

Figure S1. Protein abundance compared with mRNA-seq and ribosome profile data

Three results of RNA quantification among 55 genes were compared with the amount of the corresponding protein, and their correlation coefficients are shown. RT-PCR ($D = 0.7 \text{ h}^{-1}$) and protein quantification data ($D = 0.5$ and 0.7 h^{-1}) are as those from Ishii et al., and the values were converted to molecule/cell. RPKMcc for the genes analyzed using mRNA-seq and RP ($D = 0.6 \text{ h}^{-1}$) are those of the present study.

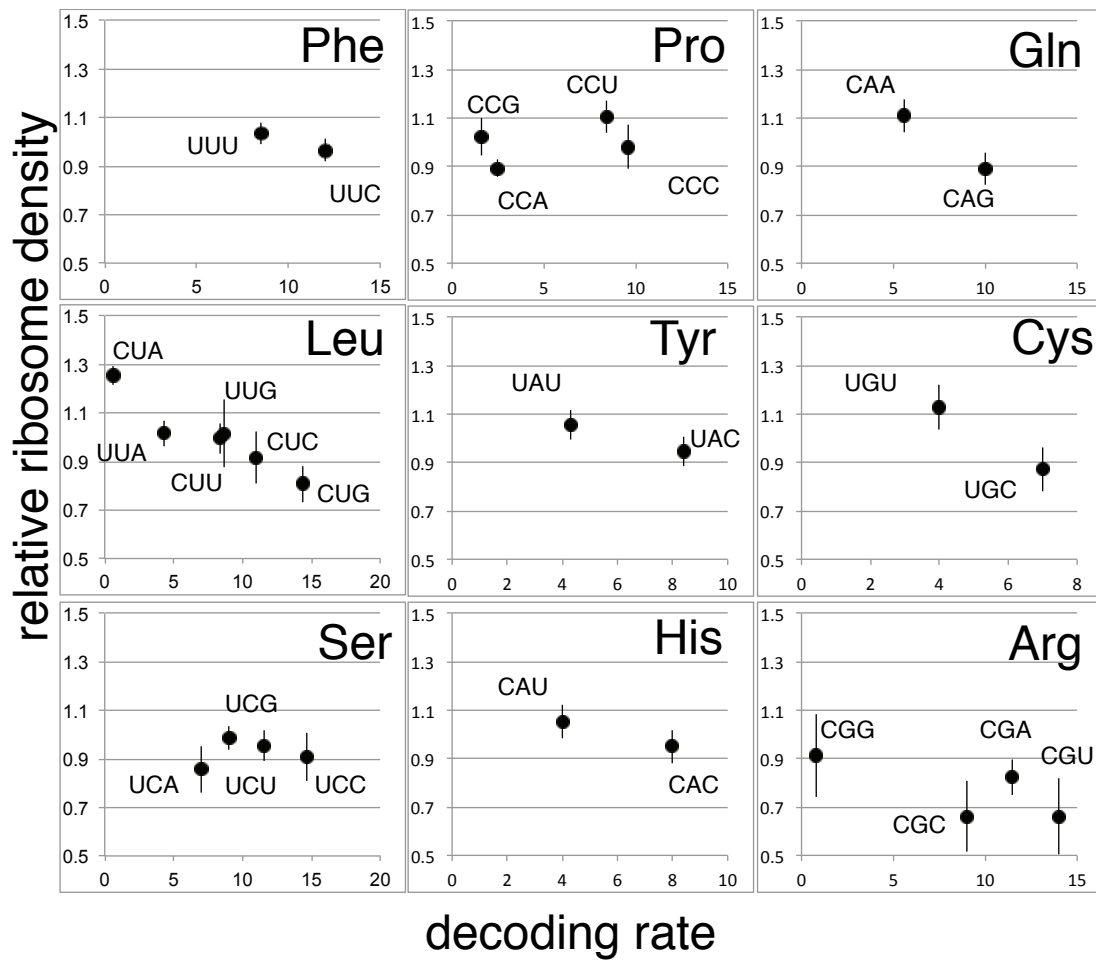


Figure S2. A-site ribosome density compared with decoding rate

The relative A-site ribosome density (shown in Figure 5, Y-axes) was compared with the RtRNA/Rshift value (decoding rate, Curran and Yarus [10], X-axes) . Only YNN codons that could be analyzed using their system are shown.

