

## Supplementary Information

### Enhanced photocurrent production by bio-dyes of photosynthetic macromolecules on designed TiO<sub>2</sub> film

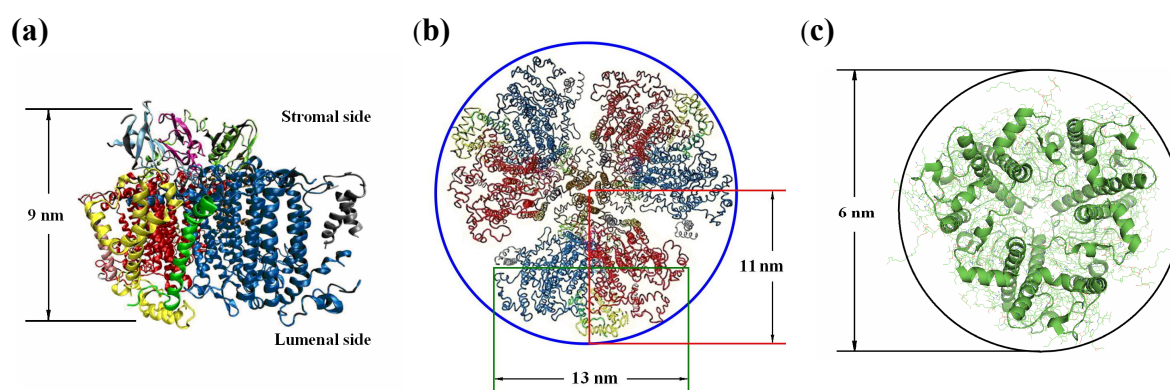
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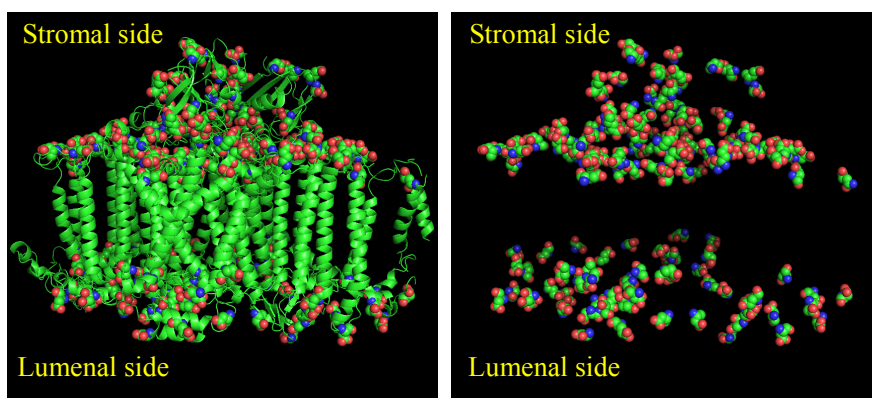
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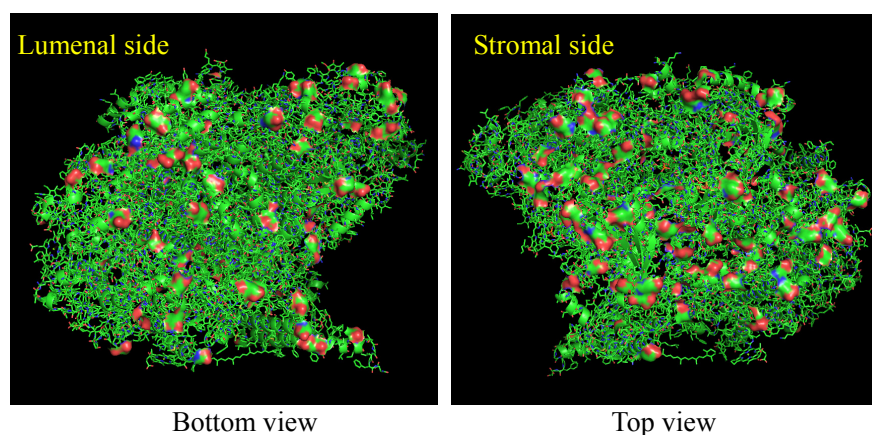
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**Figure S1** Dimensional size of PSI (PDB ID: 1JB0) and LHCII (PDB ID: 1RWT). (a) PSI monomer. (b) Luminal side of trimeric PSI complex constructed from 1JB0. (c) LHCII trimer. Marked sizes are measured by PyMOL.



