

# Analysis of the complete mitochondrial genome of *Pochonia chlamydosporia* suggests a close relationship to the invertebrate-pathogenic fungi in Hypocreales

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## Additional file 1

### Supplementary Tables

**Table S1.** Comparison of the assembled mitogenome sequences with the published *P. chlamydosporia* mt genes. **Table S2.** Gene and intergenic regions sizes from the complete mitogenomes of five Pezizomycotina fungal species. **Table S3.** The three primary contributors to mitogenome size variation. **Table S4.** Distinct codon usage for fungal mitogenome genes. **Table S5.** Codon usage bias statistics. **Table S6.** Nucleotide sequence lengths (bp) of the *rps3* gene and group I introns. **Table S7.** The dN/dS values of the *rps3* sequences. **Table S8.** Summary of the sequenced data of *P. chlamydosporia*. **Table S9.** Proteins used for phylogenetic analyses are shown.

### Supplementary Figures

**Figure S1.** Distribution of the genes and repeats in the mitogenomes of *P. chlamydosporia*, *M. anisopliae* and *L. muscarium*. **Figure S2.** Synteny between *P. chlamydosporia* and *B. bassiana*, *V. dahliae*, *A. fumigatus*. **Figure S3.** Evolutionary characteristics of *rps3*. **Figure S4.** Putative transposition events in the mitogenomes of 20 fungi detected by the use of CREx algorithm.

## Supplementary Tables

**Table S1. Comparison of the assembled mitogenome sequences with the published *P. chlamydosporia* mt genes.** BLASTN were used for alignment analyses.

Accession	Length (bp)	Identity	Matched (%)	E_value	Description
FJ973100.1	323	0.97	100.00	1.00E-170	<i>Pochonia chlamydosporia</i> strain IMI 113169 NADH dehydrogenase subunit 3 ( <i>nad3</i> ) and ATP synthase F0 subunit 9 ( <i>atp9</i> ) genes, partial cds
FJ973052.1	452	0.99	96.90	0	<i>Pochonia chlamydosporia</i> strain IMI 113169 ATP synthase F0 subunit 6 ( <i>atp6</i> ) gene, partial cds; and small subunit ribosomal RNA ( <i>rns</i> ) gene, partial sequence
AY556048.1	439	0.84	97.27	9.00E-83	<i>Pochonia chlamydosporia</i> strain IMI 113169 small subunit ribosomal RNA ( <i>rns</i> ) gene, partial sequence
AY556020.1	158	0.95	100.00	7.00E-72	<i>Pochonia chlamydosporia</i> strain IMI 113169 NADH dehydrogenase subunit 3 ( <i>nad3</i> ) gene, partial cds
AY555993.1	479	0.98	100.00	0	<i>Pochonia chlamydosporia</i> strain IMI 113169 NADH dehydrogenase subunit 1 ( <i>nad1</i> ) gene, partial cds
AY555938.1	363	0.99	100.00	0	<i>Pochonia chlamydosporia</i> strain IMI 113169 cytochrome oxidase subunit III ( <i>cox3</i> ) gene, partial cds
AY556047.1	501	0.99	100.00	0	<i>Pochonia chlamydosporia</i> strain IMI 156157 small subunit ribosomal RNA ( <i>rns</i> ) gene, partial sequence
AY556019.1	158	0.98	100.00	8.00E-84	<i>Pochonia chlamydosporia</i> strain IMI 156157 NADH dehydrogenase subunit 3 ( <i>nad3</i> ) gene, partial cds
AY555992.1	479	0.97	100.00	0	<i>Pochonia chlamydosporia</i> strain IMI 156157 NADH dehydrogenase subunit 1 ( <i>nad1</i> ) gene, partial cds
AY555937.1	363	0.99	100.00	0	<i>Pochonia chlamydosporia</i> strain IMI 156157 cytochrome oxidase subunit III ( <i>cox3</i> ) gene, partial cds
EF469008.1	633	0.99	100.00	0	<i>Cordyceps chlamydosporia</i> strain CBS 101244 ATP synthase subunit 6 ( <i>atp6</i> ) gene, partial cds
EF469040.1	614	0.98	100.00	0	<i>Pochonia chlamydosporia</i> strain CBS 504.66 ATP synthase subunit 6 ( <i>atp6</i> ) gene, partial cds

**Table S2. Gene and intergenic regions sizes from the complete mitogenomes of five Pezizomycotina fungal species.** Some genes share overlap sequences, and length of overlap sequences among coding regions (CRs) is calculated once.

Species	Length (bp)	CRs (bp)	tRNAs (bp)	rRNAs (bp)	Introns (bp)	Intergenic (bp)	Genes have Intron
<i>M. anisopliae</i>	24,673	14,562 (59.02%)	1,786 (7.24%)	6,240 (25.29%)	313 (1.27%)	3,468 (14.06%)	<i>rnl</i>
<i>P. chlamydosporia</i>	25,615	14,385 (56.16%)	1,697 (6.63%)	6,232 (24.33%)	335 (1.31%)	4,618 (18.03%)	<i>rnl</i>
<i>F. oxysporum</i>	34,477	15,399 (44.66%)	1,865 (5.41%)	6,470 (18.77%)	1,397 (4.05%)	11,074 (32.12%)	<i>rnl, nad5</i>
<i>H. jecorina</i>	42,130	19,830 (47.07%)	1,937 (4.60%)	5,904 (14.01%)	10,049 (23.85%)	7,261 (17.23%)	<i>rnl, cox2, cob, cox1</i>
<i>N. crassa</i>	64,840	27,711 (42.74%)	2,074 (3.20%)	7,179 (11.07%)	6,465 (9.97%)	23,155 (35.71%)	<i>rnl, nad3, nad4L, NCU16011, NCU16015, nad1, nad4, atp6, nad5, cob</i>

Note: 1. Overlaps between genes:

*H. jecorina*: *nad4L, nad5*: 1 bp (i.e., 1 bp of overlap sequence between *nad4L* and *nad5*); *cox1*, unknown gene: 690 bp.