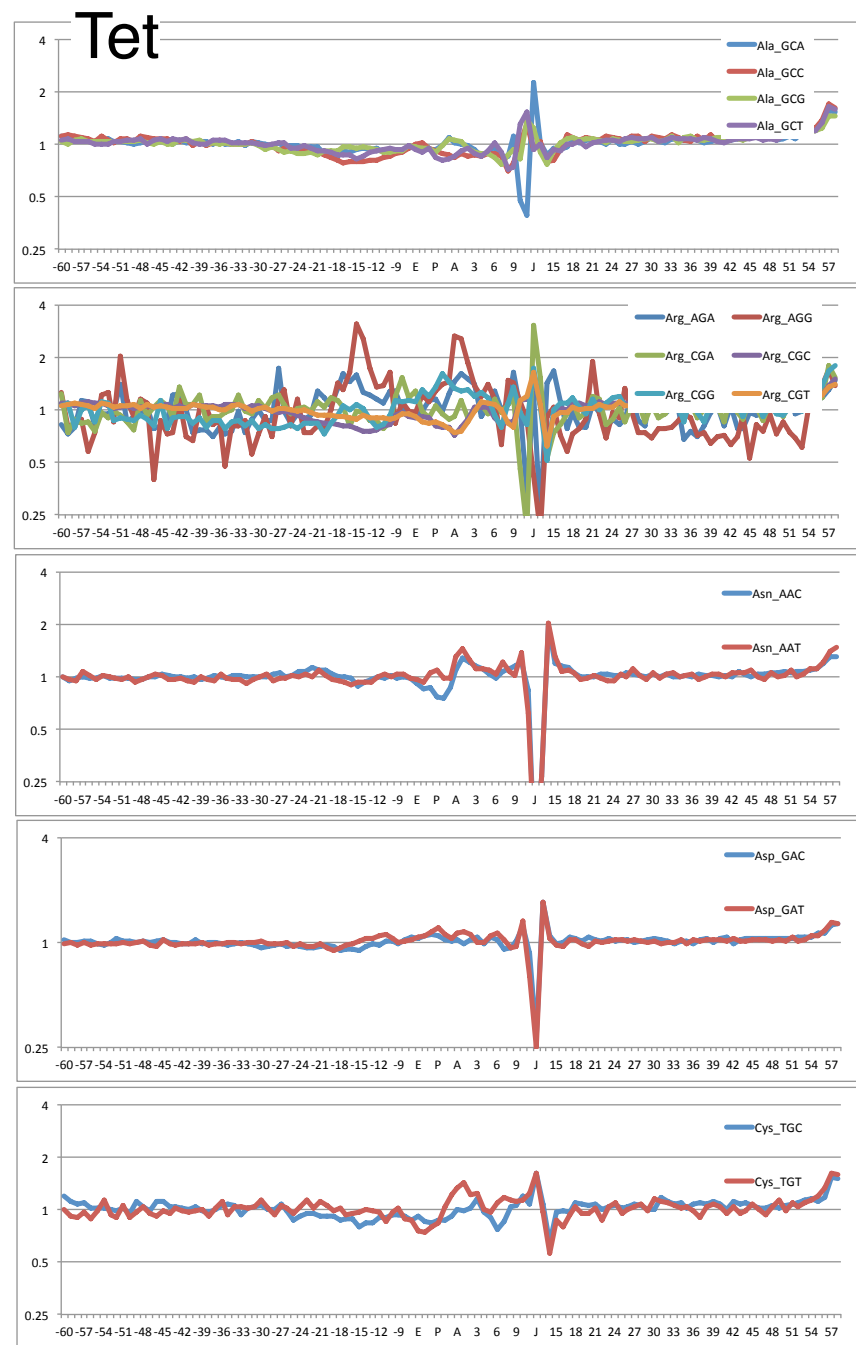
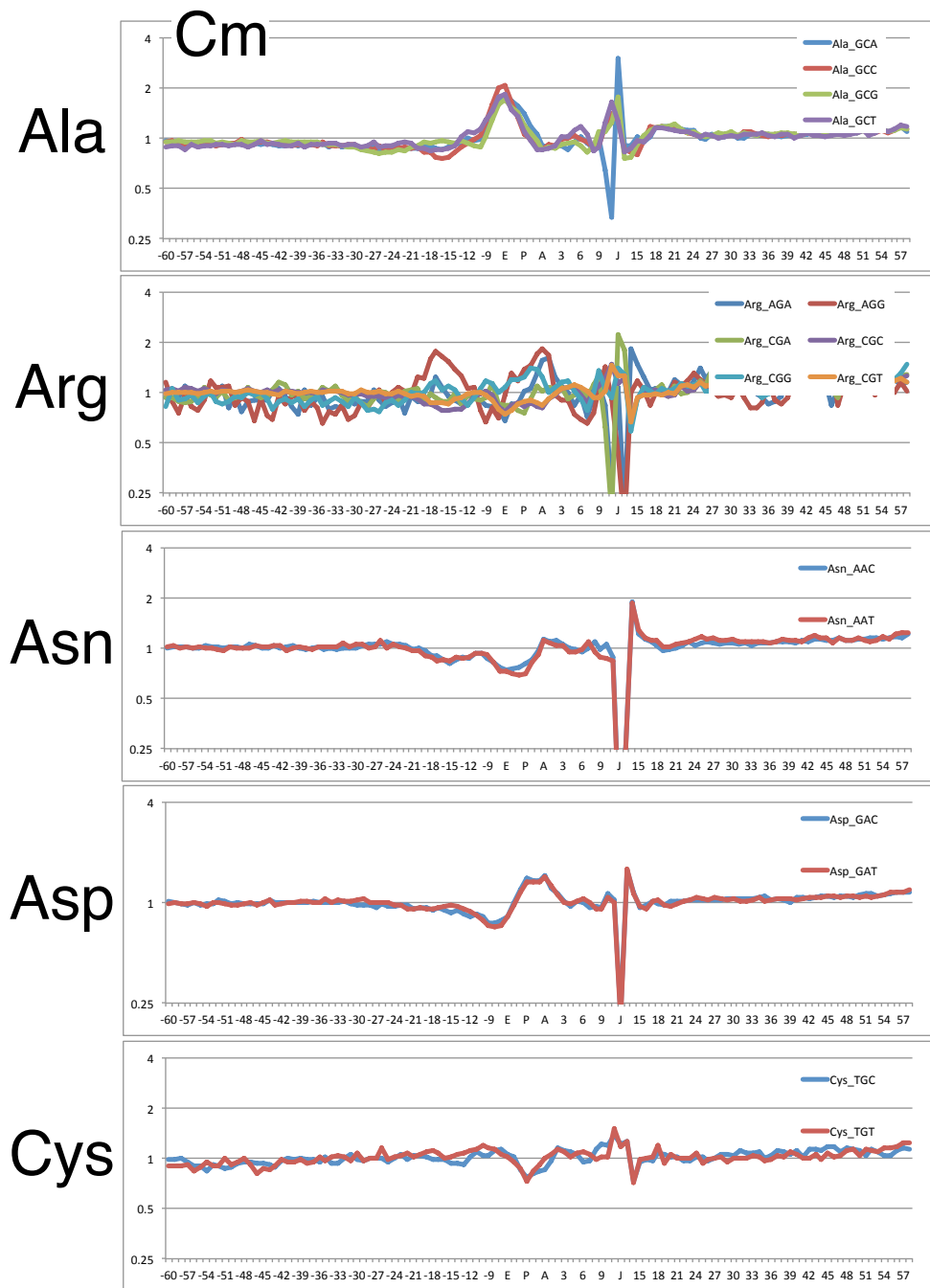


Figure S3 (1/4) Relative ribosome density of each codon at different locations.

