

1 **Supplementary methods**

2 **Cells.**

3 K562 cells (a human chronic myelogenous leukemia cell line) were cultured in Dulbecco's  
4 modified Eagle's medium (DMEM) supplemented with 10% fetal calf serum (FCS) and  
5 antibiotics.

6

7 ***In vitro* Ad gene expression analysis in K562 cells.**

8 K562 cells were seeded onto 12-well plates at  $1 \times 10^5$  cells/well. On the following day, cells  
9 were transduced with Ad vectors at an MOI of 100 and harvested 12 h after transduction. After  
10 total RNA isolation, mRNA levels of Ad genes were determined by real-time RT-PCR.

11

12

13 **Supplementary Figure S1**

14 **Suppression of the leaky expression of Ad genes in cultured cells.** K562 cells were  
15 transduced with Ad vectors at an MOI of 100 and harvested 12 h after transduction. The Ad  
16 gene expression levels were determined by real-time RT-PCR. The data are expressed as the  
17 mean values  $\pm$  S.D. (n=4). \*p<0.05 in comparison with Ad-L2.

18

19 **Supplementary Figure S2**

20 **The leaky expression of Ad genes in the mouse liver and spleen following intravenous**  
21 **administration of the Ad vectors.** (A) The Ad gene expression levels in the liver were  
22 determined 2 days following intravenous administration of Ad vectors. (B) The Ad gene  
23 expression levels in the liver and spleen were determined 2 days following intravenous  
24 administration of Ad-L2. The data are expressed as the mean values  $\pm$  S.D. (n=5-6).

25

26 **Supplementary Figure S3**

27 **Chemokine mRNA levels in the liver after administration of Ad-L2 and**  
28 **Ad-E4-122aT-L2.** C57BL/6 mice were treated with Ad vectors at  $1 \times 10^{10}$  IFU/mouse. Ten days  
29 after administration, chemokine mRNA levels in the liver were determined by real-time RT-PCR.  
30 The data are expressed as the mean values  $\pm$  S.D. (n=3-6).

