Supplementary Materials for

Room-Temperature Enantioselective C–H Iodination via Kinetic Resolution

Ling Chu, Kai-Jiong Xiao, Jin-Quan Yu†

The Scripps Research Institute, 10550 N. Torrey Pines Road, La Jolla, California 92037, USA.

[†]Corresponding author. Email: <u>yu200@scripps.edu</u>

Table of Contents

General Information	S2
Substrates	S3
Experimental Procedures and Characterization of Compounds	S4
Procedures for the Preparation of Substrates	S4
Optimization of Reaction Conditions	S12
General Procedure for Pd(OAc) ₂ -Catalyzed C–H Iodination	S13
Deprotection of Trifluoromethanesulfonamide 31	S27
Synthetic Transformations of the Iodinated Product	S27
NMR Spectra	S31
HPLC Spectra.	S89
X-ray Crystallographic Data of 31	S141
References	

General Information:

Unless otherwise noted all commercial materials were used without further purification. Solvents were obtained from Acros or Sigma-Aldrich and used directly without further purification. Nuclear magnetic resonance (NMR) spectra were recorded with Varian Inova-400, Bruker DRX-600. ¹H and ¹³C chemical shifts are reported in ppm downfield of tetramethylsilane and referenced to residual solvent peak (CHCl₃ = 7.26) unless otherwise noted. Multiplicities are reported using the following abbreviations: s = singlet, d = doublet, t = triplet, q = quartet, m = multiplet, br = broad resonance. High resolution mass spectra for new compounds were recorded on an Agilent LC/MSD TOF mass spectrometer or an Agilent 6230 APCI-TOF mass spectrometer. Enantiomeric excesses (ee) were determined on a Hitachi LaChrow Elite HPLC system using commercially available chiral columns. All amino acids were purchased from Bachem or Sigma-Aldrich or synthesized according to literature procedures. All the substrates were synthesized according to the literature procedures (*31*).

Substrates:



Experimental Procedures and Characterization of Compounds:

Procedures for the Preparation of Substrates

General procedure for preparation of substrates 1b, 1h, 1k:

To a stirred solution of the corresponding amine (5 mmol, 1.0 equiv.) in dichloromethane (20 mL) was added triethylamine (5 mmol, 1.0 equiv.) at -78 °C under nitrogen. After stirring for 10 min at -78 °C, trifluoromethanesulfonic anhydride (5.3 mmol, 1.05 equiv.) was added dropwise and the mixture was stirred for 1 h at that temperature before being quenched by water (20 mL). The organic layer was separated and the aqueous layer was extracted with dichloromethane (10 mL \times 2). The combined organic phase was washed with brine (20 mL), and then dried over MgSO₄. Evaporation and column chromatography on silica gel (EtOAc/hexane = 1:20 as eluent) afforded corresponding trifluoromethanesulfonamide **1b**, **1h**, **1k**.

General procedure for preparation of substrates 1a, 1c, 1d, 1e, 1f, 1g, 1i, 1j, 1l (31)

To a 100 mL round bottle was added the corresponding ketone, $Ti(O^{i}Pr)_{4}$ (2 equiv.) and a 2 M solution of NH_{3} in EtOH (5 equiv.) under nitrogen. The resulting solution was heated to 50 °C and stirred for 24 h. Then the mixture was cooled to room temperature and NaBH₄ was added and the reaction mixture was stirred at room temperature for 6 h. The reaction was quenched with $NH_{4}Cl$ solution and filtered through a pad of Celite. The resulting liquid was then extracted with EtOAc and the aqueous layer was washed twice with EtOAc. The organic layers were combined and extracted with 1 M HCl. The aqueous layer from this extraction was washed once with EtOAc. The organic layers were domined organic layer was washed with sat. aq. NaCl and concentrated under reduced pressure to give the crude amine, which could be used directly into the next step without purification. And the corresponding trifluoromethanesulfonamides **1a**, **1c**, **1d**, **1e**, **1f**, **1g**, **1i**, **1j**, **1i** could be synthesized using the same protocol shown above.

General procedure for preparation of substrates 4a-4g:

To a 100 mL round bottle was added the corresponding β -amino acid and methanol. The mixture was cooled to 0 °C and SOCl₂ (1.5 equiv.) was added dropwise. The solution was then stirred at room temperature for 12 h. After completion, the solution was concentrated under reduced pressure to give the crude ester. The corresponding trifluoromethanesulfonamides **4a-4g** could be synthesized using the same protocol shown above.

General procedure for preparation of substrates 7a-7d:

The amino alcohol was converted to the corresponding triflate amide according to the protocol shown above. The alcohol was protected as the TBS ether according to the protocol shown below.

To a 100 mL round bottle was added the corresponding alcohol, TBS-Cl (1.1 equiv.), DMAP (0.1 equiv.) and DCM. The mixture was cooled to 0 °C and Et₃N (1.1 equiv.) was added dropwise. The solution was warmed to room temperature and stirred for 12 h. Then the reaction was quenched with water. The organic layer was separated and the aqueous layer was washed twice with DCM. The combined organic layer was washed with sat. aq. NaCl and concentrated under reduced pressure to give the crude product, which was purified by column chromatography on silica gel (ethyl acetate/hexane= 1:30 as eluent) to afford **7a-7d**.

1a 1,1,1-Trifluoro-N-(1-(o-tolyl)ethyl)methanesulfonamide

¹H NMR (600 MHz, CDCl₃) δ 7.32 – 7.13 (m, 4H), 5.65 (br s, 1H), 5.03 (q, *J* = 6.8 Hz, 1H), 2.37 (s, 3H), 1.55 (d, *J* = 6.8 Hz, 3H).

¹³C NMR (151 MHz, CDCl₃) δ 139.63, 134.28, 130.86, 127.98, 126.77, 124.57, 119.41 (q, J = 321.1 Hz) 51.56, 23.42, 18.88.

HRMS (ESI-TOF) m/z Calcd for $C_{10}H_{11}F_3NO_2S^-[M-H]^-$ 266.0468, found 266.0469.

1b 1,1,1-trifluoro-N-(1-(o-tolyl)propyl)methanesulfonamide

¹H NMR (600 MHz, CDCl₃) δ 7.27 – 7.13 (m, 4H), 5.73 (s, 1H), 4.78 (m, 1H), 2.36 (s, 3H), 1.95 – 1.77 (m, 2H), 0.93 (t, J = 7.4 Hz, 3H).

