumor suppressor?	(64)
L3mbtl4	Het	NM_177278	Non-Synonym	1	E	G	Transcription factor, tumor suppressor	(65)
Lrig1	Het	NM_008377	Non-Synonym	1	A	T	Tumor suppressor	(66)
Mktn1	Hom	NM_018810	Non-Synonym	1	N	Y	E3 ubiquitin ligase, oncogene?	(67)
Mxd1	Hom	NM_010751	Non-Synonym	1	S	G	Tumor suppressor	(68)
NSPC1	Hom	NM_197992	Non-Synonym	1	R	C	Transcription factor, oncogenic	(69)
Pdzk1	Het	NM_001146001	Non-Synonym	2	N	D	Oncogenic	(70)
PP2Cm	Hom	NM_175523	Non-Synonym	2	S	L	Cell death/survival	(71)
Rtkn	Hom	NM_001136227	Non-Synonym	3	A	V	A scaffold protein, oncogenic?	(72)
Samd9l	Hom	NM_010156	Non-Synonym	2	I	V	oncogene	(73)
Smo	Hom	NM_176996	Non-Synonym	2	D	N	Tumor suppressor	(74)
ECRG2	Het	NM_001001803	Non-Synonym	1	C	S	Tumor suppressor	(75)
AMSH	Hom	NM_024239	Non-Synonym	1	V	I	Cytokine signaling, oncogenic	(76)
Sulf1	Het	NM_172294	Non-Synonym	1	D	N	Tumor suppressor	(77)
<b>Subtotal: 28 genes</b>								

**Supplemental Table S4.** Disease Activity Index

<b>Score</b>	<b>Weight loss (%)</b>	<b>Stool consistency</b>	<b>Occult/gross blood in stools</b>
0	$\leq 1$	Normal stool (well-formed pellets)	Normal (no blood in stools)
1	$1 < \leq 5$		
2	$5 < \leq 10$	Loose stools (pasty and semi-formed, not adhered to the anus)	Positive occult blood in stools
3	$10 < \leq 20$		
4	$20 <$	Diarrhea (liquid, adhered to the anus)	Gross bleeding in stools

**Supplemental Table S5.** Histopathological Inflammation Score

<b>Score</b>	<b>Epithelial Damage (E)</b>	<b>Infiltration (I)</b>
0	Normal morphology	No infiltrate
1	Loss of goblet cells	Infiltrate around crypt basis
2	Loss of goblet cells in large areas	Infiltrate reaching to muscularis mucosa
3	Loss of crypts	Extensive infiltration reaching the muscularis mucosa, thickening of the mucosa with abundant oedema
4	Loss of crypts in large area	Infiltration of the submucosa

**Supplemental Table S6.** Definition of histopathology.

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<b>Histopathology</b>	
Regenerative mucosa	Regenerative mucosa is usually due to damage (ulceration). In the background, there are some changes indicating the prior damages, such as thrombus formation/ hyaline thrombus in small vessels, necrosis, ulceration, granulation tissue, fibrin, hemosiderin/hemorrhage, edema in lamina propria, and inflammatory exudate (early) or stromal fibrosis (late).
Hyperplasia without dysplasia	Elongated glands/crypts usually with serrated lumen, mixed goblet cells and absorptive cells, bland cytology (no atypia) with brush borders and basal nuclei, the basement membrane is usually thickened, cells at the base of crypt are a little bit immature including nuclear elongation, crowding and increased mitotic rate, but this is not present at the top of crypt (maturation).
Mild dysplasia	Pseudostratification of nuclei that reach only half of the cell, apical mucin is present, nuclei are elongated and hyperchromatic. Mitotic activity present but atypical mitosis and loss of polarity are absent or minimal.
Moderate dysplasia	Changes are between mild and high grade dysplasia.
Severe dysplasia	Cytological changes: nuclei are rounded, hyperchromatic or vesicular with prominent nucleoli; nuclei are stratified and reach luminal surface. Architectural changes: back to back glands, cribriform, mitotic and atypical mitotic figures are prominent with reduced mucin.
Adenoma	Dome-shaped elevated lesions with mild dysplasia.

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**Supplemental Table S7.** Gene specific primers.