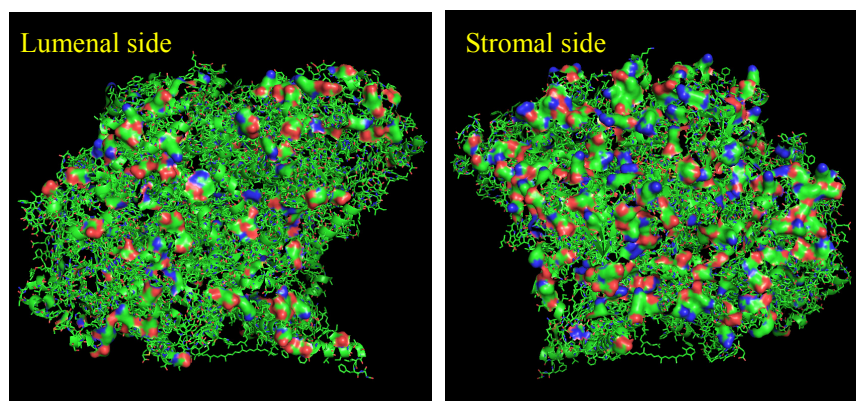
(a) Distribution of residues of aspartic acid and glutamic acid (Lateral view)



Bottom view

Top view

(b) Negative charged surface of residues of aspartic acid and glutamic acid

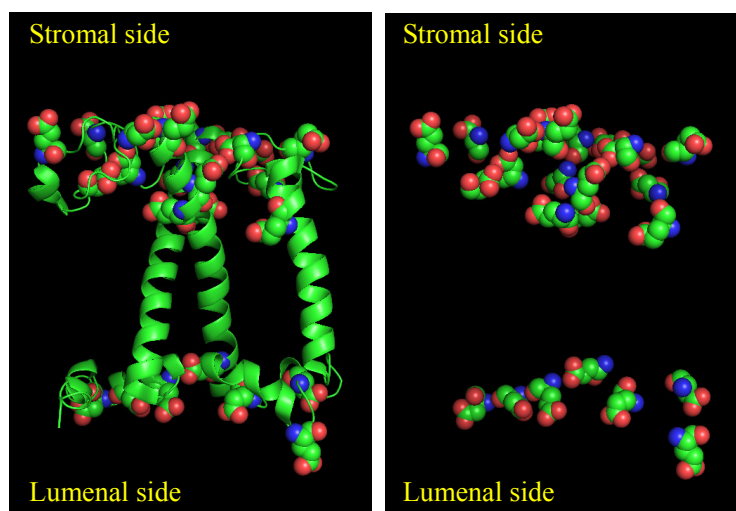


Bottom view

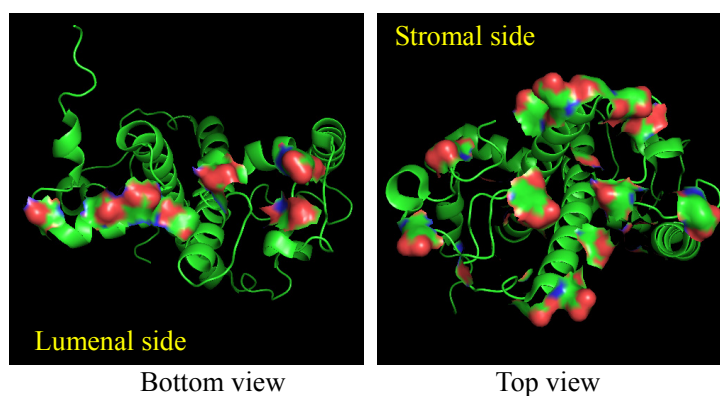
Top view

(c) Charged surface of negative residues of D and E, and positive residues of H, R, and K

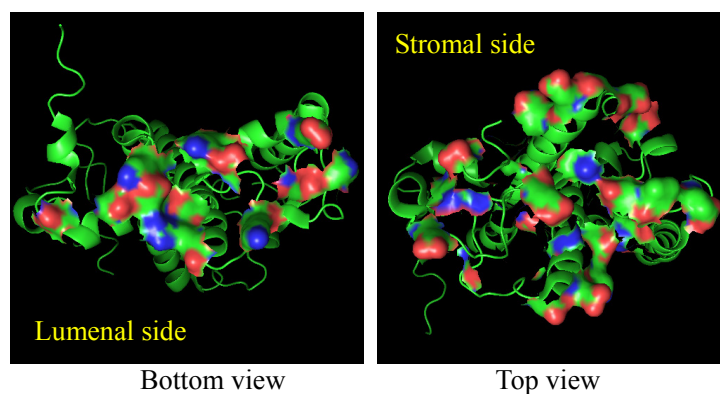
**Figure S2** PSI monomer, Deduced from PDB ID 1JB0. (a) There are 102 carboxyl residues (55E+47D) at the stromal side, and there are 58 carboxyl residues (15E+43D) at the luminal side (PDBTM 1jb0). (b) The stromal side has more carboxyl groups on the surface. (c) The stromal side has more charged groups of carboxyl and amino on the surface.