¹³C NMR (151 MHz, CDCl₃) δ 138.79, 134.65, 130.71, 127.76, 126.69, 124.78, 119.38 (q, *J* = 321.04 Hz), 57.14, 30.75, 19.04, 10.39.

HRMS (ESI-TOF) *m/z* Calcd for C₁₁H₁₃F₃NO₂S⁻ [M-H]⁻ 280.0625, found 280.0622.



1c 1,1,1-trifluoro-N-(1-(o-tolyl)pentyl)methanesulfonamide

¹H NMR (600 MHz, CDCl₃) δ 7.73 – 6.88 (m, 4H), 4.84 (t, *J* = 7.3 Hz, 1H), 2.36 (s, 3H), 1.90 – 1.72 (m, 2H), 1.47

-1.18 (m, 4H), 0.88 (t, J = 7.2 Hz, 3H).

¹³C NMR (151 MHz, CDCl₃) δ 139.16, 134.48, 130.70, 127.71, 126.70, 124.79, 119.36 (q, *J* = 321.0 Hz), 55.72, 37.46, 27.93, 22.17, 19.02, 13.75.

HRMS (ESI-TOF) *m/z* Calcd for C₁₃H₁₈F₃NO₂SNa⁺ [MNa]⁺ 332.0902, found 332.0899.

1d 1,1,1-trifluoro-N-(3-methyl-1-(o-tolyl)butyl)methanesulfonamide

¹H NMR (600 MHz, CDCl₃) δ 7.27 – 7.13 (m, 4H), 5.46 (d, J = 7.6 Hz, 1H), 4.98 – 4.90 (m, 1H), 2.37 (s, 3H), 1.75 – 1.63 (m, 2H), 1.60 – 1.50 (m, 1H), 0.98 (d, J = 6.4 Hz, 3H), 0.94 (d, J = 6.5 Hz, 3H).

¹³C NMR (151 MHz, CDCl₃) δ 139.31, 134.28, 130.83, 127.76, 126.73, 124.71, 119.30 (q, *J* = 320.9 Hz), 54.06, 47.04, 24.78, 22.80, 21.72, 18.94.

HRMS (ESI-TOF) *m*/*z* Calcd for C₁₃H₁₈F₃NO₂SNa⁺ [MNa]⁺ 322.0902, found 322.0899.

Me NHTf

$1e\ 1, 1, 1- trifluoro-N- (2- phenyl-1- (o-tolyl) ethyl) methane sulfon a mide$

¹H NMR (600 MHz, CDCl₃) δ 7.28 – 7.17 (m, 5H), 7.13 (d, J = 7.6 Hz, 1H), 7.10 (d, J = 7.4 Hz, 1H), 7.01 – 6.94 (m, 2H), 5.36 (s, 1H), 5.11 (m, 1H), 3.16 – 3.07 (m, 2H), 2.16 (s, 3H).

¹³C NMR (151 MHz, CDCl₃) δ 137.96, 135.28, 134.77, 130.69, 129.49, 128.58, 127.99, 127.25, 126.55, 125.17, 119.28 (q, *J* = 321.2 Hz), 56.53, 43.94, 18.87.

HRMS (ESI-TOF) m/z Calcd for $C_{16}H_{16}F_3NO_2SNa^+$ [MNa]⁺ 366.0746, found 366.0745.

1f N-(cyclopropyl(o-tolyl)methyl)-1,1,1-trifluoromethanesulfonamide

¹H NMR (600 MHz, CDCl₃) δ 7.29 – 7.18 (m, 3H), 7.19 – 7.14 (m, 1H), 5.56 (s, 1H), 4.52 (dd, *J* = 7.9 Hz, 7.9 Hz, 1H), 2.35 (s, 3H), 1.41 – 1.30 (m, 1H), 0.73 – 0.64 (m, 1H), 0.58 – 0.50 (m, 1H), 0.37 – 0.26 (m, 2H).

¹³C NMR (151 MHz, CDCl₃) δ 137.26, 134.89, 130.69, 127.96, 126.41, 125.95, 119.41 (q, J = 321.0 Hz), 59.19, 19.25, 17.20, 4.48, 2.70.

HRMS (ESI-TOF) m/z Calcd for $C_{12}H_{14}F_3NO_2SNa^+$ [MNa]⁺ 316.0589, found 316.0587.

OMeNHTf

1g 1,1,1-trifluoro-N-(1-(2-methoxyphenyl)ethyl)methanesulfonamide

¹H NMR (600 MHz, CDCl₃) δ 7.30 (t, *J* = 7.6 Hz, 1H), 7.13 (d, *J* = 7.6 Hz, 1H), 6.94 (m, 2H), 6.13 (d, *J* = 8.8 Hz, 1H), 4.74 (m, 1H), 3.90 (s, 3H), 1.61 (d, *J* = 7.0 Hz, 3H).

¹³C NMR (151 MHz, CDCl₃) δ 156.56, 129.41, 128.70, 128.08, 119.47 (q, *J* = 321.1 Hz), 116.28, 111.28, 55.40, 55.14, 22.95.

HRMS (ESI-TOF) *m/z* Calcd for C₁₀H₁₂F₃NO₃SNa⁺ [MNa]⁺ 306.0382, found 306.0384.



1h 1,1,1-trifluoro-N-(1-(2-fluorophenyl)ethyl)methanesulfonamide

¹H NMR (600 MHz, CDCl₃) δ 7.35 – 7.28 (m, 1H), 7.26 (dd, *J* = 7.1, 6.9 Hz, 1H), 7.14 (d, *J* = 7.1, 7.1 Hz, 1H), 7.12 – 7.00 (m, 1H), 5.78 (br s, 1H), 4.92 (q, *J* = 7.1 Hz, 1H), 1.63 (d, *J* = 7.1 Hz, 3H).

¹³C NMR (151 MHz, CDCl₃) δ 160.09 (d, J = 246.1 Hz), 129.97 (d, J = 8.5 Hz), 128.37 (d, J = 12.8 Hz), 127.68 (d, J = 4.1 Hz), 124.66 (d, J = 3.1 Hz), 119.39 (q, J = 320.8 Hz), 116.09 (d, J = 21.5 Hz), 51.83 (d, J = 1.8 Hz), 23.04 (d, J = 2.0 Hz).

HRMS (ESI-TOF) *m*/*z* Calcd for C₉H₈F₄NO₂SNa⁺ [MNa]⁺ 293.0104, found 293.0104.