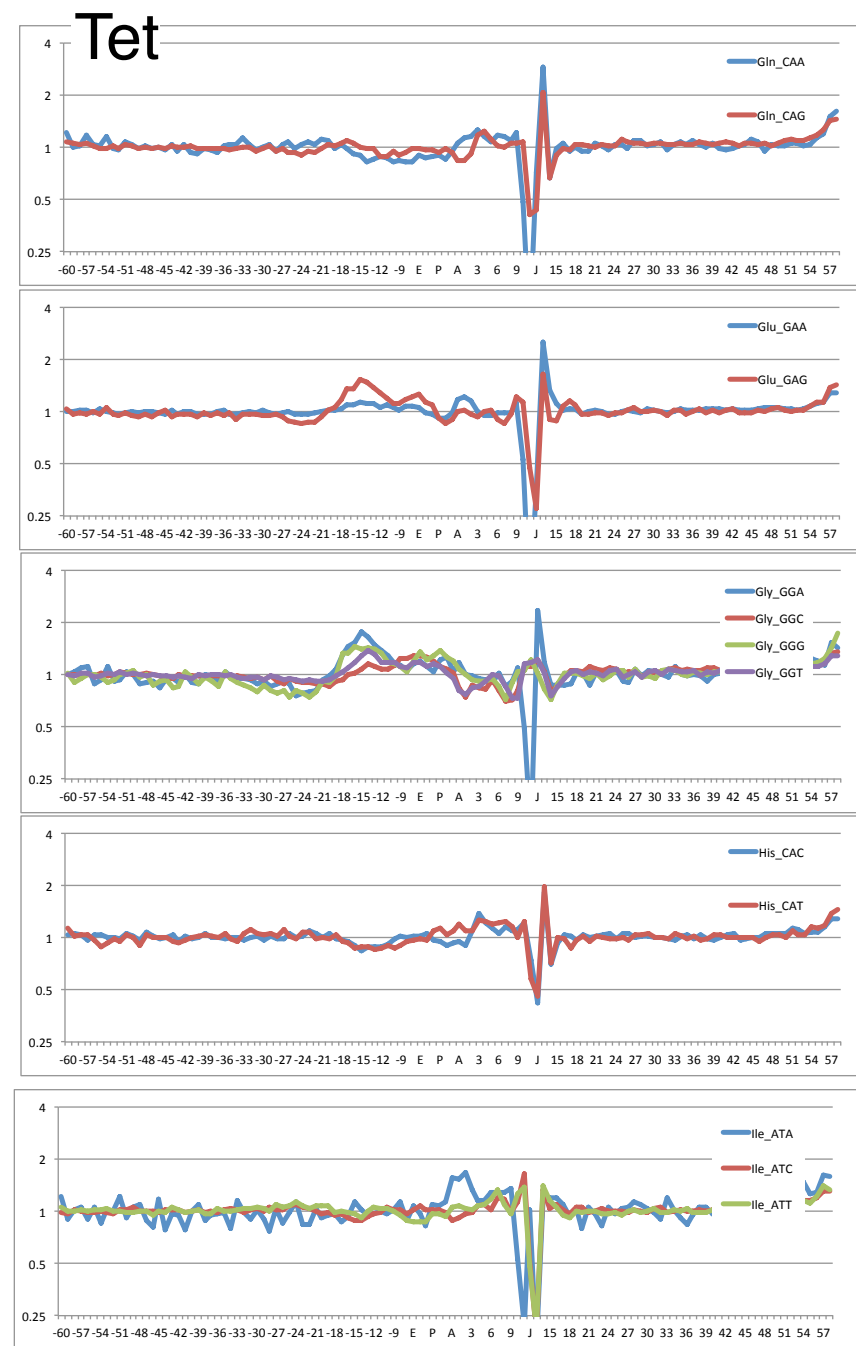
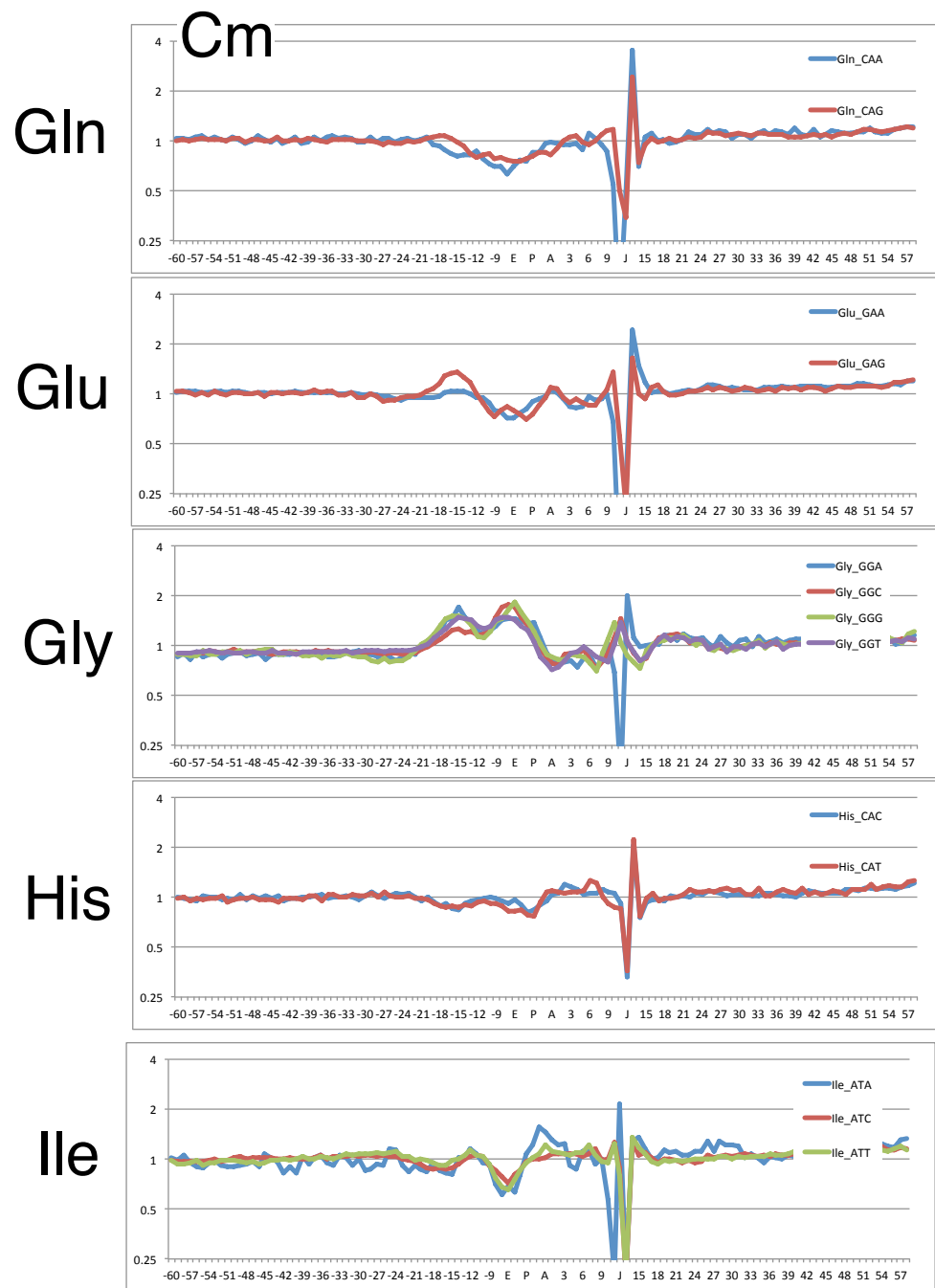


Figure S3 (2/4) Relative ribosome density of each codon at different locations.

