

Supporting Information

for

3 α ,5 α -Cyclocholestan-6 β -yl ethers as donors of the cholesterol moiety for the electrochemical synthesis of cholesterol glycoconjugates

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**Experimental section including ¹H, ¹³C NMR, and mass spectra
for all new compounds**

Experimental

Cyclic voltammograms were recorded with iR compensation at 25 °C using a three-electrode potentiostat (Princeton Applied Research, model Parstat 2273). The experiments were conducted in a 3-mL electrochemical cell with an argon-purge system. The working electrode was a Bioanalytical Systems platinum inlay (1 mm in diameter), the auxiliary electrode was a platinum mesh (contained in a glass tube with a medium porosity glassfrit), and the reference electrode was Ag/0.1 M AgNO₃ in acetonitrile. The latter was contained in a Pyrex tube with a cracked softglass tip which was placed inside a Luggin capillary. Before each experiment, the working electrode was polished using Buehler Micropolish Alumina Gamma 3B and a Buehler Microcloth polishing cloth, rinsed with dichloromethane and dried. In all of the measurements, 0.2M solution of tetrabutylammonium tetrafluoroborate (TBABF₄) from Aldrich in dichloromethane was used as a supporting electrolyte.

The preparative electrolyses were performed with a potentiostat/galvanostat (Princeton Applied Research, model Parstat 2273) under galvanostatic conditions using a current that was equal in a typical experiment to 7.5 mA and a reaction time of 4000 s. The current applied was the maximum current available for the electrolysis set-up being used (power supply and ohmic resistance). The reactions were monitored by TLC and stopped when no further increase in the concentration of the glycosylation products was observed. A divided H-cell was used in which the cathodic and anodic compartments (3.5 mL of electrolyte each) were separated by a glass frit. In all measurements, 0.1 M solution of tetrabutylammonium tetrafluoroborate (TBABF₄) from Aldrich in dichloromethane was used as a supporting electrolyte. The steroid (0.30 mmol) and sugar (0.36 mmol) substrates were introduced into the anodic compartment together with 0.3 g of 3 Å molecular sieves added to eliminate traces of water, whereas anionite (1.5–2 g, Dowex 2 × 8, 200–400 mesh, perchlorate form) was placed in the cathodic compartment to eliminate chloride ions that are formed by the reduction of dichloromethane. The solutions in both compartments were stirred during electrolysis and, additionally, a continuous flow of argon was applied in the anodic compartment. A platinum mesh was used as a cathode and a platinum plate (2 × 1.5 cm) was used as an anode. All measurements were performed at 25 °C.

The sugar (1,2:3,4-di-*O*-isopropylidene- α -D-galactopyranose; **7**) [1] and steroidal substrates, 3 α ,5 α -cyclocholestan-6 β -ol (*i*-cholesterol; **6a**) [2], 6 β -methoxy-3 α ,5 α -cyclocholestane (**6b**) [3], and 6 β -ethoxy-3 α ,5 α -cyclocholestane (**6c**) [4], were prepared according to known procedures.

Melting points were determined on a Toledo Mettler-MP70 apparatus. ¹H and ¹³C NMR (400 and 100 MHz, respectively) spectra were recorded on a Bruker Avance II spectrometer in CDCl₃ solutions with TMS as the internal standard (only selected signals in the ¹H NMR spectra are reported; sugar protons are marked with the 'prime' index). Infrared spectra were recorded on a Nicolet series II Magna-IR 550 FTIR spectrometer in chloroform solutions. Mass spectra were recorded at 70 eV with a time-of-flight (TOF) AMD-604 spectrometer with electrospray ionization (ESI) or AutoSpec Premier (Waters) (EI).

Merck Silica Gel 60, F 256 TLC aluminum sheets were applied for thin-layer chromatographic analysis. For a visualization of the products, a 5% solution of phosphomolybdic acid in ethanol was used. The reaction products were separated by column chromatography performed on a 70–230 mesh silica gel (J. T. Baker).

Synthesis of 6 β -benzyloxy-3 α ,5 α -cyclocholestane (**6e**)

To cholesteryl *p*-tosylate (1 g; 1.9 mmol) dissolved in dioxane (50 mL) freshly dried potassium acetate (0.8 g; 5.7 mmol; 3 equiv.) and benzyl alcohol (6.2 g; 57 mmol; 30 equiv.) were added. The reaction mixture was refluxed for 24 h. After cooling it was poured into water (200 mL) and extracted with benzene (3 \times 100 mL). The extract was dried over anhydrous Na₂SO₄ and evaporated to dryness *in vacuo*. The residue was subjected to silica gel column chromatography. Elution with CH₂Cl₂/hexane (1:9) afforded 6 β -benzyloxy-3 α ,5 α -cyclocholestane (**6e**; 0.43 g; 48%).

6e: Colorless oil; $[\alpha]_D^{20}$ +37.0 (*c* 1.0, CHCl₃); R_f = 0.31 (hexane-AcOEt 98:2); IR, ν_{\max} (cm⁻¹): 3064, 1496, 1093, 1067; ¹H NMR (ppm), δ : 7.35 (m, 4H, H-Ar), 7.27 (m, 1H, H-Ar), 4.66 (d, 1H, *J* = 12.5 Hz, O-CH₂Ph), 4.50 (d, 1H, *J* = 12.5 Hz, O-CH₂Ph), 2.98 (m, 1H, H-6), 1.11 (s, 3H, H-19), 0.93 (d, 3H, *J* = 6.5 Hz, H-21), 0.887 (d, 3H, *J* = 6.6 Hz, H-26 or H-27), 0.883 (d, 3H, *J* = 6.6 Hz, H-26 or H-27), 0.72 (s, 3H, H-18), 0.67 (dd, 1H, *J* = 4.9 Hz, *J* = 4.1 Hz, H-4 α), 0.41 (dd, 1H, *J* = 8.0 Hz, *J* = 4.9 Hz, H-4 β); ¹³C NMR (ppm), δ : 140.1 (C), 128.2 (CH), 127.1 (CH), 126.9 (CH), 80.2 (CH), 70.0 (CH₂), 56.6 (CH), 56.4 (CH), 48.1 (CH), 43.3 (C), 42.8 (C), 40.4 (CH₂), 39.5 (CH₂), 36.2 (CH₂), 35.8 (CH), 35.7 (C), 35.4 (CH₂), 33.4 (CH₂), 30.6 (CH), 28.3 (CH₂), 28.0 (CH), 25.0 (CH₂), 24.2 (CH₂), 23.9 (CH₂), 22.84 (CH₂), 22.81 (CH₃), 22.6 (CH₃), 21.8 (CH), 19.5 (CH₃), 18.7 (CH₃), 13.2 (CH₂), 12.3 (CH₃); EI MS, *m/z*: 476 (M⁺, 35%), 461 [(M-Me)⁺, 7%], 385 [(M-Bn)⁺, 77%], 370 [(M-Bn-Me)⁺, 67%], 91 (100%).

6 β -Isopropoxy-3 α ,5 α -cyclocholestane (**6d**) was obtained in a similar way (compound **6d** was eluted with hexane).

6d: Colorless oil; $[\alpha]_D^{20}$ +43.4 (*c* 1.0, CHCl₃); R_f = 0.39 (hexane-AcOEt 98:2); IR, ν_{\max} (cm⁻¹): 1138, 1124, 1041; ¹H NMR (ppm), δ : 3.74 (h, 1H, *J* = 6.1 Hz, O-CH(Me)₂), 2.93 (m, 1H, H-6), 1.08 (d, 6H, *J* = 6.2 Hz, (CH₃)₂CH), 1.02 (s, 3H, H-19), 0.93 (d, 3H, *J* = 6.6 Hz, H-21), 0.881 (d, 3H, *J* = 6.6 Hz, H-26 or H-27), 0.877 (d, 3H, *J* = 6.6 Hz, H-26 or H-27), 0.73 (s, 3H, H-18), 0.58 (dd, 1H, *J* = 4.9 Hz, *J* = 3.9 Hz, H-4 α), 0.33 (dd, 1H, *J* = 7.9 Hz, *J* = 4.9 Hz, H-4 β); ¹³C NMR (ppm), δ : 76.9 (CH), 67.1 (CH), 56.6 (CH), 56.4 (CH), 48.2 (CH), 43.2 (C), 42.8 (C), 40.4 (CH₂), 39.6 (CH₂), 36.4 (C), 36.2 (CH₂), 35.9 (CH), 35.8 (CH₂), 33.4 (CH₂), 30.4 (CH), 28.4 (CH₂), 28.0 (CH), 25.1 (CH₂), 24.3 (CH₂), 23.9 (CH₂), 22.9 (CH₂), 22.8 (CH₃), 22.6 (CH₃), 22.52 (CH₃), 22.47 (CH₃), 22.3 (CH), 19.6 (CH₃), 18.7 (CH₃), 12.8 (CH₂), 12.2 (CH₃); EI MS, *m/z*: 428 [M⁺, 56%], 413 [(M-Me)⁺, 20%], 386 [(M-propene)⁺, 17%], 371 [(M-Me-propene)⁺, 81%], 43 (100%).

6 β -(4-Hydroxyphenyloxy)-3 α ,5 α -cyclocholestane (**6g**) was obtained in a similar way (compound **6g** was eluted with a hexane-ethyl acetate (93:7) mixture).

6g: Beige crystalline material, mp 54-56 °C (CH₂Cl₂-hexane); $[\alpha]_D^{20}$ +24.4 (*c* 0.33, CHCl₃); R_f = 0.27 (hexane-AcOEt 9:1); IR, ν_{\max} (cm⁻¹): 3601, 3340, 1601, 1507, 1175, 827; ¹H NMR (ppm), δ : 6.79 (d, 2H, *J* = 9.0 Hz, H-Ar), 6.72 (d, 2H, *J* = 9.0 Hz, H-Ar), 4.36 (s, 1H, -OH), 3.64 (m, 1H, H-6), 1.16 (s, 3H, H-19), 0.93 (d, 3H, *J* = 6.5 Hz, H-21), 0.876 (d, 3H, *J* = 6.6 Hz, H-26 or H-27), 0.872 (d, 3H, *J* = 6.6 Hz, H-26 or H-27), 0.73 (s, 3H, H-18), 0.60 (dd, 1H, *J* = 5.1 Hz, *J* = 3.9 Hz, H-4 α), 0.34 (dd, 1H, *J* = 7.9 Hz, *J* = 5.1 Hz, H-4 β); ¹³C NMR (ppm), δ : 152.4 (C), 150.1 (C), 119.2 (CH), 116.0 (CH), 82.5 (CH), 56.39 (CH), 56.35 (CH), 47.7 (CH), 43.2 (C), 42.7 (C), 40.2 (CH₂), 39.5 (CH₂), 36.6 (C), 36.2 (CH₂), 35.8 (CH), 34.5 (CH₂), 33.4 (CH₂), 30.3 (CH), 28.2 (CH₂), 28.0 (CH), 25.0 (CH₂), 24.1 (CH₂), 23.8 (CH₂), 23.5 (CH), 22.8 (CH₃), 22.7 (CH₂), 22.5 (CH₃), 19.9 (CH₃), 18.7 (CH₃), 12.6 (CH₂), 12.1 (CH₃); EI MS, *m/z*: 478 (M⁺, 1%), 369 [(M-*p*-OH-PhO)⁺, 100%].

Synthesis of 6 β -phenyloxy-3 α ,5 α -cyclocholestane (**6f**)

Phenol (10 g; 0.1 mol, 36 equiv.) was melted at 42 °C. Then cholesteryl *p*-tosylate (1.5 g; 2.8 mmol) and freshly dried potassium acetate (0.8 g; 8.2 mmol, 3 equiv.) were added. The stirred reaction mixture was maintained at this temperature for 2 h. After cooling it was poured into 100 mL of 1 M NaOH solution and extracted with benzene (3 \times 100 mL). The extract was dried over anhydrous Na₂SO₄ and evaporated to dryness *in vacuo*. The reaction products were separated by silica gel column chromatography. 6 β -Phenyloxy-3 α ,5 α -cyclocholestane (**6f**; 0.27 g, 21%) was eluted with hexane. Further elution with a hexane/ethyl acetate (99:1) mixture afforded cholesteryl phenyl ether **12f** (0.41 g; 32%).

6f: White crystals, mp 58-61 °C (AcOEt-hexane); $[\alpha]_D^{20} +36.5$ (*c* 0.33, CHCl₃); R_f = 0.28 (hexane); IR, ν_{\max} (cm⁻¹): 3064, 1597, 1243; ¹H NMR (ppm), δ : 7.26 (m, 2H, H-Ar), 6.91 (m, 3H, H-Ar), 3.82 (m, 1H, H-6), 1.19 (s, 3H, H-19), 0.94 (d, 3H, *J* = 6.5 Hz, H-21), 0.893 (d, 3H, *J* = 6.6 Hz, H-26 or H-27), 0.889 (d, 3H, *J* = 6.6 Hz, H-26 or H-27), 0.74 (s, 3H, H-18), 0.62 (dd, 1H, *J* = 5.2 Hz, *J* = 3.9 Hz, H-4 α), 0.36 (dd, 1H, *J* = 7.9 Hz, *J* = 5.2 Hz, H-4 β); ¹³C NMR (ppm), δ : 158.6 (C), 129.2 (CH), 120.5 (CH), 116.9 (CH), 80.7 (CH), 56.4 (CH), 56.3 (CH), 47.7 (CH), 43.2 (C), 42.7 (C), 40.2 (CH₂), 39.5 (CH₂), 36.7 (C), 36.2 (CH₂), 35.8 (CH), 34.3 (CH₂), 33.3 (CH₂), 30.4 (CH), 28.2 (CH₂), 28.0 (CH), 25.1 (CH₂), 24.1 (CH₂), 23.9 (CH₂), 23.6 (CH), 22.81 (CH₃), 22.76 (CH₂), 22.6 (CH₃), 19.8 (CH₃), 18.7 (CH₃), 12.6 (CH₂), 12.2 (CH₃); EI MS, *m/z*: 462 (M⁺, 1%), 369 [(M-PhO)⁺, 100%].

Synthesis of 6 β -*t*-butyldimethylsilyloxy-3 α ,5 α -cyclocholestane (**6h**)

i-Cholesterol (1 g; 2.6 mmol), TBDMSCl (584 mg; 3.88 mmol, 1.5 equiv.), imidazole (530 mg; 7.8 mmol; 3 equiv.), and DMAP (64 mg; 0.52 mmol; 0.2 equiv.) were dissolved in 16 mL of anhydrous DMF. The reaction was carried out for 5 h (TLC control) at 80 °C under argon. The reaction was poured into water and extracted with benzene. The extract was dried with anhydrous Na₂SO₄ and the solvent was evaporated *in vacuo*. The residue was subjected to flash column chromatography with hexane as an eluent. Yield of 6 β -*t*-butyldimethylsilyloxy-3 α ,5 α -cyclocholestane (**6h**) - 688 mg (53%).

6h: White crystals, mp 70-71 °C (AcOEt-hexane); $[\alpha]_D^{20} +27.0$ (*c* 1.0, CHCl₃); R_f = 0.84 (hexane); IR, ν_{\max} (cm⁻¹): 1254, 1065; ¹H NMR (ppm), δ : 3.20 (m, 1H, H-6), 1.02 (s, 3H, H-19), 0.93 (d, 3H, *J* = 6.5 Hz, H-21), 0.89 (s, 9H, *t*-Bu), 0.881 (d, 3H, *J* = 6.6 Hz, H-26 or H-27), 0.877 (d, 3H, *J* = 6.6 Hz, H-26 or H-27), 0.72 (s, 3H, H-18), 0.48 (dd, 1H, *J* = 4.9 Hz, *J* = 3.8 Hz, H-4 α), 0.18 (dd, 1H, *J* = 7.9 Hz, *J* = 4.9 Hz, H-4 β), 0.01 (s, 3H, Si-CH₃), -0.01 (s, 3H, Si-CH₃); ¹³C NMR (ppm), δ : 74.4 (CH), 56.4 (CH), 56.3 (CH), 47.9 (CH), 43.3 (C), 42.7 (C), 40.4 (CH₂), 39.6 (CH₂), 39.1 (CH₂), 38.3 (C), 36.3 (CH₂), 35.9 (CH), 33.4 (CH₂), 30.0 (CH), 28.4 (CH₂), 28.1 (CH), 25.8 (CH₃), 25.2 (CH₂), 24.3 (CH₂), 23.9 (CH₂), 23.6 (CH), 22.9 (CH₃), 22.8 (CH₂), 22.6 (CH₃), 20.2 (CH₃), 18.8 (CH₃), 18.0 (C), 12.8 (CH₂), 12.0 (CH₃), -4.5 (CH₃), -4.8 (CH₃); ESI MS, *m/z*: 523 [(M+Na)⁺, 20%].

Typical electrochemical experiment. Anodic oxidation of 6 β -phenyloxy-3 α ,5 α -cyclocholestane (**6f**) in the presence of 1,2:3,4-di-*O*-isopropylidene-D-galactopyranose (**7**)

6 β -Phenyloxy-3 α ,5 α -cyclocholestane (138 mg; 0.30 mmol) and 1,2:3,4-di-*O*-isopropylidene-D-galactopyranose (94 mg; 0.36 mmol) were dissolved in a 0.1 M solution of tetrabutylammonium-tetrafluoroborate in dichloromethane (3.5 mL) and introduced into the anodic compartment together with 0.5 g 3 Å molecular sieves to eliminate traces of water. The same supporting electrolyte was placed in the cathodic compartment with anionite (2 g, Dowex 2x8, 200–400 mesh,

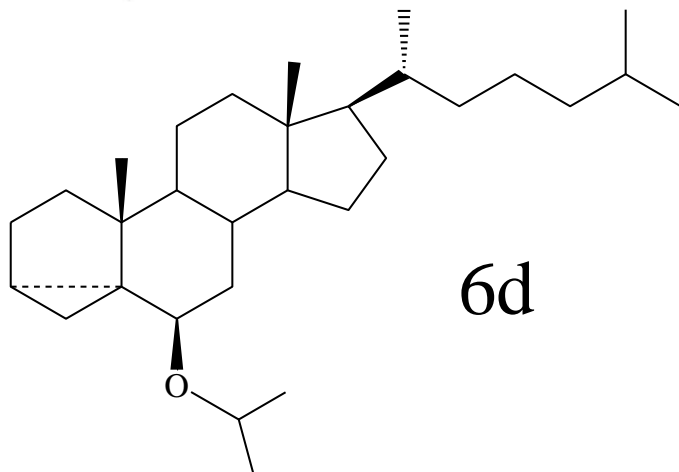
perchlorate form) added. Preparative electrolysis was carried out in a divided H-cell in which the cathodic and anodic compartments (3.5 mL of electrolytes each) were separated by a glass frit under galvanostatic conditions. A direct current 7.5 mA was run for 4000 s. A platinum mesh was used as a cathode and a platinum plate (2 × 1.5 cm) was used as an anode. Ag/0.1 M AgNO₃ in acetonitrile electrode was used as a reference. When the electrolysis was completed, the solvent was removed from the reaction mixture and the products were separated by silica gel column chromatography. The hexane elution afforded diene **13** (1 mg; 1%) and cholesteryl chloride **14** (1 mg; 1%). With the hexane-ethyl acetate mixture (96:4), cholesteryl phenyl ether **12f** (31 mg; 22%) was eluted. Further elution with hexane-ethyl acetate (93:7) afforded 3β-O-(1',2':3',4'-di-O-isopropylidene-α-D-galactopyranos-6'-yl)-cholest-5-ene **11** (108 mg; 58%), followed by cholesterol **1** (5 mg, 4%) eluted with hexane/ethyl acetate (9:1).

