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Corresponding Author:	György Buzsáki	# Main Figures:	7
Manuscript Number:	NN-A49390	# Supplementary Figures:	8
Manuscript Type:	Article	# Supplementary Tables:	1
		# Supplementary Videos:	1

Reporting Checklist for Nature Neuroscience

This checklist is used to ensure good reporting standards and to improve the reproducibility of published results. For more information, please read Reporting Life Sciences Research.

Please note that in the event of publication, it is mandatory that authors include all relevant methodological and statistical information in the manuscript.

▶ Statistics reporting, by figure

- Please specify the following information for each panel reporting quantitative data, and where each item is reported (section, e.g. Results, & paragraph number).
- Each figure legend should ideally contain an exact sample size (n) for each experimental group/condition, where n is an exact number and not a range, a clear definition of how n is defined (for example x cells from x slices from x animals from x litters, collected over x days), a description of the statistical test used, the results of the tests, any descriptive statistics and clearly defined error bars if applicable.
- · For any experiments using custom statistics, please indicate the test used and stats obtained for each experiment.
- Each figure legend should include a statement of how many times the experiment shown was replicated in the lab; the details of sample collection should be sufficiently clear so that the replicability of the experiment is obvious to the reader.
- For experiments reported in the text but not in the figures, please use the paragraph number instead of the figure number.

Note: Mean and standard deviation are not appropriate on small samples, and plotting independent data points is usually more informative. When technical replicates are reported, error and significance measures reflect the experimental variability and not the variability of the biological process; it is misleading not to state this clearly.

	TEST USED		SED	n			DESCRIPTIVE STATS (AVERAGE, VARIANCE)		P VALUE		DEGREES OF FREEDOM & F/t/z/R/ETC VALUE	
	FIGURE NUMBER	WHICH TEST?	SECTION & PARAGRAPH #	EXACT VALUE	DEFINED?	SECTION & PARAGRAPH #	REPORTED?	SECTION & PARAGRAPH #	EXACT VALUE	SECTION & PARAGRAPH #	VALUE	SECTION & PARAGRAPH #
example	1a	one-way ANOVA	Fig. legend	9, 9, 10, 15	mice from at least 3 litters/group	Methods para 8	error bars are mean +/- SEM	Fig. legend	p = 0.044	Fig. legend	F(3, 36) = 2.97	Fig. legend
example	results, para 6	unpaired t- test	Results para 6	15	slices from 10 mice	Results para 6	error bars are mean +/- SEM	Results para 6	p = 0.0006	Results para 6	t(28) = 2.808	Results para 6

		TEST US	SED		n		DESCRIPTIVE S' (AVERAGE, VARIA		P VALU	JE	DEGREES FREEDOM F/t/z/R/ETC	1 &
	FIGURE NUMBER	WHICH TEST?	SECTION & PARAGRAPH #	EXACT VALUE	DEFINED?	SECTION & PARAGRAPH #	REPORTED?	SECTION & PARAGRAPH#	EXACT VALUE	SECTION & PARAGRAPH #	VALUE	SECTION & PARAGRAPH #
+	1e	Mann- Whitney U- test	Results para 1	242,111	HD cells from ADn (6 mice) and PoS (3 animals)	Results para 1 and Methods p16	full distribution	1e	p<1e-10	Results para 1		
+	1f	Mann- Whitney U- test	Results para 1	242,111	HD cells from ADn (6 mice) and PoS (3 animals)	Results para 1 and Methods p16	full distribution	1f	p=1.4e-7	Results para 1		
+ -	За	Pearson's test	Results para 3	907,512, 92	ADn-ADn, ADn-PoS and PoS-PoS cell pairs pairs	Results para 3 and Methods p16	mean +/- SEM	3a	ADn-ADn: p <e-10 (rem)<="" (sws);="" (wake);="" 3="" adn-pos:="" brain="" for="" p="1.2e-8" p<e-10="" pos-="" pos:p<e-10="" states;="" td=""><td>Results para 3</td><td></td><td></td></e-10>	Results para 3		
+	3a	Mann- Whitney U- test	Results para 3	345, 55	ADn-ADn pairs versus PoS-PoS pairs (<60°)	Results para 3	mean +/- SEM	3a	p=1.9e-6 (wake); p=3.6e-5 (SWS); p=9.2e-7 (REM)	Results para 3		
+	4c	Kruskal- Wallis one- way analysis of variance	Results para 3	(96, 200); (77, 157); (29, 57)	ADn-ADn pairs for 0° +/- 15° and 60° +/-15°; ADn-PoS pairs; PoS-PoS pairs	Results para 3	mean +/- SEM	3d	p <e-10 0°<br="" for="">+/-15° and p=1.9e-6 for 60° +/-15°</e-10>	Results para 3		
+	5b	Mann- Whitney U test	Results para 5	86, 92	PoS HD cells predicted from ADn assemblies and ADn cells predicted from PoS assemblies	Results para 4	cumulative distribution	4b	p <e-10 (wake); P=0.018 (SWS); P=3x10-6 (REM);</e-10 	Results para 4		
+ -	Supp leme ntary Fig. 7b	Binomial test	Results para 6	3394 (14,6)	total number of cell pairs between ADN HD cells and putative pyramidal cells in the PoS (significantly connected from ADn to PoS and from PoS to ADn)	Results para 6	full distribution	7c	p=0.014	Results para 6		
+	Fig. 7d	Wilconxon's signed rank test after testing for unimodality (Rayleigh test, p=2.4e-4)	Results para 6	12	number of HD-HD cell connected from ADn to PoS.	Results para 6	full distribution	7e	p=0.68	Results para 6		