(a) Distribution of residues of aspartic acid and glutamic acid (Lateral view)



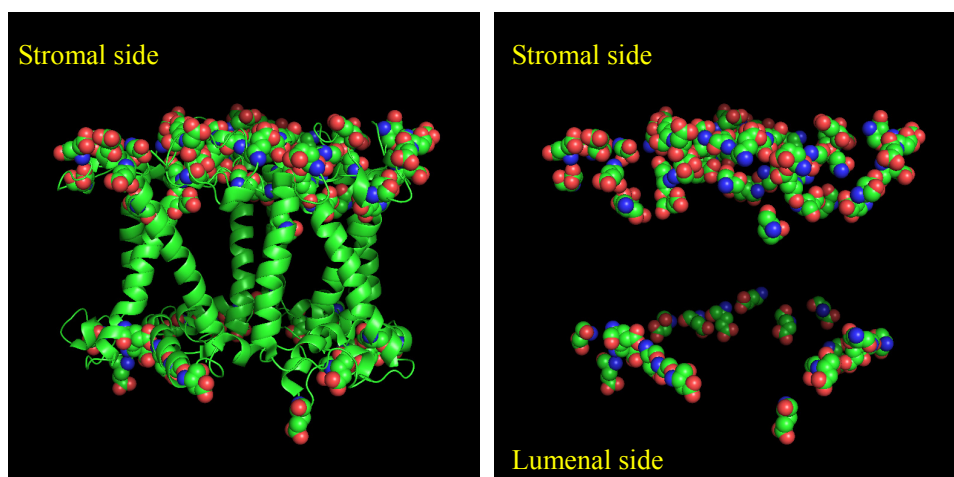
(b) Negative charged surface of residues of aspartic acid and glutamic acid



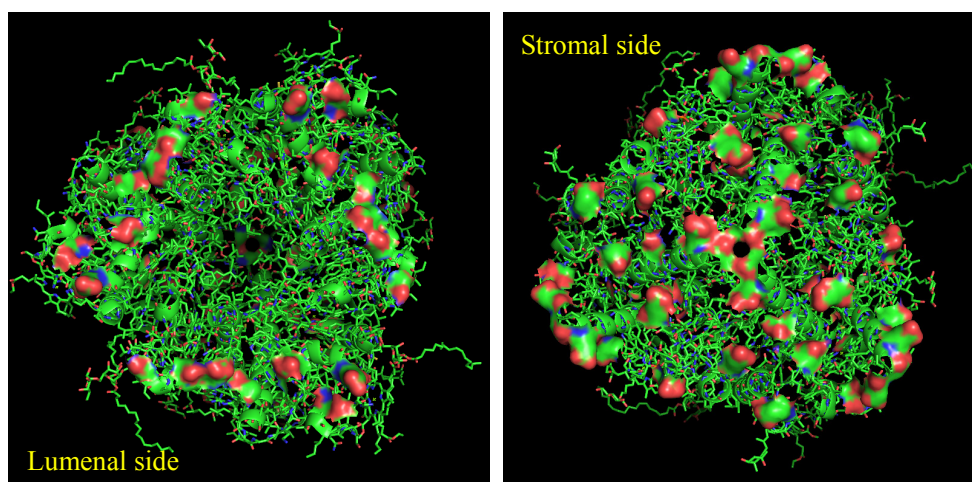
(c) Charged surface of negative residues of D and E, and positive residues of H, R, and K

**Figure S3** LHCII monomer, Deduced from PDB ID 1RWT. (a) There are 18 carboxyl residues (10E+8D) at the stromal side, and there are only 7 carboxyl residues (4E+3D) at the luminal side. (b) The stromal side has more carboxyl groups on the surface. (c) The stromal side has more charged groups of carboxyl and amino on the surface.