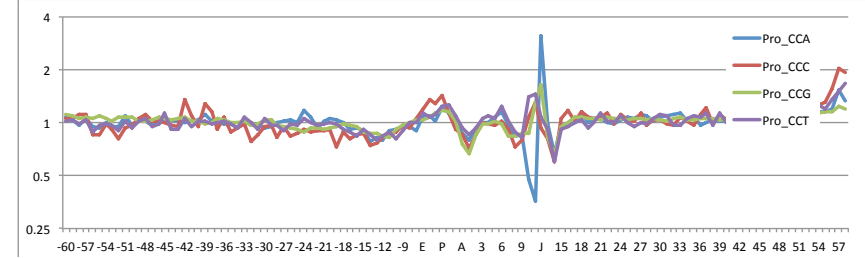
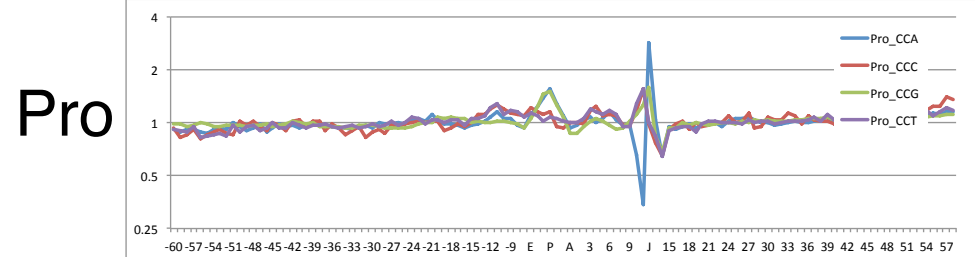
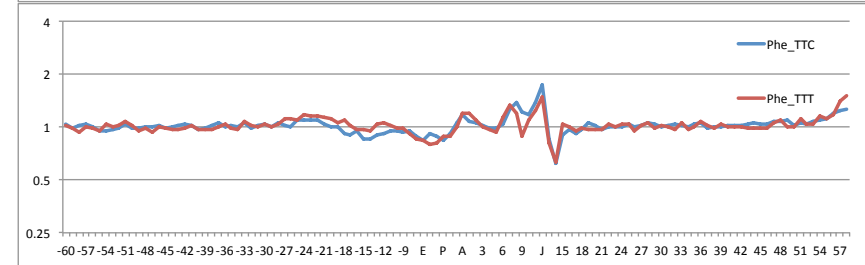
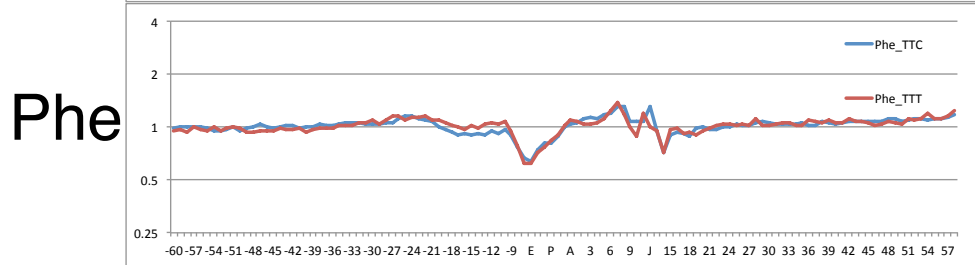
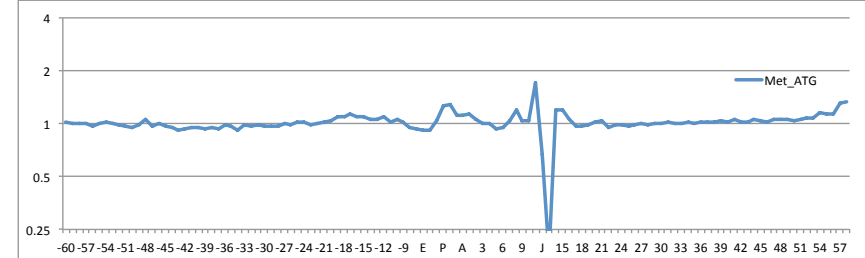