Glycosylation product **11** was described in our previous report [5]. Also, other products of the electrochemical reactions (compounds **2**, **13**, **14**, and **15**) were described in our previous papers [6-9]. The isomerization products, i.e., 3β-cholesteryl ethers **12b** [10], **12c** [11], **12e** [12], **12f** [12], **12g** [9], and **12h** [13], are known compounds, except for **12d** which was obtained during electrochemical reaction of 3α,5α-cyclocholestan-6β-yl isopropyl ether (**6d**).

12d: White crystals, mp 122-124 °C (AcOEt-hexane); $[\alpha]_D^{20}$ -24.1 (c 1.0, CHCl₃); R_f = 0.44 (hexane-AcOEt 95:5); IR, ν_{max} (cm⁻¹): 1125, 1064; ¹H NMR (ppm), δ: 5.35 (m, 1H, H-6), 3.73 (h, 1H, J = 6.1 Hz, O-CH(Me)₂), 3.22 (m, 1H, H-6), 1.151 (d, 3H, J = 6.1 Hz, CH₃-isopropyl), 1.148 (d, 3H, J = 6.1 Hz, CH₃-isopropyl), 1.01 (s, 3H, H-19), 0.93 (d, 3H, J = 6.6 Hz, H-21), 0.877 (d, 3H, J = 6.6 Hz, H-26 or H-27), 0.873 (d, 3H, J = 6.6 Hz, H-26 or H-27), 0.67 (s, 3H, H-18); ¹³C NMR (ppm), δ: 141.4 (C), 121.3 (CH), 76.3 (CH), 68.5 (CH), 56.8 (CH), 56.2 (CH), 50.3 (CH), 42.3 (C), 40.0 (CH₂), 39.8 (CH₂), 39.5 (CH₂), 37.5 (CH₂), 36.9 (C), 36.2 (CH₂), 35.8 (CH), 32.0 (CH₂), 31.9 (CH), 29.3 (CH₂), 28.2 (CH₂), 28.0 (CH), 24.3 (CH₂), 23.8 (CH₂), 23.03 (CH₃), 22.95 (CH₃), 22.8 (CH₃), 22.6 (CH₃), 21.1 (CH₂), 19.4 (CH₃), 18.7 (CH₃), 11.9 (CH₃); EI MS, m/z: 428 [M⁺, 15%], 370 [(M-i-Pr-Me)⁺, 63%], 329 (100%).

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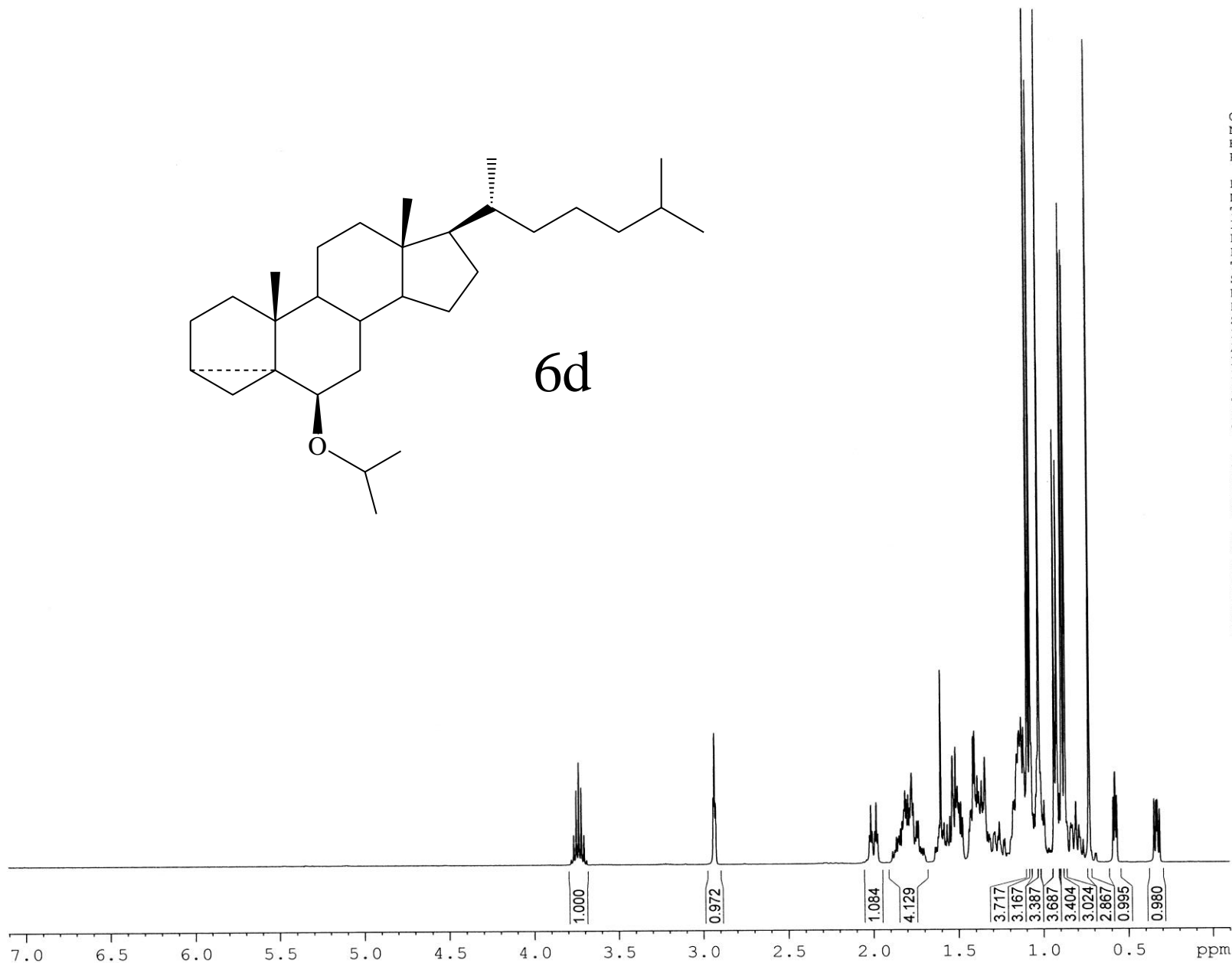


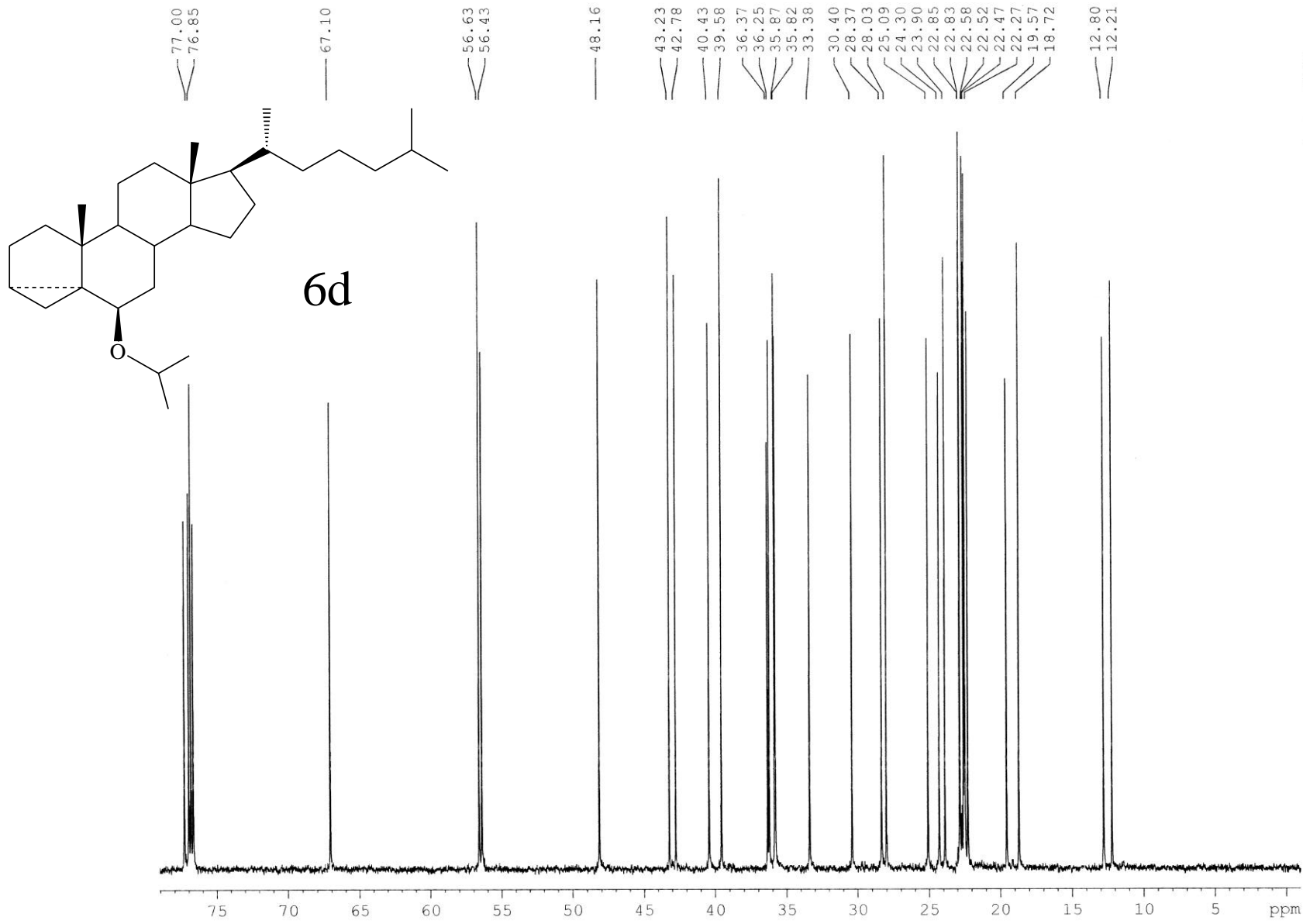
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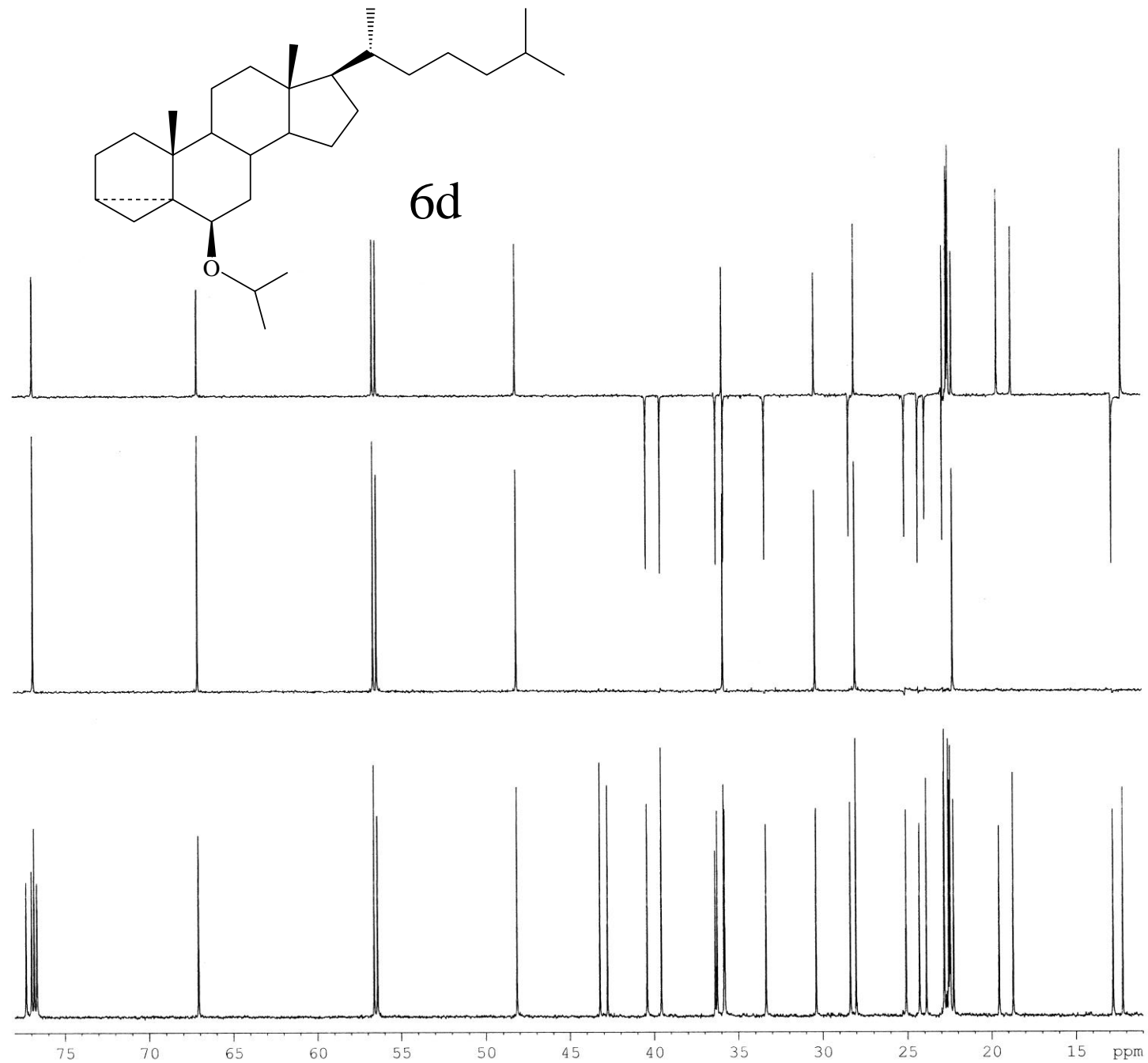
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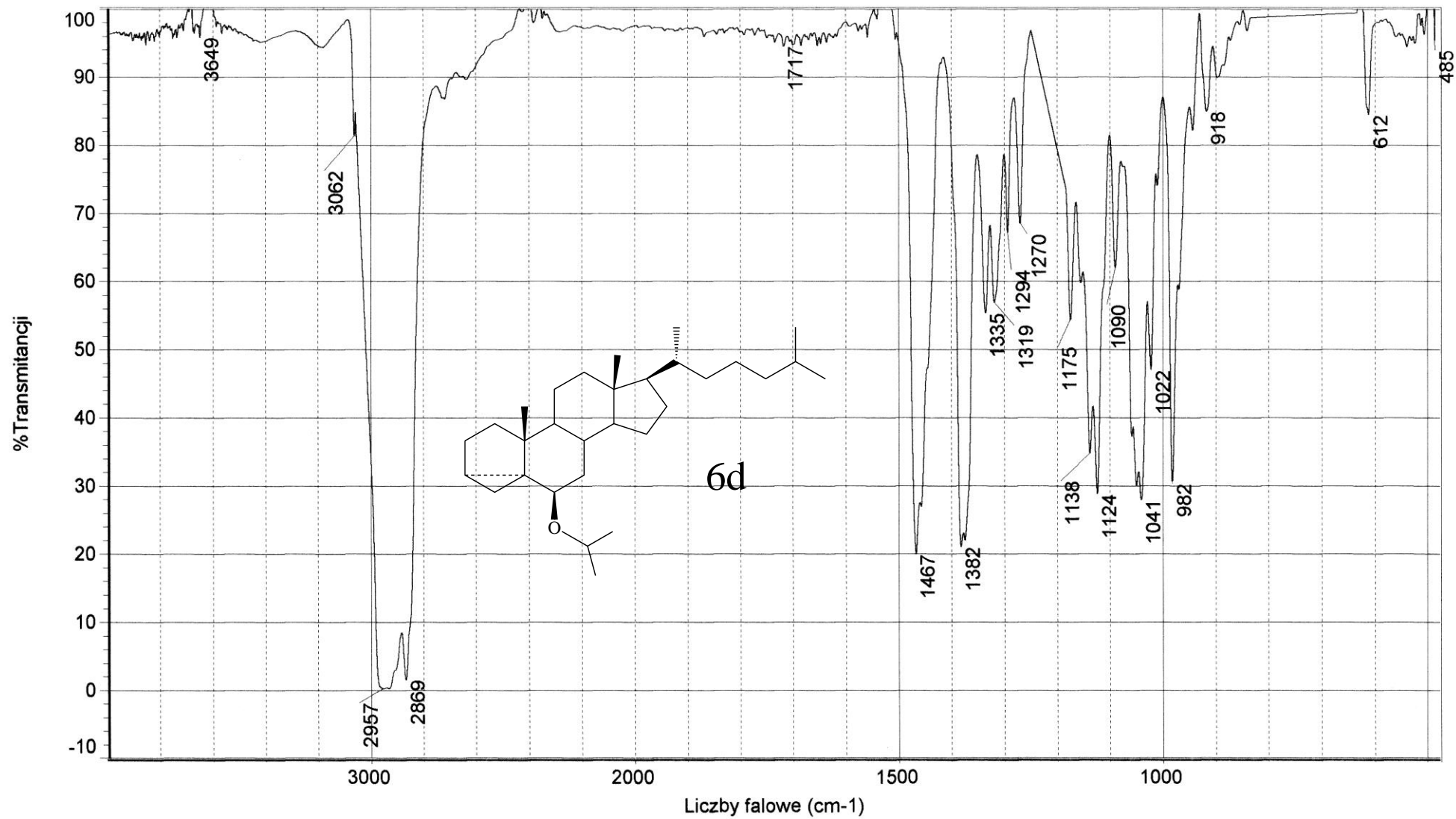
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SSB            0
LB             1.00 Hz
GB            0
PC            0.20