<b>Gene Description</b>	<b>Primers</b>	<b>Sequence (5' → 3')</b>
AKR1B3	Forward primer	CAA CAG GAA CTG GAG GGT GTG
	Reverse primer	GAA CAG GTG CAA GCC ACT TG
AKR1B7	Forward primer	GAT CCC CAA GTC TGT GAC AC
	Reverse primer	CAG TCC TTG CAT CCA ACA G
AKR1B8	Forward primer	CAT CCT TAC CTC ACC CAG GA
	Reverse primer	GAT CAC CAC CAC GTT CCT CT
AKR1C12	Forward primer	GAC ACA TGG GAG AGG TTG GAG
	Reverse primer	GTC CTG GCT TGT TGA GGA TTC
AKR1C13	Forward primer	CCT TGT TGA CCA CCC AGA G
	Reverse primer	CTG TCC ACA CAC AGG GAC AC
AKR1C14	Forward primer	GGA GGC CAT GGA AAA GTG TA
	Reverse primer	GAT GGC ATT CTA CCT GGT TGC
IL-1 $\beta$	Forward primer	GCA ACT GTT CCT GAA CTC AAC
	Reverse primer	ATC TTT TGG GGT CCG TCA ACT
IL-6	Forward primer	TAG TCC TTC CTA CCC CAA TTT CC
	Reverse primer	TTG GTC CTT AGC CAC TCC TTC
IFN $\gamma$	Forward primer	GCG TCA TTG AAT CAC ACC TG
	Reverse primer	GAC CTG TGG GTT GTT GAC C
IKK $\beta$	Forward primer	CTG AAG ATC GCC TGT AGC AAA
	Reverse primer	TCC ATC TGT AAC CAG CTC CAG

## References

50. Unoki M, Nakamura Y. Growth-suppressive effects of BPOZ and EGR2, two genes involved in the PTEN signaling pathway. *Oncogene*. 2001;20:4457-65.
51. Bagchi A, Papazoglu C, Wu Y, Capurso D, Brodt M, Francis D, et al. CHD5 is a tumor suppressor at human 1p36. *Cell*. 2007;128:459-75.
52. Senchenko VN, Krasnov GS, Dmitriev AA, Kudryavtseva AV, Anedchenko EA, Braga EA, et al. Differential expression of CHL1 gene during development of major human cancers. *PloS one*. 2011;6:e15612.
53. Bruinsma SP, Baranski TJ. Beta2-chimaerin in cancer signaling: connecting cell adhesion and MAP kinase activation. *Cell cycle (Georgetown, Tex)*. 2007;6:2440-4.
54. Van Rossom S, Op de Beeck K, Franssens V, Swinnen E, Schepers A, Ghillebert R, et al. The splicing mutant of the human tumor suppressor protein DFNA5 induces programmed cell death when expressed in the yeast *Saccharomyces cerevisiae*. *Frontiers in oncology*. 2012;2:77.
55. Xu J, Testa JR. DLX5 (distal-less homeobox 5) promotes tumor cell proliferation by transcriptionally regulating MYC. *J Biol Chem*. 2009;284:20593-601.
56. Mercier PL, Bachvarova M, Plante M, Gregoire J, Renaud MC, Ghani K, et al. Characterization of DOK1, a candidate tumor suppressor gene, in epithelial ovarian cancer. *Mol Oncol*. 2011;5:438-53.
57. Xie H, Hu J, Pan H, Lou Y, Lv P, Chen Y. Adenovirus vector-mediated FAM176A overexpression induces cell death in human H1299 non-small cell lung cancer cells. *BMB reports*. 2014;47:104-9.
58. Wang Z, Liu P, Inuzuka H, Wei W. Roles of F-box proteins in cancer. *Nat Rev Cancer*. 2014;14:233-47.