$1 i {\it N-(1-(4-chloro-2-methylphenyl)ethyl)-1,1,1-trifluoromethanesulfonamide}$

¹H NMR (600 MHz, CDCl₃) δ 7.20 (s, 2H), 7.17 (s, 1H), 5.75 (s, 1H), 5.02 – 4.95 (m, 1H), 2.35 (s, 3H), 1.53 (d, J = 6.9 Hz, 3H).

¹³C NMR (151 MHz, CDCl₃) δ 138.31, 136.22, 133.47, 130.64, 126.85, 126.11, 119.35 (q, J = 321.1 Hz), 51.11, 23.32, 18.73.

HRMS (ESI-TOF) *m*/*z* Calcd for C₁₀H₁₀ClF₃NO₂S⁻ [M-H]⁻ 300.0078, found 300.0080.

1j 1,1,1-trifluoro-N-(1-(m-tolyl)ethyl)methanesulfonamide

¹H NMR (600 MHz, CDCl₃) δ 7.27 (dd, *J* = 7.5, 7.5 Hz, 1H), 7.18 – 7.07 (m, 3H), 5.63 – 5.37 (br s, 1H), 4.80 – 4.71 (m, 1H), 2.37 (s, 3H), 1.61 (d, *J* = 6.9 Hz, 3H).

¹³C NMR (151 MHz, CDCl₃) δ 140.91, 138.76, 129.03, 128.88, 126.54, 122.81, 119.49 (q, J = 321.0 Hz), 55.32, 23.47, 21.37.

HRMS (ESI-TOF) m/z Calcd for $C_{10}H_{12}F_3NO_2SNa^+$ [MNa]⁺ 290.0433, found 290.0431.

1k 1,1,1-trifluoro-N-(1-(p-tolyl)ethyl)methanesulfonamide

¹H NMR (600 MHz, CDCl₃) δ 7.22 – 7.13 (m, 4H), 5.35 (s, 1H), 4.74 (m, 1H), 2.34 (s, 3H), 1.59 (d, *J* = 7.0 Hz, 3H).

¹³C NMR (151 MHz, CDCl₃) δ 138.13, 137.99, 129.62, 125.75, 119.49 (q, J = 320.8 Hz), 55.14, 23.42, 21.01. HRMS (ESI-TOF) m/z Calcd for C₁₀H₁₂F₃NO₂SNa⁺ [MNa]⁺ 290.0433, found 290.0435.



$11\ 1, 1, 1-trifluoro-N-(1-(naphthalen-2-yl)ethyl) methanesulfon a mide$

¹H NMR (600 MHz, CDCl₃) δ 7.87 – 7.78 (m, 3H), 7.73 (s, 1H), 7.54 – 7.46 (m, 2H), 7.38 (d, *J* = 8.5 Hz, 1H), 5.43 (d, *J* = 7.5 Hz, 1H), 4.99 – 4.89 (m, 1H), 1.68 (d, *J* = 6.9 Hz, 3H).

¹³C NMR (151 MHz, CDCl₃) δ 138.18, 133.14, 132.97, 129.05, 128.01, 127.68, 126.62, 126.49, 124.74, 123.61, 119.50 (q, *J* = 321.0 Hz), 55.42, 23.37.

HRMS (ESI-TOF) m/z Calcd for $C_{13}H_{12}F_3NO_2SNa^+$ [MNa]⁺ 326.0433, found 326.0434.



1m 1,1,1-trifluoro-N-(1-phenylpentyl)methanesulfonamide

¹H NMR (600 MHz, CDCl₃) δ 7.41 – 7.35 (m, 2H), 7.35 – 7.30 (m, 1H), 7.27 – 7.20 (m, 2H), 5.30 (br s, 1H), 4.55 (ddd, J = 7.7, 7.7, 7.7 Hz, 1H), 2.00 – 1.81 (m, 2H), 1.39 – 1.27 (m, 3H), 1.27 – 1.15 (m, 1H), 0.87 (t, J = 7.2 Hz, 3H).

¹³C NMR (151 MHz, CDCl₃) δ 140.26 , 128.97 , 128.24 , 126.09 , 119.39 (q, J = 321.1 Hz), 60.02 , 37.46 , 27.97 , 22.12 , 13.76 .

HRMS (ESI-TOF) m/z Calcd for $C_{12}H_{15}F_3NO_2S^{-}$ [M-H]⁻ 294.0781, found 294.0783.



1n 1,1,1-trifluoro-N-(2-methyl-1-phenylpropyl)methanesulfonamide

1H NMR (600 MHz, CDCl3) δ 7.42 – 7.27 (m, 3H), 7.18 (d, J = 7.7 Hz, 2H), 5.54 (br, J = 37.2 Hz, 1H), 4.30 (d, J = 7.6 Hz, 1H), 2.10 – 1.96 (m, 1H), 1.02 (d, J = 6.7 Hz, 3H), 0.86 (d, J = 6.7 Hz, 3H).

13C NMR (151 MHz, CDCl3) δ 139.55, 128.68, 127.97, 126.38, 119.33 (q, J = 320.9 Hz), 77.21, 77.00, 76.79, 65.76, 34.63, 19.45, 18.49.

HRMS (ESI-TOF) m/z Calcd for C₁₁H₁₃F₃NO₂S⁻ [M-H]⁻ 280.0625, found 280.0626.

4a methyl 3-phenyl-3-((trifluoromethyl)sulfonamido)propanoate

¹H NMR (600 MHz, CDCl₃) δ 7.40 – 7.34 (m, 2H), 7.34 – 7.26 (m, 3H), 6.85 (s, 1H), 5.03 (m, 1H), 3.65 (s, 3H), 2.96 (d, J = 5.5 Hz, 2H).

¹³C NMR (151 MHz, CDCl₃) δ 171.19, 138.70, 128.93, 128.38, 125.84, 119.36 (q, J = 320.9 Hz), 55.52, 52.25, 41.22.

HRMS (ESI-TOF) m/z Calcd for C₁₁H₁₂F₃NO₄SNa⁺ [MNa]⁺ 334.0331, found 334.0326.

4b methyl 3-(o-tolyl)-3-((trifluoromethyl)sulfonamido)propanoate

¹H NMR (600 MHz, CDCl₃) δ 7.32 (d, J = 7.2 Hz, 1H), 7.29 – 7.20 (m, 2H), 7.17 (d, J = 7.0 Hz, 1H), 6.81 (s, 1H), 5.27 (dd, J = 6.7 Hz, 5.7 Hz, 1H), 3.67 (s, 3H), 2.91 (dd, J = 15.8, 5.7 Hz, 1H), 2.84 (dd, J = 15.8, 6.7 Hz, 1H), 2.39 (s, 3H).