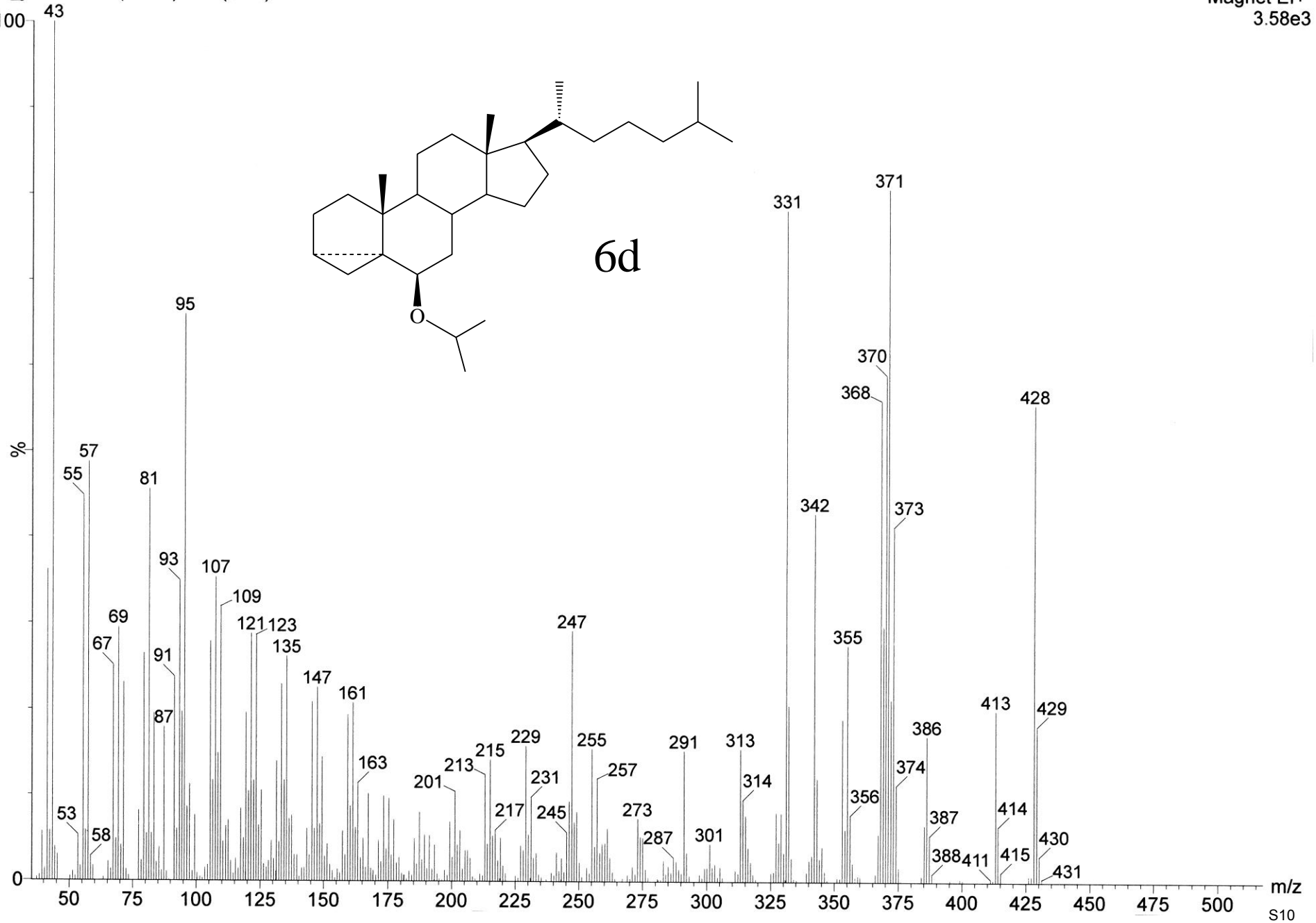
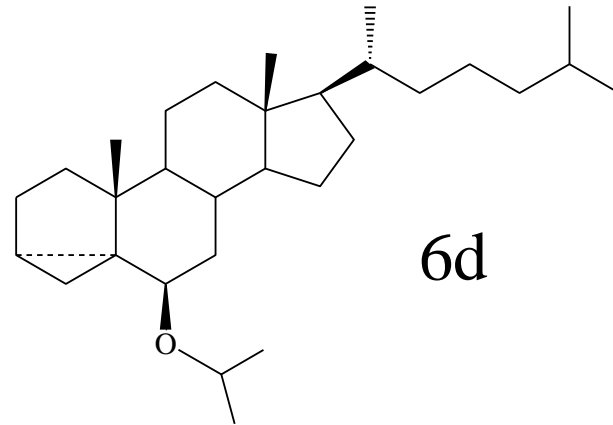
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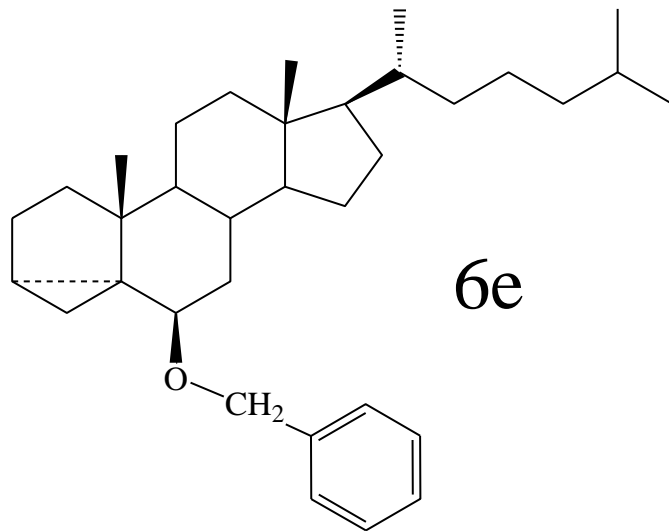



AT214

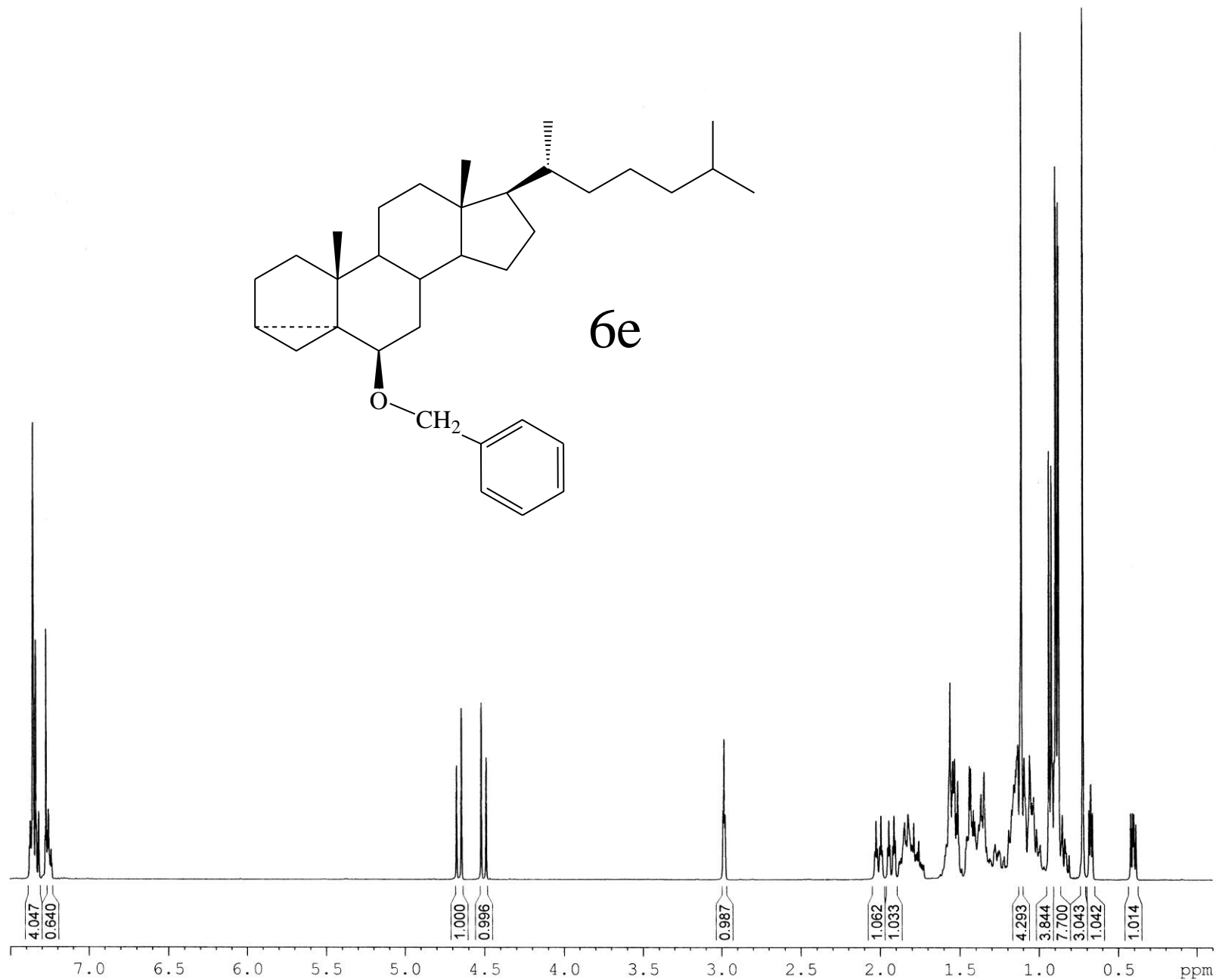
ub_jm3232 12 (0.952) Cm (9:17)

Magnet EI+
3.58e3





6e

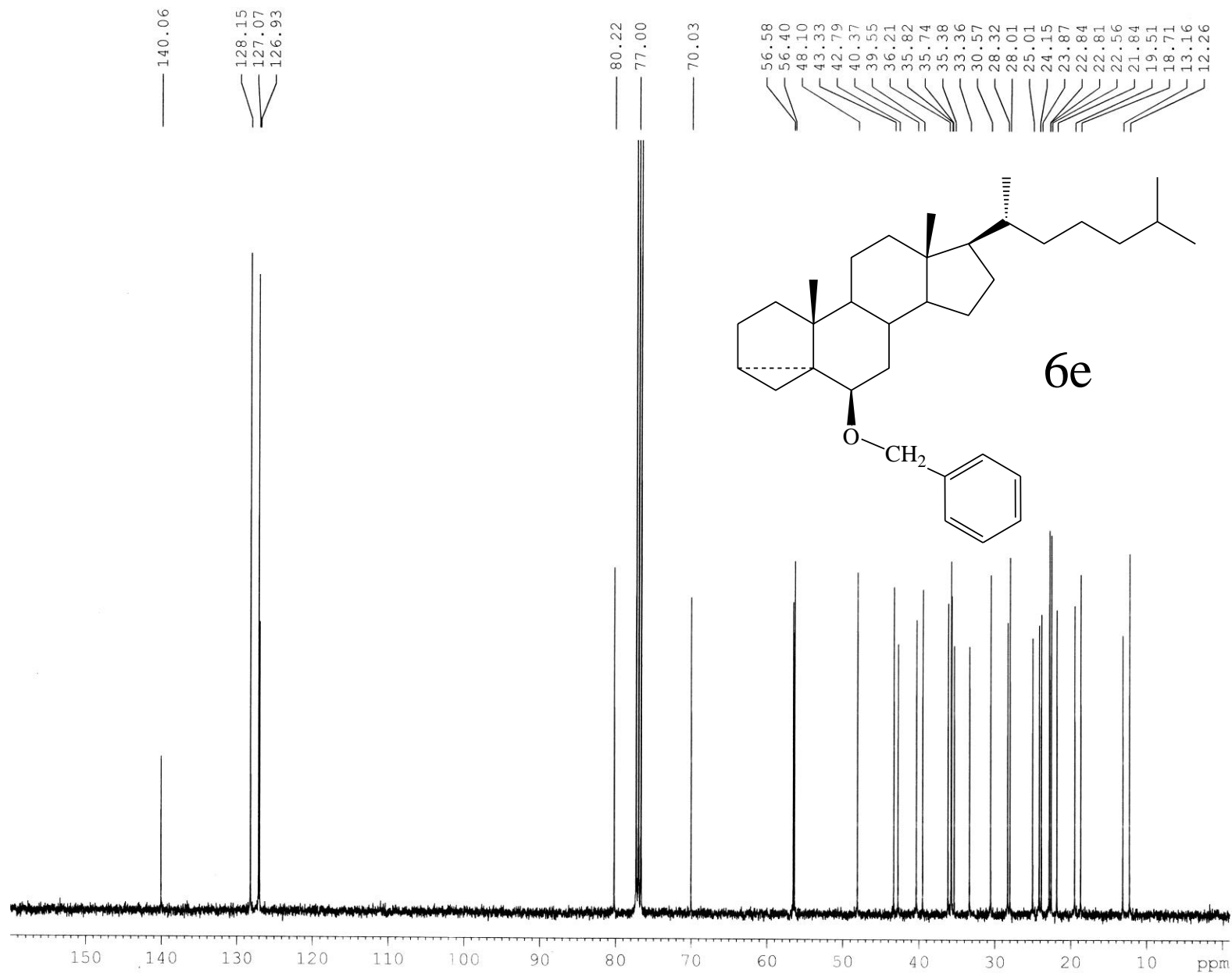


Current Data Parameters
 NAME AT benzytowy W6f1
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20131129
 Time_ 14.28
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3
 NS 128
 DS 0
 SWH 8223.685 Hz
 FIDRES 0.125483 Hz
 AQ 3.9846387 sec
 RG 114
 DW 60.800 usec
 DE 8.00 usec
 TE 673.2 K
 D1 1.00000000 sec
 TD0 1

==== CHANNEL f1 =====
 NUC1 1H
 P1 11.15 usec
 PL1 -3.00 dB
 SFO1 400.1524711 MHz

F2 - Processing parameters
 SI 32768
 SF 400.1500000 MHz
 WDW GM
 SSB 0
 LB -0.20 Hz
 GB 0.2
 PC 1.00



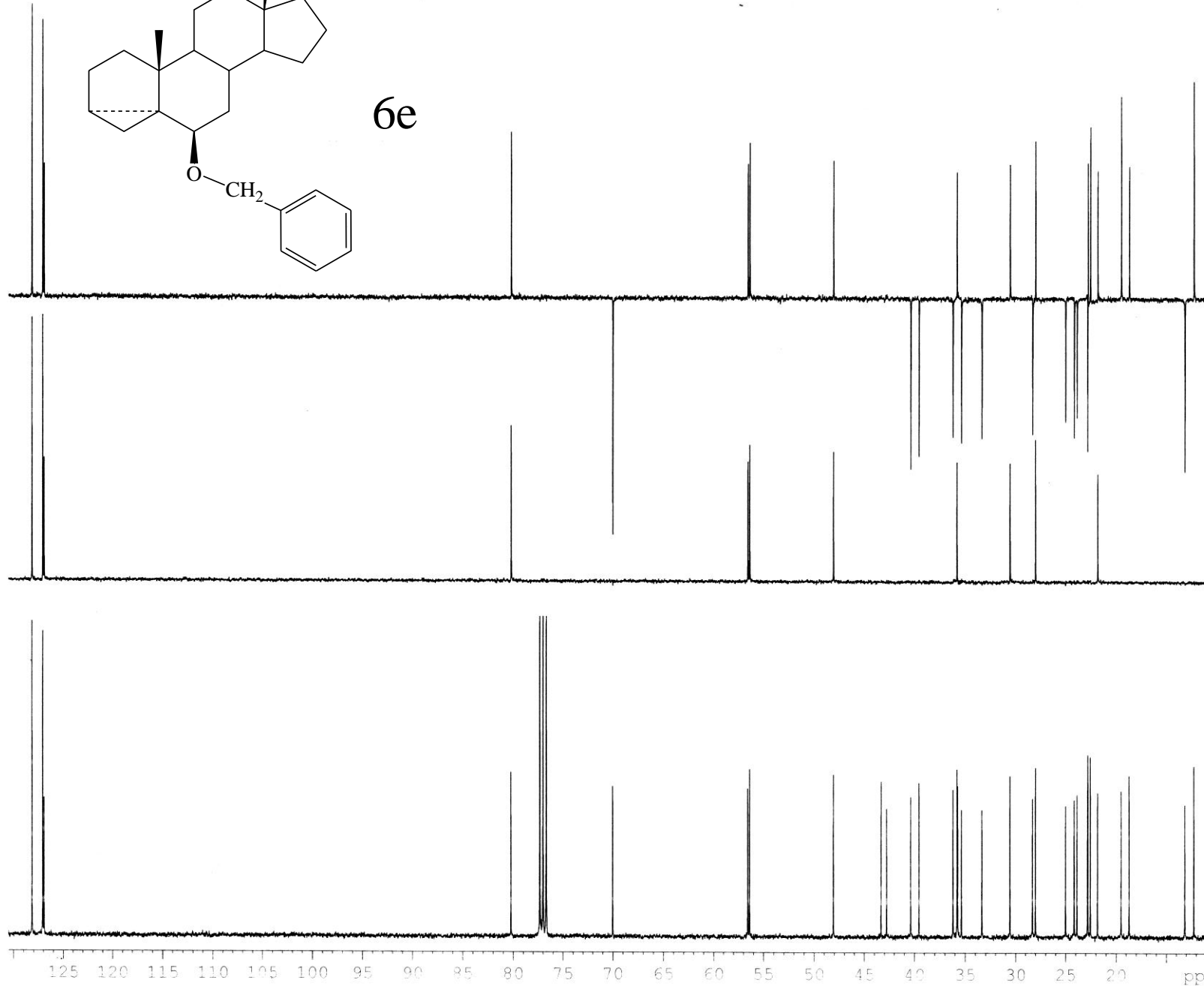
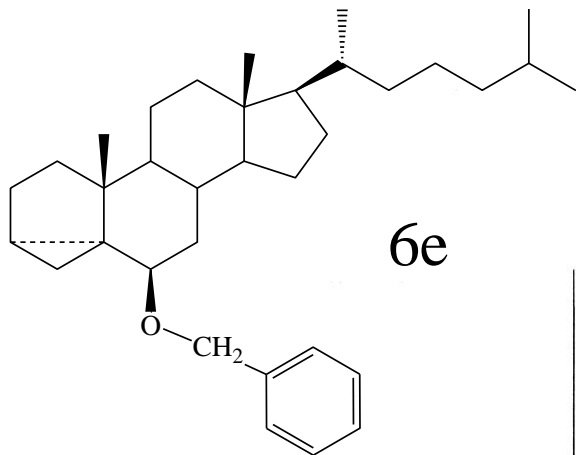
Current Data Parameters
 NAME EM Evel8
 EXPNO 2
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20130314
 Time_ 15.16
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 384
 DS 4
 SWH 27173.912 Hz
 FIDRES 0.414641 Hz
 AQ 1.2059124 sec
 RG 2050
 DW 18.400 usec
 DE 6.00 usec
 TE 303.0 K
 D1 2.0000000 sec
 d11 0.0300000 sec
 DELTA 1.8999999 sec
 TD0 1

===== CHANNEL f1 =====
 NUC1 13C
 P1 12.25 usec
 PL1 -1.00 dB
 SFO1 100.6288660 MHz

===== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 100.00 usec
 PL2 -3.00 dB
 PL12 13.65 dB
 PL13 18.00 dB
 SFO2 400.1516006 MHz

F2 - Processing parameters
 SI 32768
 SF 100.6177986 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 EC 1.40



