

## SUPPORTING INFORMATION

**Supplemental Figure 1.** Hyperproliferation of PC1 depleted cells is dependent on  $\text{Int}\beta 1$ . Growth of Pkd1 knockdown IMCD3 (A) or MDCK (B) cells is suppressed by the inhibition of  $\text{Int}\beta 1$  expression. Constitutive specific targeting of *Pkd1* and *Int\beta 1* was achieved following transduction of IMCD3 with the using the lentivectors VIRHD/P/siPkd1<sub>10071</sub> or VIRHD/HY/siInt\beta 1<sub>2363</sub> and MDCK with the lentivectors VIRHD/P/siPkd1<sub>3211</sub> or VIRHD/HY/siInt\beta 1<sub>451</sub> followed by selection with and puromycin (2\mu g/ml) or hygromycin (400\mu g/ml), respectively<sup>5,6,55</sup>. shLuc from the VIRHD/P/Luc850 lentivector was used as control. Subscript numbers indicate the position of the first nucleotide of the 19-mer sequence in the shRNA on the target mRNA with respect to the starting ATG. Cell proliferation assays were performed using crystal violet dye. Statistical comparison by two-way ANOVA with Bonferroni post-test: \* $P < 0.0001$ .

**Supplemental Figure 2.** Establishment of immortalized mouse F1 collecting duct cells. The F1 cell line was established following the isolation of renal epithelial cells from kidney papillae of *Pkd1*<sup>fl/fl</sup> mice (B6.129S4-Pkd1<sup>tm2Ggg</sup>/J, Jackson Laboratory) by immortalization with the lentiviral vector VVPW/mTert expressing the murine telomerase reverse transcriptase (mTert) (kindly provided by Dr. Ronald DePinho), similarly to what described by Steele et al.<sup>36</sup>. After three passages, these cells were subjected to high osmolarity medium (700mOsm/kg) for 24 hours to enrich for collecting duct cells. Multiple clones obtained by limiting dilution. (A) Immunodetection of Aqp2 and Tamm-Horsfall glycoprotein (uromodulin: UMOD) in clones 1-12. Positive expression of Aqp2

and absence of UMOD confirmed the collecting duct origin of the cells. Actin expression was determined as loading control on the stripped membrane. B: bulk culture before cloning; I: IMCD3 cells; T: 293T cells; C: UMOD positive urine control. One representative clone (F1) was chosen for subsequent studies. (B) F1 cells form a monolayer with high transepithelial resistance on transwell membrane ( $>1000$  Ohm/cm<sup>2</sup>) characteristics of renal collecting duct epithelia.

**Supplemental Figure 3.** Determination of *Pkd1*<sup>fl/fl</sup> allele inactivation in kidneys. The genomic structure of the first 5 exons of floxed *Pkd1* allele of *Pkd1*<sup>fl/fl</sup> (B6.129S4-*Pkd1*<sup>tm2Ggg/J</sup>) mice is shown indicating the position of the loxP sites at the 5' of exon 2 and exon 5 and the primers used for the PCR detection of the inactivated alleles. The corresponding transcripts originated before (wild-type) or after (excised) the Cre-mediated excision of the floxed region are shown. Semi-quantitative nested PCR. Total RNA was isolated from kidneys of 3-week old mice with the indicated genotypes and cDNA was prepared using the EcoDry Premix (Clontech) according to manufacturer's instructions. A first round PCR was performed for the amplification of the region spanning exon 1 to exon 5 of *Pkd1* using primers E1/F (5'-CGTCAATTGCTCCGGCC-3') and E5/R (5'-TCATGGGCAAAGTAGAAGGG-3'). A second nested PCR was performed on the 1/20<sup>th</sup> of the product of the first PCR to amplify the region spanning exon 1 and the spliced E1-E5 exons using the E1/F and the E1-5/R primers (E1-5/R: 5'-GACATATTCCTCACAGCGCGGT-3'; underlined sequence is on exon 5, italicized sequence is on exon 1). Amplification of the aquaporin-2 gene transcript (using primers: mAqp2/F2: 5'-GCATTGGCACCCCTGGTTCA-3' and mAqp2/R1: 5'-