¹³C NMR (151 MHz, CDCl₃) δ 171.08, 137.23, 134.25, 130.91, 128.28, 126.73, 125.16, 119.32 (q, *J* = 320.8 Hz), 52.28, 52.04, 40.74, 18.88.

HRMS (ESI-TOF) m/z Calcd for C₁₂H₁₄F₃NO₄SNa⁺ [MNa]⁺ 348.0488, found 348.0488.

4c methyl 3-(*m*-tolyl)-3-((trifluoromethyl)sulfonamido)propanoate

¹H NMR (600 MHz, CDCl₃) δ 7.25 (t, *J* = 7.5 Hz, 1H), 7.16 – 7.06 (m, 3H), 6.69 (br s, 1H), 4.98 (m, 1H), 3.66 (s, 3H), 3.01 – 2.91 (m, 2H), 2.35 (s, 3H).

¹³C NMR (151 MHz, CDCl₃) δ 171.21, 138.73, 129.15, 128.83, 126.57, 122.80, 119.41 (q, J = 320.7 Hz), 55.45, 52.22, 41.11, 21.41.

HRMS (ESI-TOF) m/z Calcd for $C_{12}H_{14}F_3NO_4SNa^+$ [MNa]⁺ 348.0488, found 348.0490.

4d methyl 3-(4-methoxyphenyl)-3-((trifluoromethyl)sulfonamido)propanoate

¹H NMR (600 MHz, CDCl₃) δ 7.23 (d, *J* = 7.6 Hz, 2H), 6.88 (d, *J* = 7.6 Hz, 2H), 6.67 (s, 1H), 4.98 (m, 1H), 3.79 (s, 3H), 3.66 (s, 3H), 2.95 (s, 2H).

¹³C NMR (151 MHz, CDCl₃) δ 171.20, 159.45, 130.74, 127.18, 119.39 (q, J = 320.8, 320.2 Hz), 114.26, 55.23, 55.05, 52.22, 41.15.

HRMS (ESI-TOF) m/z Calcd for $C_{12}H_{14}F_3NO_5SNa^+$ [MNa]⁺ 364.0437, found 364.0434.

4e methyl 3-(4-fluorophenyl)-3-((trifluoromethyl)sulfonamido)propanoate

¹H NMR (600 MHz, CDCl₃) δ 7.40 – 7.24 (m, 2H), 7.16 – 6.99 (m, 2H), 6.79 (s, 1H), 5.16 – 4.94 (m, 1H), 3.67 (s, 3H), 3.04 – 2.87 (m, 2H).

¹³C NMR (151 MHz, CDCl₃) δ 171.17 , 162.46 (d, J = 247.8 Hz), 134.58 (d, J = 3.3 Hz), 127.74 (d, J = 8.4 Hz), 119.41 (q, J = 320.8 Hz), 115.93 (d, J = 21.8 Hz), 54.79 , 52.35 , 40.97 .

HRMS (ESI-TOF) m/z Calcd for C₁₁H₁₁F₄NO₄SNa⁺ [MNa]⁺ 352.0237, found 352.0237.

4f methyl 3-(4-chlorophenyl)-3-((trifluoromethyl)sulfonamido)propanoate

¹H NMR (600 MHz, CDCl₃) δ 7.35 (d, J = 8.2 Hz, 2H), 7.26 (d, J = 8.2 Hz, 2H), 6.82 (br s, 1H), 4.99 (t, J = 4.8 Hz, 1H), 3.67 (s, 3H), 3.02 – 2.86 (m, 2H).

¹³C NMR (151 MHz, CDCl₃) δ 171.12, 137.22, 134.34, 129.16, 127.31, 119.34 (q, J = 320.6 Hz), 54.76, 52.38, 40.78.

HRMS (ESI-TOF) m/z Calcd for C₁₁H₁₁ClF₃NO₄SNa⁺ [MNa]⁺ 367.9942, found 367.9943.

4g methyl 3-(4-trifluoromethylphenyl)-3-((trifluoromethyl)sulfonamide)propanoate

¹H NMR (600 MHz, CDCl₃) δ 7.65 (d, J = 8.4 Hz, 2H), 7.46 (d, J = 8.4 Hz, 2H), 6.83 (s, 1H), 5.06 (dd, J = 5.3, 4.8 Hz, 1H), 3.67 (s, 3H), 3.05 (dd, J = 16.7, 5.3 Hz, 1H), 2.97 (dd, J = 16.7, 4.8 Hz, 1H).

¹³C NMR (151 MHz, CDCl₃) δ 171.12, 142.55, 130.69 (q, *J* = 32.8 Hz), 126.35, 126.01 (q, *J* = 3.6 Hz), 123.75 (q,

J = 272.2 Hz), 119.37 (q, J = 320.5 Hz), 54.72 , 52.46 , 40.33 .HRMS (ESI-TOF) *m/z* Calcd for C₁₂H₁₁F₆NO₄SNa⁺ [MNa]⁺ 402.0205, found 402.0206.

7a N-(2-((tert-butyldimethylsilyl)oxy)-1-phenylethyl)-1,1,1-trifluoromethanesulfonamide

¹H NMR (600 MHz, CDCl₃) δ 7.40 – 7.35 (m, 2H), 7.35 – 7.29 (m, 3H), 5.96 (br s, 1H), 4.73 (m, 1H), 3.97 (dd, J = 10.4, 4.1 Hz, 1H), 3.80 (dd, J = 10.4, 4.7 Hz, 1H), 0.90 – 0.79 (m, 9H), -0.01 – -0.10 (m, 6H).

¹³C NMR (151 MHz, CDCl₃) δ 138.01, 128.56, 128.19, 126.56, 119.50 (q, *J* = 320.9 Hz), 66.71, 60.12, 25.66, 18.20, -5.78, -5.87.

HRMS (ESI-TOF) *m*/*z* Calcd for C₁₅H₂₅F₃NO₃SSi⁺ [MH]⁺ 384.1271, found 384.1270.