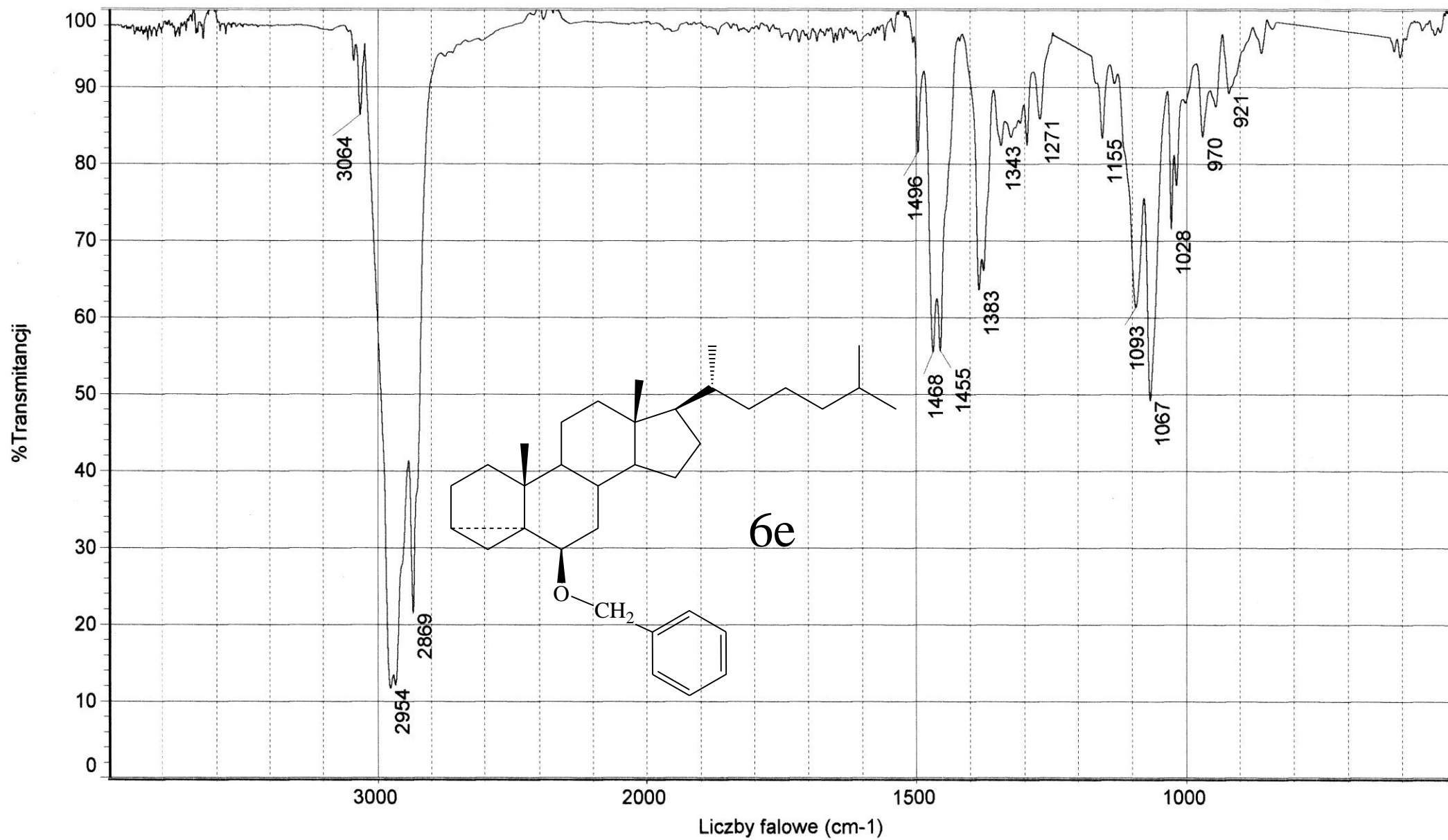
Current Data Parameters
 NAME EM Evel8
 EXPNO 4
 PROCNO 1

F2 - Acquisition Parameters
 Date 20130314
 Time 15.53
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG dept135
 TD 65536
 SOLVENT CDCl3
 NS 192
 DS 4
 SWH 24038.461 Hz
 FIDRES 0.366798 Hz
 AQ 1.3631988 sec
 RG 2050
 DW 20.800 usec
 DE 6.00 usec
 TE 303.0 K
 CNST2 145.000000
 D1 2.00000000 sec
 d2 0.00344828 sec
 d12 0.00002000 sec
 DELTA 0.00001560 sec
 TD0 1

==== CHANNEL f1 =====
 NUC1 13C
 P1 12.25 usec
 p2 24.50 usec
 PL1 -1.00 dB
 SFO1 100.6278593 MHz

==== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
 P3 14.70 usec
 p4 29.40 usec
 PCPD2 100.00 usec
 PL2 -3.00 dB
 PL12 13.65 dB
 SFO2 400.1516006 MHz

F2 - Processing parameters
 SI 32768
 SF 100.6177985 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40



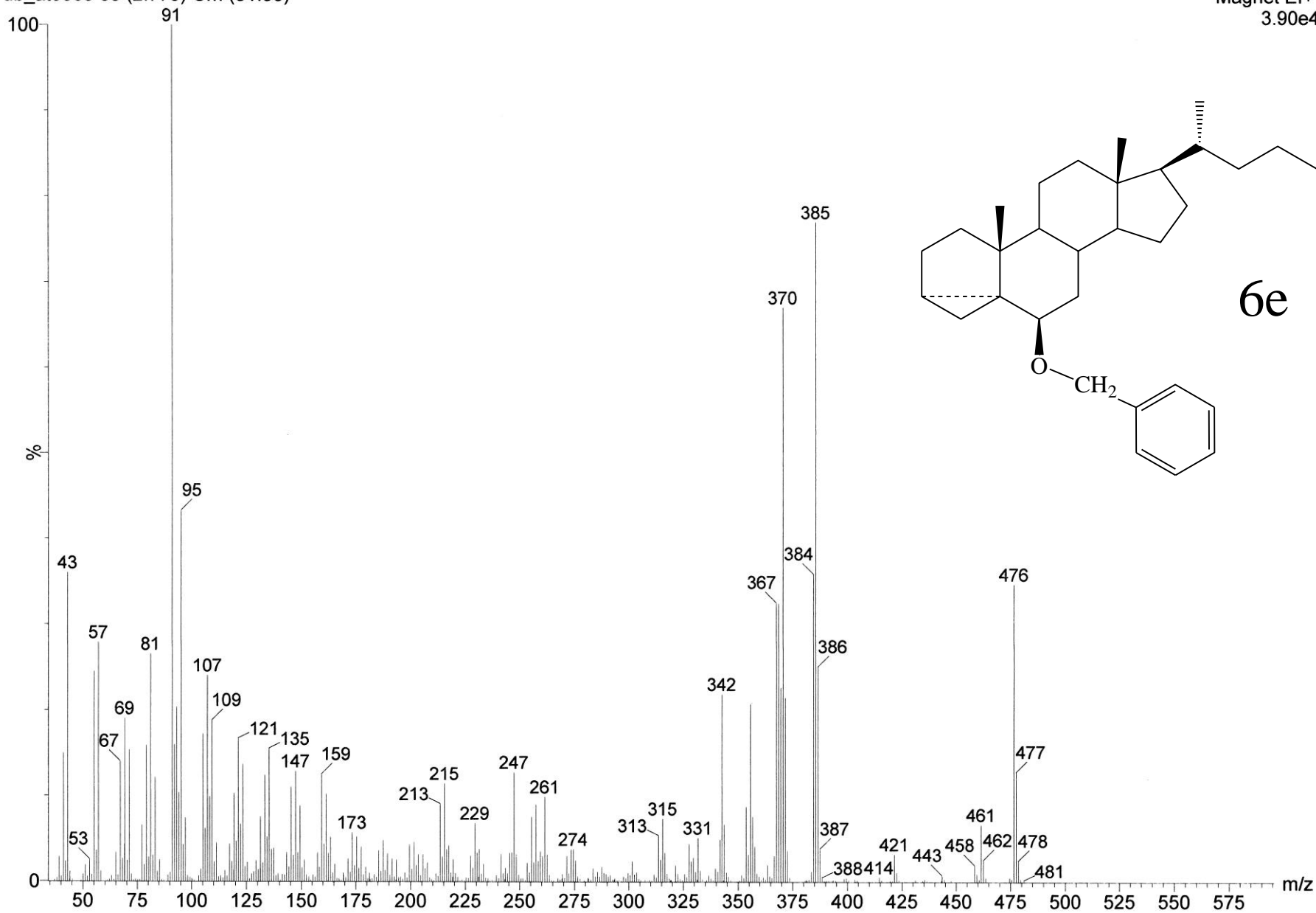
A. Tomkiel
Eve19

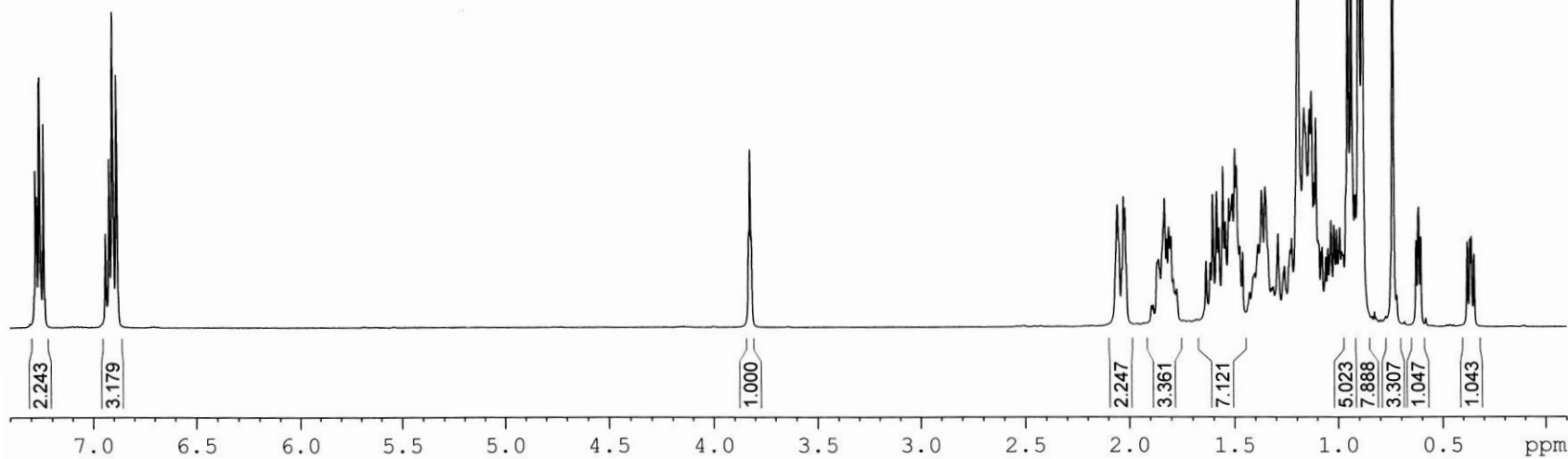
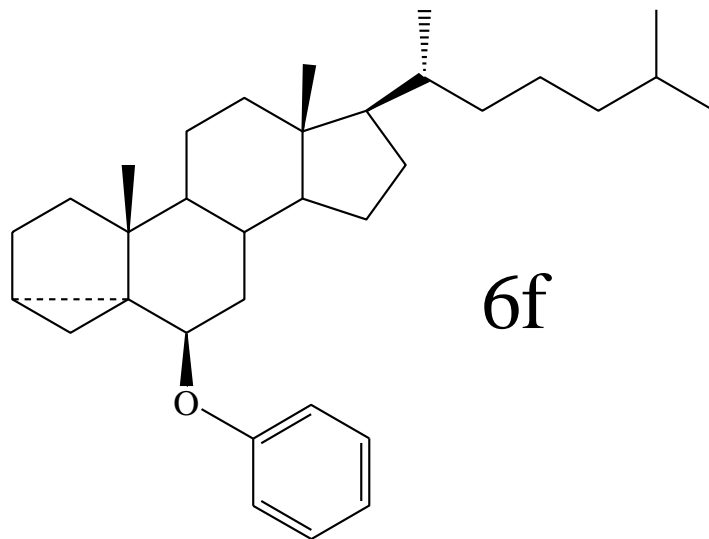
AUTOSPEC

19-Mar-2013 08:51:05

ub_at0865 35 (2.775) Cm (31:38)

Magnet EI+
3.90e4



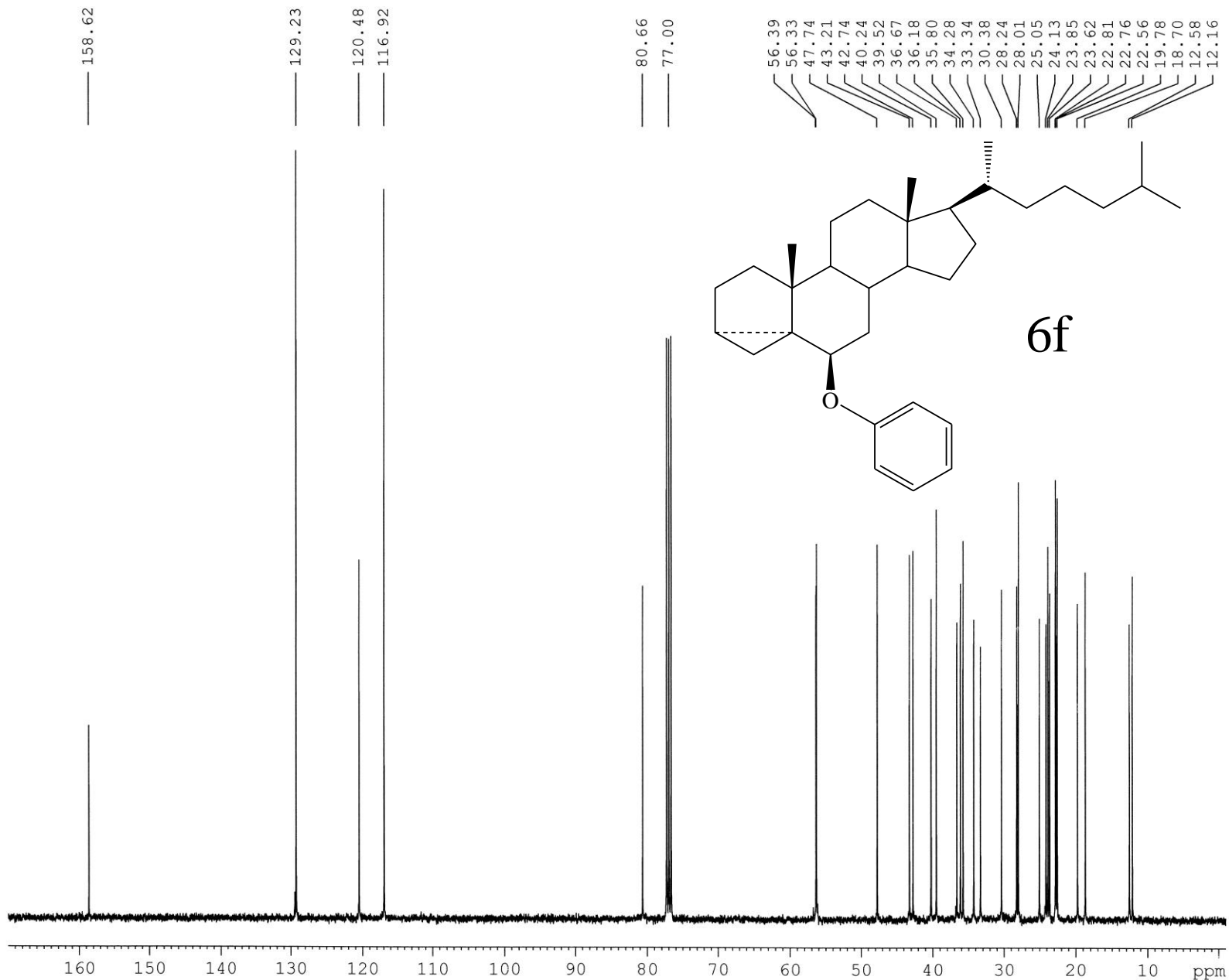


Current Data Parameters
 NAME ABd XI-1
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20110120
 Time_ 15.00
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zg30
 TD 65536
 SOLVENT CDC13
 NS 32
 DS 0
 SWH 8223.685 Hz
 FIDRES 0.125483 Hz
 AQ 3.9846387 sec
 RG 57
 DW 60.800 usec
 DE 8.00 usec
 TE 297.6 K
 D1 1.0000000 sec
 TD0 1

===== CHANNEL f1 =====
 NUC1 1H
 P1 11.15 usec
 PL1 -3.00 dB
 SFO1 400.1524711 MHz

F2 - Processing parameters
 SI 32768
 SF 400.1500000 MHz
 WDW GM
 SSB 0
 LB -0.20 Hz
 GB 0.2
 PC 1.00



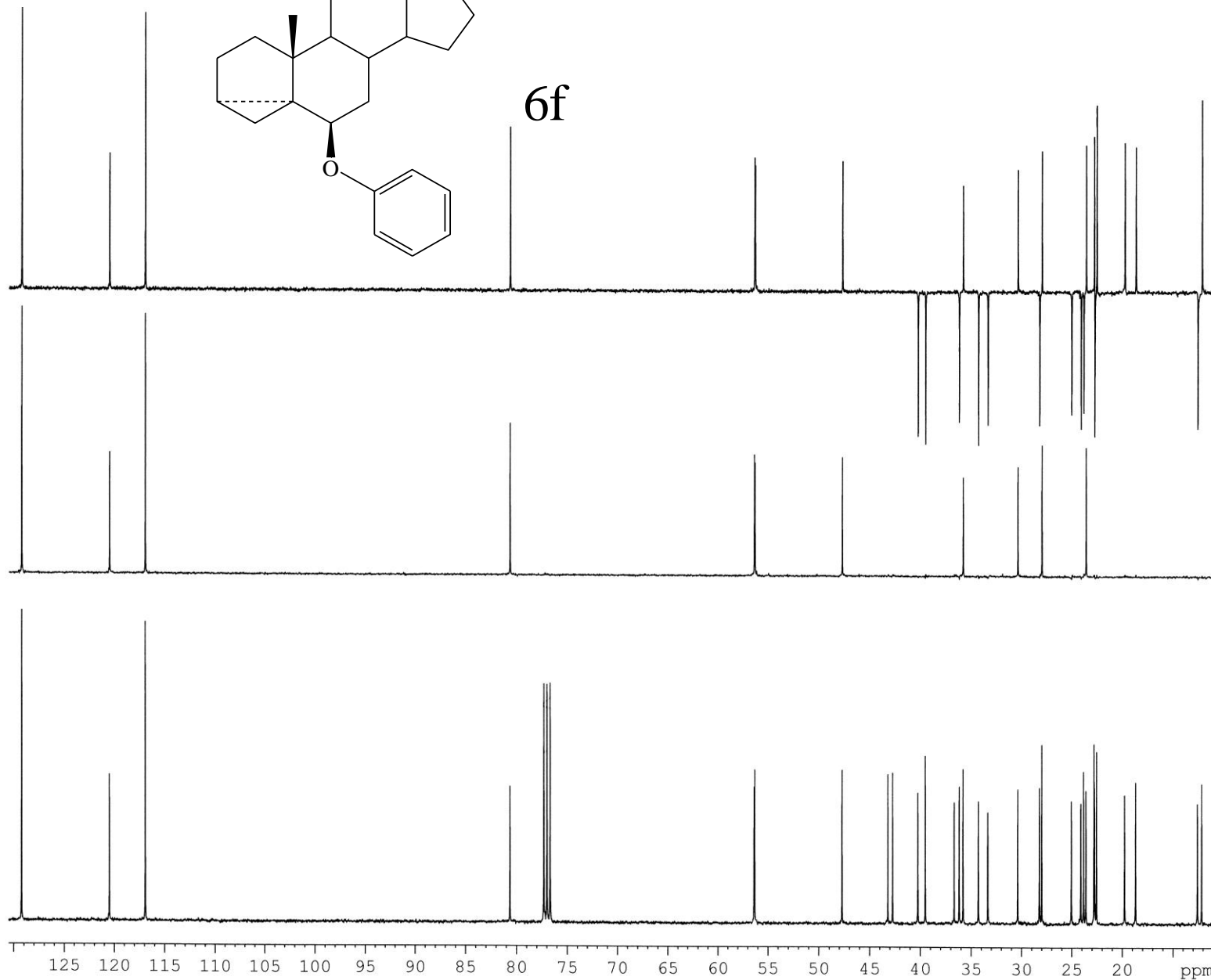
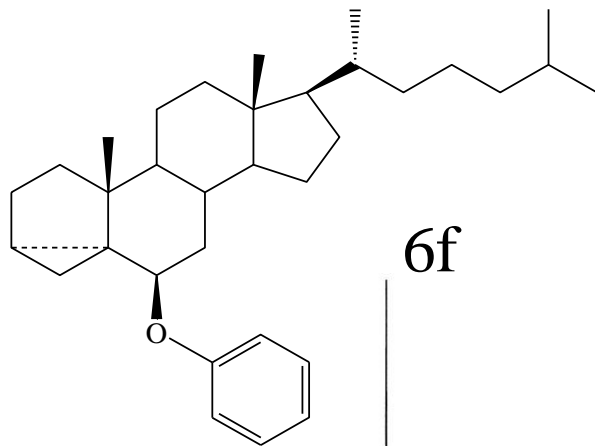
Current Data Parameters
 NAME ABd XI-1
 EXPNO 2
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20110329
 Time_ 12.56
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 576
 DS 4
 SWH 27173.912 Hz
 FIDRES 0.414641 Hz
 AQ 1.2059124 sec
 RG 1030
 DW 18.400 usec
 DE 6.00 usec
 TE 299.1 K
 D1 2.00000000 sec
 d11 0.03000000 sec
 DELTA 1.89999998 sec
 TD0 1