*N. crassa*: *nad2*, NCU16002: 1 bp; *nad4L*, NCU16009: 1359 bp; *nad4L*, NCU16010: 1 bp; *nad4L*, NCU16011: 1 bp; *nad4L, nad5*: 1 bp; NCU16010, NCU16011: 1602; NCU16010, *nad5*: 1602 bp; NCU16011, *nad5*: 3047; *cob*, NCU16014: 1 bp; *cob*, NCU16015: 1 bp; NCU16014, NCU16015: 1347; *cox1*, NCU16017: 38 bp; *nad1*, NCU16019: 1416 bp; *nad4*, NCU16021: 2007 bp; *atp6*, NCU16026: 1443 bp.



**Table S4. Distinct codon usage for fungal mitogenome genes.** MA, *M. anisopliae*; PC, *P. chlamydosporia*; HJ, *H. jecorina*; FO, *F. oxysporum*; VD, *V. dahliae*; NC, *N. crassa*; AF, *A. fumigatus*.

AA	Codon	MA (%)	PC (%)	HJ (%)	FO (%)	VD (%)	NC (%)	AF (%)
F	TTC	3.96	4.19	2.78	2.57	2.65	1.73	2.52
I	ATA	7.52	7.3	6.93	7.91	6.5	5.33	5.6
L	CTC	0.04	0	0.11	0.04	0.02	0.27	0.03
	CTG	0.04	0	0.18	0.18	0	0.33	0.03
	TTA	12.73	12.51	9.95	11.09	12.28	8.29	12.27
R	AGG	0.19	0	0.02	0.06	0.04	0.33	0.07
	CGA	0	0.02	0.02	0	0.04	0.23	0
	CGC	0.02	0	0.02	0	0	0.09	0
	CGG	0.06	0	0.05	0.02	0.02	0.07	0.02
U	TAG	0	0.02	0.08	0.06	0.02	0.1	0.05
V	GTA	4.18	4.28	3.27	3.37	3.45	2.5	3.52
W	TGG	0.12	0	0.08	0.04	0	0.18	0.1

**Table S5. Codon usage bias statistics.** MA, *M. anisopliae*; PC, *P. chlamydosporia*; HJ, *H. jecorina*; FO, *F. oxysporum*; VD, *V. dahliae*; NC, *N. crassa*; AF, *A. fumigatus*.

Codon	MA (%)	PC (%)	HJ (%)	FO (%)	VD (%)	NC (%)	AF (%)
A/T	83.05	84.27	85.47	85.28	88.78	81.42	90.17

**Table S6. Nucleotide sequence lengths (bp) of the *rps3* gene and group I introns.**

<b>Species</b>	<b><i>rps3</i></b>	<b>Group I intron</b>
<i>C. brongniartii</i>	1,416	1,926
<i>B. bassiana</i>	1,389	1,876
<i>B. pseudobassiana</i>	1,389	1,887
<i>C. militaris</i>	1,323	1,828
<i>L. muscarium</i>	1,332	1,709
<i>P. chlamydosporia</i>	1,317	1,652
<i>M. anisopliae</i>	1,383	1,696
<i>H. jecorina</i>	1,425	1,535
<i>F. solani</i>	1,536	2,061
<i>F. graminearum</i>	1,482	1,992
<i>F. oxysporum</i>	1,341	1,728
<i>F. verticillioides</i>	1,545	2,445
<i>V. dahlia</i>	1,425	1,774
<i>N. crassa</i>	1,527	2,266
<i>A. fumigatus</i>	1,233	1,601
<i>A. tubingensis</i>	1,218	1,794
<i>A. niger</i>	1,218	1,800
<i>P. brasiliensis</i>	1,212	1,685
<i>T. rubrum</i>	1,140	1,539
<i>E. floccosum</i>	1,140	1,547

**Table S7. The dN/dS values of the *rps3* sequences.** The dN/dS values are generated based on comparative analysis with the *rps3* of *T. rubrum*. FO, *F. oxysporum*; MA, *M. anisopliae*; PC, *P. chlamydosporia*; HJ, *H. jecorina*; FS, *F. solani*; FG, *Fusarium graminearum*; FV, *F. verticillioides*; CM, *C. militaris*; LM, *L. muscarium*; CB, *C. brongniartii*; BB, *B. bassiana*; BP, *B. pseudobassiana*; VD, *V. dahliae*; NC, *N. crassa*; AF, *A. fumigatus*; AT, *A. tubingensis*; AN, *A. niger*; EF, *E. floccosum*; TR, *T. rubrum*; PB, *P. brasiliensis*.

Seq1	Seq2	CODEML (version 4.7a)			DNASP (version 5.10.1)		
		dN	dS	dN/dS	dN	dS	dN/dS
TR	FO	0.7752	0.5873	1.3199	0.7804	0.5742	1.3591
TR	MA	0.8847	0.5919	1.4947	0.8917	0.5772	1.5449
TR	PC	0.7551	0.5438	1.3885	0.7599	0.5324	1.4273
TR	HJ	0.7726	0.5837	1.3236	0.7779	0.5706	1.3633
TR	FV	0.7499	0.5520	1.3584	0.7556	0.5383	1.4037
TR	FG	0.7327	0.5748	1.2748	0.7380	0.5611	1.3153
TR	FS	0.7460	0.5227	1.4272	0.7517	0.5100	1.4739
TR	BB	0.7873	0.4746	1.6589	0.7926	0.4650	1.7045
TR	BP	0.8002	0.4974	1.6089	0.8061	0.4863	1.6576
TR	CB	0.8178	0.4914	1.6642	0.8252	0.4783	1.7253
TR	CM	0.8278	0.5704	1.4512	0.8337	0.5578	1.4946
TR	LM	0.7917	0.5268	1.5030	0.7977	0.5142	1.5513
TR	VD	0.7866	0.4585	1.7158	0.7922	0.4488	1.7652
TR	NC	0.7281	0.5904	1.2333	0.7308	0.5830	1.2535
TR	AN	0.3703	0.3908	0.9474	0.3718	0.3846	0.9667
TR	AT	0.3776	0.3860	0.9783	0.3792	0.3798	0.9984
TR	AF	0.3701	0.4511	0.8205	0.3724	0.4397	0.8469
TR	EF	0.0401	0.1309	0.3061	0.0402	0.1297	0.3099
TR	PB	0.4697	0.5382	0.8728	0.4730	0.5229	0.9046