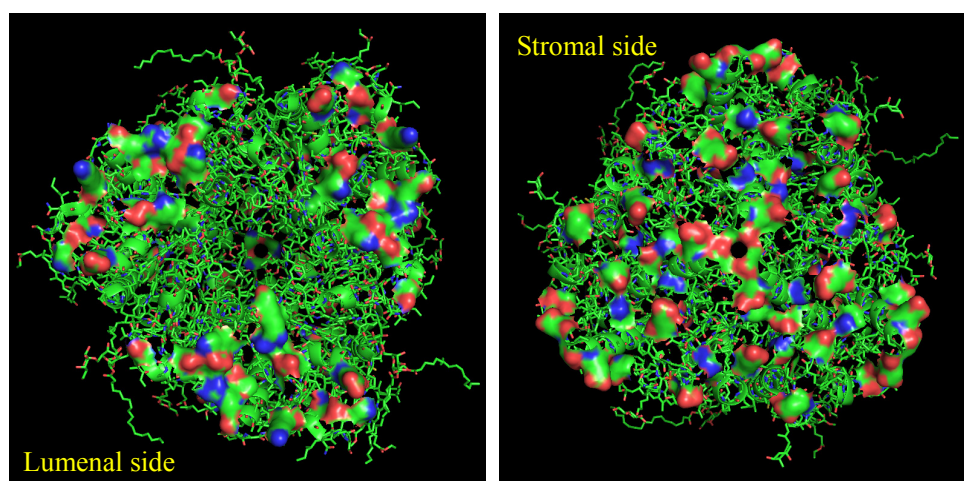




(a) Distribution of residues of aspartic acid and glutamic acid (Lateral view)

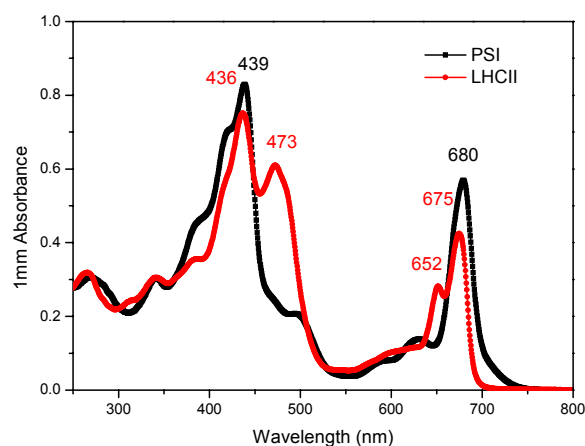


Bottom view Top view  
(b) Negative charged surface of residues of aspartic acid and glutamic acid

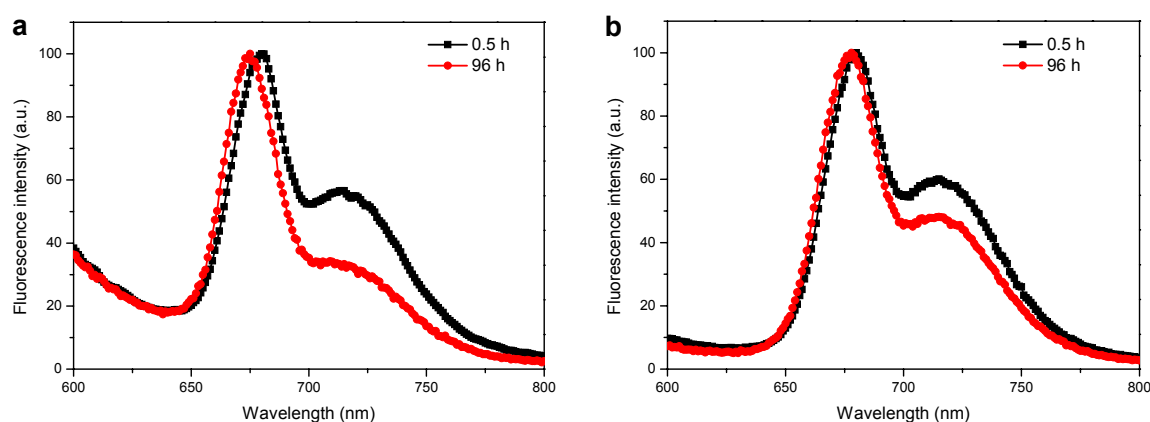


Bottom view Top view  
(c) Charged surface of negative residues of D and E, and positive residues of H, R, and K