==== CHANNEL f1 =====
 NUC1 13C
 P1 11.00 usec
 PL1 -1.00 dB
 SFO1 100.6288660 MHz

==== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 100.00 usec
 PL2 -3.00 dB
 PL12 13.65 dB
 PL13 18.00 dB
 SFO2 400.1516006 MHz

F2 - Processing parameters
 SI 32768
 SF 100.6178008 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 0.10



```

Current Data Parameters
NAME      ABd XI-1
EXPNO     4
PROCNO    1

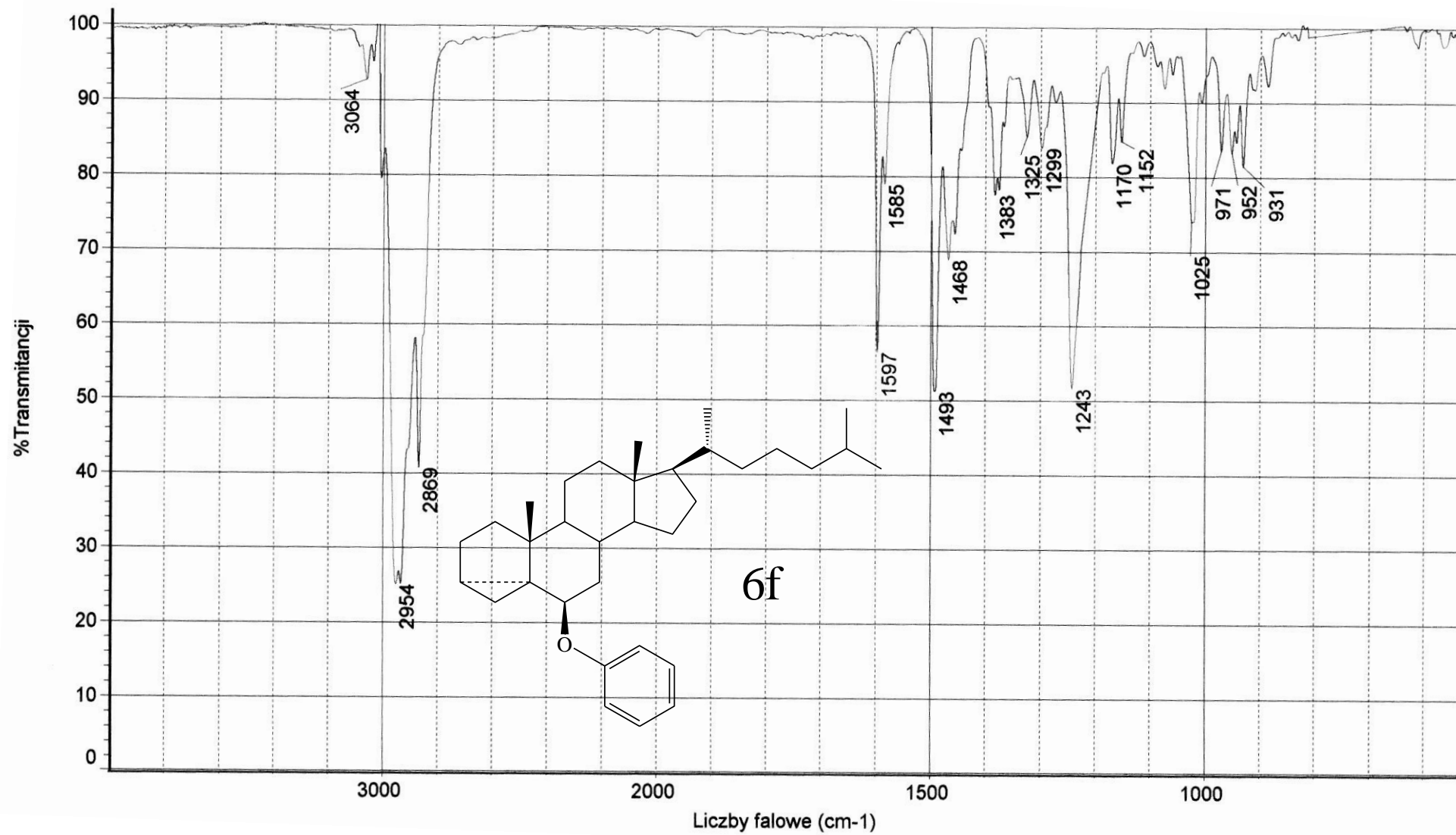
F2 - Acquisition Parameters
Date_     20110329
Time      13.41
INSTRUM   spect
PROBHD    5 mm PABBO BB-
PULPROG   dept135
TD         65536
SOLVENT   CDCl3
NS         192
DS         4
SWH        24038.461 Hz
FIDRES     0.366798 Hz
AQ         1.3631988 sec
RG         2050
DW         20.800 usec
DE         6.00 usec
TE         299.7 K
CNST2     145.0000000
D1         2.00000000 sec
d2         0.00344828 sec
d12        0.00002000 sec
DELTA     0.00001401 sec
TD0        1

===== CHANNEL f1 =====
NUC1       13C
P1         11.00 usec
p2         22.00 usec
PL1        -1.00 dB
SFO1       100.6278593 MHz

===== CHANNEL f2 =====
CPDPRG2    waltz16
NUC2       1H
P3         14.70 usec
p4         29.40 usec
PCPD2     100.00 usec
PL2        -3.00 dB
PL12       13.65 dB
SFO2       400.1516006 MHz

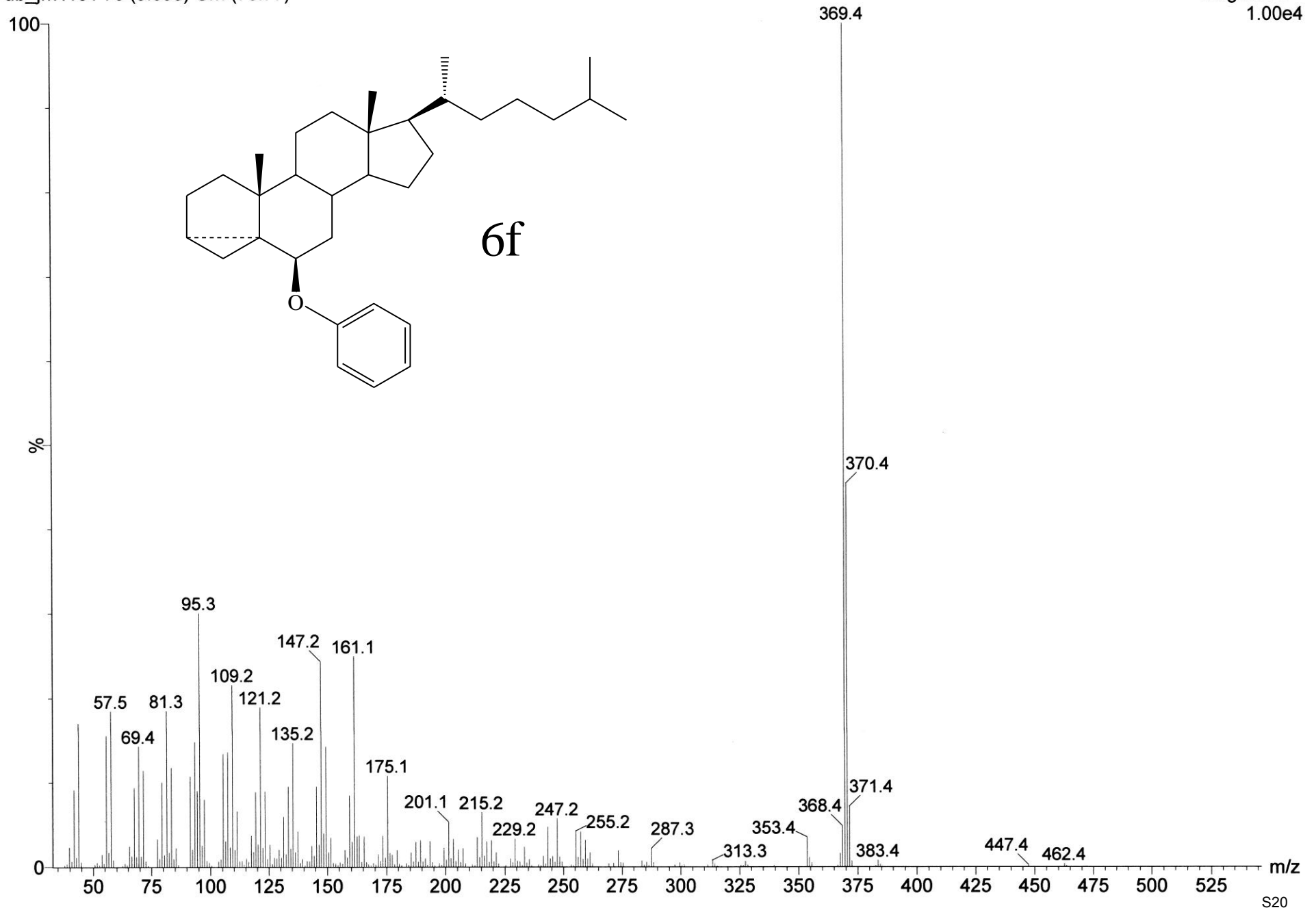
F2 - Processing parameters
SI         32768
SF         100.6178008 MHz
WDW        EM
SSB        0
LB         1.00 Hz
GB         0
PC         0.10

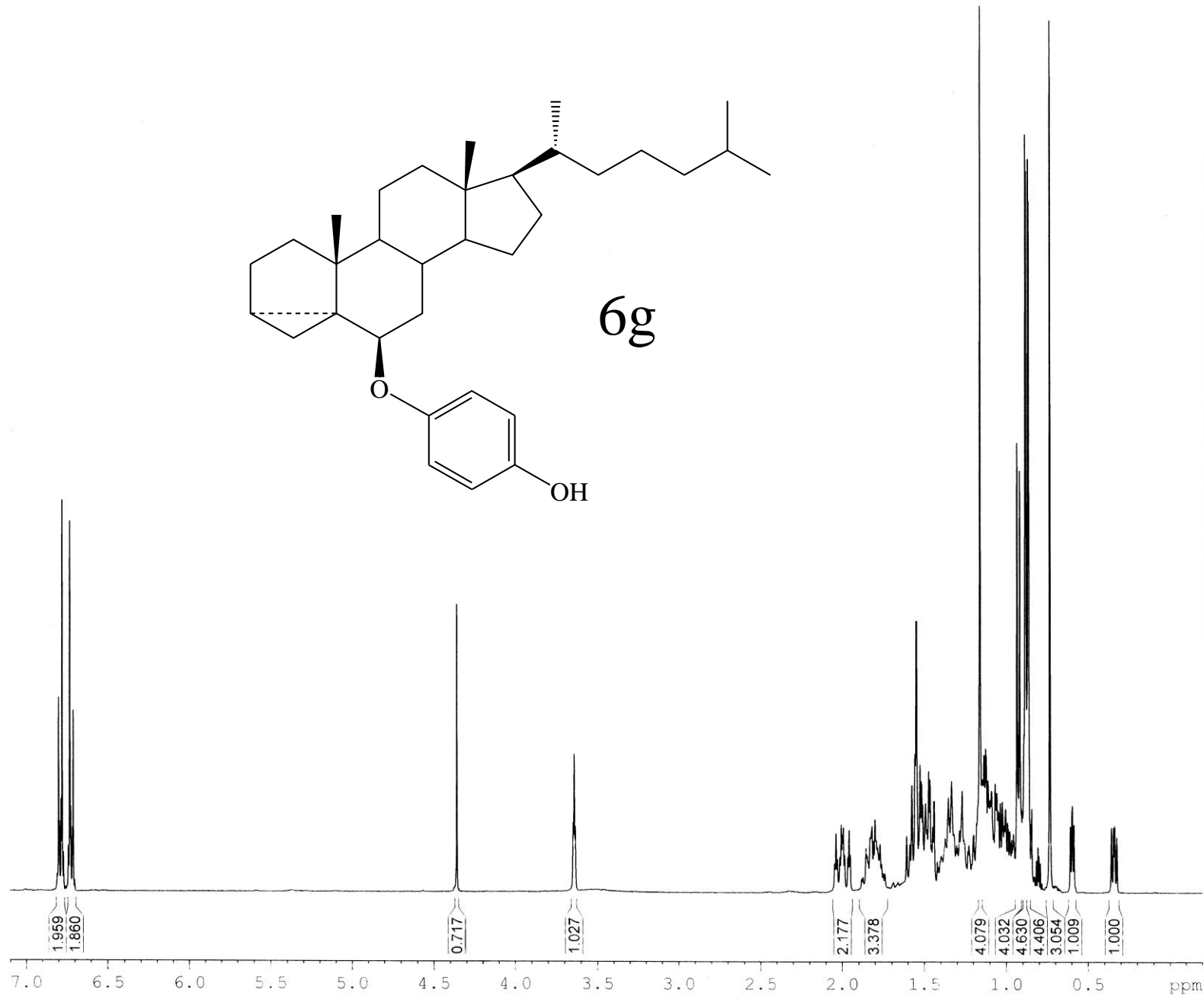
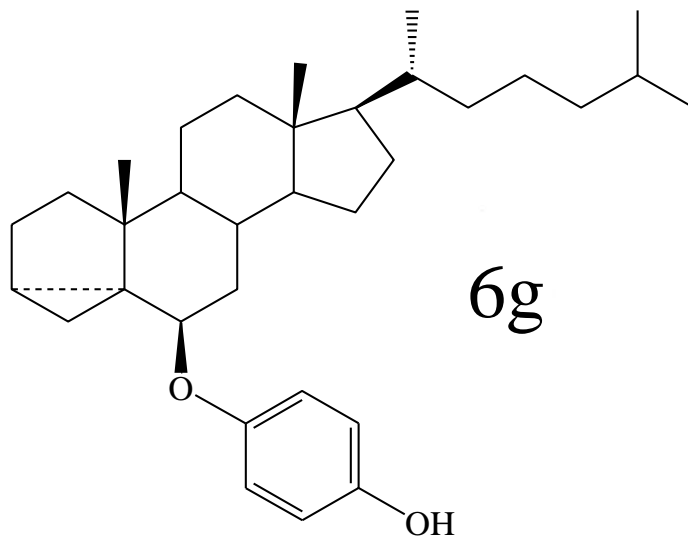
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eter fenylowy w6

ub_jm1101 70 (5.698) Cm (70:77)

Magnet EI+
1.00e4

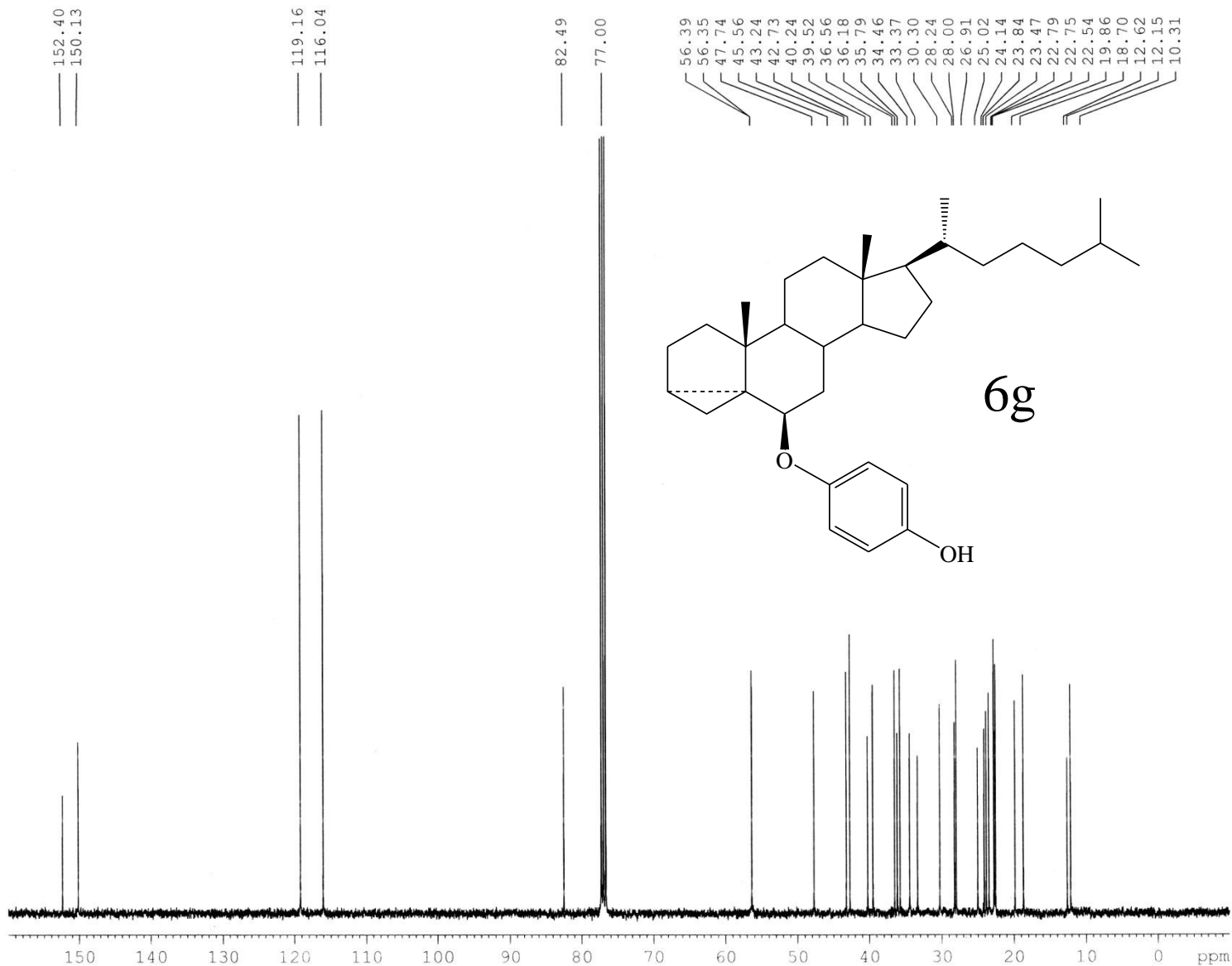


Current Data Parameters
 NAME EM 6-et.h.chinowoy cz.
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20130411
 Time 11.53
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3
 NS 192
 DS 0
 SWH 8223.685 Hz
 FIDRES 0.125483 Hz
 AQ 3.9846387 sec
 RG 287
 DW 60.800 usec
 DE 8.00 usec
 TE 673.2 K
 D1 1.00000000 sec
 TD0 1

===== CHANNEL f1 =====
 NUC1 1H
 P1 30.00 usec
 PL1 -3.00 dB
 SF01 400.1524711 MHz

F2 - Processing parameters
 SI 32768
 SF 400.1500000 MHz
 WDW GM
 SSB 0
 LB -0.20 Hz
 GB 0.2
 PC 1.00



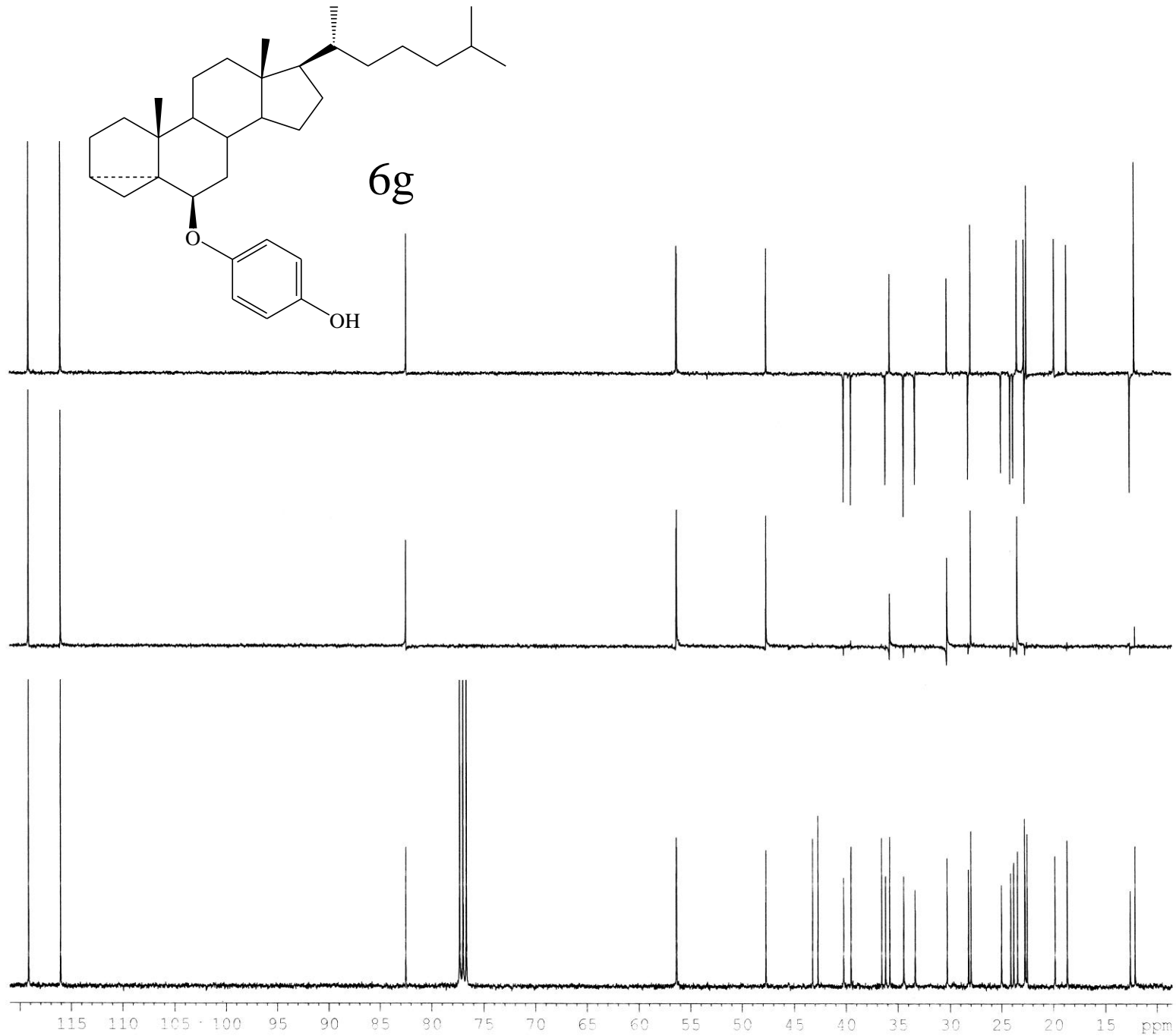
Current Data Parameters
 NAME EM Evel9czysty
 EXPNO 2
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20130315
 Time_ 13.38
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 800
 DS 4
 SWH 27173.912 Hz
 FIDRES 0.414641 Hz
 AQ 1.2059124 sec
 RG 2050
 DW 18.400 usec
 DE 6.00 usec
 TE 303.2 K
 D1 2.00000000 sec
 d11 0.03000000 sec
 DELTA 1.89999998 sec
 TD0 1

==== CHANNEL f1 =====
 NUC1 13C
 P1 12.25 usec
 PL1 -1.00 dB
 SFO1 100.6288660 MHz

==== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 100.00 usec
 PL2 -3.00 dB
 PL12 13.65 dB
 PL13 18.00 dB
 SFO2 400.1516006 MHz

F2 - Processing parameters
 SI 32768
 SF 100.6177986 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 0.10



6g

