

## Figure S1. LytM-mediated release of protein A by S. aureus Newman.

A) Protein A on the surface of Newman *sbi* and Newman *sbi lytM* was detected using FITC-labelled rabbit IgG and the fluorescence intensity was measured by flow cytometry. Values are plotted as a percentage of the mean fluorescence intensity measured for NM *sbi* grown to an  $OD_{600}$  of 0.3. Bars represent the mean values and error bars indicate the SEM of three independent experiments.

B) Protein was captured from culture supernatants using chicken anti-SpA polyclonal IgY and detected using biotin-conjugated mouse monoclonal anti-SpA IgG followed by streptavidin-HRP in an ELISA. Values are expressed as a percentage of total released SpA measured for Newman *sbi* grown to an  $OD_{600} = 0.3$ . Bars represent the mean percentage release from four independent experiments. Error bars represent the SEM. \*\* p = 0.003, \* p = 0.014, n.s. = not significant, p > 0.05.



## Figure S2. Replacing the SpA sorting signal with the sorting signal from SdrE reduces release of SpA by S. *aureus* Newman

- A) Protein A on the surface of Newman *sbi* and Newman *sbi* [SpA $\Omega$ SdrESS] was detected using FITC-labelled rabbit IgG and the fluorescence intensity was measured by flow cytometry. Values are plotted as a percentage of the mean fluorescence intensity measured for Newman *sbi* grown to an OD<sub>600</sub> of 0.3. Bars represent the mean values and error bars indicate the SEM of three independent experiments.
- B) Protein was captured from culture supernatants using chicken anti-SpA polyclonal IgY and detected using biotinconjugated mouse monoclonal anti-SpA IgG followed by streptavidin-HRP in an ELISA. Values are expressed as a percentage of total released SpA measured for Newman *sbi* grown to an  $OD_{600} = 0.3$ . Bars represent the mean percentage release from four independent experiments. Error bars represent the SEM. \*\*\* p = <0.001, \*\* p = 0.02, n.s. = not significant, p > 0.05.

Oligonucleotide name	5' - 3' sequence <sup>a, b</sup>
pSLF	GAATTCACTGGCCGTCGTTTTACAACGTCG
pSLR	GAGCTCAGATCTGTTAACGGTACCATCATA
SpAF	TATGATGGTACCGTTAACAGATCTGAGCTCTACAGGGGGGTATTAATTTG
SpAR	CGACGTTGTAAAACGACGGCCAGTGAATTCTTATAGTTCGCGACGACG
RESF	CAGATGCTAACAAAGCTCAAGCATTACCAGAAACAGGTAGTG
RESR	<u>GTAAAACGACGGCCAGTGAATTC</u> TTATTTGTTTTGTTTT
DWrR	TGGCGCCACTTTCTTTCAGCATC
DSpF	GATGCTGAAAAGAAAGTGGCGCCATTACCAGAAACTGGTGAAG
DSdF	GATGCTGAAAAGAAAGTGGCGCCATTACCAGAAACAGGTAGTG
SEQF	ATTCAGGCTGCGCAAC
SEQR	TTGTTGACATTATATCATTG
lytm-A	GGAGGTACCCAATCACTATACGATGTATCAGAC
lytM-B	CATGTATAAAACATCCTCCATTAAAG
lytM-C	TTAATGGAGGATGTTTTATACATGTAATACAGAAAATCCCAAGTTG
lytM-D	GGAGAGCTCAAAGACAAATACAAAATCACATC
pSRER	TTATTTGTTTTGTTTTTGCGAC
SpADSF	<u>CGTCGCAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAA</u>
SpADSR	CGACGTTGTAAAACGACGGCCAGTGAATTCATTCAACAAGCGGTTCATTAC
pIREF	CCTCACTAAAGGGAACAAAAGCTGGGTACCAAAGCCTTAAAGACGATC
pIRER	CGACTCACTATAGGGCGAATTGGAGCTCTCAACAAGCGGTTCATTAC
pIMAYF	GGTACCCAGCTTTTGTTCCCTTTAGTGAGG
pIMAYR	GAGCTCCAATTCGCCCTATAGTGAGTCG

 TABLE S1. Oligonucleotides used in this study.

<sup>a</sup>\* indicates a 5' phosphorylation modification

<sup>b</sup>5' extensions with homology to vector sequences were added to facilitate sequence and ligase-independent cloning (underlined).