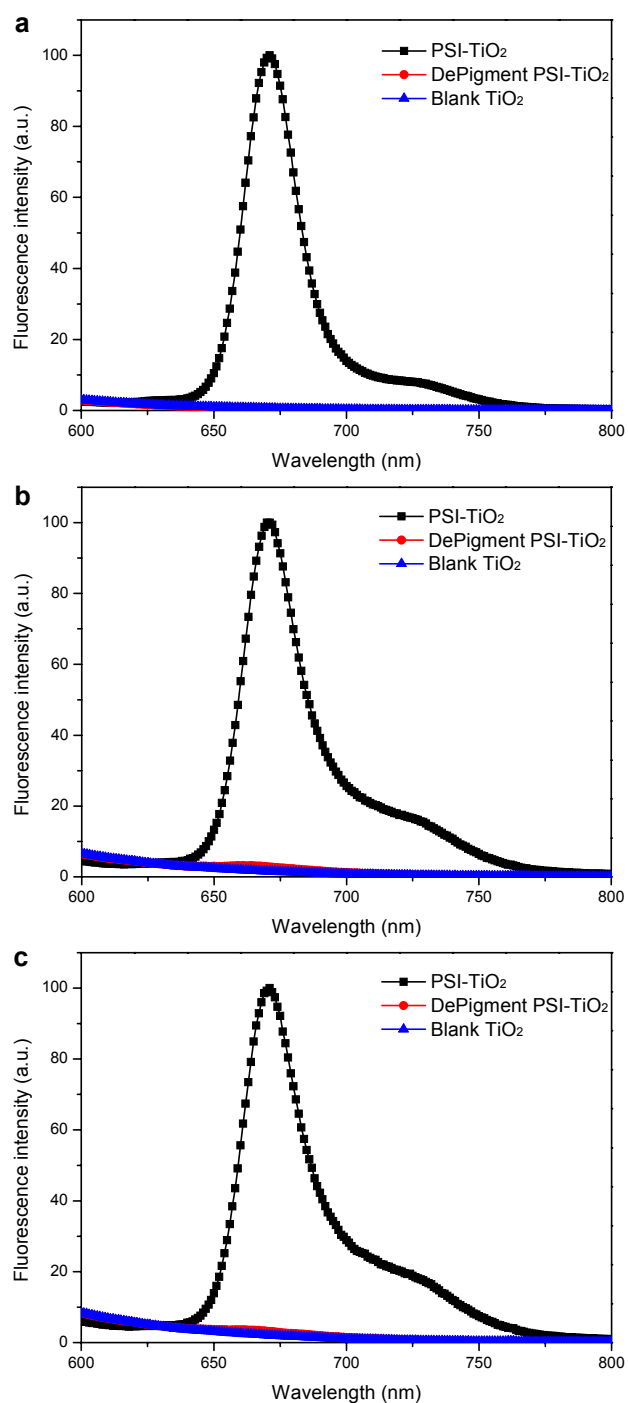
**Figure S4** LHCII trimer, Deduced from PDB ID 1RWT. (a) There are 54 carboxyl residues (30E+24D) at the stromal side, and there are only 21 carboxyl residues (12E+9D) at the luminal side. (b) The stromal side has more carboxyl groups on the surface. (c) The stromal side has more charged groups of carboxyl and amino on the surface.



**Figure S5** Absorption spectra of *A. platensis* PSI trimer and spinach LHCII trimer in solutions. Spinach LHCII, 80 $\mu$ g Chl/mL, was solubilized by 1.0 mM DDM in 20 mM MES buffer at pH=6.0 with 8 mM sucrose. *A. platensis* PSI, 80 $\mu$ g Chl/mL, was solubilized by 1.0 mM DDM in 20 mM MES buffer at pH=6.5. Steady-state UV/Vis absorption spectra were scanned at 0.5 nm resolution by a Shimadzu (Kyoto, Japan) UV/Vis 2450 spectrometer at room temperature. The maxima of LHCII in the Qy region are around 675 nm and 652 nm, and in the Soret region around 436 nm and 473 nm. The maxima of PSI in the Qy region and Soret region are at about 680 nm and 439 nm respectively, and these are characteristic peaks of Chl *a* assembled in PSI complexes, while the shoulder peak at about 495 nm is contributed by carotenoids assembled in PSI complexes.