```

Current Data Parameters
NAME      EM Evel9czysty
EXPNO     4
PROCNO    1

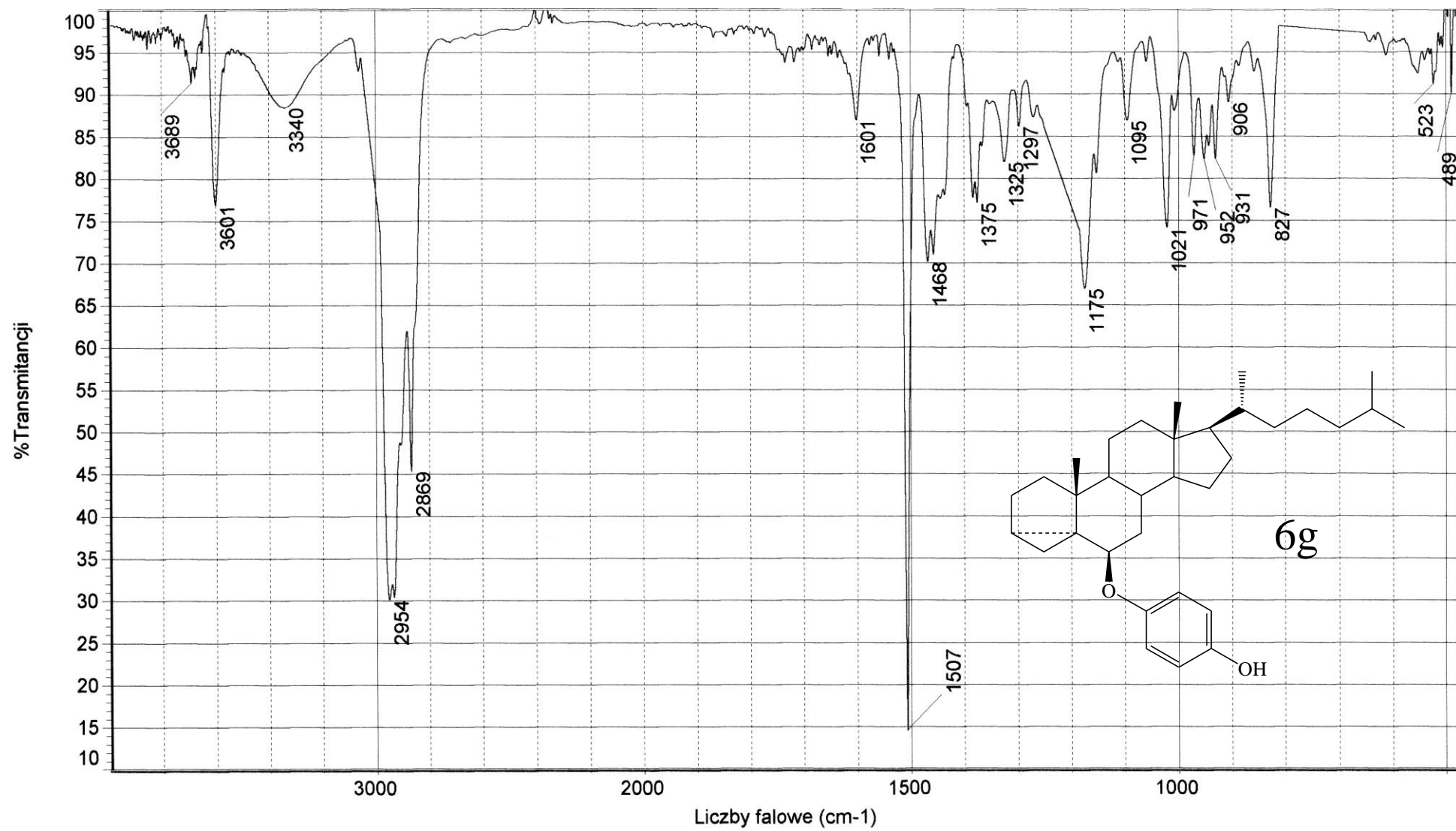
F2 - Acquisition Parameters
Date_     20130315
Time      15.06
INSTRUM   spect
PROBHD    5 mm PABBO BB-
PULPROG   dept135
TD         65536
SOLVENT   CDCl3
NS         320
DS         4
SWH        24038.461 Hz
FIDRES     0.366798 Hz
AQ         1.3631988 sec
RG         2050
DW         20.800 usec
DE         6.00 usec
TE         303.0 K
CNST2     145.0000000
D1         2.0000000 sec
d2         0.00344828 sec
dI2        0.00002000 sec
DELTA     0.00001560 sec
TD0        1

===== CHANNEL f1 =====
NUC1       13C
P1         12.25 usec
p2         24.50 usec
PL1        -1.00 dB
SFO1       100.6278593 MHz

===== CHANNEL f2 =====
CPDPRG2    waltz16
NUC2       1H
P3         14.70 usec
p4         29.40 usec
PCPD2      100.00 usec
PL2         -3.00 dB
PL12       13.65 dB
SFO2       400.1516006 MHz

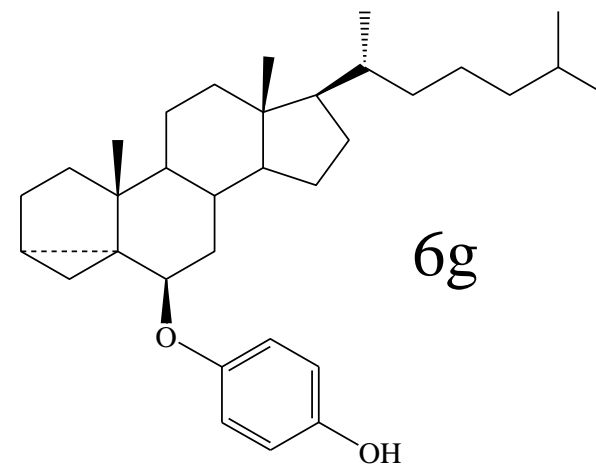
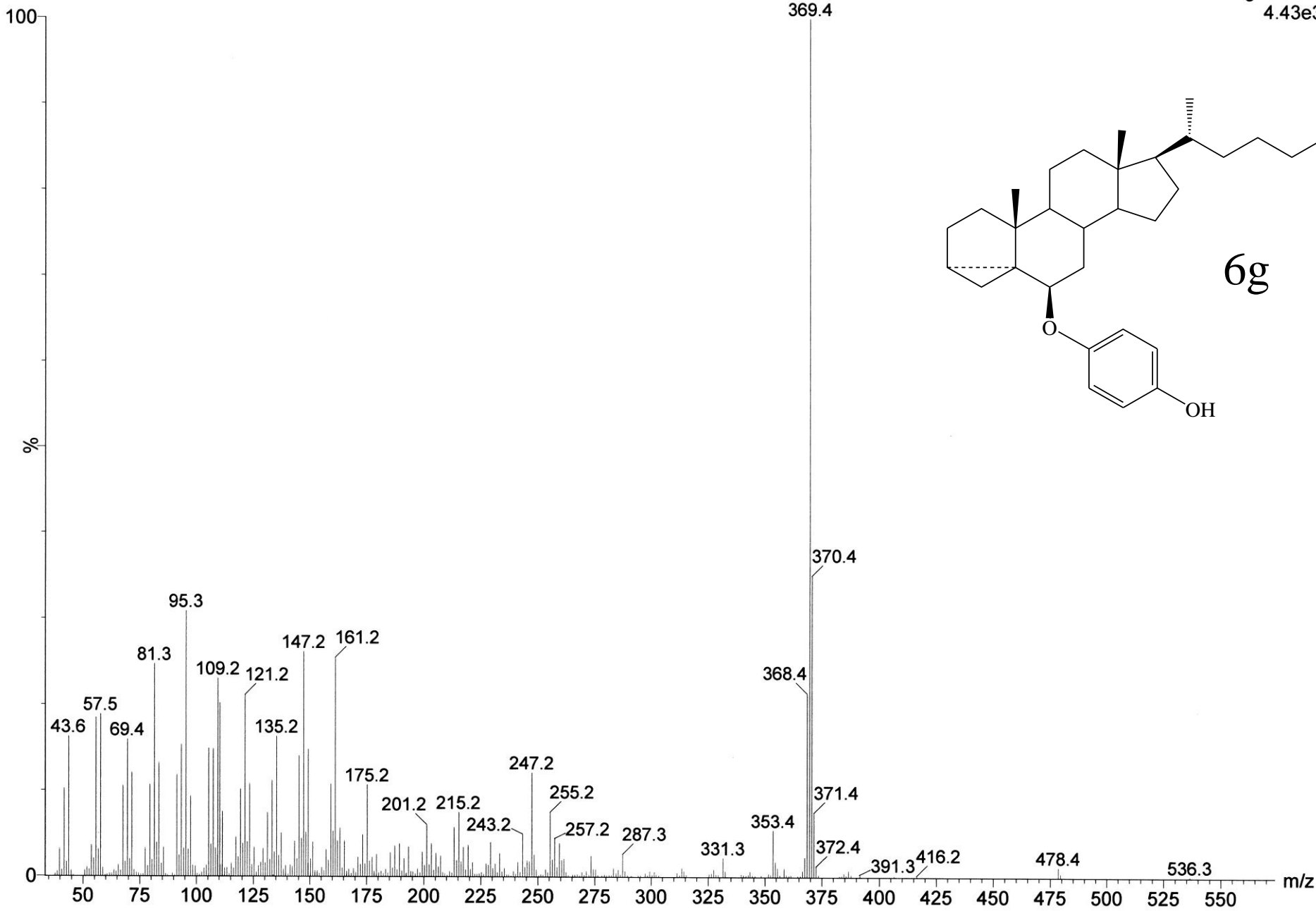
F2 - Processing parameters
SI         32768
SF         100.6177987 MHz
WDW        EM
SSB        0
LB         1.00 Hz
GB         0
PC         0.10

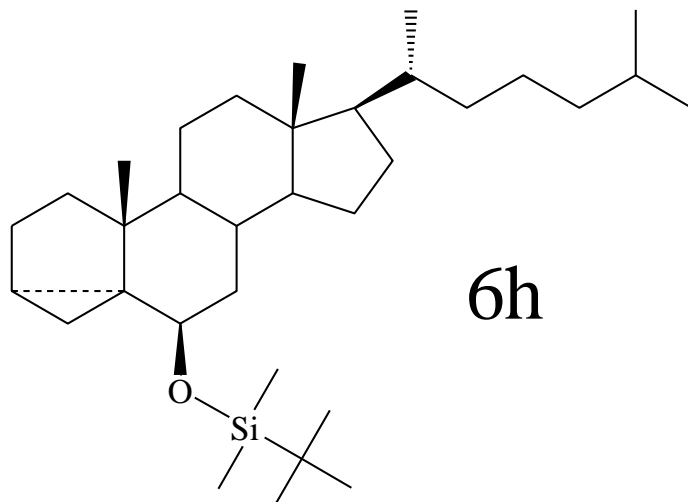
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eter chinonowy w6
ub_jm1102 19 (1.546) Cm (19:27-3:6)