**Table S8. Summary of the sequenced data of *P. chlamydosporia*.** Three libraries were constructed and sequenced using Illumina HiSeq 2000 technology.

Insert Size (bp)	# of reads	Reads Length (bp)	# of Bases
165	42,331,564	(90, 90)	3,809,840,760
760	47,699,680	(90, 90)	4,292,971,200
4,261	24,517,892	(49, 49)	1,201,376,708
Total			9,304,188,668

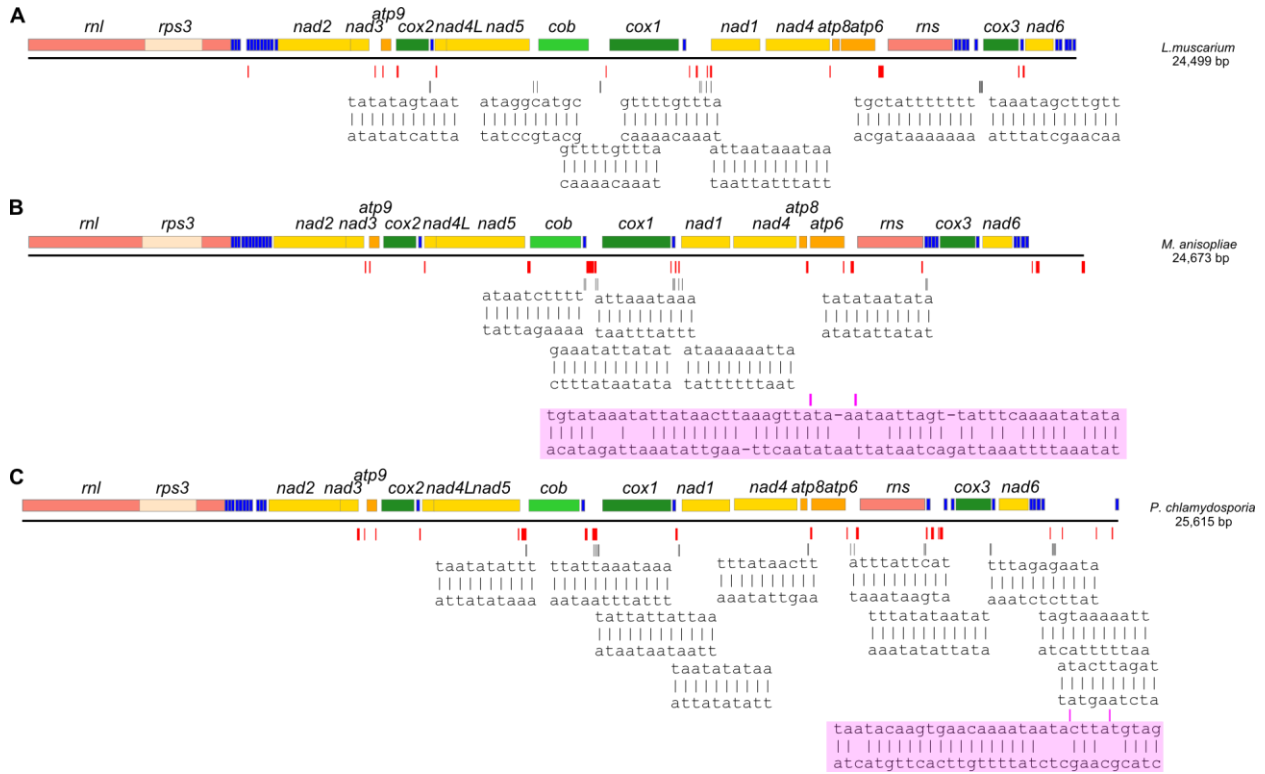
**Table S9. Proteins used for phylogenetic analyses are shown.** The accession numbers of these proteins are obtained from the GenBank files introduced in Table 1. BB, *B. bassiana*; BP, *B. pseudobassiana*; CB, *C. brongniartii*; LM, *L. muscarium*; CM, *C. militaris*; MA, *M. anisopliae*; PC, *P. chlamydosporia*; HJ, *H. jecorina*; FS, *F. solani*; FG, *F. graminearum*; FO, *F. oxysporum*; FV, *F. verticillioides*; VD, *V. dahliae*; NC, *N. crassa*; AN, *A. niger*; AT, *A. tubingensis*; AF, *A. fumigatus*; PB, *P. brasiliensis*; TR, *T. rubrum*; EF, *E. floccosum*; CP, *C. parapsilosis*.