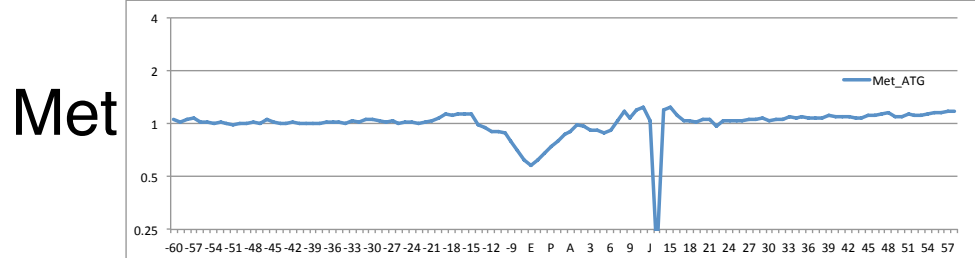
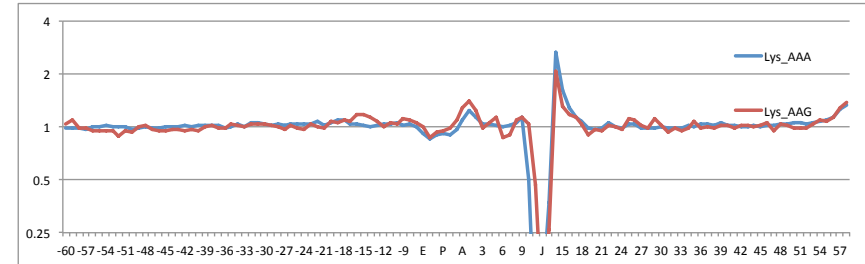
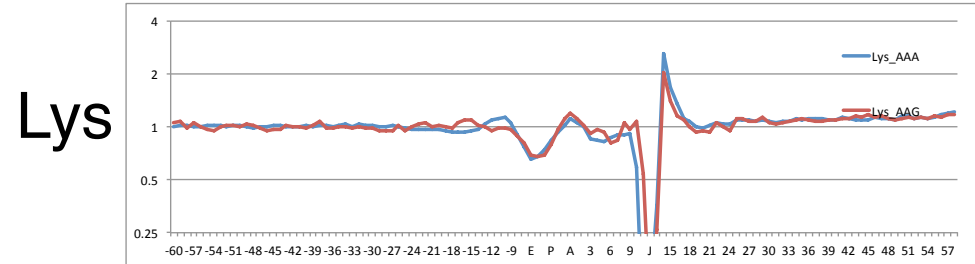
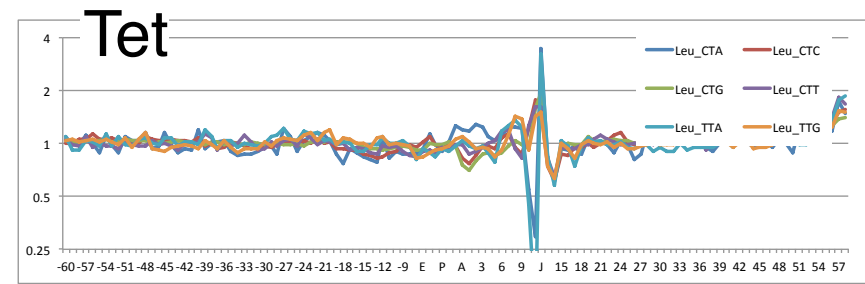
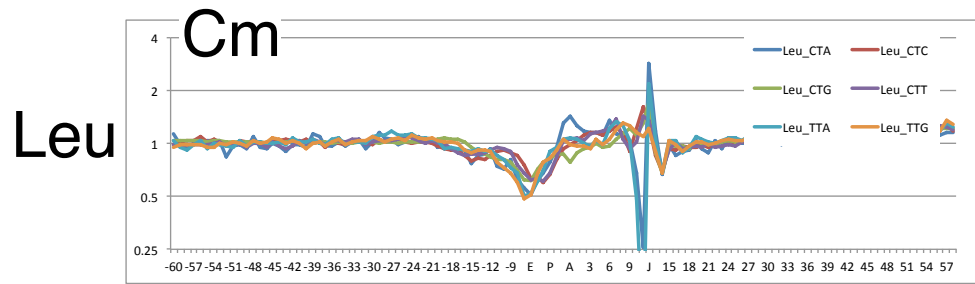


