

Figure S1. LytM-mediated release of protein A by S. aureus Newman.

A) Protein A on the surface of Newman *sbi* and Newman *sbi lytM* was detected using FITC-labelled rabbit IgG and the fluorescence intensity was measured by flow cytometry. Values are plotted as a percentage of the mean fluorescence intensity measured for NM *sbi* grown to an OD_{600} of 0.3. Bars represent the mean values and error bars indicate the SEM of three independent experiments.

B) Protein was captured from culture supernatants using chicken anti-SpA polyclonal IgY and detected using biotin-conjugated mouse monoclonal anti-SpA IgG followed by streptavidin-HRP in an ELISA. Values are expressed as a percentage of total released SpA measured for Newman *sbi* grown to an $OD_{600} = 0.3$. Bars represent the mean percentage release from four independent experiments. Error bars represent the SEM. ** p = 0.003, * p = 0.014, n.s. = not significant, p > 0.05.



Figure S2. Replacing the SpA sorting signal with the sorting signal from SdrE reduces release of SpA by S. *aureus* Newman

- A) Protein A on the surface of Newman *sbi* and Newman *sbi* [SpA Ω SdrESS] was detected using FITC-labelled rabbit IgG and the fluorescence intensity was measured by flow cytometry. Values are plotted as a percentage of the mean fluorescence intensity measured for Newman *sbi* grown to an OD₆₀₀ of 0.3. Bars represent the mean values and error bars indicate the SEM of three independent experiments.
- B) Protein was captured from culture supernatants using chicken anti-SpA polyclonal IgY and detected using biotinconjugated mouse monoclonal anti-SpA IgG followed by streptavidin-HRP in an ELISA. Values are expressed as a percentage of total released SpA measured for Newman *sbi* grown to an $OD_{600} = 0.3$. Bars represent the mean percentage release from four independent experiments. Error bars represent the SEM. *** p = <0.001, ** p = 0.02, n.s. = not significant, p > 0.05.

| Oligonucleotide name | 5' - 3' sequence ^{a, b} |
|----------------------|---|
| | |
| pSLF | GAATTCACTGGCCGTCGTTTTACAACGTCG |
| pSLR | GAGCTCAGATCTGTTAACGGTACCATCATA |
| SpAF | TATGATGGTACCGTTAACAGATCTGAGCTCTACAGGGGGGTATTAATTTG |
| SpAR | CGACGTTGTAAAACGACGGCCAGTGAATTCTTATAGTTCGCGACGACG |
| RESF | CAGATGCTAACAAAGCTCAAGCATTACCAGAAACAGGTAGTG |
| RESR | <u>GTAAAACGACGGCCAGTGAATTC</u> TTATTTGTTTTGTTTT |
| DWrR | TGGCGCCACTTTCTTTCAGCATC |
| DSpF | <u>GATGCTGAAAAGAAAGTGGCGCCA</u> TTACCAGAAACTGGTGAAG |
| DSdF | GATGCTGAAAAGAAAGTGGCGCCATTACCAGAAACAGGTAGTG |
| SEQF | ATTCAGGCTGCGCAAC |
| SEQR | TTGTTGACATTATATCATTG |
| lytm-A | GGAGGTACCCAATCACTATACGATGTATCAGAC |
| lytM-B | CATGTATAAAACATCCTCCATTAAAG |
| lytM-C | TTAATGGAGGATGTTTTATACATGTAATACAGAAAATCCCAAGTTG |
| lytM-D | GGAGAGCTCAAAGACAAATACAAAATCACATC |
| pSRER | TTATTTGTTTTGTTTTTGCGAC |
| SpADSF | <u>CGTCGCAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAA</u> |
| SpADSR | CGACGTTGTAAAACGACGGCCAGTGAATTCATTCAACAAGCGGTTCATTAC |
| pIREF | CCTCACTAAAGGGAACAAAAGCTGGGTACCAAAGCCTTAAAGACGATC |
| pIRER | CGACTCACTATAGGGCGAATTGGAGCTCTCAACAAGCGGTTCATTAC |
| pIMAYF | GGTACCCAGCTTTTGTTCCCTTTAGTGAGG |
| pIMAYR | GAGCTCCAATTCGCCCTATAGTGAGTCG |

 TABLE S1. Oligonucleotides used in this study.

^a* indicates a 5' phosphorylation modification

^b5' extensions with homology to vector sequences were added to facilitate sequence and ligase-independent cloning (underlined).