Supplementary materials

Supplementary Figure S1: Distribution of full length LTR-type retrotransposons across the nine chromosomes of foxtail millet.

Supplementary Figure S2: Genome-wide prediction of Full length LTR retrotransposon from foxtail genome using LTR finder software.

Supplementary Figure S3: BLAST analysis showing unique junction region for designing the junction based primer.

Supplementary Figure S4: BLAST analysis showing repetitive region in the sequence (highlighted red in color) to demarcate the junction site. Arrow shows one primer from junction site and other primer from other region of the sequence.

Supplementary Figure S5: PCR amplification profile of Repeat Junction Marker (a) dna13398, (b) dna16027, (c) dna19096 and (d) LINE2650. The detailed information for 24 accessions (SN. 25-48) of *Setaria italica* are listed in Supplementary Table S1.

Supplementary Figure S6: Multiple sequence alignment of 88 accessions of *Setaria italica* accessions along with reference genome amplified with LINE2650 primer. The detailed information for 88 accessions of *Setaria italica* are listed in Supplementary Table S1.

Supplementary Figure S7: Multiple sequence alignment of 11 accessions of *Setaria italica* accessions along with reference genome amplified with RJM7 primer. The detailed information for 11 accessions (RJM07:reference genome; IC480117, IC403579, cv. Prasad, cv. Lepakshi, IC403476, GS464, IC404178, IC403521, EC539248, EC539291) of *Setaria italica* are listed in Supplementary Table S1.

Supplementary Figure S8: Distribution of different types of nested TE subclasses.

Supplementary Figure S9: An un-rooted neighbor-joining tree for the 99 individuals of *Setaria italica* alongwith three wild species.

Supplementary Figure S10: Graphical representation showed the detection of the true number of groups *K** resulted after structure analysis.

Supplementary Figure S11: Bar plot of *Setaria italica* of 99 accessions were generated by STRUCTURE 2.3.4 using the admixture model with correlated allele frequencies. Groups are represented by colours.

Supplementary Figure S12: Tutorial of Foxtail millet Transposable Element-based Marker Database.

Supplementary Table S1: List of plant materials used in the present study

Supplementary Table S2: Details of *Copia*-type retrotransposons identified in foxtail millet

Supplementary Table S3: Details of *Gypsy*-type retrotransposons identified in foxtail millet

Supplementary Table S4: Details of partially or solo *Copia*-type retrotransposons identified in foxtail millet

Supplementary Table S5: Details of partially or solo *Gypsy*-type retrotransposons identified in foxtail millet

Supplementary Table S6: Details of *LINEs* (Non-LTR type) identified in foxtail millet

Supplementary Table S7: Details of *SINE*s (Non-LTR type) identified in foxtail millet

Supplementary Table S8: Estimation of insertion time of *Copia*-type transposable elements

Supplementary Table S9: Estimation of insertion time of *Gypsy*-type transposable elements

Supplementary Table S10: Details of DNA transposons identified in foxtail millet

Supplementary Table S11: Presence of transcriptionally active *Copia*-type retrotransposons in various tissues of foxtail millet

Supplementary Table S12: Presence of transcriptionally active *Gypsy*-type retrotransposons in various tissues of foxtail millet

Supplementary Table S13: Presence of transcriptionally active non-LTR type retrotransposons (LINE) in various tissues of foxtail millet

Supplementary Table S14: Presence of transcriptionally active non-LTR type retrotransposons (SINE) in various tissues of foxtail millet

Supplementary Table S15: Presence of transcriptionally active DNA transposons in various tissues of foxtail millet

Supplementary Table S16: Details of RBIP primers designed in foxtail millet

Supplementary Table S17: Details of IRAP primers designed in foxtail millet

Supplementary Table S18: Details of RJM primers designed in foxtail millet

Supplementary Table S19: Details of RJJM primers designed in foxtail millet

Supplementary Table S20: Details of ISBP primers designed in foxtail millet

Supplementary Table S21: Details of REMAP primers designed in foxtail millet

Supplementary Table S22: Primer sequences of 134 transposable elements junction based markers validated in foxtail millet

Supplementary Table S23: The raw STRUCTURE output used for determination of true number of K

Supplementary Table S24: Determination of K using Evanno's method

Supplementary Table S25 Summary of distribution of pure ancestral *Setaria italica* individuals

Supplementary Table S26: Syntenic relationships between transposable elements of foxtail millet and sorghum

Supplementary Table S27: Syntenic relationships between transposable elements of foxtail millet and maize

Supplementary Table S28: Syntenic relationships between transposable elements of foxtail millet and rice **Supplementary Table S29:** Syntenic relationships between transposable elements of foxtail millet and *Brachypodium*

 $\overline{\mathbf{A}}^{\text{E}}$ junction/Fragment

901 TTGATGACATCAGCTAGTTTATCGAATGCTTCAGG

HOME PAGE

Welcome to FMTEMDB

Transposable elements (TE) constitute a significant fraction of plant genomes and are considered to be one of the major forces driving genome evolution. TEs have capability to change its position in the genome and called as "jumping gene" by Barbara McClintock, a renowned geneticist. Each transposition event generates new variability by creating mutations and altering cell's genome size. Retrotransposons are the class I elements which produce RNA intermediates and by the action of reverse transcriptase, they are copied into DNA and then inserted into new locations within the genome, while other TEs class II TEs, or DNA transposons move directly by a "cut and paste" mechanism. Activation of TEs in genomes generate a range of effects, including evolution, structural and functional alterations in gene expression, gene deletion and insertion. Therefore, inactivation of TEs can be crucial for the survival of the host organisms.

Foxtail millet (Setaria italica L.), the second largest cultivated millet species has been recently considered to be a tractable model crop for studying functional genomics, as it possesses a small genome (~515 Mb), short life-cycle, rich genetic diversity (~6,000 varieties), inbreeding nature and closely-related to several bioenergy grasses.

NAVIGATION BAR

The 'SEARCH' tab provides the user a wide-range of search features including searching by chromosome, type of TE, type of primer, ID of TE, chromosomal location and TE family etc.

'SEARCH BY TE ID'

'SEARCH BY TE ID'

'SEARCH BY TE ID'

 $-$ Hag $+$ $1-42,100,000$ bp

'SEARCH BY CHROMOSOME'

'SEARCH BY CHROMOSOME'

-LTR Start

-LTR Length

-LTR Length

Position <u>View</u> <u>R Region Similarit</u>

2147483647

16160720

16166525

Primers

5⁻LTR End

²-LTR Start

3⁻-LTR Length

MOPGI

16160283

 438

16166976

'SEARCH BY CHROMOSOME'

HOW TO VIEW THE DETAILS

HOW TO VIEW THE DETAILS

VIEWING THE DETAILS

Primers Details: Part-L. TE ID: conja1000

Chromosome-wise Map

Comparative map

na1565

TE-marker based comparative mapping data is available for visualization with sorghum, maize, rice and *Brachypodium*

Comparative map

The Syntenic TEs are displayed in RED and the blue lines indicate the orthologs

Download Data

The complete TE & marker data of foxtail millet are available for download in this section.