Magnet EI+
4.43e3



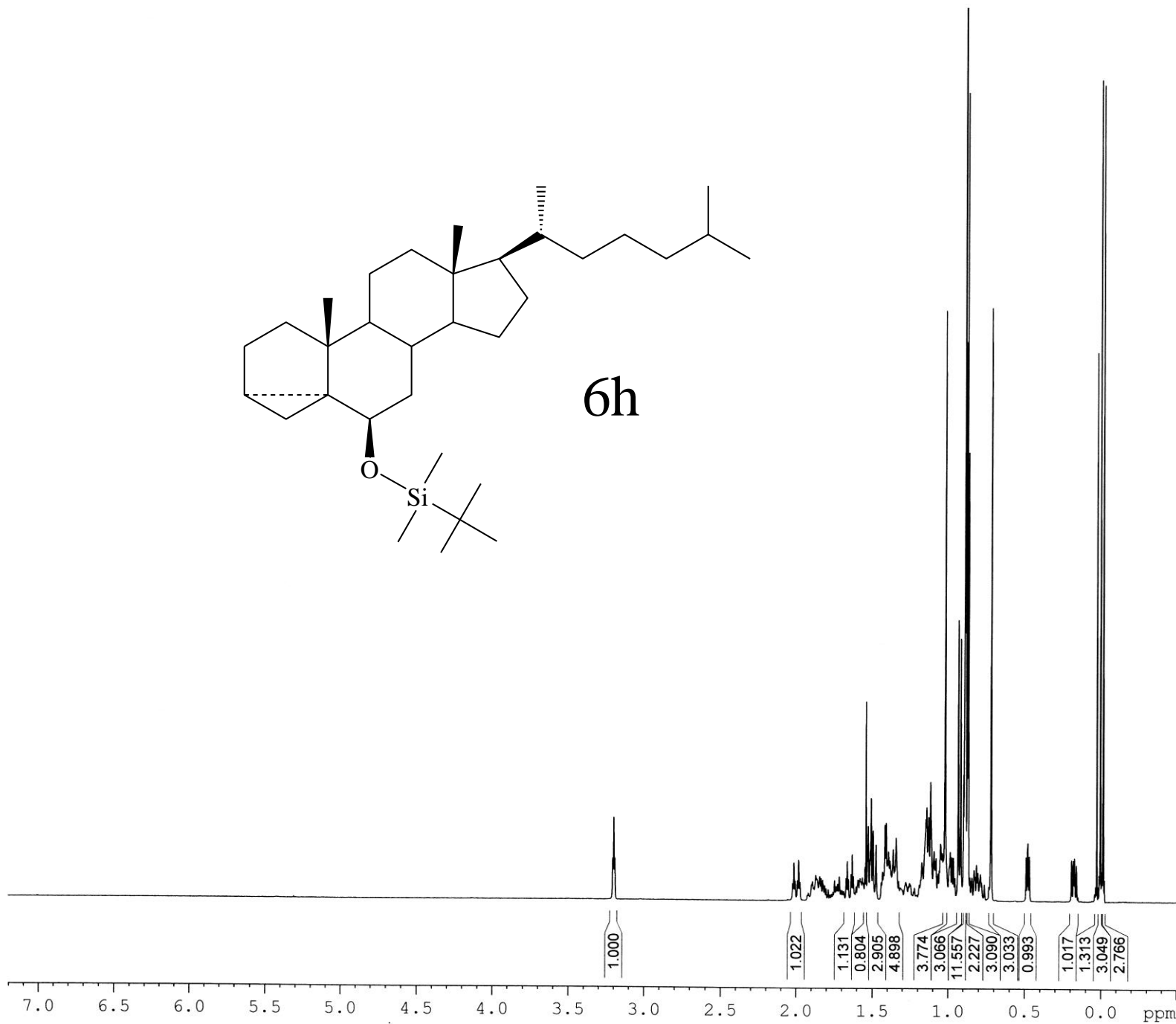


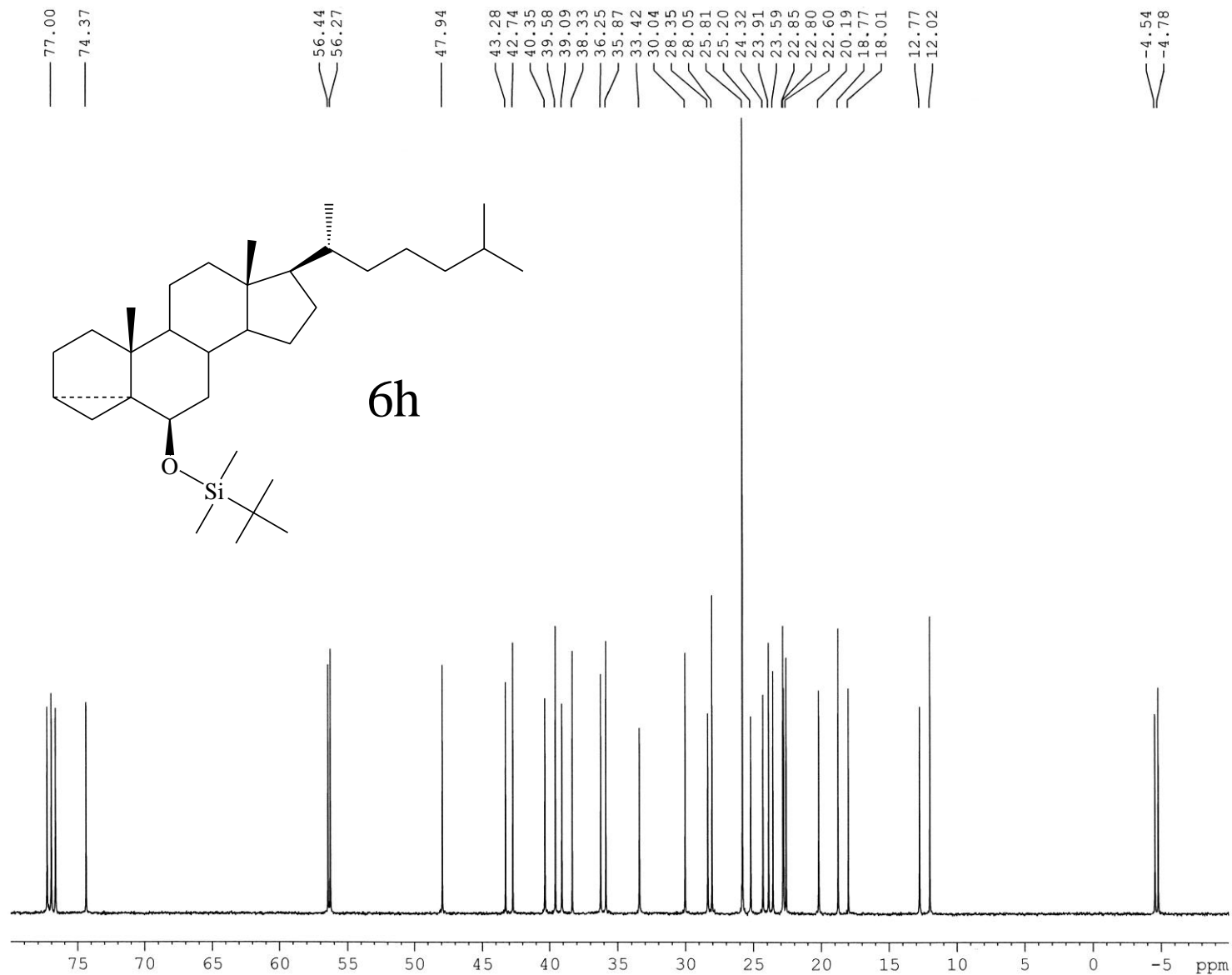
Current Data Parameters
 NAME ABd VIII
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20101220
 Time_ 11.19
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3
 NS 128
 DS 0
 SWH 8223.685 Hz
 FIDRES 0.125483 Hz
 AQ 3.9846387 sec
 RG 228
 DW 60.800 usec
 DE 8.00 usec
 TE 299.4 K
 D1 1.00000000 sec
 TD0 1

==== CHANNEL f1 =====
 NUC1 1H
 P1 11.15 usec
 PL1 -3.00 dB
 SFO1 400.1524711 MHz

F2 - Processing parameters
 SI 32768
 SF 400.1500000 MHz
 WDW GM
 SSB 0
 LB -0.20 Hz
 GB 0.2
 PC 1.00





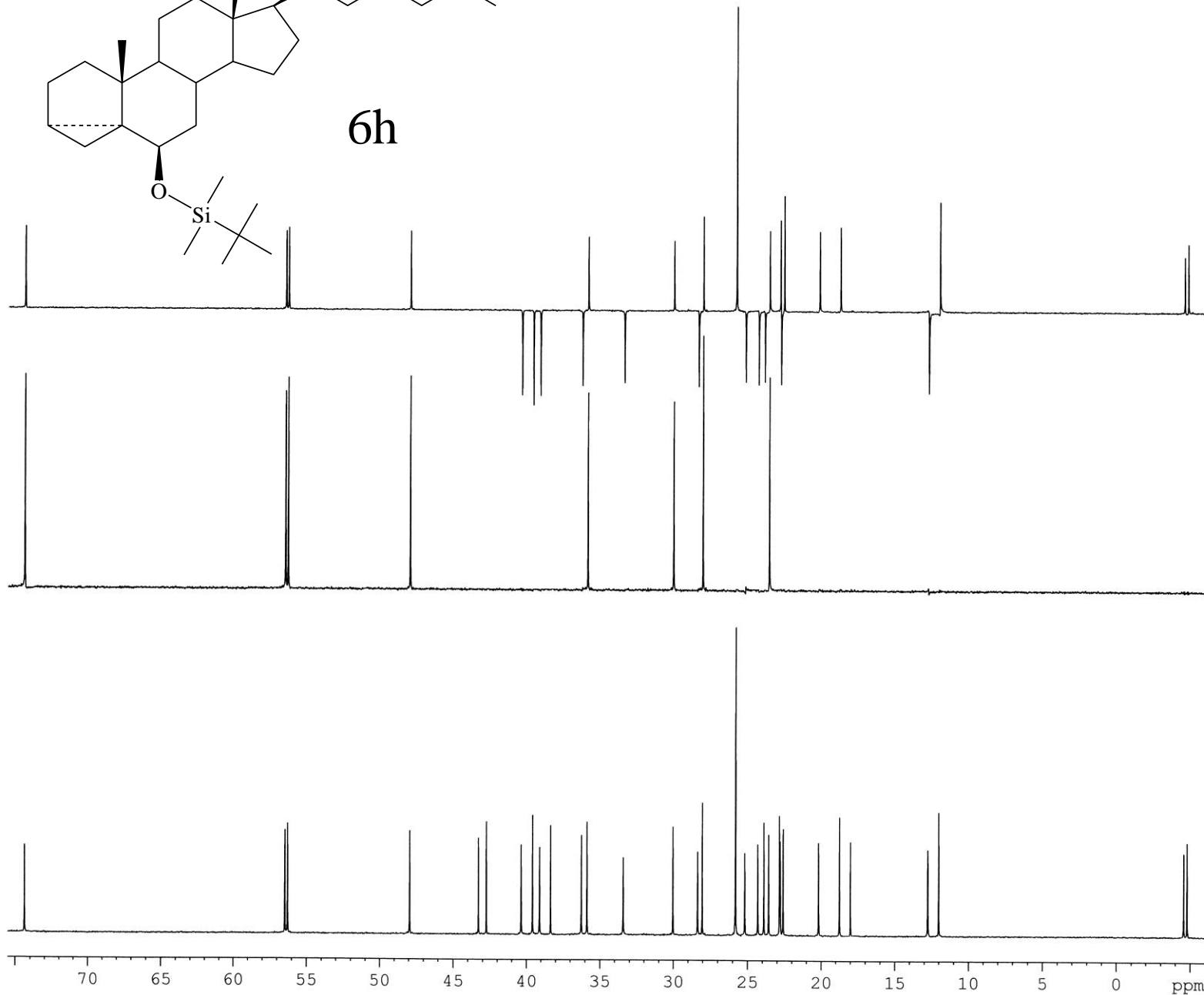
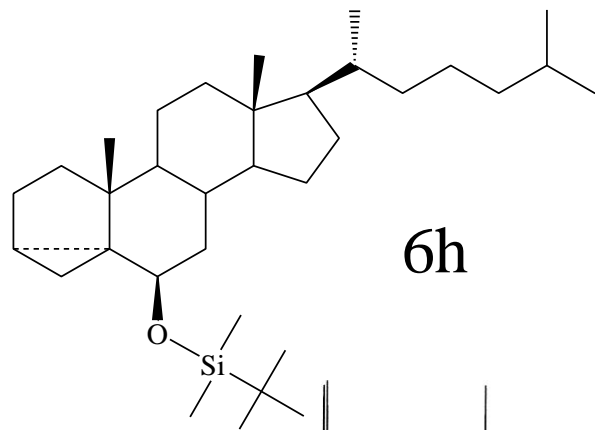
Current Data Parameters
 NAME Abd VIII
 EXPNO 2
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20110309
 Time_ 16.48
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 448
 DS 4
 SWH 27173.912 Hz
 FIDRES 0.414641 Hz
 AQ 1.2059124 sec
 RG 1030
 DW 18.400 usec
 DE 6.00 usec
 TE 299.9 K
 D1 2.00000000 sec
 d11 0.03000000 sec
 DELTA 1.89999998 sec
 TD0 1

===== CHANNEL f1 =====
 NUC1 13C
 P1 11.00 usec
 PL1 -1.00 dB
 SFO1 100.6288660 MHz

===== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 100.00 usec
 PL2 -3.00 dB
 PL12 13.65 dB
 PL13 18.00 dB
 SFO2 400.1516006 MHz

F2 - Processing parameters
 SI 32768
 SF 100.6177950 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40



```

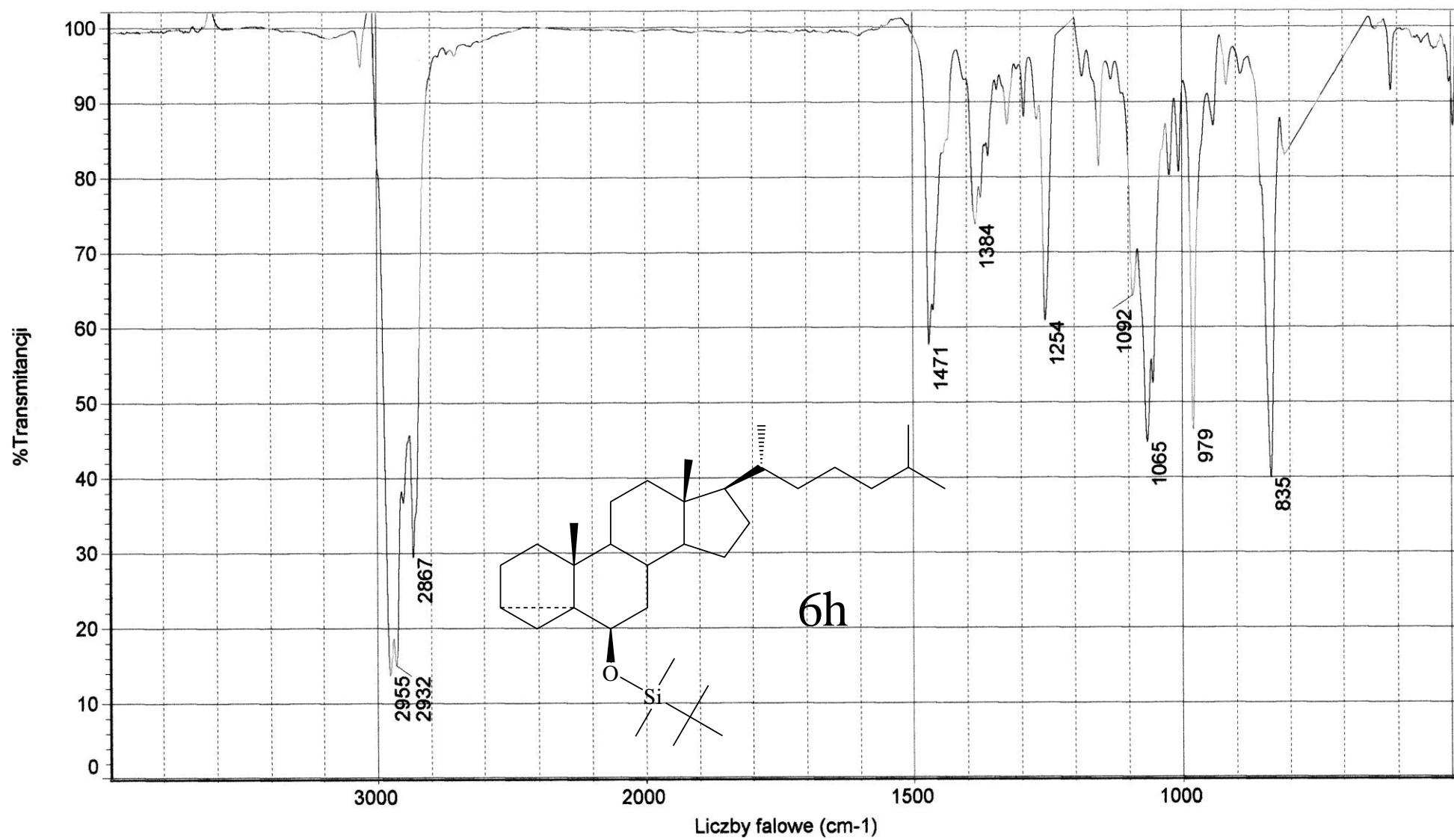
Current Data Parameters
NAME      Abd VIII
EXPNO    4
PROCNO   1

F2 - Acquisition Parameters
Date_    20110309
Time     17.25
INSTRUM  spect
PROBHD   5 mm PABBO BB-
PULPROG  dept135
TD       65536
SOLVENT  CDC13
NS       128
DS       4
SWH      24038.461 Hz
FIDRES   0.366798 Hz
AQ       1.3631988 sec
RG       1030
DW       20.800 usec
DE       6.00 usec
TE       299.8 K
CNST2    145.0000000
D1       2.00000000 sec
d2       0.00344828 sec
d12      0.00002000 sec
DELTA    0.00001401 sec
TD0      1

===== CHANNEL f1 =====
NUC1     13C
P1       11.00 usec
p2       22.00 usec
PL1      -1.00 dB
SFO1     100.6278593 MHz

===== CHANNEL f2 =====
CPDPRG2  waltz16
NUC2     1H
P3       14.70 usec
p4       29.40 usec
PCPD2    100.00 usec
PL2      -3.00 dB
PL12     13.65 dB
SFO2     400.1516006 MHz

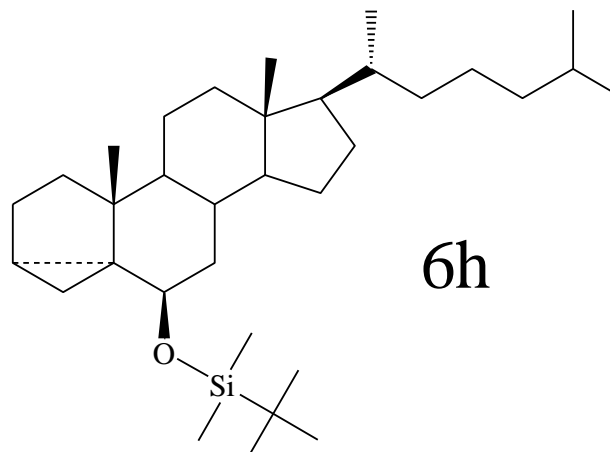
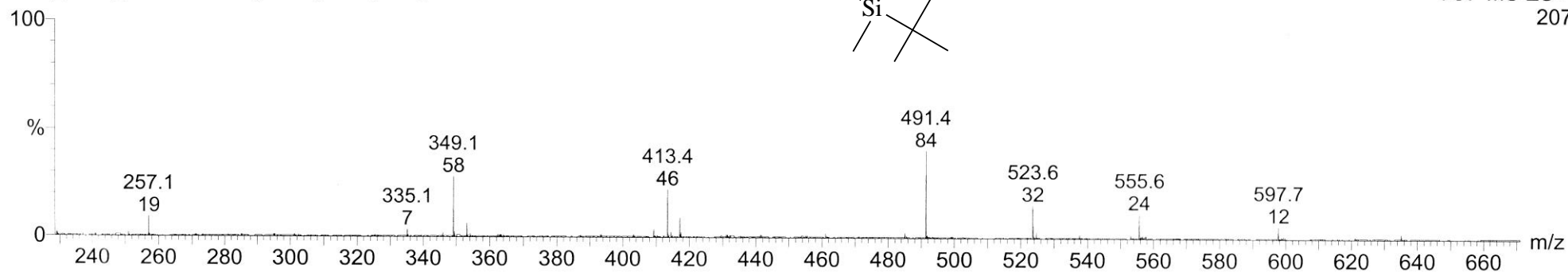
F2 - Processing parameters
SI       32768
SF       100.6177949 MHz
WDW      EM
SSB      0
LB       1.00 Hz
GB       0
PC       0.50
  