+	5d-e	Wilconxon's signed rank test	Results para 5	86	number of PoS cells predicted by ADn assemblies	Results para 4	mean +/- SEM	4f	p <e-10 all<br="" in="">brain states</e-10>	Results para 4	
+	5d-e	Mann- Whitney U test	Results para 5	86, 62	number of PoS (or ADn) cells predicted by ADn (or PoS) assemblies	Results para 4	mean +/- SEM	4f	p=3e-5 (wake); p <e-10 (sws);<br="">p=1.1e-6 (REM)</e-10>	Results para 4	
+	4e	Mann- Whitney U test	Results para 4	20,8	number of ADn (or PoS) sessions where 6 or more HD cells were simulaneously recorded	Results para 4	full distribution	4e	p=0.0088	Results para 4	
+	6b	Kruskal- Wallis test	Result para 6	970	number of ADn neuronal pairs	Result para 6	mean +/- SEM	6b	p<10-10	Result para 6	
+ -	7e	Mann- Whitney U- test	Result para 6	11 (170)	number of putative pyramidal cells in the PoS postsynaptic to a recorded ADn HD cell (or not connected)	Result para 6	full distribution	7e	p=0.011	Result para 6	
+	7e	Mann- Whitney U- test	Result para 6	13 (79)	number of putative interneurons in the PoS postsynaptic to a recorded ADn HD cell (or not connected)	Result para 6	full distribution	7e	p=0.38	Result para 6	

▶ Representative figures

1.	Are any representative images shown (including Western blots and
	immunohistochemistry/staining) in the naner?

If so, what figure(s)?

6. Figure 7a

2. For each representative image, is there a clear statement of how many times this experiment was successfully repeated and a discussion of any limitations in repeatability?

If so, where is this reported (section, paragraph #)?

N/A

Figure 1a,b,c,d
 Figure 2c
 Figure 4a

4. Figure 5a,c5. Figure 6a

▶ Statistics and general methods

1. Is there a justification of the sample size?

If so, how was it justified?

Where (section, paragraph #)?

Even if no sample size calculation was performed, authors should report why the sample size is adequate to measure their effect size.

2. Are statistical tests justified as appropriate for every figure?

Where (section, paragraph #)?