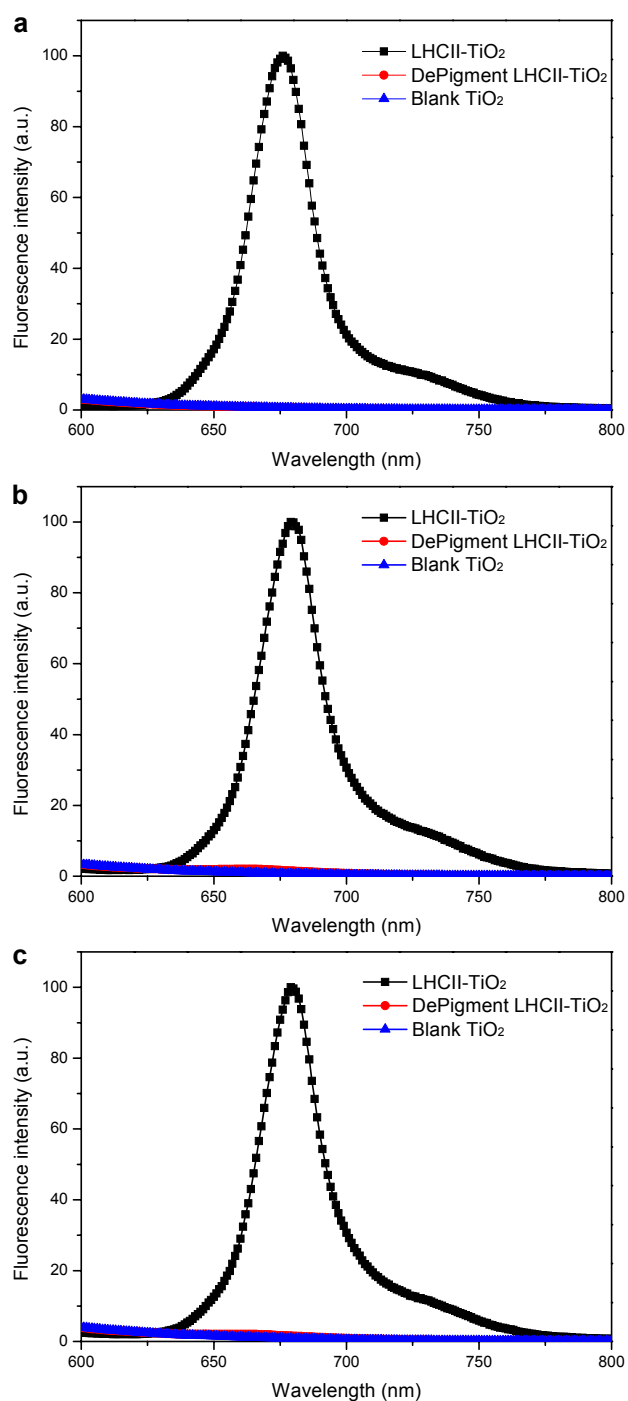


**Figure S6** TIRF spectra of *A. Platensis* PSI solubilized by 1.0 mM DDM in 20 mM MES buffer at pH 6.5. **(a)** PSI, 80  $\mu$ g Chl/mL. **(b)** PSI, 400  $\mu$ g Chl/mL. TIRF spectra were measured with an accessory TA1004 on a FluoroMax-4 fluorescence spectrometer (Horiba Jobin Yvon) from 600 nm to 800 nm at room temperature excited at 436 nm, and slit width of excitation and emission were set at 5 nm and 5 nm respectively. The TA1004 accessory allows for detecting molecules at the TIRF surface in the presence of relatively high concentration of the same molecules in the bulk solution.