**Supplementary Table S1 Oligonucleotide sequences.**

<b>Ad-E2A-122aT</b>	
E2A-3'-UTR-F	5'-accagcacactgggtt-3'
E2A-3'-UTR-R	5'-aaccagtgtgctggt-3'
E2A-miR-122aT-BstXI-S3	5'- <u>acaaacaccattgtcacactccacagcac</u> aaacaccattgtcacactccattaattaacagactagttgtg-3'
E2A-miR-122aT-BstXI-AS3	5'-actagtctgttaattaatgagtgtagacaatggtgtttgtgctgtggagtgtgacaatggtgtttgtcaca-3'
E2A-miR-122aT-BstXI-S4	5'- <u>acaaacaccattgtcacactccag</u> gacacaaacaccattgtcacactcca-3'
E2A-miR-122aT-BstXI-AS4	5'-ctagtgagtgtagacaatggtgtttgtgcctggagtgtgacaatggtgtttgat-3'
<b>Ad-E4-122aT</b>	
E4-3'-UTR-F1	5'-aatttcaagtcattttcattcagtagtatagccccaccaccacatagcttatacagatcaccgtaccttaatca aactaggtaccacctgccacc-3'
E4-3'-UTR-R1	5'-gggaggtggcaggtgttacctagtttgattaaggtacggtgatctgtataagctatgtggtggggctata ctactgaatgaaaaatgactga-3'
E4-3'-UTR-F2	5'-catgcccgcgtatcacagaaccctagtattcaacctgccacc-3'
E4-3'-UTR-R2	5'-gggaggtggcaggttgaataactagggttctgtgatagcggccgcaggttac-3'
E4-miR-122aT-S1	5'-ggcc <u>acaaacaccattgtcacactccacag</u> cacaaacaccattgtcacactccattaattaagcggtag-3'
E4-miR-122aT-AS1	5'-cgcttaattaatgagtgtagacaatggtgtttgtgctgtggagtgtgacaatggtgtttgt-3'
E4-miR-122aT-S2	5'- <u>acaaacaccattgtcacactccag</u> gacacaaacaccattgtcacactccagtag-3'
E4-miR-122aT-AS2	5'- <u>tggagtgtgacaatggtgtttgtgcctg</u> gagtgtgacaatggtgtttgat-3'
<b>Ad-pIX-122aT</b>	
pIX-miR-122aT-S1	5'-ctagctaaacaaacaccattgtcacactccacagcacaaacaccattgtcacactccagaattccagggtacc-3'
pIX-miR-122aT-AS1	5'-ctagggtaccctggaattctgagtgtagacaatggtgtttgtgctgtggagtgtgacaatggtgtttgttag-3'
pIX-miR-122aT-S2	5'-aattgacaaacaccattgtcacactccagacacaaacaccattgtcacactccaggtac-3'
pIX-miR-122aT-AS2	5'-ctggagtgtgacaatggtgtttgtgcctggagtgtgacaatggtgtttgtc-3'
<b>Ad-E2A-142-3pT</b>	
E2A-miR-142-3pT-BstXI-S3	5'- <u>tccataaagtag</u> gaaacactacacagctccataaagtaggaaacactacattaattaacagactagttgtg-3'
E2A-miR-142-3pT-BstXI-AS3	5'-actagtctgttaattaatgtagtgtttcctactttatggagctgtgtagtgtttcctactttatggacaca-3'
E2A-miR-142-3pT-BstXI-S4	5'- <u>tccataaagtag</u> gaaacactacagactccataaagtaggaaacactaca-3'
E2A-miR-142-3pT-BstXI-AS4	5'-ctagtgtagtgtttcctactttatggagctcctgtagtgtttcctactttatggaat-3'
<b>Ad-E4-142-3pT</b>	
E4-miR-142-3pT-S1	5'-ggcctccataaagtaggaaacactacacagctccataaagtaggaaacactacattaattaagcggtag-3'
E4-miR-142-3pT-AS1	5'-cgcttaattaatgtagtgtttcctactttatggagctgtgtagtgtttcctactttatgga-3'
E4-miR-142-3pT-S2	5'- <u>tccataaagtag</u> gaaacactacagactccataaagtaggaaacactacagtag-3'
E4-miR-142-3pT-AS2	5'-tgtagtgtttcctactttatggagctcctgtagtgtttcctactttatggaat-3'

<b>Ad-pIX-142-3pT</b>	
pIX-miR-142-3pT-S1	5'-ctagctaat <u>ccataaagtaggaaacactacacagctccataaagtaggaaacactacagaattccaggtacc</u> -3'
pIX-miR-142-3pT-AS1	5'-ctaggtaccctggaattctgtagtgttctactttatggagctgtagtgttctactttatggattag-3'
pIX-miR-142-3pT-S2	5'-aattgcataaagtaggaaacactacaggactccataaagtaggaaacactacaggtac-3'
pIX-miR-142-3pT--AS2	5'-ctgtagtgttctactttatggagtctgtagtgttctactttatggac-3'

Underlines indicate miRNA-targeted sequences.