Figure S3 (3/4) Relative ribosome density of each codon at different locations.

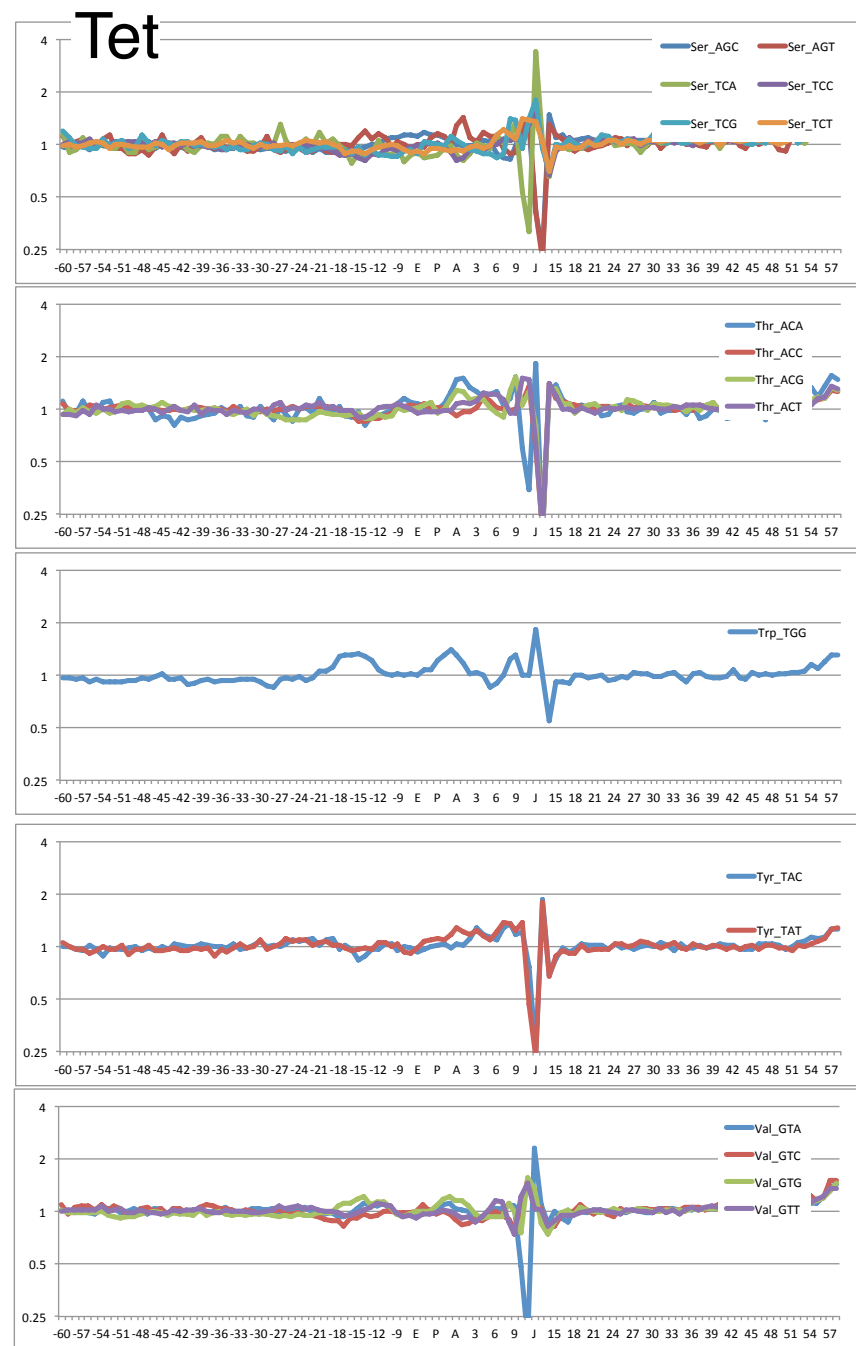
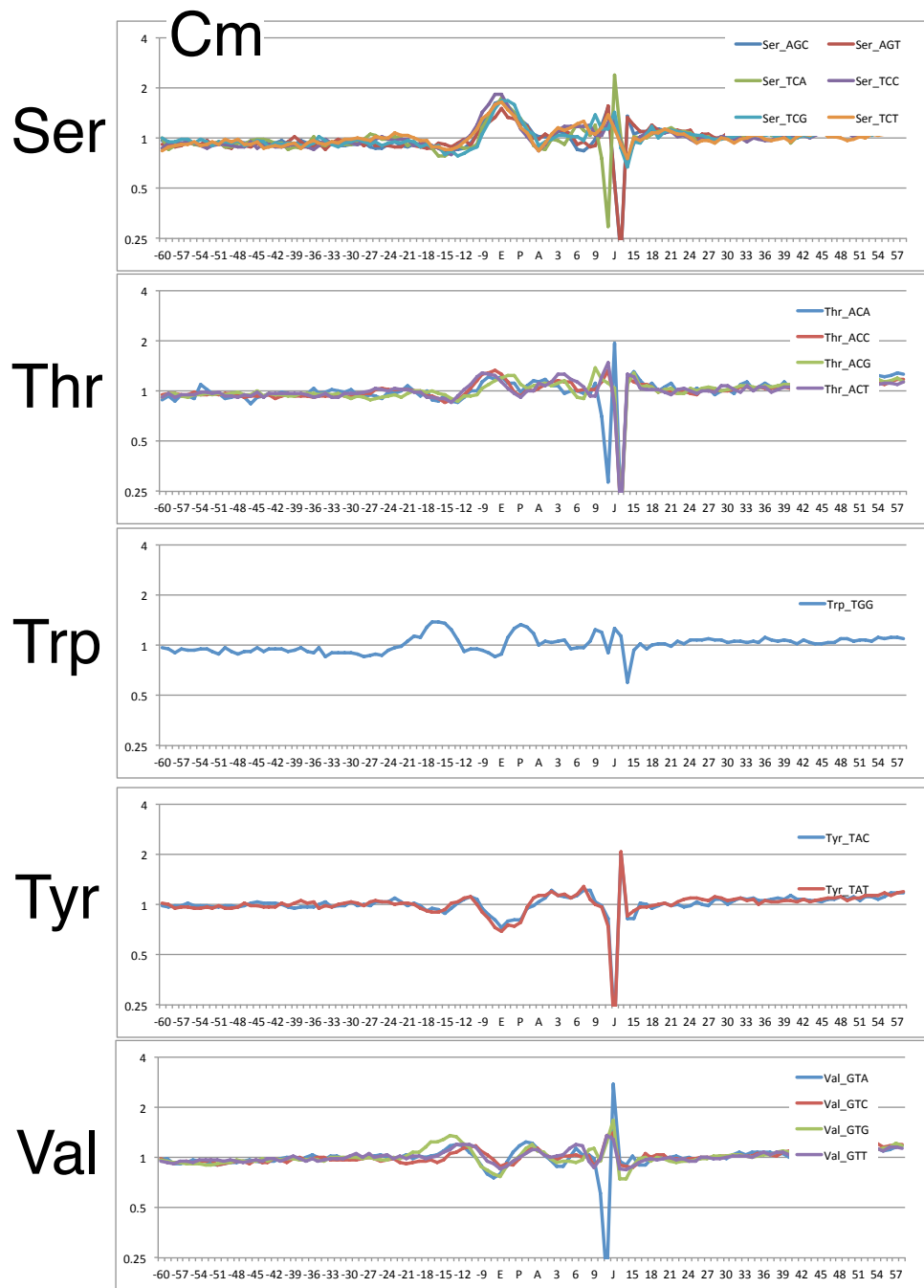