$7b\ N-(2-((tert-butyldimethylsilyl) oxy)-1-(o-tolyl) ethyl)-1, 1, 1-trifluoromethan esulf on a mide normalized and the second statement of the secon$

¹H NMR (600 MHz, CDCl₃) δ 7.32 – 7.24 (m, 1H), 7.24 – 7.18 (m, 2H), 7.18 – 7.13 (m, 1H), 5.80 (s, 1H), 5.02 – 4.90 (m, 1H), 3.88 (dd, J = 10.5, 4.4 Hz, 1H), 3.67 (dd, J = 10.5, 5.5 Hz, 1H), 2.35 (s, 3H), 0.85 (s, 10H), -0.03 – -0.09 (m, 6H).

¹³C NMR (151 MHz, CDCl₃) δ 135.95, 134.44, 130.61, 128.02, 126.33, 126.21, 119.46 (q, *J* = 321.2 Hz), 65.61, 56.40, 25.70, 19.07, 18.23, -5.70, -5.85.

HRMS (ESI-TOF) m/z Calcd for $C_{16}H_{27}F_3NO_3SSi^+$ [MH]⁺ 398.1427, found 398.1428.

7c N-(2-((tert-butyldimethylsilyl)oxy)-1-(2-fluorophenyl)ethyl)-1,1,1-trifluoromethanesulfonamide

¹H NMR (600 MHz, CDCl₃) δ 7.42 – 7.30 (m, 2H), 7.18 (t, *J* = 7.5 Hz, 1H), 7.13 – 7.06 (m, 1H), 6.08 (d, *J* = 8.3 Hz, 1H), 5.04 (ddd, *J* = 8.3, 4.7 Hz, 4.4 Hz, 1H), 3.99 (dd, *J* = 10.4, 4.4 Hz, 1H), 3.86 (dd, *J* = 10.4, 4.7 Hz, 1H), 0.86 (s, 9H), -0.03 (s, 3H), -0.04 (s, 3H).

¹³C NMR (151 MHz, CDCl₃) δ 159.77 (d, *J* = 245.7 Hz), 129.87 (d, *J* = 8.5 Hz), 128.64 (d, *J* = 3.6 Hz), 125.19 (d, *J* = 13.0 Hz), 124.23 (d, *J* = 3.3 Hz), 119.47 (q, *J* = 321.0 Hz), 115.58 (d, *J* = 21.8 Hz), 65.60 , 55.35 , 25.60 , 18.15 , -5.89 , -5.97.

HRMS (ESI-TOF) m/z Calcd for $C_{15}H_{24}F_4NO_3SSi^+$ [MH]⁺ 402.1177, found 402.1179.



7d *N*-(**2**-((*tert*-butyldimethylsilyl)oxy)-**1**-(naphthalen-2-yl)ethyl)-**1**,**1**,**1**-trifluoromethanesulfonamide ¹H NMR (600 MHz, CDCl₃) δ 7.87 – 7.79 (m, 3H), 7.77 (s, 1H), 7.54 – 7.46 (m, 2H), 7.40 (d, *J* = 8.5 Hz, 1H), 5.86 (d, *J* = 7.4 Hz, 1H), 4.93 – 4.83 (m, 1H), 4.03 (dd, *J* = 10.4, 3.8 Hz, 1H), 3.87 (dd, *J* = 10.4, 4.3 Hz, 1H), 0.85 (s, 9H), -0.04 (s, 3H), -0.07 (s, 3H).

¹³C NMR (151 MHz, CDCl₃) δ 135.38, 133.05, 133.03, 128.52, 127.96, 127.67, 126.48, 126.35, 125.89, 124.20, 119.52 (q, *J* = 321.2 Hz), 66.59, 60.12, 25.69, 18.21, -5.68, -5.78.

HRMS (ESI-TOF) m/z Calcd for $C_{19}H_{27}F_3NO_3SSi^+$ [MH]⁺ 434.1427, found 434.1424.

Optimization of the Reaction Conditions

Me NH	ITf	M	e NHTf	Me NHTf	<u> </u>	R ₂
	Me[Pd] / Ligan I ₂		Me +	Me		
1a			2a	3a		igund
entry	ligand		– conv. (%) [*]	ee [†]		- s‡
	R ₁	R ₂		2a	3a	3
1	C_6H_5	ⁿ Pr	38	50	83	17.6
2	C_6H_5	ⁿ Bu	48	77	83	24.8
3	C ₆ H₅	ⁱ Bu	49	86	88	50
4	C_6H_5	neopentyl	33	31	63	5.9
5	C_6H_5	ⁱ Pr	48	79	87	30.8
6	C_6H_5	^t Bu	25	23	70	6.8
7	$2-CF_3C_6H_4$	ⁱ Bu	48	80	88	34.2
8	3,5-(CF ₃) ₂ C ₆ H ₃	ⁱ Bu	45	39	47	4.1
9	4-OMeC ₆ H ₄	ⁱ Bu	57	90	73	15.5
10	$4-FC_6H_4$	ⁱ Bu	46	70	83	21.2
11	1-naphthyl	ⁱ Bu	39	50	81	13.3
12	2-naphthyl	ⁱ Bu	47	77	88	32.9
13	Me	ⁱ Bu	24	15	48	3.2
14	CF_3	ⁱ Bu	5	1	21	1.5
15	^t BuO	ⁱ Bu	38	24	39	2.9
16 [§]	C_6H_5	ⁱ Bu	50	89	90	62.0
17 ¹¹	C ₆ H₅	ⁱ Bu	50 (50 [¶])	92 (89 [¶])	92 (89 [¶])	78.8 (51.2 [¶])
18 [#]	C_6H_5	ⁱ Bu	54	94.5	80	33.0

Table S1. Optimization of enantioselective C-H iodination

Reaction conditions: 10 mol% Pd(OAc)₂, 40 mol% Ligand, 3 eq. Na₂CO₃, 3 eq. CsOAc, 3 eq. I₂,15eq. DMSO, 1mL ^tamyl-OH, air, 20°C, 24h ^{*}Calculated conversion, c = ee_{2a} / ($ee_{2a} + ee_{3a}$).[†]Determined by chiral HPLC analysis. [‡]Selectivity(s) = (rate of fast-reacting enantiomer) / (rate of slow-reacting enantiomer). [§]0.5 mL ^tamyl-OH. ^{II}0.5 mL ^tamyl-OH/DMSO (5 : 2.2). [¶]Reaction conditions: 2 mol% Pd(OAc)₂, 10 mol% Bz-Leu-OH, 3 equiv. CsOAc, 3 equiv. Na₂CO₃, 3 equiv. I₂, 0.5 mL ^tamyl-OH/DMSO (5 : 2.2), air, 20°C, 24h, then 3 equiv. I₂, 20°C, 24h. [#]20 mol% Ligand.