**Supplementary Table S2 Sequences of primers for IFN- $\gamma$  and chemokines.**

<b>Gene</b>	<b>Forward primer</b>	<b>Reverse primer</b>	<b>Amplicon size</b>
IFN- $\gamma$	5'-ATG AAC GCT ACA CAC TGC ATC-3'	5'-TCT AGG CTT TCA ATG ACT GTG C-3'	92 bp
CCL2	5'-TTA AAA ACC TGG ATC GGA ACC AA-3'	5'-GCA TTA GCT TCA GAT TTA CGG GT-3'	121 bp
CCL3	5'-TTC TCT GTA CCA TGA CAC TCT GC-3'	5'-CGT GGA ATC TTC CGG CTG TAG-3'	100 bp
CCL4	5'-TTC CTG CTG TTT CTC TTA CAC CT-3'	5'-CTG TCT GCC TCT TTT GGT CAG-3'	121 bp
CCL5	5'-GCT GCT TTG CCT ACC TCT CC-3'	5'-TCG AGT GAC AAA CAC GAC TGC-3'	104 bp
CXCL2	5'-CGC TGT CAA TGC CTG AAG AC-3'	5'-ACA CTC AAG CTC TGG ATG TTC TTG-3'	62 bp
CX <sub>3</sub> CL	5'-ACG AAA TGC GAA ATC ATG TGC-3'	5'-CTG TGT CGT CTC CAG GAC AA-3'	120 bp
CXCL10	5'-CCA AGT GCT GCC GTC ATT TTC-3'	5'-TCC CTA TGG CCC TCA TTC TCA-3'	133 bp

# Figure S1

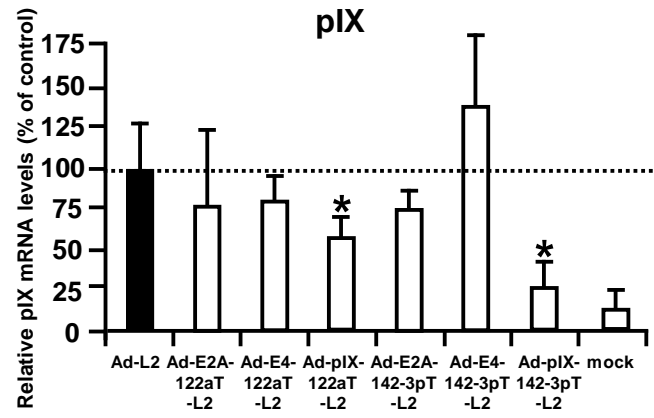
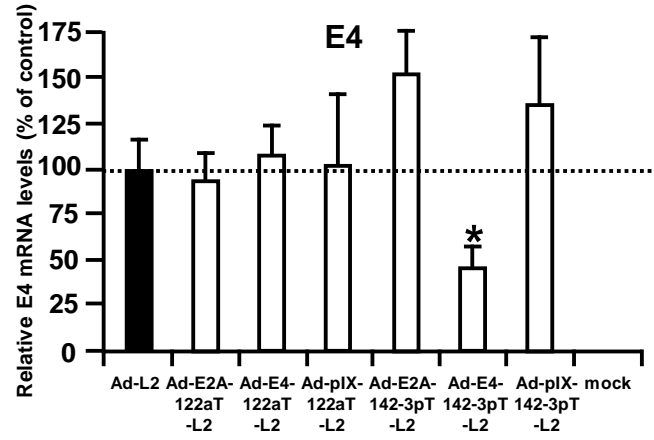
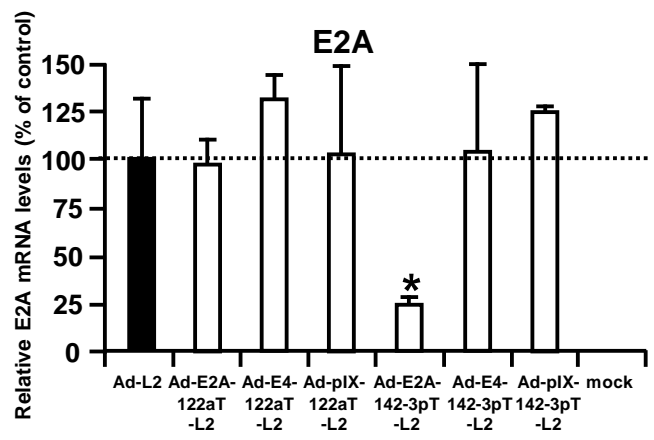