Yes, methods, section "statistical analysis"

yes, methods, section "statistical analysis"

	a.	If there is a section summarizing the statistical methods in the methods, is the statistical test for each experiment clearly defined?	yes, methods, section "statistical analysis"
	b.	Do the data meet the assumptions of the specific statistical test you chose (e.g. normality for a parametric test)?	all tests are non-parametric
		Where is this described (section, paragraph #)?	
	C.	Is there any estimate of variance within each group of data?	all tests are non-parametric
		Is the variance similar between groups that are being statistically compared?	
		Where is this described (section, paragraph #)?	
	d.	Are tests specified as one- or two-sided?	all tests are two-sided (methods, section "statistical analysis")
	e.	Are there adjustments for multiple comparisons?	no
	С.	The there adjustments for matter companisons.	
3.	Are crite	ria for excluding data points reported?	N/A
	Was this	criterion established prior to data collection?	
	Where is	this described (section, paragraph #)?	
	D (; '1		(,
4.		ne method of randomization used to assign subjects (or to the experimental groups and to collect and process data.	N/A
	If no rand	domization was used, state so.	
	Where d	oes this appear (section, paragraph #)?	
5.		ement of the extent to which investigator knew the group n during the experiment and in assessing outcome included?	N/A
	If no blin	ding was done, state so.	
	Where (s	section, paragraph #)?	
6.		riments in live vertebrates, is a statement of compliance with uidelines/regulations included?	yes, methods, section "Electrodes, surgery and data acquisition"
	Where (s	section, paragraph #)?	
7.	Is the spe	ecies of the animals used reported?	yes, methods, section "Electrodes, surgery and data acquisition"
	Where (s	section, paragraph #)?	
8.		ain of the animals (including background strains of KO/ic animals used) reported?	yes, methods, section "Electrodes, surgery and data acquisition"
	Where (s	section, paragraph #)?	
9.	Is the sex	x of the animals/subjects used reported?	no, 2 females and 5 males
	Where (s	section, paragraph #)?	

10.	Is the age of the animals/subjects reported?	no, aged 3-12 months
	Where (section, paragraph #)?	
11.	For animals housed in a vivarium, is the light/dark cycle reported?	yes, methods, section "Recording sessions and behavioral
	Where (section, paragraph #)?	paradigm"
	where (section, paragraph #7:	
12	For animals housed in a vivarium, is the housing group (i.e. number of	no, housed individually after surgery.
12.	animals per cage) reported?	no, noused marvidually after surgery.
	Where (section, paragraph #)?	
	where (section, paragraph #7:	
13	For behavioral experiments, is the time of day reported (e.g. light or	yes, methods, section "Recording sessions and behavioral
15.	dark cycle)?	paradigm"
	Where (section, paragraph #)?	
	where (section, paragraph #7:	
14	Is the previous history of the animals/subjects (e.g. prior drug	N/A
17.	administration, surgery, behavioral testing) reported?	
	Where (section, paragraph #)?	
	where (section, paragraph n).	
	a. If multiple behavioral tests were conducted in the same	N/A
	group of animals, is this reported?	
	Where (section, paragraph #)?	
15.	If any animals/subjects were excluded from analysis, is this reported?	N/A
	Where (section, paragraph #)?	
	a. How were the criteria for exclusion defined?	N/A
		IN/A
	Where is this described (section, paragraph #)?	
	 Specify reasons for any discrepancy between the number of animals at the beginning and end of the study. 	N/A
	Where is this described (section, paragraph #)?	
F	Reagents	
1	Have antibodies been validated for use in the system under study.	
Ι.	Have antibodies been validated for use in the system under study (assay and species)?	
	a. Is antibody catalog number given?	
	Where does this appear (section, paragraph #)?	

b. Where were the validation data reported (citation supplementary information, Antibodypedia)?	١,
Where does this appear (section, paragraph #)?	
2. If cell lines were used to reflect the properties of a particula disease state, is their source identified?	ir tissue or
Where (section, paragraph #)?	
 a. Were they recently authenticated? 	
Where is this information reported (section, parag	graph #)?
▶ Data deposition	
Data deposition in a public repository is mandatory for:	
a. Protein, DNA and RNA sequences b. Macromolecular structures	
c. Crystallographic data for small molecules d. Microarray data	
	or which structured public repositories exist; more details on our data policy are
	a in supplementary information or in unstructured repositories such as Figshare
1. Are accession codes for deposit dates provided?	N/A
Where (section, paragraph #)?	
> Commutan and a fathering	
► Computer code/software	
Any custom algorithm/software that is central to the methods m time of publication. However, referees may ask for this informat	nust be supplied by the authors in a usable and readable form for readers at the tion at any time during the review process.
Identify all custom software or scripts that were required to	o conduct N/A
the study and where in the procedures each was used.	
2. Is computer source code/software provided with the paper deposited in a public repository? Indicate in what form this	
or how it can be obtained.	is provided
▶ Human subjects	
Which IRB approved the protocol?	
Where is this stated (section, paragraph #)?	
2. Is demographic information on all subjects provided?	
2. Is defined up in a matter of an subjects provided.	