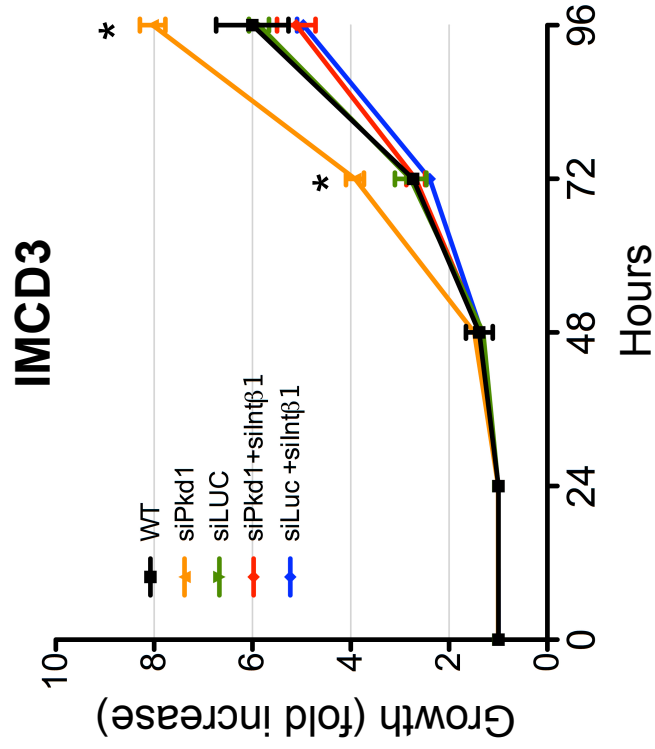
CATTGTTGTGGAGAGCATTGAC-3' was performed in parallel from the same cDNAs to normalize for the number of collecting duct cells. Separation of the amplified products on 2% TBE agarose gel revealed comparable levels of amplified 85bp and the 221bp bands from the excised Pkd1 and the aquaporin-2 gene, respectively, indicating an equally efficient inactivation of the Pkd1 gene in kidneys from single Pkd1-KO and DKO animals.

**Supplemental Figure 4.** Representative images of PAS-stained kidneys of Pkd1-KO and DKO at 6 weeks (top panel) and at 7 months (bottom panel), showing much less variability of cystic phenotype in DKO in comparison to the Pkd1-KO kidneys.

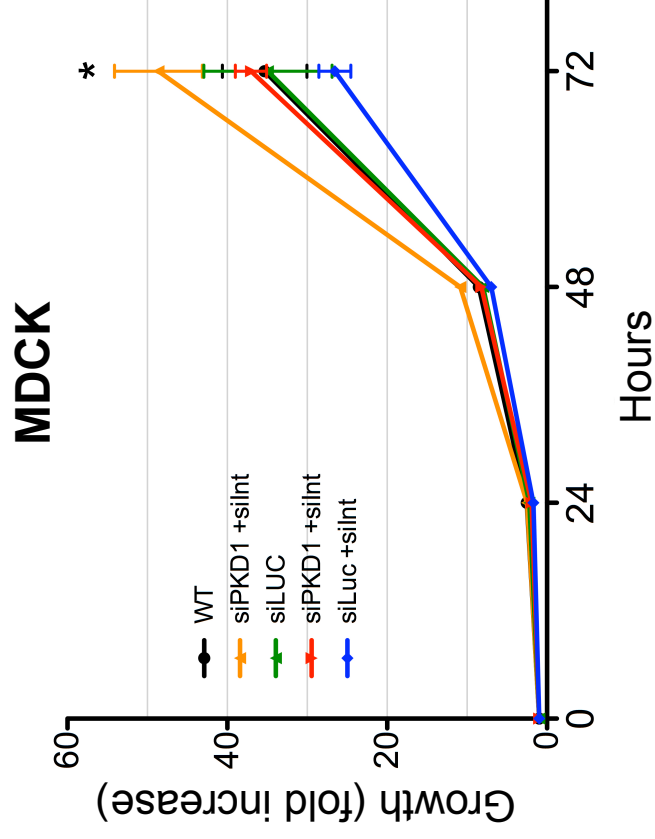
**Supplemental Figure 5.** Cysts form in the collecting ducts. Co-staining with Aqp-2, DBA, LTA, or Tamm-Horsfall (THP) show co-localization of Aqp-2 and DBA with cystic cells but not LTA or THP. Counts were performed on four to eight chosen fields/slide at a magnification of 200X. Over 6 slides per kidney from each genotype (n=3) at 4 weeks of age were scored. Of a count >60 (Pkd1-KO) and >10 (DKO) cysts all were DBA positive.