Figure S2

Figure S2A

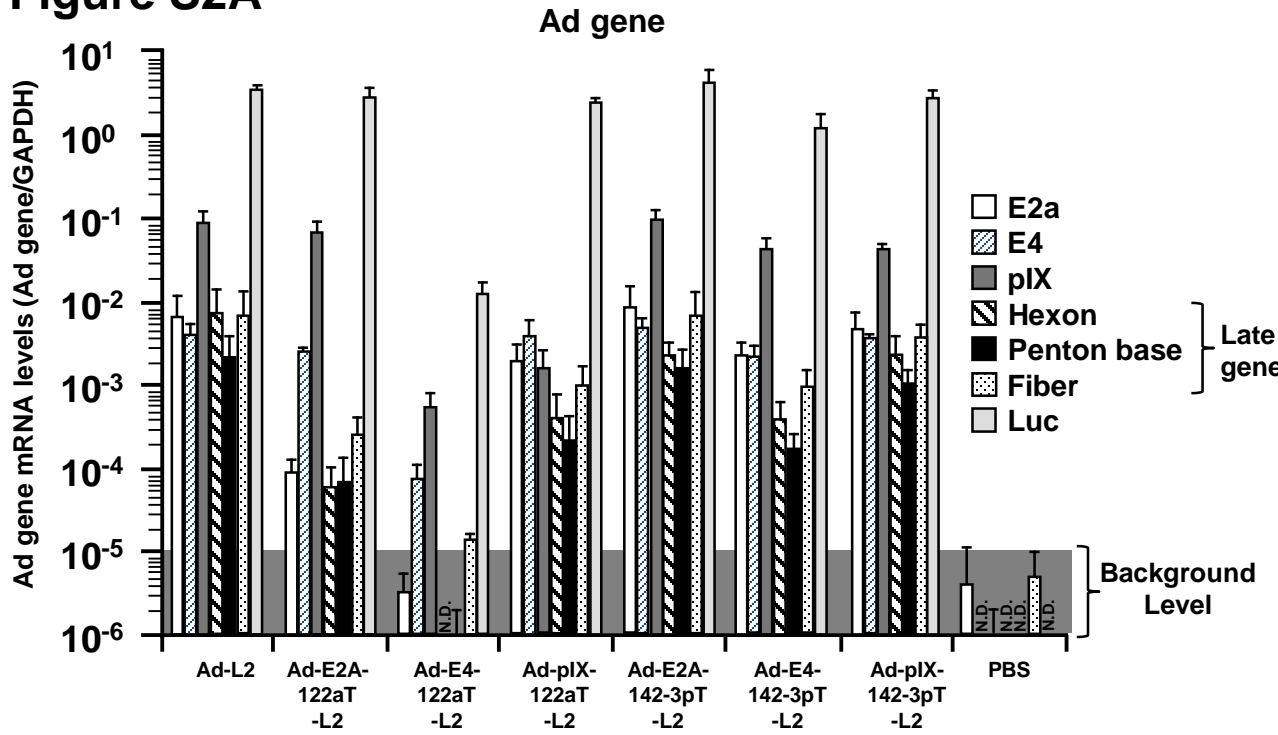


Figure S2B

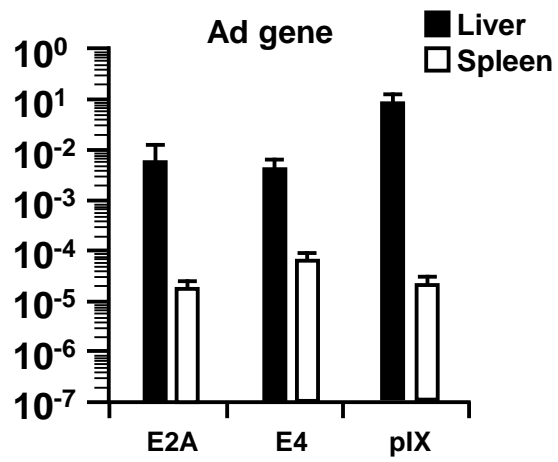


Figure S3