3.	Is the number of human subjects, their age and sex clearly defined?	
	Where (section, paragraph #)?	
4.	Are the inclusion and exclusion criteria (if any) clearly specified?	
	Where (section, paragraph #)?	
5.	How well were the groups matched?	
	Where is this information described (section, paragraph #)?	
6.	Is a statement included confirming that informed consent was obtained from all subjects?	
	Where (section, paragraph #)?	
	Where (seedon, paragraph n/).	
7.	For publication of patient photos, is a statement included confirming	
	that consent to publish was obtained?	
	Where (section, paragraph #)?	
	fMRI studies	
	r papers reporting functional imaging (fMRI) results please ensure that th ormation is clearly provided in the methods:	ese minimal reporting guidelines are met and that all this
1.	Were any subjects scanned but then rejected for the analysis after the data was collected?	
	a. If yes, is the number rejected and reasons for rejection described?	
	Where (section, paragraph #)?	
2.	Is the number of blocks, trials or experimental units per session and/ or subjects specified?	
	Where (section, paragraph #)?	
	where (section, paragraph #):	
3.	Is the length of each trial and interval between trials specified?	
	·	
4.	Is a blocked, event-related, or mixed design being used? If applicable, please specify the block length or how the event-related or mixed design was optimized.	
5.	Is the task design clearly described?	
	Where (section, paragraph #)?	
	Coccom barabraku nj.	
6.	How was behavioral performance measured?	
	Is an ANOVA or factorial design being used?	

8.	For data acquisition, is a whole brain scan used?	
	If not, state area of acquisition.	
	a. How was this region determined?	
0		
9. 1	Is the field strength (in Tesla) of the MRI system stated?	
	 a. Is the pulse sequence type (gradient/spin echo, EPI/spiral) stated? 	
	b. Are the field-of-view, matrix size, slice thickness, and TE/TR/ flip angle clearly stated?	
10.	Are the software and specific parameters (model/functions, smoothing kernel size if applicable, etc.) used for data processing and pre-processing clearly stated?	
11.	Is the coordinate space for the anatomical/functional imaging data clearly defined as subject/native space or standardized stereotaxic space, e.g., original Talairach, MNI305, ICBM152, etc? Where (section, paragraph #)?	
12.	If there was data normalization/standardization to a specific space template, are the type of transformation (linear vs. nonlinear) used and image types being transformed clearly described? Where (section, paragraph #)?	
13.	How were anatomical locations determined, e.g., via an automated labeling algorithm (AAL), standardized coordinate database (Talairach daemon), probabilistic atlases, etc.?	
14.	Were any additional regressors (behavioral covariates, motion etc) used?	
15.	Is the contrast construction clearly defined?	
16.	Is a mixed/random effects or fixed inference used?	
	a. If fixed effects inference used, is this justified?	
17.	Were repeated measures used (multiple measurements per subject)?	
	a. If so, are the method to account for within subject correlation and the assumptions made about variance clearly stated?	
18.	If the threshold used for inference and visualization in figures varies, is this clearly stated?	
19.	Are statistical inferences corrected for multiple comparisons?	

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a. If not, is this labeled as uncorrected?	
20. Are the results based on an ROI (region of interest) analysis?	
a. If so, is the rationale clearly described?	
b. How were the ROI's defined (functional vs anatomical localization)?	
21. Is there correction for multiple comparisons within each voxel?	
22. For cluster-wise significance, is the cluster-defining threshold and the corrected significance level defined?	
▶ Additional comments	
Additional Comments	