pooled to knock-down SNTA or SNTB2, respectively. To knock-down both syntrophins, cells were transfected with the six different siRNAs. EndoPorter was used for transfection as recommended by the company. Cells were harvested 48 h after transfection.

Suppl. Table 1: Sequences of the siRNAs which were pooled to knock-down SNTA, SNTB2 or both in Hepa1-6 cells.

Identification number	**Sequence (5´- 3´)**
SNTA siRNAs	
s74113	CAA GAU GCC UAU UCU CAU Utt
s74114	CGA UGG UCU UUA UCA UCC Att
s74115	UGA AGG AGG UCU CAC CCU Att
SNTB2 siRNAs	
s74119	ACC UUG AUC UUA CGC UGC Att
s74120	GAA GAU GUC UGC UGA UGA Utt
s74121	GCA AGG AAG UCA AGC AUA Utt

Monitoring of Gene Expression by Real-time RT-PCR

Total RNA was isolated with TRIzol reagent from Life Technologies GmbH (Darmstadt, Germany) and 1 µg RNA was reverse transcribed using the Promega Reverse Transcription System (Promega, Madison, WI) in a volume of 40 µl; 2 µl of the cDNA was used for amplification in glass capillaries (LightCycler) with the primers listed in Suppl. Table 2. These oligonucleotides were synthesized by Metabion (Planegg-Martinsried, Germany). Real-time PCR was performed using the LightCycler FastStart DNA Master SYBR Green I kit (Roche, Mannheim, Germany) and the specificity of the PCRs was confirmed by sequencing of the amplified DNA fragments (Geneart, Regensburg, Germany). For quantification of the results, RNA of liver tissue was reverse transcribed, cDNA was serially diluted and used to

create a standard curve for each of the genes analyzed. The second derivative maximum method was used for quantification with the LightCycler software. Values were normalized to cyclophilin mRNA expression.

Suppl. Table 2: Sequences of the primers used in real-time PCR

Gene	**Forward Primer (5´- 3´)**	**Reverse Primer (5´- 3´)**
ABCA1	GATGAACCAACCACAGGCAT	GAGATGACAAGGAGGATGGAA
ApoA-I	CTTGGGCCAACAGCTGAA	TCCTGTCTCACCCAATCTGTT
ApoB-100	TGAATGTCTACTTCCACCCACA	GGGAGCCTAGCAATCTGG A
Cyclophilin	AACACAAACGGTTCCCAGTT	TTGAAGGGGAATGAGGAAAA
Cyp7A1	CACATAAAGCCC GGGAAA G	GGCTGCTTTCATTGCTTC A
HMG-CoA-R	TGTGCTTGGGGCTTCTGTA	CCACGTTCATGAGTTTCCATT
LDL-R	GATGGCTATACCTACCCCTCAA	CCTTTTCTGTCCCCAGACAA
SMPD3	CCCTGACTGGAAGGCTGA	TGGCATGGCTGGTCTATG
SMS2	TTACCTGTGCCCGGAATG	CGCTACGAGAATGCAGATGA
SR-BI	TGCCCCAGGTTCTTCACTAC	CCAAAACAAAAAGCATTTCTCC
SREBP2	CCCTATTCCATTGACTCTGAGC	GAGTCCGGTTCATCCTTGAC

Immunohistochemistry

Immunohistochemical studies for the expression of PDZK1 utilized the EnVision+ Kit (DAKO, Glostrup, Denmark) based on a HRP labelled polymer which is conjugated with a secondary antibody. Three μm sections were cut from formalin-fixed and paraffin-embedded mouse liver tissues. After deparaffinization for 15 min in Histol, tissue sections were rehydrated in descending ethanol series following antigen retrieval (microwave oven for 20 min at 800 W in sodium citrate buffer). Endogenous peroxidase activity was eliminated by subsequent incubation with 0.3% hydrogen peroxide for 10 min. After washing in TBS, 0.5% Tween 20 slides were incubated for 1 h in a protein-blocking solution (DAKO). Incubation