59. Nakhaei-Nejad M, Haddad G, Zhang QX, Murray AG. Facio-genital dysplasia-5 regulates matrix adhesion and survival of human endothelial cells. *Arteriosclerosis, thrombosis, and vascular biology*. 2012;32:2694-701.
60. Niu C, Yan Z, Cheng L, Zhu J, Zhang H, Xu X, et al. Downregulation and antiproliferative role of FHL3 in breast cancer. *IUBMB life*. 2011;63:764-71.
61. Mitsui K, Nakanishi M, Ohtsuka S, Norwood TH, Okabayashi K, Miyamoto C, et al. A novel human gene encoding HECT domain and RCC1-like repeats interacts with cyclins and is potentially regulated by the tumor suppressor proteins. *Biochem Biophys Res Commun*. 1999;266:115-22.
62. Chu MC, Selam FB, Taylor HS. HOXA10 regulates p53 expression and matrigel invasion in human breast cancer cells. *Cancer biology & therapy*. 2004;3:568-72.
63. Gu ZD, Shen LY, Wang H, Chen XM, Li Y, Ning T, et al. HOXA13 promotes cancer cell growth and predicts poor survival of patients with esophageal squamous cell carcinoma. *Cancer Res*. 2009;69:4969-73.
64. Chen H, Chung S, Sukumar S. HOXA5-induced apoptosis in breast cancer cells is mediated by caspases 2 and 8. *Mol Cell Biol*. 2004;24:924-35.
65. Addou-Klouche L, Adelaide J, Finetti P, Cervera N, Ferrari A, Bekhouche I, et al. Loss, mutation and deregulation of L3MBTL4 in breast cancers. *Mol Cancer*. 2010;9:213.
66. Johansson M, Oudin A, Tiemann K, Bernard A, Golebiewska A, Keunen O, et al. The soluble form of the tumor suppressor Lrig1 potently inhibits in vivo glioma growth irrespective of EGF receptor status. *Neuro-oncology*. 2013;15:1200-11.

67. Ko A, Shin JY, Seo J, Lee KD, Lee EW, Lee MS, et al. Acceleration of gastric tumorigenesis through MKRN1-mediated posttranslational regulation of p14ARF. *J Natl Cancer Inst.* 2012;104:1660-72.
68. Wu Q, Yang Z, An Y, Hu H, Yin J, Zhang P, et al. MiR-19a/b modulate the metastasis of gastric cancer cells by targeting the tumour suppressor MXD1. *Cell death & disease.* 2014;5:e1144.
69. Gong Y, Yue J, Wu X, Wang X, Wen J, Lu L, et al. NSPc1 is a cell growth regulator that acts as a transcriptional repressor of p21Waf1/Cip1 via the RARE element. *Nucleic Acids Res.* 2006;34:6158-69.
70. Kim H, Abd Elmageed ZY, Ju J, Naura AS, Abdel-Mageed AB, Varughese S, et al. PDZK1 is a novel factor in breast cancer that is indirectly regulated by estrogen through IGF-1R and promotes estrogen-mediated growth. *Molecular medicine (Cambridge, Mass.* 2013;19:253-62.
71. Lu G, Ren S, Korge P, Choi J, Dong Y, Weiss J, et al. A novel mitochondrial matrix serine/threonine protein phosphatase regulates the mitochondria permeability transition pore and is essential for cellular survival and development. *Genes Dev.* 2007;21:784-96.
72. Wang S, Bian C, Yang Z, Bo Y, Li J, Zeng L, et al. miR-145 inhibits breast cancer cell growth through RTKN. *Int J Oncol.* 2009;34:1461-6.
73. Nagamachi A, Matsui H, Asou H, Ozaki Y, Aki D, Kanai A, et al. Haploinsufficiency of SAMD9L, an endosome fusion facilitator, causes myeloid malignancies in mice mimicking human diseases with monosomy 7. *Cancer Cell.* 2013;24:305-17.

74. Brastianos PK, Horowitz PM, Santagata S, Jones RT, McKenna A, Getz G, et al. Genomic sequencing of meningiomas identifies oncogenic SMO and AKT1 mutations. *Nat Genet.* 2013;45:285-9.
75. Cui Y, Wang J, Zhang X, Lang R, Bi M, Guo L, et al. ECRG2, a novel candidate of tumor suppressor gene in the esophageal carcinoma, interacts directly with metallothionein 2A and links to apoptosis. *Biochem Biophys Res Commun.* 2003;302:904-15.
76. Tanaka N, Kaneko K, Asao H, Kasai H, Endo Y, Fujita T, et al. Possible involvement of a novel STAM-associated molecule "AMSH" in intracellular signal transduction mediated by cytokines. *J Biol Chem.* 1999;274:19129-35.
77. Lai JP, Sandhu DS, Shire AM, Roberts LR. The tumor suppressor function of human sulfatase 1 (SULF1) in carcinogenesis. *Journal of gastrointestinal cancer.* 2008;39:149-58.