Species	<i>rps3</i>	<i>nad1</i>	<i>nad2</i>	<i>nad3</i>	<i>nad4</i>	<i>nad4L</i>	<i>nad5</i>	<i>nad6</i>	<i>atp6</i>	<i>atp8</i>	<i>atp9</i>	<i>cox1</i>	<i>cox2</i>	<i>cox3</i>	<i>cob</i>
BB	ABY61 762.1	ABY61 755.1	ABY61 748.1	ABY61 749.1	ABY61 756.1	ABY61 752.1	ABY61 761.1	ABY61 760.1	ABY61 758.1	ABY61 757.1	ABY61 750.1	ABY61 754.1	ABY61 751.1	ABY61 759.1	ABY61 753.1
BP	YP_00 875805 4.1	YP_00 875806 3.1	YP_00 875805 5.1	YP_00 875805 6.1	YP_00 875806 4.1	YP_00 875805 9.1	YP_00 875806 0.1	YP_00 875806 8.1	YP_00 875806 6.1	YP_00 875806 5.1	YP_00 875805 7.1	YP_00 875806 2.1	YP_00 875805 8.1	YP_00 875806 7.1	YP_00 875806 1.1
CB	YP_00 221359 3.1	YP_00 221360 5.1	YP_00 221359 4.1	YP_00 221359 5.1	YP_00 221360 7.1	YP_00 221359 8.1	YP_00 221359 9.1	YP_00 221361 1.1	YP_00 221360 9.1	YP_00 221360 8.1	YP_00 221360 6.1	YP_00 221359 2.1	YP_00 221360 7.1	YP_00 221359 0.1	YP_00 221361 0.1
LM	AAO14 657.1	AAL27 687.1	AAO14 658.1	AAO14 659.1	AAO14 666.1	AAO14 662.1	AAO14 663.1	AAO14 669.1	AAO14 668.1	AAO14 667.1	AAO14 660.1	AAO14 665.1	AAO14 661.1	AAO14 688.1	AAL27 664.1
CM	YP_00 881551 7.1	YP_00 881552 6.1	YP_00 881551 8.1	YP_00 881551 9.1	YP_00 881552 7.1	YP_00 881552 2.1	YP_00 881552 3.1	YP_00 881553 1.1	YP_00 881552 9.1	YP_00 881552 8.1	YP_00 881552 8.1	YP_00 881552 5.1	YP_00 881552 1.1	YP_00 881553 0.1	YP_00 881552 4.1
MA	AAW5 8815.1	AAW5 8824.1	AAW5 8816.1	AAW5 8817.1	AAW5 8825.1	AAW5 8820.1	AAW5 8821.1	AAW5 8829.1	AAW5 8827.1	AAW5 8826.1	AAW5 8818.1	AAW5 8823.1	AAW5 8819.1	AAW5 8828.1	AAW5 8822.1
PC	AGY95 315.1	AGY95 324.1	AGY95 316.1	AGY95 317.1	AGY95 310.1	AGY95 320.1	AGY95 321.1	AGY95 314.1	AGY95 312.1	AGY95 311.1	AGY95 318.1	AGY95 323.1	AGY95 319.1	AGY95 313.1	AGY95 322.1
HJ	AAL74 166.1	AAL74 169.1	AAL74 165.1	AAL74 174.1	AAL74 179.1	AAL74 175.1	AAL74 164.1	AAL74 172.1	AAL74 171.1	AAL74 177.1	AAL74 176.1	AAL74 182.1	AAL74 180.1	AAL74 170.1	AAL74 181.1
FS	YP_00 508809 8.1	YP_00 508811 9.1	YP_00 508810 0.1	YP_00 508810 2.1	YP_00 508812 0.1	YP_00 508810 5.1	YP_00 508810 6.1	YP_00 508812 7.1	YP_00 508812 3.1	YP_00 508812 2.1	YP_00 508812 3.1	YP_00 508810 9.1	YP_00 508810 4.1	YP_00 508812 5.1	YP_00 508810 8.1
FG	YP_00 124929 7.1	YP_00 124933 7.1	YP_00 124929 9.1	YP_00 124930 4.1	YP_00 124933 9.1	YP_00 124931 2.1	YP_00 124931 4.1	YP_00 124934 6.1	YP_00 124934 1.1	YP_00 124934 0.1	YP_00 124930 6.1	YP_00 124932 3.1	YP_00 124930 8.1	YP_00 124934 3.1	YP_00 124931 6.1
FO	AAX21 823.1	AAX21 833.1	AAX21 824.1	AAX21 825.1	AAX21 834.1	AAX21 828.1	AAX21 829.1	AAX21 838.1	AAX21 836.1	AAX21 835.1	AAX21 826.1	AAX21 832.1	AAX21 827.1	AAX21 837.1	AAX21 831.1
FV	YP_00 508821 2.1	YP_00 508822 6.1	YP_00 508821 4.1	YP_00 508821 6.1	YP_00 508822 8.1	YP_00 508821 9.1	YP_00 508822 0.1	YP_00 508823 2.1	YP_00 508823 0.1	YP_00 508822 9.1	YP_00 508821 7.1	YP_00 508822 2.1	YP_00 508821 8.1	YP_00 508823 1.1	YP_00 508822 1.1
VD	ABC60 419.1	ABC60 422.1	ABC60 420.1	ABC60 421.1	ABC60 423.1	ABC60 430.1	ABC60 431.1	ABC60 427.1	ABC60 425.1	ABC60 424.1	ABC60 428.1	ABC60 433.1	ABC60 429.1	ABC60 426.1	ABC60 432.1
NC*	NCU16 005	NCU16 018	NCU16 006	NCU16 007	NCU16 020	NCU16 008	NCU16 012	NCU16 004	NCU16 025	NCU16 024	NCU16 027	NCU16 016	NCU16 028	NCU16 003	NCU16 013
AN	rps3_A N**	YP_33 7877.1	YP_33 7891.1	YP_33 7887.1	YP_33 7880.1	YP_33 7889.1	YP_33 7890.1	YP_33 7883.1	YP_33 7882.1	YP_33 7881.1	YP_33 7886.1	YP_33 7885.1	YP_33 7888.1	YP_33 7884.1	YP_33 7876.1
AT	rps3_A T***	YP_39 8768.1	YP_39 8782.1	YP_39 8778.1	YP_39 8771.1	YP_39 8780.1	YP_39 8781.1	YP_39 8774.1	YP_39 8773.1	YP_39 8772.1	YP_39 8777.1	YP_39 8776.1	YP_39 8779.1	YP_39 8775.1	YP_39 8767.1
AF	AFE02 877.1	AFE02 871.1	AFE02 889.1	AFE02 885.1	AFE02 872.1	AFE02 887.1	AFE02 888.1	AFE02 875.1	AFE02 874.1	AFE02 873.1	AFE02 884.1	AFE02 880.1	AFE02 886.1	AFE02 876.1	AFE02 870.1
PB	YP_53 7113.1	YP_53 7107.1	YP_53 7103.1	YP_53 7106.1	YP_53 7108.1	YP_53 7118.1	YP_53 7102.1	YP_53 7111.1	YP_53 7110.1	YP_53 7109.1	YP_53 7116.1	YP_53 7114.1	YP_53 7117.1	YP_53 7112.1	YP_53 7104.1
TR	YP_00 297077 6.1	YP_00 297078 7.1	YP_00 297078 4.1	YP_00 297078 6.1	YP_00 297078 8.1	YP_00 297078 2.1	YP_00 297078 3.1	YP_00 297079 0.1	YP_00 297078 9.1	YP_00 297077 5.1	YP_00 297078 0.1	YP_00 297077 9.1	YP_00 297078 1.1	YP_00 297077 4.1	YP_00 297078 5.1
EF	YP_31 3620.1	YP_31 3638.1	YP_31 3633.1	YP_31 3637.1	YP_31 3639.1	YP_31 3629.1	YP_31 3630.1	YP_31 3641.1	YP_31 3640.1	YP_31 3619.1	YP_31 3627.1	YP_31 3625.1	YP_31 3628.1	YP_31 3618.1	YP_31 3634.1
CP	--	NP_94 3644.1	NP_94 3631.1	NP_94 3630.1	NP_94 3641.2	NP_94 3646.1	NP_94 3647.1	NP_94 3643.1	NP_94 3649.1	NP_94 3648.1	NP_94 3642.1	NP_94 3636.1	NP_94 3635.1	NP_94 3645.1	NP_94 3632.1