Figure S3 (4/4) Relative ribosome density of each codon at different locations.

Figure S3. Local ribosome densities of codons located at each position
Local ribosome density of each of 61 in-frame coding codons when that is located at each position was shown. One panel shows codons encoding an amino acid and samples taken from cultures treated with Cm or Tet. The x-axis shows the nucleotide position, and the first position of codon at A-site is designated as zero. Positions corresponding to ribosomal A, P, and E-site and the 3' end of the sequence read (J) are marked.

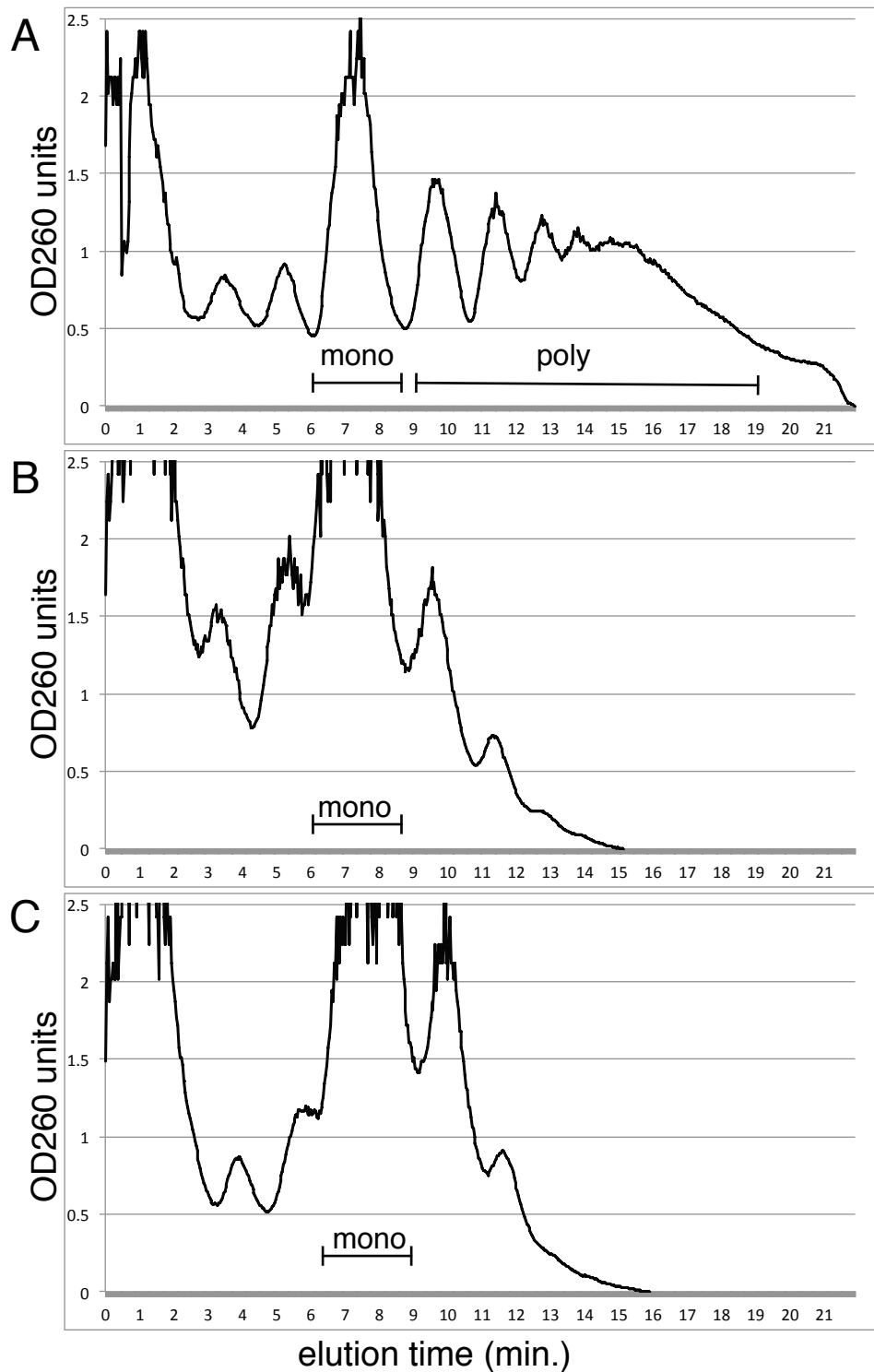


Figure S4. Sucrose density gradient pattern of ribosome fraction

Sucrose density gradient pattern of ribosome fraction without RNase treatment (A), or treated by RNaseI (B) or MNase (C). After the sucrose gradient centrifugation, solution was eluted from the top (light) and OD260 was monitored. Monosome (mono) and polysome (poly) sections were indicated.

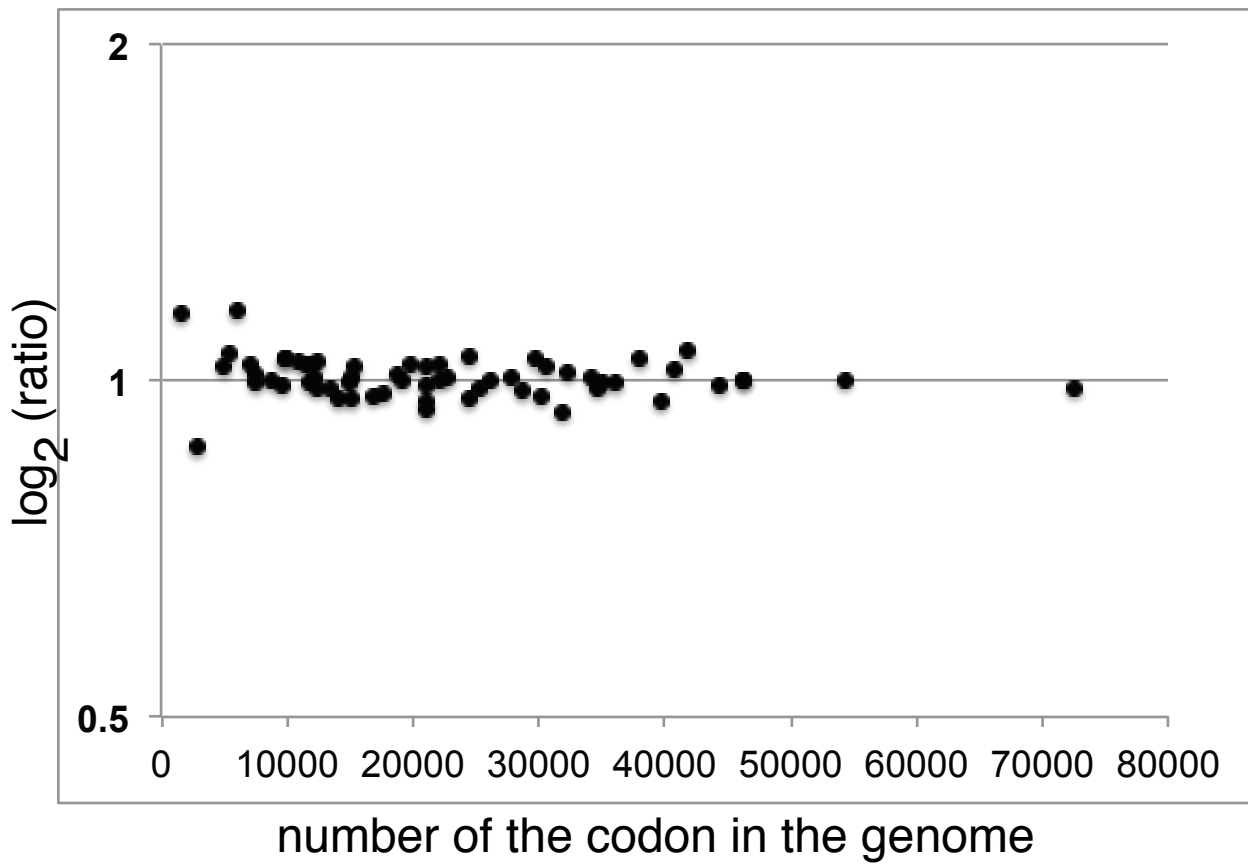


Figure S5. Ribosome density of each codon at A-site was calculated using all the sequence reads or reads whose 3'-end was not mapped to the position followed by A. Then the ratio of the two results for a codon was plotted against usage of the codon in the genome.

