

Table S2**Primers used in this study**

#	Primer Name	Sequence	Description
1	DTO13	catgtcaagtctcgagGCATAATGATTGTTTGATTCTTA ATGAAAC	3' <i>NRG1</i> 2.9 kb promoter primer for cloning into placbasal
2	DTO33	catgtgcaattgtactgcagCTTGCAACATTGTGTAGTTG C	5' <i>NRG1</i> 2.9 kb promoter primer for cloning into placbasal
3	DTO228	agtactgaactgcagCAAAACCCCAATTGCTTGGC	5' <i>NRG1</i> 2.5 kb promoter primer for cloning into placbasal
4	DTO30	catgtgcaattgtactgcagCACAATAGCATACTATGCA GGC	5' <i>NRG1</i> 2.1 kb promoter primer for cloning into placbasal
5	DTO229	agtactgaactgcagGAGATGCACTTGGTGGTATAACC	5' <i>NRG1</i> 1.5 kb promoter primer for cloning into placbasal
6	DTO83	agtactgaactgcagGCACATAGCCATGCATTAGCC	5' <i>NRG1</i> 902 bp promoter primer for cloning into placbasal
7	DTO84	agtactgaactgcagGCTAGCAATGAACCAAACGG	5' <i>NRG1</i> 570 bp promoter primer for cloning into placbasal
8	DTO154	agtactgaactgcagCGTCCAGGTTACTATTTTCC	5' <i>NRG1</i> 362 bp promoter primer for cloning into placbasal
9	DTO182	agtactgaactgcagCATTGTTCAAGTATCTTCC	5' <i>NRG1</i> 122 bp promoter primer for cloning into placbasal
10	DTO181	agtactgaactgcagCAATAAAACATCGTTATCC	5' <i>NRG1</i> 74 bp promoter primer for cloning into placbasal
11	DTO156	agtactgaactgcagCATCTCAAATTTTCCCTGC	5' <i>NRG1</i> 50 bp promoter primer for cloning into placbasal
12	DTO230	agtactgaactgcagGATGAGAAACAGGATAACG	3' <i>NRG1</i> 45 bp promoter primer for cloning into

			placbasal
13	DTO237	agtactgaactcgagGGAAAATAGTAACCTGGACG	3' <i>NRG1</i> 342 bp promoter primer for cloning into placbasal
14	DTO233	agtactgaactcgagGGTTCATTGCTAGCTTGTTAGG	3' <i>NRG1</i> 556 bp promoter primer for cloning into placbasal
15	DTO234	agtactgaactcgagCTAATGCATGGCTATGTGCC	3' <i>NRG1</i> 883 bp promoter primer for cloning into placbasal
16	DTO235	agtactgaactcgagGACCACCTCAAGCCTTGG	3' <i>NRG1</i> 1.9 kb promoter primer for cloning into placbasal
17	DTO250*	agtactgaactgcagcatgcTTATACAGTGGATTA AATAAACTAATTGGAAGTAAATTGAGG	3' <i>UME6</i> promoter downstream primer for cloning into placbasal
18	DTO90*	CTGTAAAGTTGTTGTGGTAGG	3' <i>UME6</i> promoter primer downstream of native HindIII site for cloning into placbasal
19	DTO89*	GGTCCAGTCGACCAAGAACAATCCATTCTCG	5' <i>UME6</i> promoter primer upstream of native HindIII site for cloning into placbasal
20	DTO243*	agtactgaactgcagcatgcTTATACAGTGGATTA AATAAACTAATTGGAAGTAAATTGAGG	5' 6.0 kb <i>UME6</i> promoter upstream primer for cloning into placbasal
21	DTO255	agtactgaactgcagGAGGCAAATGGCAAGCACC	5' 7.0 kb <i>UME6</i> promoter upstream primer for cloning into placbasal
22	DTO110	CAACATTATCACCTAGACCC	3' <i>UME6</i> promoter primer downstream of native XhoI site for cloning into placbasal
23	DTO123	GGTTACCAAAACATATCTCG	5' <i>UME6</i> promoter primer upstream of native XhoI site for cloning into placbasal
24	DTO254	agtactgaactgcagGTTGAAATGCACATATCGG	5' 5.5 kb <i>UME6</i> promoter upstream primer for

			cloning into placbasal
25	DTO93	GCAACTCCCGGGAAATTCTA	3' <i>UME6</i> promoter primer downstream of native HindIII and SmaI sites for cloning into placbasal
26	DTO51	caatcgattctgcagGATATGTCTTTGTGTTGC	5' 5.0 kb <i>UME6</i> promoter upstream primer for cloning into placbasal
27	DTO252	agtactgaactgcaggcatgcCCTTACTTTTCTAGTTCC	3' 3.0 kb <i>UME6</i> promoter downstream primer for cloning into placbasal
28	DTO251	agtactgaactgcaggcatgcGAATGAAGTTGGAGTGGC	3' 4.0 kb <i>UME6</i> promoter downstream primer for cloning into placbasal
29	DTO260	agtactgaagcatgcCATGCAGCAAGTAATTTGC	3' 4.9 kb <i>UME6</i> promoter downstream primer for cloning into placbasal
30	DTO169*	CGTCTTTGCAGATCGTACCC	5' <i>lacZ</i> primer for qRT-PCR analysis
31	DTO170*	CTGGGAATGTTGCTTCTTCG	3' <i>lacZ</i> primer for RT-qPCR analysis
32	DTO85*	TTGCTCCAGAAGAACATCCAG	5' <i>ACT1</i> primer for RT-qPCR analysis
33	DTO86*	AGTAACACCATCACCAGAATCC	3' <i>ACT1</i> primer for RT-qPCR analysis
34	DTO155	agtactgaactgcagCAATTTGCTTTACAAGAGAACC	5' <i>NRG1</i> 145 bp promoter primer for cloning into placbasal
35	DTO270	agtactgaactcgagGTGTTTGTCTTAGATTGCG	3' <i>NRG1</i> 2.5 kb promoter primer for cloning into placbasal

Note: Lowercase bases indicate flanking sequences within primers that contain restriction sites for cloning.

*previously described in Childers DS, Mundodi V, Banerjee M, Kadosh D (2014) A 5' UTR-mediated translational efficiency mechanism inhibits the *Candida albicans* morphological transition. Mol Microbiol 92:570-85.