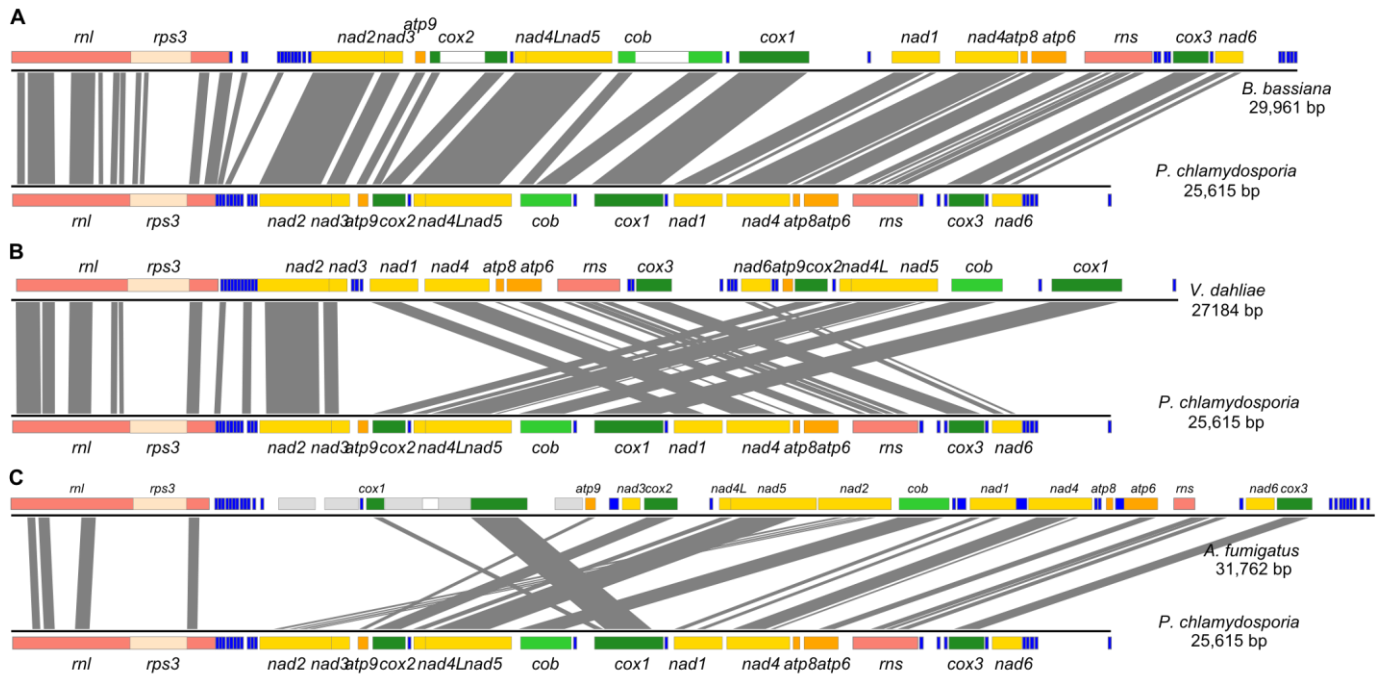
Note: \*, the protein IDs are obtained from the Broad Institute; \*\*, \*\*\*, these two *rps3* genes (*rps3\_AN* and *rps3\_AT*) are predicted based on sequences of homologous *rps3* gene (AFE02877.1) in AF. The location of *rps3\_AN* and *rps3\_AT* in the AN mitogenome (NC\_007445) and AT mitogenome (NC\_007597) are from 16,403 to 17,620 bp and from 16,536 to 17,753 bp, respectively.



