Supplementary Figure 1

IL-1R deficiency causes decreased inflammatory cell recruitment to the lung. Infected lungs 5 dpi were dissociated into single cell suspensions and analyzed via flow cytometry to determine numbers of specific cell populations. A) CD45+ cell counts. B) Alveolar macrophage cell count as defined by $CD11c^{+}$, $SiglecF^{+}$, autofluorescent cells. C) Dendritic cell count as defined by $CD11c^{+}$, $CD11b^{+}$, MHC Class II^{high} cells. D) Monocyte cell count as defined by $CD11c^{-}$, $CD11b^{+}$, side-scatter low, $Gr\text{-}1^{+}$ cells. E) Neutrophil cell count as defined by $CD11c^{-}$, $CD11b^{+}$, side-scatter high, $Gr\text{-}1^{+}$ cells. *p<0.01, p values were determined using ANOVA analysis.

Supplementary Figure 2

Representative FACS plot of CD4+ T cells that are CD69 positive. Lung cells were dissociated and stained with appropriate antibodies. This plot shows CD4+ cells that are CD69+. Similar gating strategies were used to define CD69+ populations on CD8+ T cells, γδ TCR+ cells and NK cells.

Supplementary Figure 3

IL-1R deficiency does not affect cytokine signaling in bone marrow derived dendritic cells *in vitro*. Bone-marrow derived dendritic cells from C57Bl/6J and $IL\text{-}1R^{-/-}$ mice were harvested and infected *in vitro* with *Histoplasma* at an MOI of 1. Supernatants from infected cells were collected in triplicate at 48 hours post-infection and evaluated for cytokine levels. Significance was determined by ANOVA analysis.

Supplementary Figure 4

In the absence of T cells, *Histoplasma* grows similarly in wild-type and MyD88$^{-/-}$ BMDCs. BMDCs were infected with wild-type *Histoplasma* (MOI=1). Cells were lysed and CFUs counted at the indicated timepoints. Timepoints are the mean +SD of three independent samples. All results are representative of at least three independent experiments.