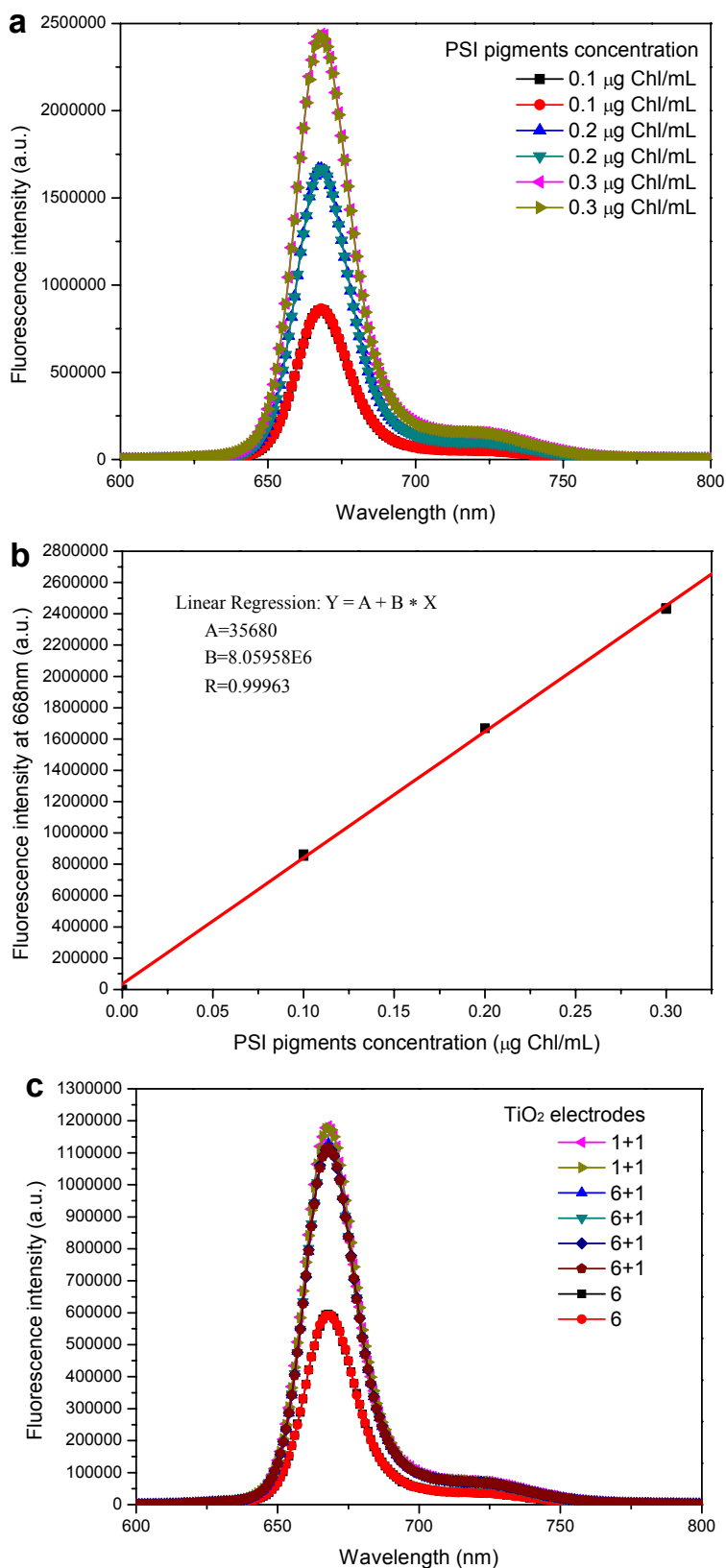
**Figure S7** Fluorescence spectra of PSI-TiO<sub>2</sub> electrodes before and after extraction of pigments by 80% acetone/water. (a) 6 type, (b) 6+1 type, and (c) 1+1 type TiO<sub>2</sub> electrodes.

Fluorescence emission spectra were recorded by a FluoroMax-4 fluorescence spectrometer (Horiba Jobin Yvon) from 600 nm to 800 nm at room temperature excited at 436 nm, and slit width of excitation and emission were set at 3 nm and 1 nm respectively. Dried TiO<sub>2</sub> electrode was fixed onto the solid sample holder at 30° angle excitation with a 515 nm cutoff filter.