General Procedure for Pd(OAc)₂-Catalyzed C-H Iodination

To a 10 mL sealed tube was added substrates (1 equiv., 0.2 mmol), $Pd(OAc)_2$ (10 mol%, 0.02 mmol), Bz-Leu-OH (40 mol%, 0.08 mmol), CsOAc (3 equiv., 0.6 mmol), I₂ (3 equiv., 0.6 mmol), Na₂CO₃ (3 equiv., 0.6 mmol) and ^{*t*}amyl-OH / DMSO (1 mL, v / v = 5 : 2.2). The mixture was stirred at 20 °C for 24-48 h. The resulting mixture was diluted by EtOAc, filtered through a pad of celite and washed with Na₂S₂O₃. The organic layer was dried over MgSO₄, evaporated and purified by prep-TLC (EtOAc: hexanes= 1:5-1:15) to give the product and ee was determined on a Hitachi LaChrow HPLC system using commercially available chiral columns as described below.



3a 1,1,1-trifluoro-N-(1-(2-iodo-6-methylphenyl)ethyl)methanesulfonamide

The compound **3a** was prepared according to the general procedure and was purified by prep-TLC to give a colorless oil in 48 % yield (37 mg). ¹H NMR (600 MHz, CDCl₃) δ 7.76 (d, *J* = 7.8 Hz, 1H), 7.18 (d, *J* = 7.3 Hz, 1H), 6.87 (t, *J* = 7.7 Hz, 1H), 6.52 (d, *J* = 8.7 Hz, 0.8H), 5.53 (s, 0.2H), 5.35 – 5.21 (m, 1H), 2.51 (s, 0.7H), 2.42 (s, 2.3H), 1.73 (d, *J* = 7.0 Hz, 2.3H), 1.62 (s, 0.7H). ¹³C NMR (151 MHz, CDCl₃) δ 140.11, 139.37, 138.10, 131.28, 129.50, 119.32 (q, *J* = 321.2 Hz), 51.86, 20.76, 20.51 . HPLC chiralcel AS-H column (5 % isopropanol in hexanes, 0.5 mL/min) t_r= 12.340 min (major), 16.660 min (minor): 91 % ee. HRMS (ESI-TOF) *m/z* Calcd for C₁₀H₁₀F₃INO₂S^{*} [M-H]^{*} 391.9435, found 391.9436.



3b 1,1,1-trifluoro-N-(1-(2-iodo-6-methylphenyl)propyl)methanesulfonamide

The compound **3b** was prepared according to the general procedure and was purified by prep-TLC to give a colorless oil in 49% yield (40 mg). ¹H NMR (600 MHz, CDCl₃) δ 7.76 (d, *J* = 7.9 Hz, 1H), 7.18 (d, *J* = 7.5 Hz, 0.8H), 7.14 (d, *J* = 7.5 Hz, 0.2H), 6.91 – 6.83 (m, 1H), 6.44 (d, *J* = 9.7 Hz, 0.8H), 5.36 (m, 0.2H), 5.29 (m, 0.2H), 5.01 (m, 0.8H), 2.47 (s, 0.6H), 2.40 (s, 2.4H), 2.25 – 2.16 (m, 0.8H), 2.16 – 2.06 (m, 0.8H), 1.97 – 1.91 (m, 0.4H), 1.09 (t, *J* = 7.4 Hz, 0.6H), 0.99 (t, *J* = 7.4 Hz, 2.4H). ¹³C NMR (151 MHz, CDCl₃) δ 140.15, 139.45, 138.91, 138.54, 138.52, 135.76, 132.98, 131.23, 129.48, 129.36, 119.29 (q, *J* = 321.1 Hz), 101.96, 93.59, 66.79, 57.48, 27.84, 27.28, 21.08, 20.90, 11.02, 10.85. HPLC chiralcel AS-H column (5 % isopropanol in hexanes, 0.5 mL/min) t_r= 11.153 min (major), 13.567 min (minor): 89 % ee. HRMS (ESI-TOF) *m*/*z* Calcd for C₁₁H₁₃F₃INaNO₂S⁺ [MNa]⁺ 429.9556, found 429.9553.



3c 1,1,1-trifluoro-N-(1-(2-iodo-6-methylphenyl)pentyl)methanesulfonamide

The compound **3c** was prepared according to the general procedure and was purified by prep-TLC to give a colorless oil in 45 % yield (39 mg). ¹H NMR (600 MHz, CDCl₃) δ 7.76 (d, *J* = 7.9 Hz, 1H), 7.18 (d, *J* = 7.5 Hz, 0.8H), 7.14 (d, *J* = 7.6 Hz, 0.2H), 6.90 – 6.82 (m, 1H), 6.43 (s, 0.8H), 5.38 – 5.32 (m, 0.2H), 5.08 (t, *J* = 7.4 Hz, 0.8H), 2.48 (s, 0.6H), 2.40 (s, 2.4H), 2.26 – 2.16 (m, 0.8H), 2.06 – 1.98 (m, 0.8H), 1.94 – 1.84 (m, 0.4H), 1.51 – 1.30 (m, 3H), 1.30 – 1.19 (m, 1H), 0.96 – 0.88 (m, 3H). ¹³C NMR (151 MHz, CDCl₃) δ 140.14, 139.70, 138.86, 138.74, 138.56, 135.75, 132.99, 131.24, 129.44, 129.33, 119.26 (q, *J* = 321.1 Hz), 101.71, 93.66, 65.46, 56.03, 34.32, 33.78, 28.35, 28.25, 22.23, 22.20, 21.14, 20.87, 13.85, 13.84. HPLC chiralcel AS-H column (5 % isopropanol in hexanes, 0.3 mL/min) t_r= 17.227 min (major), 19.080 min (minor): 87 % ee. HRMS (ESI-TOF) *m/z* Calcd for C₁₃H₁₇F₃INaNO₂S⁺ [MNa]⁺ 457.9869, found 457.9868.