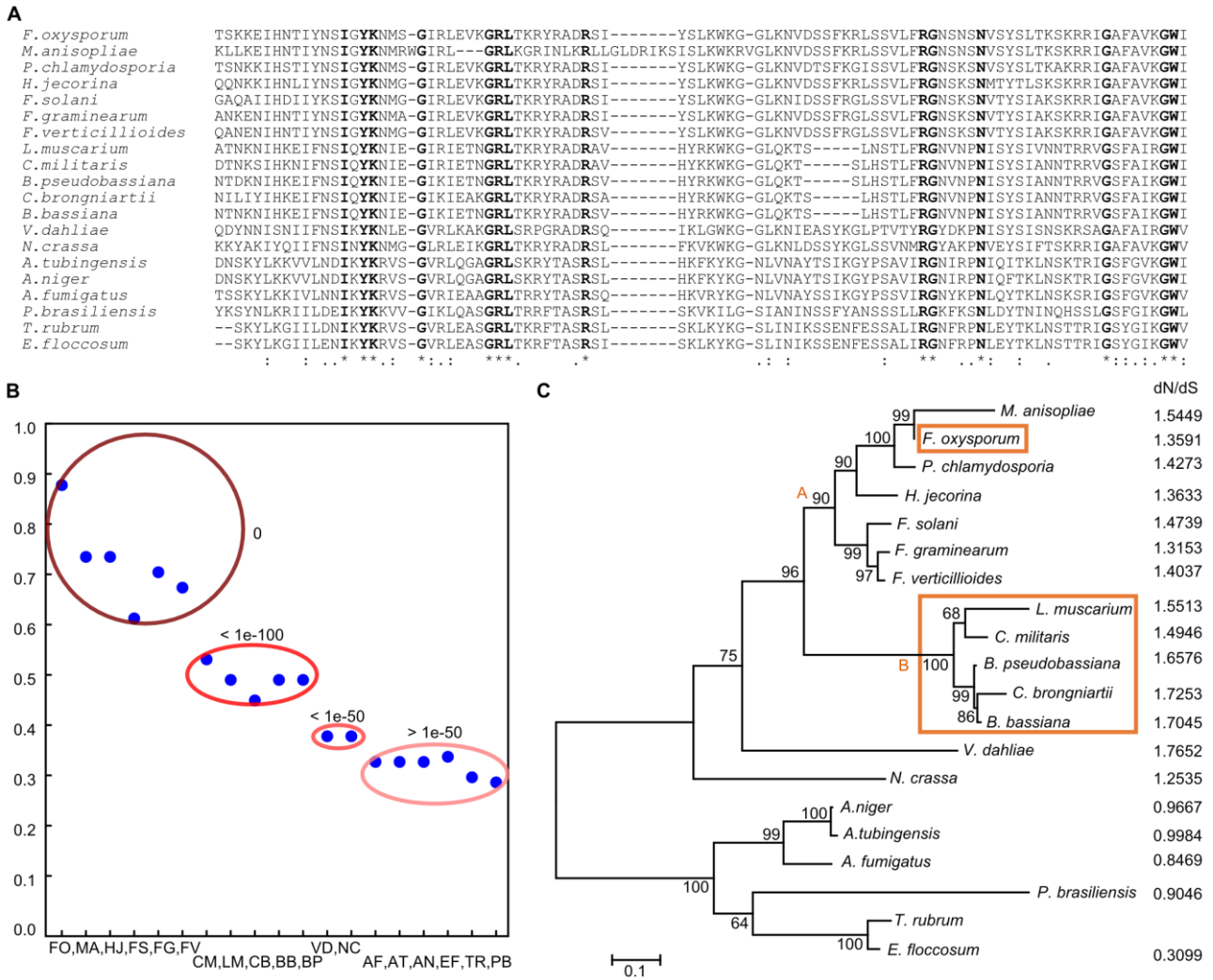
## Supplementary Figures



**Figure S1. Distribution of the genes and repeats in the mitogenomes of *P. chlamydosporia*, *M. anisopliae* and *L. muscarium*.** *trn* genes, tandem repeats, inverted repeats and palindromes are marked in blue, red, pink and grey, respectively. The inverted repeat sequences (such as the pair sequences of “tatatagtaat” from 9,382 bp to 9,392 and “atatatcatta” from 9,403 bp to 9,393 bp) and palindromic sequences are shown. In *L. muscarium*, tandem repeats and palindromes account for 532 bp and 156 bp, respectively; and there are 24 overlaps between tandem repeats and palindromes. In *M. anisopliae*, tandem repeats, inverted repeats and palindromes account for 854 bp, 109 bp and 108 bp, respectively; and 86 bp overlaps between tandem repeats and palindromes are found. In *P. chlamydosporia*, tandem repeats, inverted repeats and palindromes account for 948 bp, 64 bp and 172 bp, respectively; and there are 123 bp overlap sequences.



**Figure S2. Synteny between *P. chlamydosporia* and *B. bassiana*, *V. dahliae*, *A. fumigatus*.** The syntenic regions are identified by BLASTN analysis using E-value cutoff of  $1e^{-5}$ . Alignment blocks in size  $< 100$  bp are not shown. The blue and gray blocks represent *trn* genes and predicted ORFs, respectively.