Supplemental Figure 1

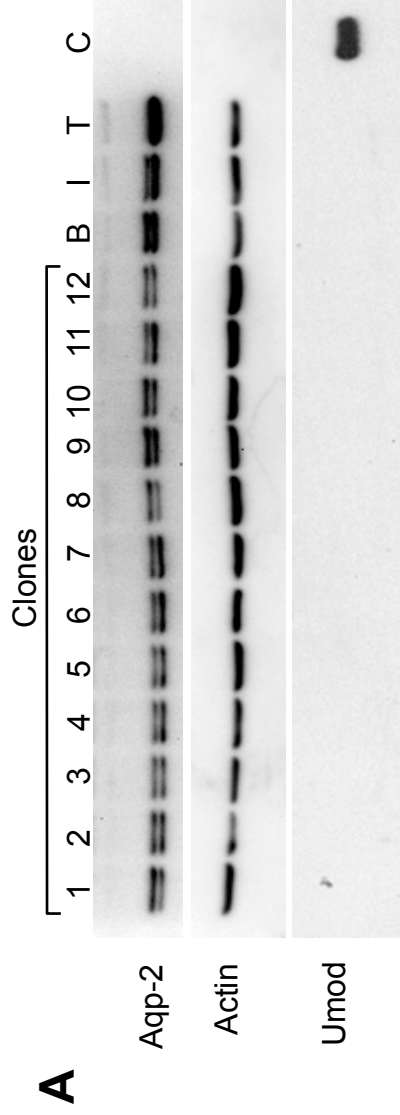
**A**



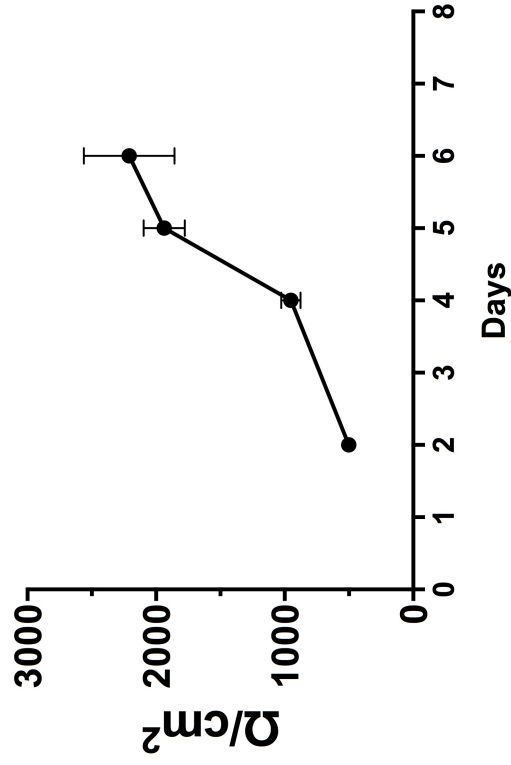
**B**



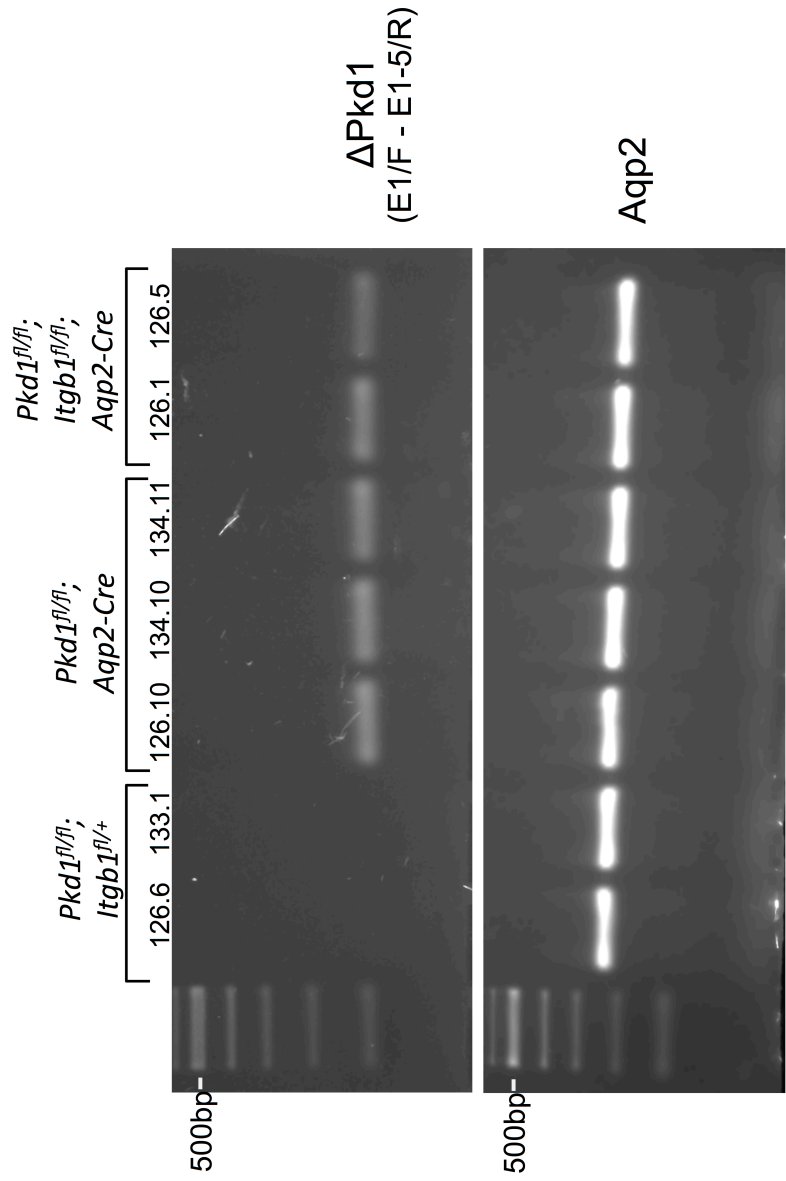
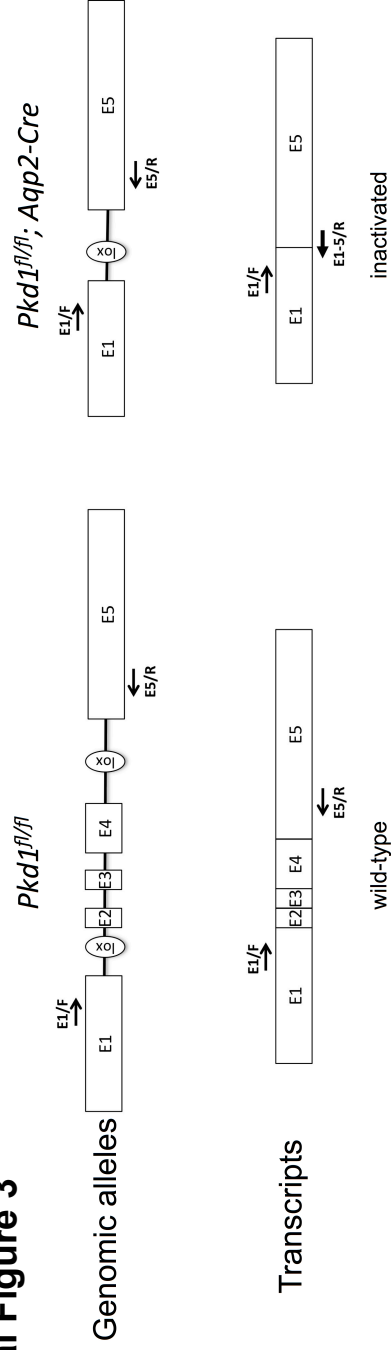
Supplemental Figure 2