3d 1,1,1-trifluoro-N-(1-(2-iodo-6-methylphenyl)-3-methylbutyl)methanesulfonamide

The compound **3d** was prepared according to the general procedure and was purified by prep-TLC to give a colorless oil in 42 % yield (37 mg). ¹H NMR (600 MHz, CDCl₃) δ 7.75 (d, *J* = 6.0 Hz, 1H), 7.20 – 7.10 (m, 1H), 6.86 (dd, *J* = 6.9, 6.9 Hz, 1H), 6.42 (d, *J* = 8.4 Hz, 0.8H), 5.42 (s, 0.2H), 5.38 (s, 0.2H), 5.17 (s, 0.8H), 2.48 (s, 0.6H), 2.40 (s, 2.4H), 1.98 – 1.67 (m, 2H), 1.57 – 1.49 (m, 1H), 1.01 (d, *J* = 5.4 Hz, 6H). ¹³C NMR (151 MHz, CDCl₃) δ 140.14, 139.89, 139.25, 138.61, 138.33, 135.91, 133.01, 131.27, 129.38, 129.30, 119.20 (q, *J* = 321.0 Hz), 101.07, 93.76, 63.95, 54.36, 43.23, 42.90, 25.01, 24.63, 23.52, 23.31, 21.37, 21.22, 20.74. HPLC chiralcel AS-H column (5 % isopropanol in hexanes, 0.5 mL/min) t_r= 8.427 min (major), 9.567 min (minor): 81 % ee. HRMS (ESI-TOF) *m/z* Calcd for C₁₃H₁₇F₃INaNO₂S⁺ [MNa]⁺ 457.9869, found 457.9870.



3e 1,1,1-trifluoro-N-(1-(2-iodo-6-methylphenyl)-2-phenylethyl)methanesulfonamide

The compound **3e** was prepared according to the general procedure and was purified by prep-TLC to give a colorless oil in 44 % yield (41 mg). ¹H NMR (500 MHz, CDCl₃) δ 7.81 (d, *J* = 7.8 Hz, 0.8H), 7.76 (d, *J* = 7.4 Hz, 0.2H), 7.39 – 7.14 (m, 4H), 7.04 (d, *J* = 8.1 Hz, 2H), 6.92 – 6.80 (m, 1H), 6.53 (d, *J* = 10.0 Hz, 0.8H), 5.65 – 5.57 (m, 0.2H), 5.32 – 5.24 (m, 0.8H), 5.22 (s, 0.2H), 3.58 (dd, *J* = 13.5, 9.1 Hz, 0.8H), 3.33 (dd, *J* = 13.5, 7.1 Hz, 0.8H), 3.25 (dd, *J* = 14.2, 5.5 Hz, 0.2H), 3.03 (dd, *J* = 14.2, 10.3 Hz, 0.2H), 2.50 (s, 0.4H), 2.01 (s, 2.6H). ¹³C NMR

(151 MHz, CDCl₃) δ 140.06, 139.58, 138.66, 138.54, 137.49, 136.21, 135.86, 135.27, 133.12, 131.07, 129.71, 129.54, 129.33, 129.23, 128.99, 128.54, 127.65, 127.20, 119.20 (q, J = 321.2 Hz), 101.52, 94.12, 66.47, 57.22, 40.45, 40.24, 21.45, 20.48. HPLC chiralcel AS-H column (5% isopropanol in hexanes, 0.5 mL/min) t_r= 14.480 min (major), 18.213 min (minor): 75 % ee. HRMS (ESI-TOF) m/z Calcd for C₁₆H₁₅F₃INaNO₂S⁺ [MNa]⁺ 491.9713, found 491.9708.

3f N-(cyclopropyl(2-iodo-6-methylphenyl)methyl)-1,1,1-trifluoromethanesulfonamide

The compound **3f** was prepared according to the general procedure and was purified by prep-TLC to give a colorless oil in 42 % yield (35 mg). ¹H NMR (600 MHz, CDCl₃) δ 7.80 (d, *J* = 7.8 Hz, 0.7H), 7.74 (d, *J* = 7.9 Hz, 0.3H), 7.18 (d, *J* = 7.6 Hz, 0.7H), 7.16 (d, *J* = 7.5 Hz, 0.3H), 6.88 (dd, *J* = 7.8, 7.7 Hz, 0.7H), 6.84 (dd, *J* = 7.8, 7.7 Hz, 0.3H), 4.64 (d, *J* = 9.3 Hz, 0.3H), 4.29 (d, *J* = 9.7 Hz, 0.7H), 2.56 (s, 1H), 2.35 (s, 2H), 2.05 – 1.96 (m, 0.7H), 1.46 – 1.39 (m, 0.3H), 0.85 – 0.41 (m, 4H). ¹³C NMR (151 MHz, CDCl₃) δ 140.13, 139.41, 138.32, 138.21, 136.75, 132.87, 131.30, 129.40, 129.19, 119.39 (q, *J* = 321.2 Hz), 102.12, 94.49, 68.88, 60.76, 21.02, 20.98, 16.58, 15.69, 5.90, 5.26, 5.14, 4.79. HPLC chiralcel AS-H column (5 % isopropanol in hexanes, 0.5 mL/min) t_r= 13.733 min (major), 19.760 min (minor): 95 % ee. HRMS (ESI-TOF) *m*/*z* Calcd for C₁₂H₁₃F₃INaNO₂S⁺ [MNa]⁺ 441.9556, found 441.9556.



3g 1,1,1-trifluoro-*N*-(1-(2-iodo-6-methoxyphenyl)ethyl)methanesulfonamide

The compound **3g** was prepared according to the general procedure and was purified by prep-TLC to give a colorless oil in 46 % yield (38 mg). ¹H NMR (600 MHz, CDCl₃) δ 7.49 (d, *J* = 7.6 Hz, 1H), 7.00 – 6.90 (m, 2H), 6.51 (d, *J* = 9.7 Hz, 1H), 5.27 – 5.18 (m, 1H), 3.92 (s, 3H), 1.58 – 1.53 (m, 3H). ¹³C NMR (151 MHz, CDCl₃) δ 156.69, 132.69, 131.31, 130.28, 119.38 (q, *J* = 321.1 Hz), 111.82, 99.22, 60.27, 55.94, 21.36. HPLC chiralcel AS-H column (5 % isopropanol in hexanes, 0.8 mL/min) t_r= 9.387 min (major), 21.127 min (minor): 97 % ee. HRMS (ESI-TOF) *m/z* Calcd for C₁₀H₁₁F₃INaNO₃S⁺ [MNa]⁺ 431.9349, found 431.9351.