```



A. Bd-VIII +HCOONa

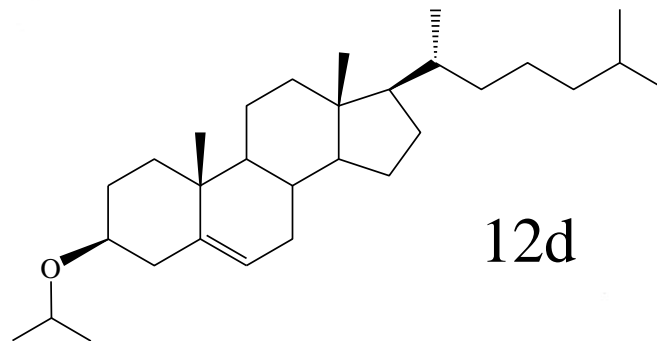
LCT

9742_JWM_HCOONa 74 (1.233) Cm (1:94)



18-Jan-2011

TOF MS ES+
207

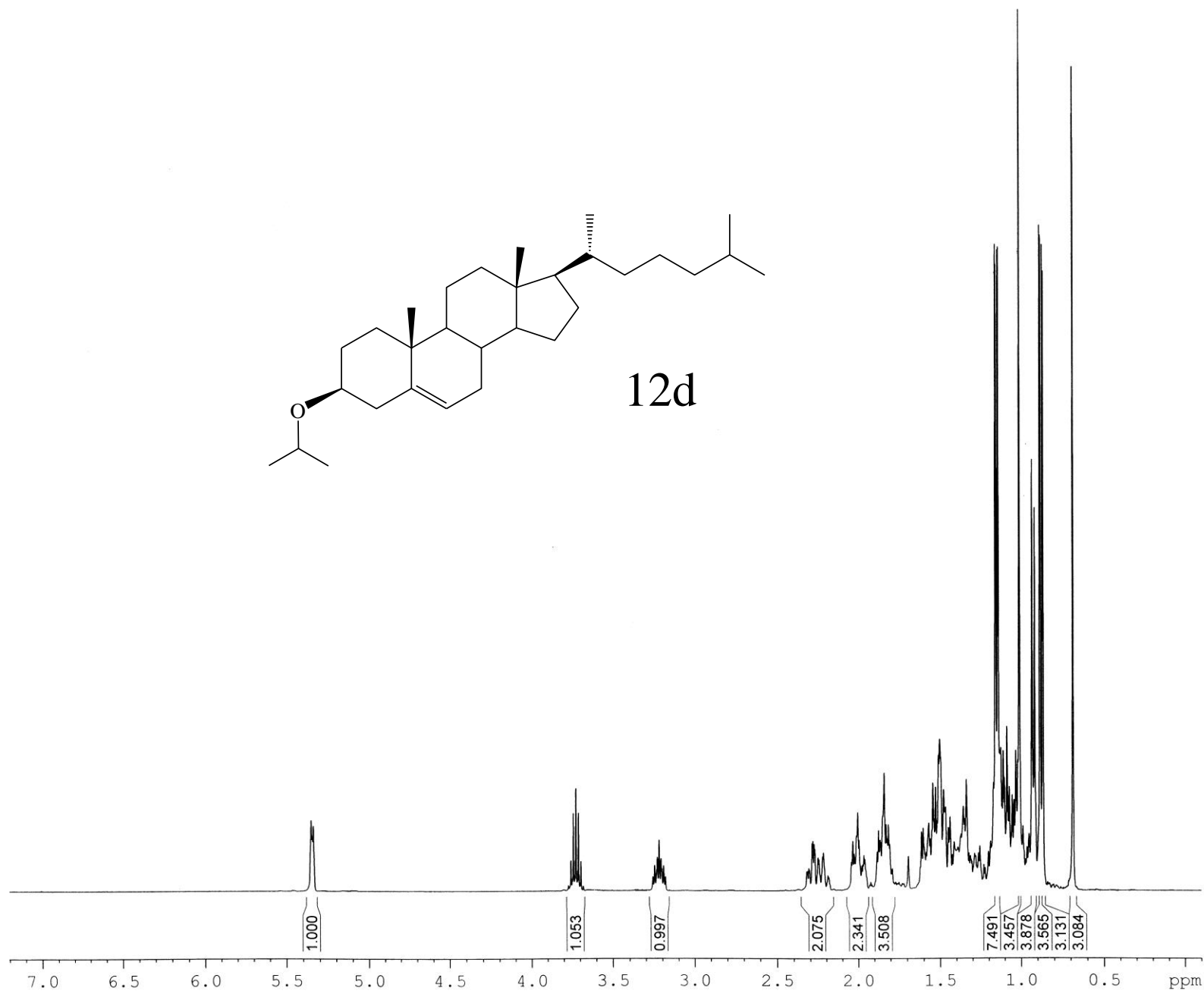


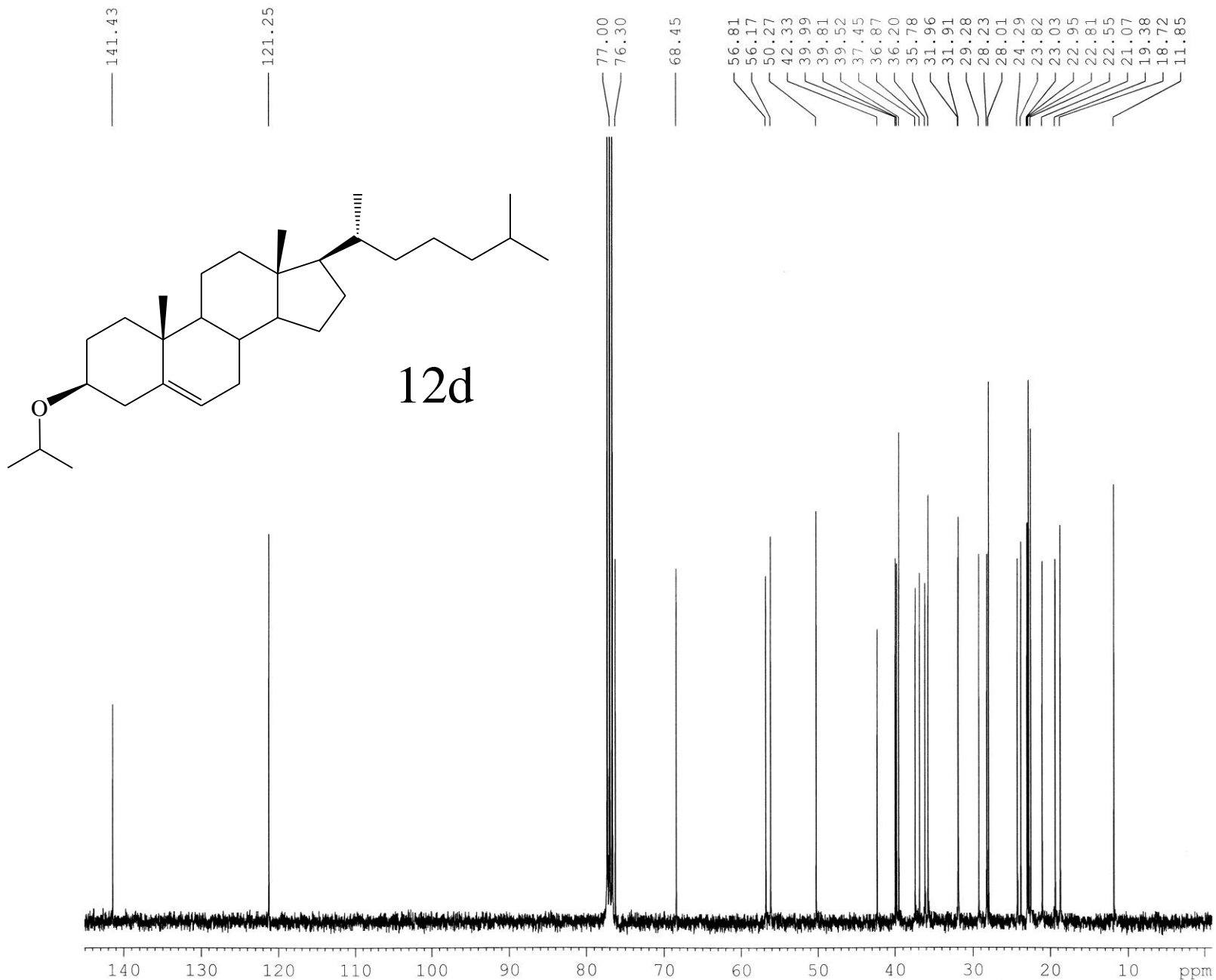
Current Data Parameters
 NAME AT eterizopropylowyw3
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20140425
 Time 16.10
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3
 NS 64
 DS 0
 SWH 8223.685 Hz
 FIDRES 0.125483 Hz
 AQ 3.9846387 sec
 RG 144
 DW 60.800 usec
 DE 8.00 usec
 TE 298.6 K
 D1 1.00000000 sec
 TDO 1

===== CHANNEL f1 =====
 NUC1 1H
 P1 11.15 usec
 PL1 -3.00 dB
 SFO1 400.1524711 MHz

F2 - Processing parameters
 SI 32768
 SF 400.1500000 MHz
 WDW GM
 SSB 0
 LB -0.20 Hz
 GB 0.2
 PC 1.00





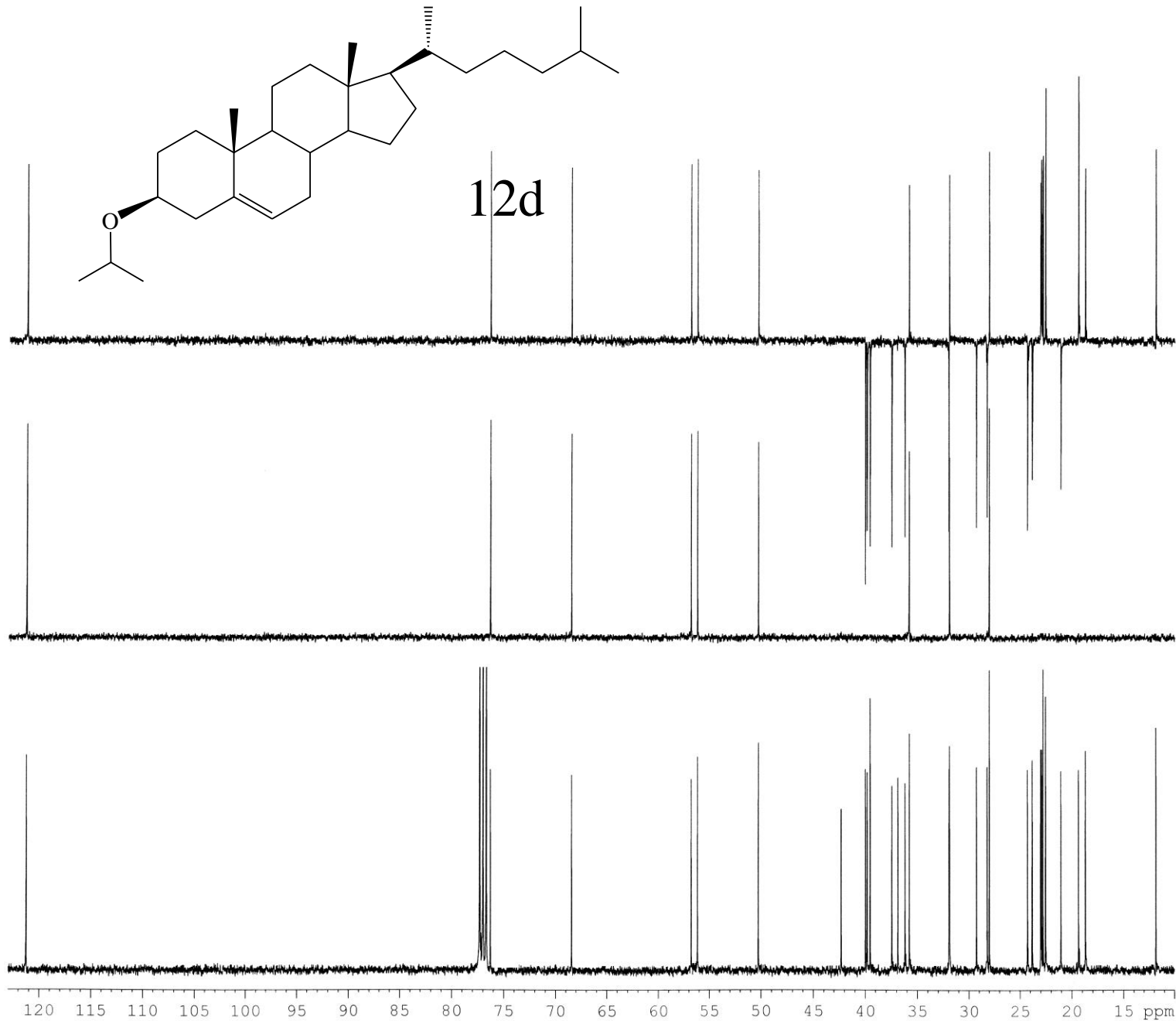
Current Data Parameters
 NAME AT eterizopropylowyw3
 EXPNO 2
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20140425
 Time_ 16.16
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zgpg30
 TD 65536
 SOLVENT CDC13
 NS 768
 DS 4
 SWH 27173.912 Hz
 FIDRES 0.414641 Hz
 AQ 1.2059124 sec
 RG 2050
 DW 18.400 usec
 DE 6.00 usec
 TE 299.1 K
 D1 2.00000000 sec
 d11 0.03000000 sec
 DELTA 1.89999998 sec
 TD0 1

==== CHANNEL f1 =====
 NUC1 13C
 P1 12.25 usec
 PL1 -1.00 dB
 SFO1 100.6288660 MHz

==== CHANNEL f2 =====
 CPDPRG2 waitz16
 NUC2 1H
 PCPD2 100.00 usec
 PL2 -3.00 dB
 PL12 13.65 dB
 PL13 18.00 dB
 SFO2 400.1516006 MHz

F2 - Processing parameters
 SI 32768
 SF 100.6177982 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40



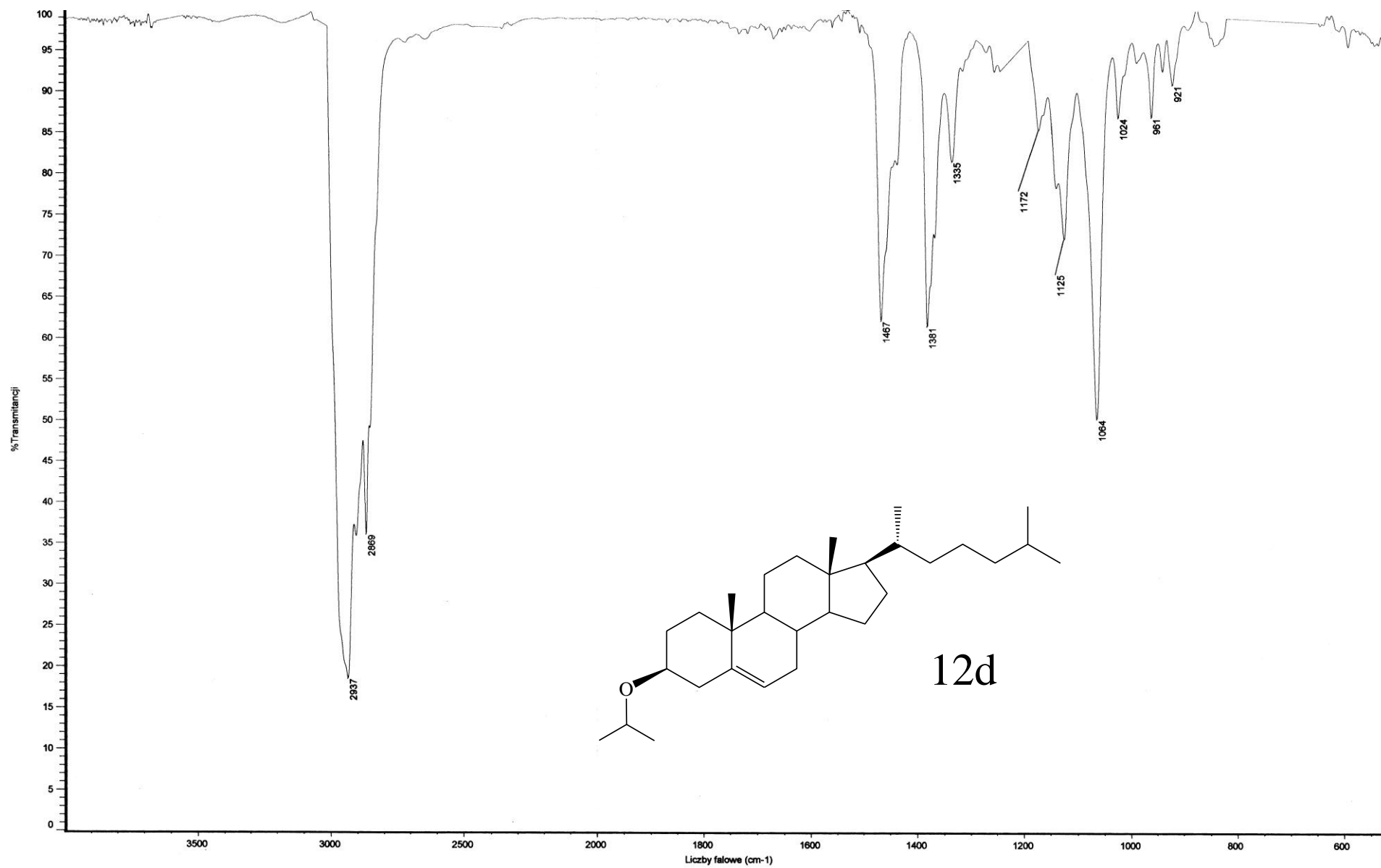
Current Data Parameters
 NAME AT eterizopropylowy3
 EXPNO 4
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20140425
 Time 17.28
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG dept135
 TD 65536
 SOLVENT CDCl3
 NS 320
 DS 4
 SWH 24038.461 Hz
 FIDRES 0.366798 Hz
 AQ 1.3631988 sec
 RG 2050
 DW 20.800 usec
 DE 6.00 usec
 TE 299.4 K
 CNST2 145.0000000
 D1 2.0000000 sec
 d2 0.00344828 sec
 d12 0.00002000 sec
 DELTA 0.00001560 sec
 TD0 1

===== CHANNEL f1 =====
 NUC1 13C
 P1 12.25 usec
 p2 24.50 usec
 PL1 -1.00 dB
 SFO1 100.6278593 MHz

===== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
 P3 14.70 usec
 p4 29.40 usec
 PCPD2 100.00 usec
 PL2 -3.00 dB
 PL12 13.65 dB
 SFO2 400.1516006 MHz

F2 - Processing parameters
 SI 32768
 SF 100.6177983 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40



J. Morzycki
eter izopropylowy w3
ub_jm1100 54 (4.396) Cm (54:72-5:8)

AUTOSPEC

14-Apr-2014 10:30:58
operator: Malgorzata Grela
Magnet EI+
5.01e3

