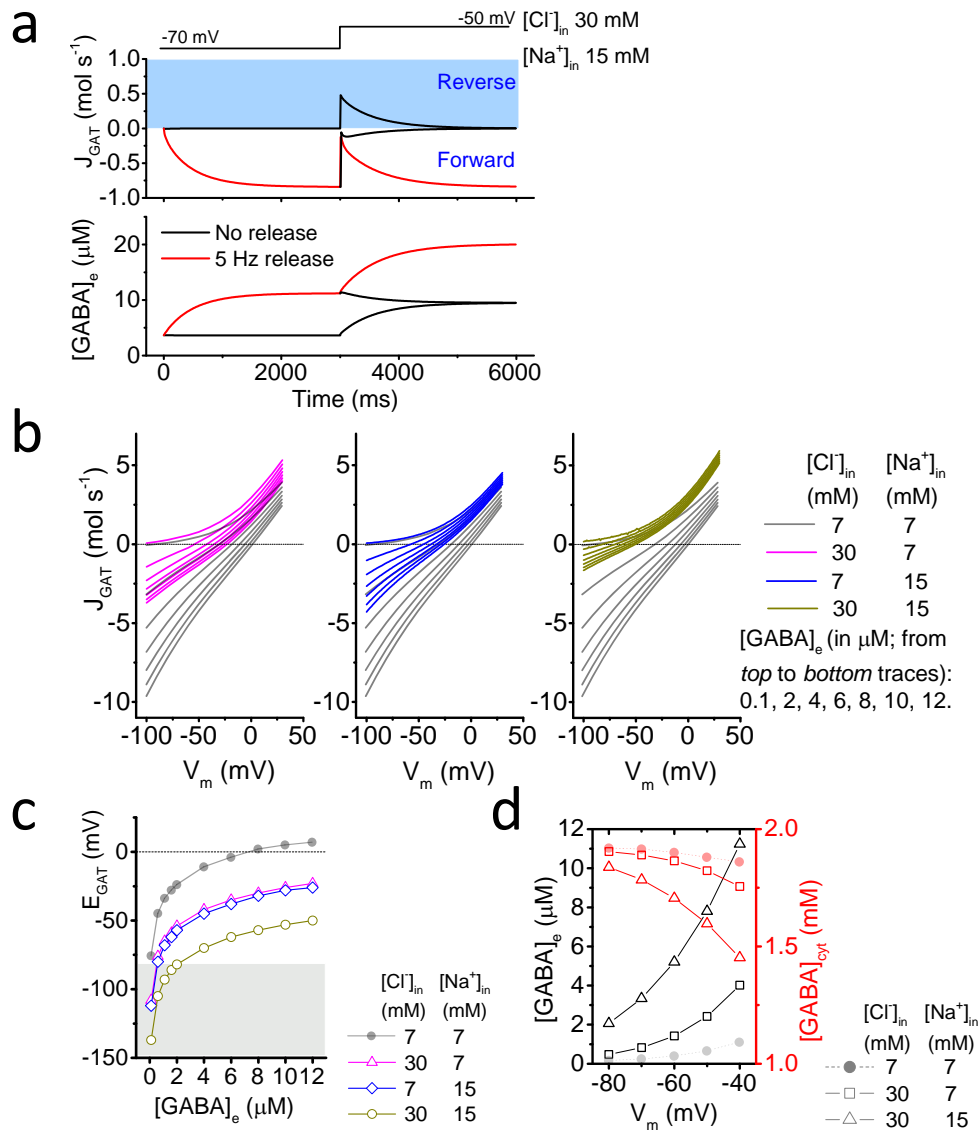
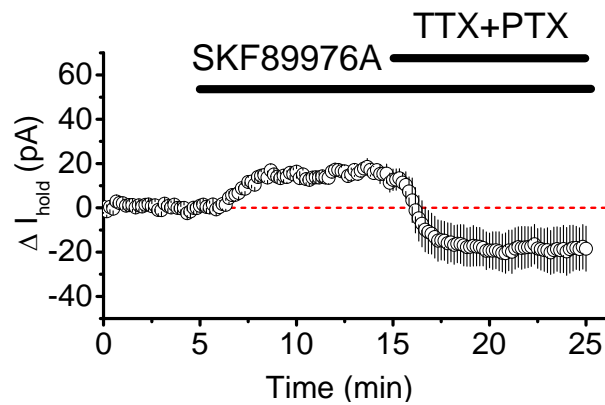