**Figure S3. Evolutionary characteristics of *rps3*.** A). Alignment of C-terminal domain of ribosomal protein S3. “\*” indicates identical aa residues, which are marked in bold; “.” indicates highly conserved aa residues (scoring > 0.5); and “.” shows weakly similar properties (scoring = 0.5). B). High *rps3* sequence similarity in four distinct groups of fungi. Four groups of E-values are displayed in four circles. FO, *F. oxysporum*; MA, *M. anisopliae*; HJ, *H. jecorina*; FS, *F. solani*; FG, *Fusarium graminearum*; FV, *F. verticillioides*; CM, *C. militaris*; LM, *L. muscarium*; CB, *C. brongniartii*; BB, *B. bassiana*; BP, *B. pseudobassiana*; VD, *V. dahliae*; NC, *N. crassa*; AF, *A. fumigatus*; AT, *A. tubingensis*; AN, *A. niger*; EF, *E. floccosum*; TR, *T. rubrum*; PB, *P. brasiliensis*. C). The Maximum Likelihood (ML) tree of *rps3* from 20 fungal mt genomes. The branches representing *F. oxysporum* and clade B are different from the genomic tree. The dN/dS values are calculated using DNASP (version version 5.10.1) based on comparative analysis with the *rps3* of *T. rubrum*.



t22 (1)	rms Y N cox3 K G D S nad6 V W atp9 cox2 R nad4L nad5 cob P cox1 P rnl T E M M L A F L Q H M nad2 nad3 I S R nad1 nad4 atp8 atp6 rms Y N cox3 K G D S W nad6 V I S P rnl T E M M L A F L Q H M nad2 nad3 nad4L nad5 cob C cox1 R nad1 nad4 atp8 atp6 atp9 cox2 R M nad2b C
t23 (2)	rms Y N cox3 K G D S nad6 V W atp9 cox2 R nad4L nad5 cob P cox1 P rnl T E M M L A F L Q H M nad2 nad3 I S R nad1 nad4 atp8 atp6 rms Y nad6 cox3 K G D S W I S P rnl T E V M M L A F L Q M H cox1 atp9 nad3 cox2 R nad4L nad5 nad2 cob nad1 nad4 R N atp8 atp6 G
t24 (3)	rms Y N cox3 K G D S nad6 V W atp9 cox2 R nad4L nad5 cob P cox1 P rnl T E M M L A F L Q H M nad2 nad3 I S R nad1 nad4 atp8 atp6 rms Y nad6 cox3 K G D S W I S P rnl Y E V M M L A F L Q M H cox1 atp9 nad3 cox2 R nad4L nad5 nad2 cob nad1 nad4 R N atp8 atp6 G
t25 (1)	rms Y N cox3 K G D S nad6 V W atp9 cox2 R nad4L nad5 cob P cox1 P rnl T E M M L A F L Q H M nad2 nad3 I S R nad1 nad4 atp8 atp6 rms Y nad6 cox3 atp8 K G D S I W S P rnl T E V M M L A F L Q M H cox1 R R N atp9 cox2 nad4L nad5 nad2 cob nad3 C nad1 nad4 atp6
t26 (1)	rms Y N cox3 K G D S nad6 V W atp9 cox2 R nad4L nad5 cob P cox1 P rnl T E M M L A F L Q H M nad2 nad3 I S R nad1 nad4 atp8 atp6 rms Y nad6 cox3 I K G D S W S P rnl T E V M M L A F L Q M H R R N cox1 atp9 cox2 nad4L nad5 nad2 cob nad3 C nad1 nad4 atp8 atp6
t27 (2)	rms Y N cox3 K G D S W nad6 V I S P rnl T E M M L A F L Q H M nad2 nad3 nad4L nad5 cob C cox1 R nad1 nad4 atp8 atp6 atp9 cox2 R M nad2b C rms Y nad6 cox3 K G D S W I S P rnl T E V M M L A F L Q M H cox1 atp9 nad3 cox2 R nad4L nad5 nad2 cob nad1 nad4 R N atp8 atp6 G
t28 (3)	rms Y N cox3 K G D S W nad6 V I S P rnl T E M M L A F L Q H M nad2 nad3 nad4L nad5 cob C cox1 R nad1 nad4 atp8 atp6 atp9 cox2 R M nad2b C rms Y nad6 cox3 K G D S W I S P rnl Y E V M M L A F L Q M H cox1 atp9 nad3 cox2 R nad4L nad5 nad2 cob nad1 nad4 R N atp8 atp6 G
t29 (1)	rms Y N cox3 K G D S W nad6 V I S P rnl T E M M L A F L Q H M nad2 nad3 nad4L nad5 cob C cox1 R nad1 nad4 atp8 atp6 atp9 cox2 R M nad2b C rms Y nad6 cox3 K G D S W I S P rnl T E V M M L A F L Q M I H cox1 atp9 A nad3 cox2 R nad4L nad5 nad2 cob C N nad1 I nad4 R N atp8 I atp6
t30 (1)	rms Y nad6 cox3 K G D S W I S P rnl T E V M M L A F L Q M H cox1 atp9 nad3 cox2 R nad4L nad5 nad2 cob nad1 nad4 R N atp8 atp6 G rms Y nad6 cox3 K G D S W I S P rnl Y E V M M L A F L Q M H cox1 atp9 nad3 cox2 R nad4L nad5 nad2 cob nad1 nad4 R N atp8 atp6 G
t31 (1)	rms Y nad6 cox3 K G D S W I S P rnl T E V M M L A F L Q M H cox1 atp9 nad3 cox2 R nad4L nad5 nad2 cob nad1 nad4 R N atp8 atp6 G rms Y nad6 cox3 K G D S W I S P rnl T E V M M L A F L Q M I H cox1 atp9 A nad3 cox2 nad4L nad5 nad2 cob R C N nad1 I nad4 R N atp8 I atp6
t32 (1)	rms Y nad6 cox3 K G D S W I S P rnl T E V M M L A F L Q M H cox1 atp9 nad3 cox2 R nad4L nad5 nad2 cob nad1 nad4 R N atp8 atp6 G rms Y nad6 cox3 atp8 K G D S I W S P rnl T E V M M L A F L Q M H cox1 R R N atp9 cox2 nad4L nad5 nad2 cob nad3 C nad1 nad4 atp6
t33 (1)	rms Y nad6 cox3 K G D S W I S P rnl T E V M M L A F L Q M H cox1 atp9 nad3 cox2 R nad4L nad5 nad2 cob nad1 nad4 R N atp8 atp6 G rms Y nad6 cox3 I K G D S W S P rnl T E V M M L A F L Q M H R R N cox1 atp9 cox2 nad4L nad5 nad2 cob nad3 C nad1 nad4 atp8 atp6
t34 (2)	rms Y nad6 cox3 K G D S W I S P rnl Y E V M M L A F L Q M H cox1 atp9 nad3 cox2 R nad4L nad5 nad2 cob nad1 nad4 R N atp8 atp6 G rms Y nad6 cox3 K G D S W I S P rnl T E V M M L A F L Q M I H cox1 atp9 A nad3 cox2 nad4L nad5 nad2 cob R C N nad1 I nad4 R N atp8 I atp6
t35 (2)	rms Y nad6 cox3 K G D S W I S P rnl Y E V M M L A F L Q M H cox1 atp9 nad3 cox2 R nad4L nad5 nad2 cob nad1 nad4 R N atp8 atp6 G rms Y nad6 cox3 atp8 K G D S I W S P rnl T E V M M L A F L Q M H cox1 R R N atp9 cox2 nad4L nad5 nad2 cob nad3 C nad1 nad4 atp6
t36 (2)	rms Y nad6 cox3 K G D S W I S P rnl Y E V M M L A F L Q M H cox1 atp9 nad3 cox2 R nad4L nad5 nad2 cob nad1 nad4 R N atp8 atp6 G rms Y nad6 cox3 I K G D S W S P rnl T E V M M L A F L Q M H R R N cox1 atp9 cox2 nad4L nad5 nad2 cob nad3 C nad1 nad4 atp8 atp6
t37 (2)	rms Y nad6 cox3 K G D S W I S P rnl T E V M M L A F L Q M I H cox1 atp9 A nad3 cox2 nad4L nad5 nad2 cob R C N nad1 I nad4 R N atp8 I atp6 rms Y nad6 cox3 atp8 K G D S I W S P rnl T E V M M L A F L Q M H cox1 R R N atp9 cox2 nad4L nad5 nad2 cob nad3 C nad1 nad4 atp6
t38 (1)	rms Y nad6 cox3 K G D S W I S P rnl T E V M M L A F L Q M I H cox1 atp9 A nad3 cox2 nad4L nad5 nad2 cob R C N nad1 I nad4 R N atp8 I atp6 rms Y nad6 cox3 I K G D S W S P rnl T E V M M L A F L Q M H R R N cox1 atp9 cox2 nad4L nad5 nad2 cob nad3 C nad1 nad4 atp8 atp6
t39 (2)	rms Y nad6 cox3 atp8 K G D S I W S P rnl T E V M M L A F L Q M H cox1 R R N atp9 cox2 nad4L nad5 nad2 cob nad3 C nad1 nad4 atp6 rms Y nad6 cox3 I K G D S W S P rnl T E V M M L A F L Q M H R R N cox1 atp9 cox2 nad4L nad5 nad2 cob nad3 C nad1 nad4 atp8 atp6