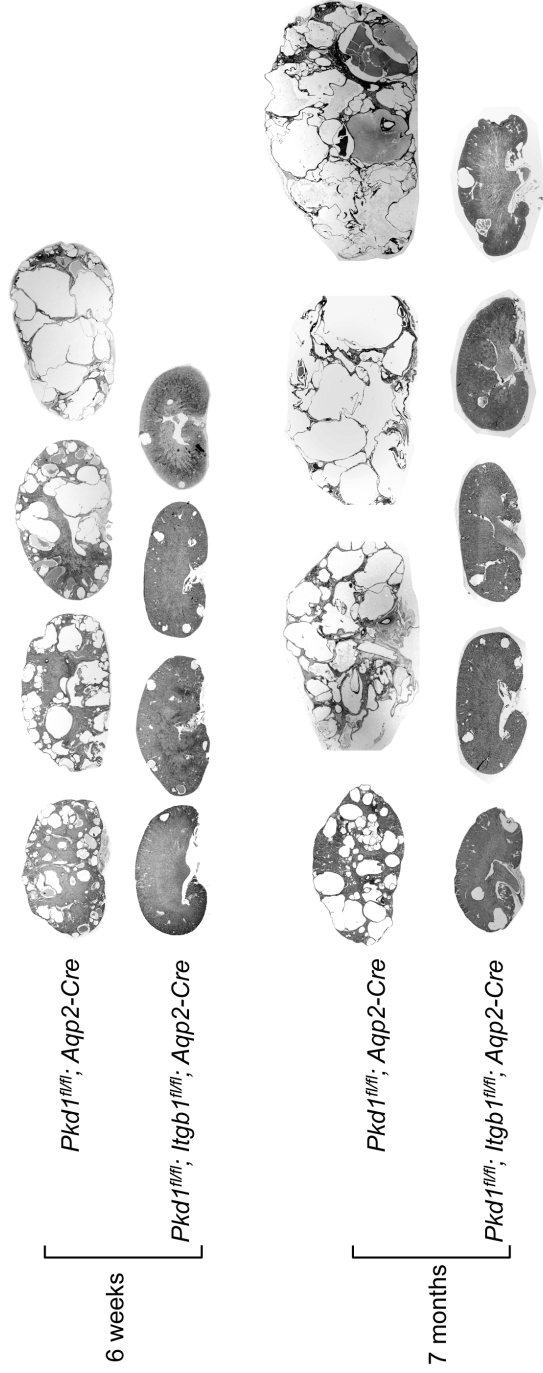
**B** Trans-epithelial resistance of F1 cells



# Supplemental Figure 3



Supplemental Figure 4



Supplemental Figure 5