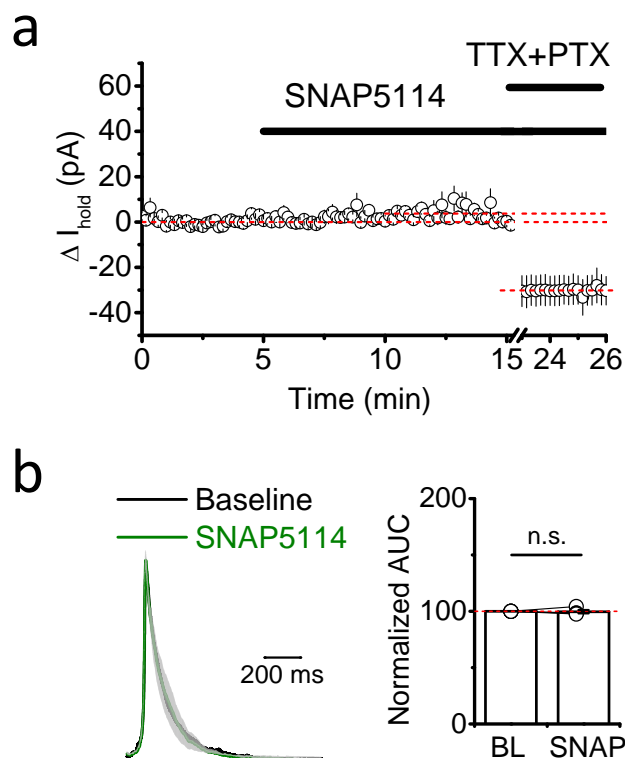
Supplementary Figure 1. GABA concentrations at different GAT-1 densities. Computer simulations showing membrane potential dependence of the steady-state extracellular GABA concentrations ($[\text{GABA}]_e$; *black*) and cytosolic GABA concentrations ($[\text{GABA}]_{\text{cyt}}$; *red*) at different GAT-1 densities. Synaptic release rate is 2 Hz; filled symbols show corresponding values from Fig. 1d for comparison.



Supplementary Figure 2. GAT-1 activity and GABA concentrations at high levels of internal Cl⁻ and Na⁺. (a) Computer simulations showing the effect of depolarization on the dynamics of GAT-1 operation and extracellular GABA concentration ($[GABA]_e$) with (red) and without (black) synaptic GABA release. (b) Current-voltage relationships of the steady-state GAT-1-mediated current (J_{GAT} , molecules per s; negative values – forward mode, positive values – reverse mode) at different concentrations of extracellular GABA ($[GABA]_e$) as in Fig. 1a). A family of traces from Fig. 1a ($[Cl^-]_{in} = 7$ mM; $[Na^+]_{in} = 7$ mM) is shown in gray for comparison. (c) Dependence of GAT-1 reversal potential (E_{GAT}) on the concentration of extracellular GABA at different values of internal Cl⁻ and Na⁺. White area, physiologically relevant membrane potentials. (d) Membrane potential (V_m) dependence of the steady-state cytosolic ($[GABA]_{cyt}$) and extracellular GABA at different values of internal Cl⁻ and Na⁺ in the absence of synaptic release (filled symbols show corresponding values from Fig. 1c for comparison).