**Figure S4. Putative transposition events in the mitogenomes of 20 fungi detected by the use of CREx algorithm.** Gene orders of *B. bassiana* (BB), *B. pseudobassiana* (BP), *C. brongniartii* (CB), *L. muscarium* (LM), *C. militaris* (CM), *M. anisopliae* (MA), *P. chlamydosporia* (PC) and *H. jecorina* (HJ) are similar. *F. graminearum* (FG), *F. oxysporum* (FO) and *F. verticillioides* (FV) show similar gene orders, and other two species, *T. rubrum* (TR) and *E. floccosum* (EF), display similar gene order. t1: “BB, BP, CB, LM, CM, MA, PC, HJ” vs *F. solani* (FS); t2: “BB, BP, CB, LM, CM, MA, PC, HJ” vs “FG, FO, FV”; t3: “BB, BP, CB, LM, CM, MA, PC, HJ” vs *N. crassa*

(NC); t4: “BB, BP, CB, LM, CM, MA, PC, HJ” vs *A. niger* (AN); t5: “BB, BP, CB, LM, CM, MA, PC, HJ” vs *A. tubingensis* (AT); t6: “BB, BP, CB, LM, CM, MA, PC, HJ” vs *A. fumigatus* (AF); t7: “BB, BP, CB, LM, CM, MA, PC, HJ” vs “TR, EF”; t8: “BB, BP, CB, LM, CM, MA, PC, HJ” vs *P. brasiliensis* (PB); t9: FS vs “FG, FO, FV”; t10: FS vs NC; t11: FS vs AN; t12: FS vs AT; t13: FS vs AF; t14: FS vs “TR, EF”; t15: FS vs PB; t16: “FG, FO, FV” vs NC; t17: “FG, FO, FV” vs AN; t18: “FG, FO, FV” vs AT; t19: “FG, FO, FV” vs AF; t20: “FG, FO, FV” vs “TR, EF”; t21: “FG, FO, FV” vs PB; t22: VD vs NC; t23: VD vs AN; t24: VD vs AT; t25: VD vs “TR, EF”; t26: VD vs PB; t27: NC vs AN; t28: NC vs AT; t29: NC vs AF; t30: AN vs AT; t31: AN vs AF; t32: AN vs “TR, EF”; t33: AN vs PB; t34: AT vs AF; t35: AT vs “TR, EF”; t36: AT vs PB; t37: AF vs “TR, EF”; t38: AF vs PB; t39: “TR, EF” vs PB. There are 21 transposition events in all, and transpositions were marked, such as two events (1 and 15) in t1: “G nad6 V I S W P rnl T E M M L” vs “nad6 V I S W P rnl T E M M L G”, and “K L Q H M” vs “M K L Q H”.