Table S1 RNA and protein abundance

| Gene | RNA (qRT-PCR, Ishii et al, D=0.7/h, molecule/cell)*1 | RNA (mRNA-seq, DPB, D=0.6/h)*2 | RNA(RP, DPB, D=0.6/h)*2 | protein(molecule/cell, ishii et al, D=0.5/h)*1 | protein(molecule/cell, ishii et al, D=0.7/h)*1 |
|-------------|--|--------------------------------|-------------------------|--|--|
| <i>aceA</i> | 73.57 | 2011.1 | 2268.6 | 23607.3 | 11076.0 |
| <i>aceB</i> | 73.81 | 2226.0 | 1627.6 | 10445.3 | 9376.7 |
| <i>aceE</i> | 69.88 | 1944.9 | 1626.7 | 3212.6 | 2490.8 |
| <i>aceF</i> | 23.53 | 1375.2 | 1390.2 | 1015.0 | 1193.4 |
| <i>ackA</i> | 10.75 | 192.4 | 408.8 | 6253.7 | 6899.4 |
| <i>acnA</i> | 9.22 | 143.1 | 189.8 | ND *3 | 209.7 |
| <i>acnB</i> | 48.77 | 559.9 | 1311.5 | 834.9 | 938.2 |
| <i>acs</i> | 6.02 | 46.8 | 54.6 | ND | 2399.7 |
| <i>adhE</i> | 46.35 | 534.8 | 1333.6 | 192.1 | 705.6 |
| <i>adk</i> | 47.70 | 1030.0 | 1749.0 | 719.2 | 1339.9 |
| <i>eda</i> | 8.68 | 365.6 | 874.8 | 7629.5 | 6396.5 |
| <i>eno</i> | 66.38 | 2685.6 | 4290.7 | 15018.2 | 27470.3 |
| <i>fbaA</i> | 46.20 | 2029.0 | 2787.7 | 3929.3 | 7244.8 |
| <i>fbp</i> | 7.18 | 239.4 | 254.0 | 1152.2 | 1472.6 |
| <i>frdA</i> | 1.73 | 39.6 | 38.4 | 531.3 | 129.9 |
| <i>fumA</i> | 19.15 | 121.6 | 183.2 | 758.5 | 588.9 |
| <i>fumB</i> | 0.35 | 3.7 | 6.1 | 85.9 | 656.9 |
| <i>fumC</i> | 2.98 | 54.4 | 78.1 | 223.1 | 387.6 |
| <i>galM</i> | 8.27 | 386.1 | 318.3 | 371.9 | 616.7 |
| <i>gapA</i> | 143.33 | 3693.2 | 12608.2 | 33987.3 | 47941.4 |
| <i>glcB</i> | 3.26 | 54.6 | 46.5 | 397.3 | 181.7 |
| <i>gltA</i> | 75.85 | 991.6 | 1284.2 | 3870.6 | 2494.3 |
| <i>gnd</i> | 53.37 | 2305.5 | 2267.0 | 1593.2 | 1852.3 |
| <i>gpmA</i> | 31.80 | 2650.2 | 3411.7 | 9979.3 | 10017.4 |
| <i>icd</i> | 122.67 | 3943.4 | 5741.2 | 8619.3 | 12662.7 |
| <i>ldhA</i> | 4.00 | 137.0 | 194.1 | 716.8 | 1163.4 |
| <i>lldD</i> | 1.25 | 38.9 | 64.6 | 630.1 | 1233.9 |
| <i>lpd</i> | 67.61 | 1574.7 | 2074.7 | 7291.0 | 8810.4 |
| <i>mdh</i> | 73.01 | 1755.4 | 3783.5 | ND | 2440.9 |
| <i>pck</i> | 4.81 | 84.6 | 136.9 | 1650.8 | 533.9 |
| <i>pfkB</i> | 6.96 | 57.7 | 325.5 | 385.8 | 1880.1 |
| <i>pflA</i> | 2.94 | 103.8 | 162.1 | 96.5 | 224.7 |
| <i>pflB</i> | 49.08 | 796.6 | 1185.2 | 1647.1 | 1291.0 |
| <i>pgi</i> | 20.49 | 650.4 | 796.6 | 1097.5 | 2164.9 |
| <i>pgk</i> | 46.83 | 1126.6 | 2337.3 | 5043.8 | 6850.6 |
| <i>pgm</i> | 12.47 | 358.1 | 328.5 | 861.4 | 473.1 |
| <i>ppc</i> | 40.05 | 581.9 | 767.0 | 1152.3 | 3597.3 |
| <i>pps</i> | 8.08 | 175.1 | 169.3 | 258.7 | 418.3 |
| <i>prpC</i> | 0.04 | 38.9 | 64.6 | 297.4 | 222.5 |
| <i>pta</i> | 3.63 | 148.1 | 311.1 | 206.1 | 398.5 |
| <i>ptsH</i> | 85.61 | 3005.3 | 7825.4 | 66387.7 | 86935.8 |
| <i>ptsI</i> | 44.94 | 1458.7 | 1247.4 | 1686.8 | 2992.0 |
| <i>pykA</i> | 9.06 | 200.6 | 427.3 | 943.7 | 584.0 |
| <i>pykF</i> | 24.91 | 723.2 | 1425.3 | 1958.7 | 2862.1 |
| <i>rpiA</i> | 7.52 | 681.9 | 1010.9 | 633.6 | 469.2 |
| <i>sdhA</i> | 39.26 | 496.8 | 439.8 | 1890.3 | 3383.7 |
| <i>sucB</i> | 65.25 | 624.7 | 1031.6 | 3214.0 | 3438.2 |
| <i>sucC</i> | 65.32 | 846.4 | 1125.4 | 3589.7 | 4206.5 |
| <i>sucD</i> | 18.06 | 833.5 | 1092.1 | 1387.3 | 1977.7 |
| <i>talA</i> | 8.68 | 259.7 | 318.6 | 1425.0 | 2563.0 |
| <i>talB</i> | 41.09 | 616.9 | 2016.6 | 2984.4 | 3127.9 |
| <i>tktA</i> | 28.29 | 543.9 | 1011.5 | 726.8 | 1265.0 |
| <i>tktB</i> | 7.48 | 212.0 | 184.5 | 1136.0 | 1574.2 |
| <i>tpiA</i> | 13.22 | 1341.5 | 2077.8 | 2136.0 | 3304.8 |
| <i>ybhE</i> | 11.13 | 171.2 | 395.3 | 281.1 | 270.2 |

*1 : Ishii N, et al. 2007. *Science* 316(5824): 593-597.

*2 : this work

*3 : not detected

Table S3 Sequenced samples and number of reads mapped to CDS

| Name | source strain | library* ¹ | CDS reads* ² |
|----------|---------------------|-----------------------|-------------------------|
| GAI05_3 | BW25113 <i>smpB</i> | RP(Cm) | 2,538,972 |
| GAI05_5 | BW25113 | RP(Cm) | 3,959,115 |
| GAI_9_4 | BW25113 | RP(Cm) | 2,124,985 |
| GAI_9_2 | BW25113 | RP(Cm)(Mnase) | 6,562,793 |
| GAI06_1 | BW25113 <i>smpB</i> | RNA-seq | 5,127,408 |
| GAI05_8 | BW25113 | RNA-seq | 1,611,231 |
| GAI_8_10 | BW25113 | RNA-seq | 5,967,668 |
| GAI05_4 | BW25113 | RP(Tet) | 1,027,370 |
| GAI07_4 | BW25113 | RP(Tet) | 3,357,792 |
| GAI07_5 | BW25113 | RP(Tet) | 4,498,133 |

*1 Type of library used (RNA-seq or RP), antibiotics used to stop translation (Cm or Tet), and nuclease used for polysome digestion (MNase: if specified, or RNaseI: if not indicated) are shown

*2 Total number of the sequence reads mapped to CDSs