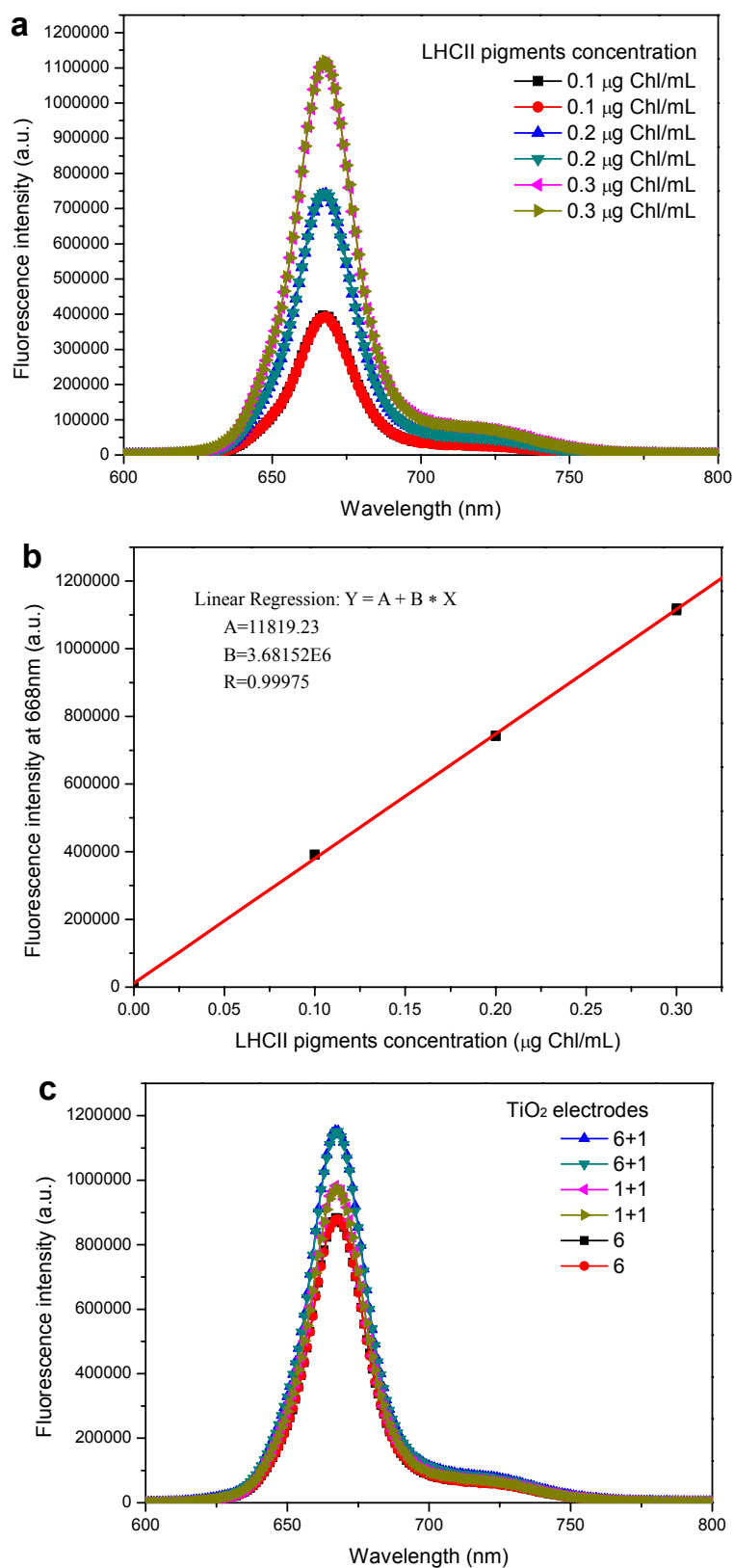
**Figure S8** Fluorescence spectra of LHCII-TiO<sub>2</sub> electrodes before and after extraction of pigments by 80% acetone/water. (a) 6 type, (b) 6+1 type, and (c) 1+1 type TiO<sub>2</sub> electrodes.

Fluorescence emission spectra were recorded by a FluoroMax-4 fluorescence spectrometer (Horiba Jobin Yvon) from 600 nm to 800 nm at room temperature excited at 436 nm, and slit width of excitation and emission were set at 3 nm and 1 nm respectively. Dried TiO<sub>2</sub> electrode was fixed onto the solid sample holder at 30° angle excitation with a 515 nm cutoff filter.

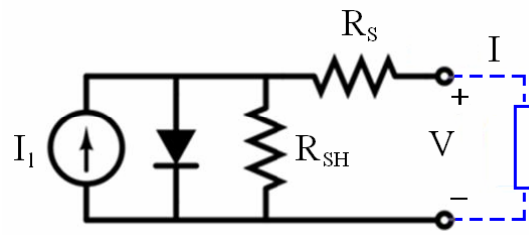


**Figure S9** Adsorbance determination of PSI on TiO<sub>2</sub> electrodes. (a) Fluorescence spectra of standard solutions of PSI pigments in 80% acetone/water. (b) The linear relationship of fluorescence intensity at 668 nm with concentration of PSI pigments. (c) Fluorescence spectra of extracted pigments from PSI-TiO<sub>2</sub> electrodes for quantitative analysis.





**Figure S10** Adsorbance determination of LHCII on TiO<sub>2</sub> electrodes. **(a)** Fluorescence spectra of standard solutions of LHCII pigments in 80% acetone/water. **(b)** The linear relationship of fluorescence intensity at 668 nm with concentration of LHCII pigments. **(c)** Fluorescence spectra of extracted pigments from LHCII-TiO<sub>2</sub> electrodes for quantitative analysis.



**Figure S11** Simplified equivalent circuit model for a solar cell.  $R_S$  and  $R_{SH}$  represent the series and shunt resistance. <http://www.ni.com/white-paper/7230/en/>