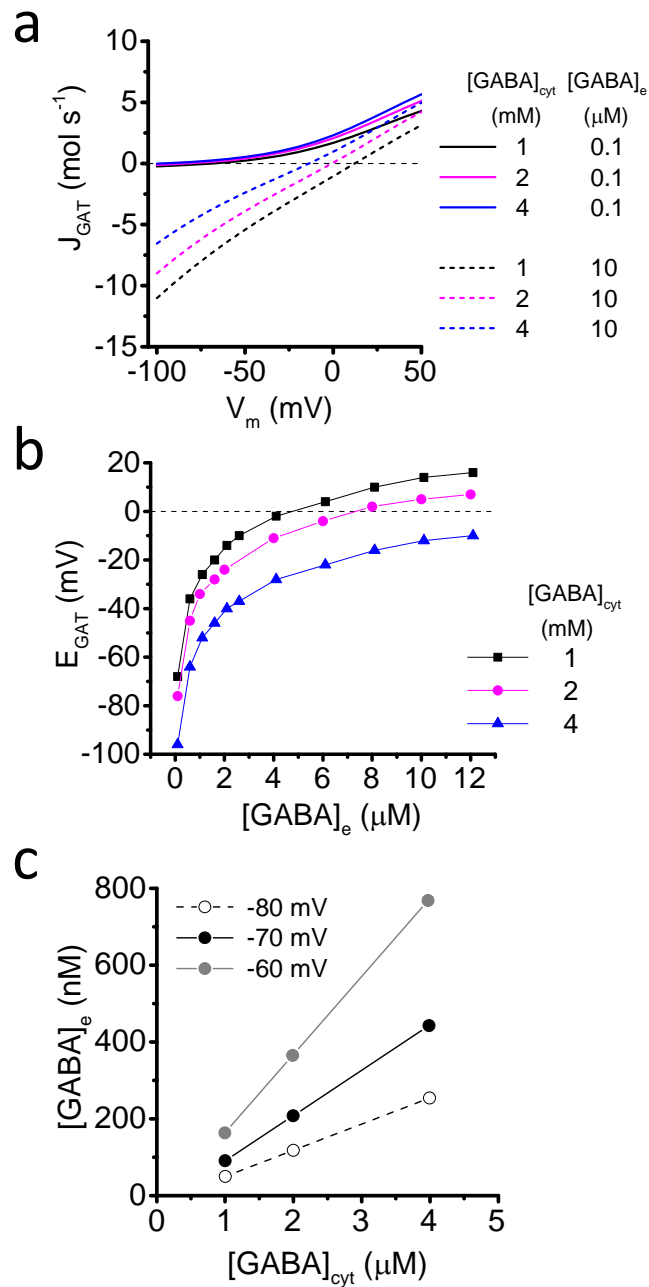


Supplementary Figure 3. Inhibition of GAT-1 increases tonic GABA_AR-mediated currents. Holding current (I_{hold}) changes in CA1 pyramidal neurons induced by application of SKF899976A in the absence of epileptiform activity (normal Mg^{2+} aCSF; $V_{\text{hold}} = 0$ mV; low Cl^- intracellular recording solution; c.f. Fig. 4a; $n = 5$). TTX, tetrodotoxin; PTX, picrotoxin. Error bars, s.e.m.



Supplementary Figure 4. GAT-3 impact on GABA_AR currents during epileptiform

activity. (a) Tonic GABA_A receptor-mediated currents in CA1 pyramidal neurons following application of SNAP5114 (100 μ M) during ongoing epileptiform activity (n = 5). (b) Mean normalized traces of GABA_AR transients (*gray*: s.e.m.) show lack of effect of GAT-3 inhibition on the kinetics of burst-associated GABA_AR transients in pyramidal neurons (area under the curve, AUC, baseline: 60.9 ± 14.8 ms, in SNAP5114: 60.3 ± 14.1 ms; n = 4; p = 0.54, paired t-test). TTX, tetrodotoxin; PTX, picrotoxin. Bars, mean; error bars, s.e.m.; circles, individual experiments.



Supplementary Figure 5. GAT-1 simulations at different cytosolic GABA

concentrations. (a) Current-voltage relationship of the steady-state GAT-1-mediated current (J_{GAT} , molecules per s; negative values – forward mode, positive values – reverse mode) at different concentrations of the cytosolic ($[GABA]_{cyt}$) and extracellular ($[GABA]_e$) GABA. (b) Dependence of GAT-1 reversal potential (E_{GAT}) on the concentration of extracellular GABA. (c) Steady-state extracellular GABA concentrations vs. cytosolic GABA at different subthreshold membrane potentials.