3h 1,1,1-trifluoro-N-(1-(2-fluoro-6-iodophenyl)ethyl)methanesulfonamide

The compound **3h** was prepared according to the general procedure and was purified by prep-TLC to give a colorless oil in 47 % yield (37 mg). ¹H NMR (600 MHz, CDCl₃) δ 7.67 (d, *J* = 7.9 Hz, 1H), 7.21 – 7.08 (m, 1H), 7.08 – 6.98 (m, 1H), 5.77 (s, 1H), 5.37 – 5.17 (m, 1H), 1.61 (d, *J* = 7.0 Hz, 3H). ¹³C NMR (151 MHz, CDCl₃) δ 160.13 (d, *J* = 250.5 Hz), 136.00 (d, J = 2.2 Hz), 131.45 (d, *J* = 15.3 Hz), 130.97 (d, *J* = 9.8 Hz), 119.30 (q, *J* = 320.7 Hz), 116.77 (d, *J* = 23.0 Hz), 99.57 , 58.93 , 21.80 . HPLC chiralcel AS-H column (5 % isopropanol in hexanes, 0.8 mL/min) t_r= 8.447 min (major), 12.593 min (minor): 96 % ee. HRMS (ESI-TOF) *m/z* Calcd for C₉H₇F₄INO₂S⁻ [M-H]⁻ 395.9184, found 395.9189.



$\label{eq:single-sing$

The compound **3i** was prepared according to the general procedure and was purified by prep-TLC to give a colorless oil in 49 % yield (42 mg). ¹H NMR (600 MHz, CDCl₃) δ 7.78 (s, 1H), 7.19 (s, 1H), 6.39 (d, *J* = 8.2 Hz, 1H), 5.33 – 5.19 (m, 1H), 2.59 – 2.27 (m, 3H), 1.82 – 1.57 (m, 3H). ¹³C NMR (151 MHz, CDCl₃) δ 139.18, 138.49, 138.07, 136.35, 133.99, 133.49, 131.14, 130.70, 126.84, 126.20, 119.43 (q, *J* = 321.1 Hz), 99.57, 93.16, 51.46, 51.00, 40.72, 23.43, 20.70, 20.40, 18.84. HPLC chiralcel AS-H column (5 % isopropanol in hexanes, 0.5 mL/min) t_r= 12.047 min (major), 15.640 min (minor): 93 % ee. HRMS (ESI-TOF) *m*/*z* Calcd for C₁₀H₉ClF₃INO₂S⁻ [M-H]⁻425.9045, found 425.9047.

3j 1,1,1-trifluoro-N-(1-(2-iodo-5-methylphenyl)ethyl)methanesulfonamide

The compound **3j** was prepared according to the general procedure and was purified by prep-TLC to give a colorless oil in 34 % yield (27 mg). ¹H NMR (600 MHz, CDCl₃) δ 7.69 (d, *J* = 8.0 Hz, 1H), 7.08 (s, 1H), 6.83 (d, *J* = 9.4 Hz, 1H), 5.47 (s, 1H), 5.03 (m, 1H), 2.31 (s, 3H), 1.55 (d, *J* = 6.9 Hz, 3H). ¹³C NMR (151 MHz, CDCl₃) δ 143.20, 139.91, 139.19, 130.84, 127.14, 119.36 (q, *J* = 321.3 Hz), 92.83, 59.07, 23.46, 21.03. HPLC chiralcel AS-H column (5 % isopropanol in hexanes, 0.8 mL/min) t_r= 7.967 min (major), 15.753 min (minor): 92 % ee. HRMS (ESI-TOF) *m*/*z* Calcd for C₁₀H₁₁F₃INaNO₂S⁺ [MNa]⁺ 415.9400, found 415.9392.



3k 1,1,1-trifluoro-N-(1-(2-iodo-4-methylphenyl)ethyl)methanesulfonamide

The compound 3k was prepared according to the general procedure and was purified by prep-TLC to give

a colorless oil in 35 % yield (28 mg). ¹H NMR (600 MHz, CDCl₃) δ 7.69 (s, 1H), 7.20 (d, *J* = 7.9 Hz, 1H), 7.16 (d, *J* = 7.9 Hz, 1H), 5.05 (q, *J* = 6.8 Hz, 1H), 2.30 (s, 3H), 1.55 (d, *J* = 6.8 Hz, 3H). ¹³C NMR (151 MHz, CDCl₃) δ 140.61, 140.56, 140.01, 129.83, 125.95, 119.36 (q, *J* = 321.2 Hz), 96.92, 58.93, 23.55, 20.42. HPLC chiralcel AS-H column (10 % isopropanol in hexanes, 0.3 mL/min) t_r= 17.080 min (major), 41.727 min (minor): 95 % ee. HRMS (ESI-TOF) *m/z* Calcd for C₁₀H₁₁F₃INaNO₂S⁺ [MNa]⁺ 415.9400, found 415.9392.



3l 1,1,1-trifluoro-N-(1-(3-iodonaphthalen-2-yl)ethyl)methanesulfonamide

The compound **31** was prepared according to the general procedure and was purified by prep-TLC to give a colorless oil in 37 % yield (32 mg). ¹H NMR (600 MHz, CDCl₃) δ 8.42 (s, 1H), 7.83 – 7.70 (m, 3H), 7.57 – 7.49 (m, 2H), 5.57 (d, *J* = 6.9 Hz, 1H), 5.26 – 5.18 (m, 1H), 1.68 (d, *J* = 6.9 Hz, 3H). ¹³C NMR (151 MHz, CDCl₃) δ 139.90, 139.86, 134.30, 132.72, 127.78, 127.29, 127.26, 126.51, 125.45, 119.38 (q, *J* = 321.1 Hz), 93.64, 58.97, 23.86. HPLC chiralcel AS-H column (5 % isopropanol in hexanes, 0.5 mL/min) t_r= 17.880 min (major), 36.760 min (minor): 95 % ee. HRMS (ESI-TOF) *m/z* Calcd for C₁₃H₁₁F₃INaNO₂S⁺ [MNa]⁺ 451.9400, found 451.9403.



3m-mono 1,1,1-trifluoro-N-(1-(2-iodophenyl)pentyl)methanesulfonamide

The compound **3m-mono** was prepared according to the general procedure and was purified by prep-TLC to give a colorless oil in 17 % yield (14 mg). ¹H NMR (600 MHz, CDCl₃) δ 7.85 (dd, *J* = 7.9, 1.2 Hz, 1H), 7.38 (td, *J* = 7.5, 1.2 Hz, 1H), 7.21 (dd, *J* = 7.8, 1.6 Hz, 1H), 7.05 – 6.95 (m, 1H), 5.45 (s, 1H), 4.91 (ddd, *J* = 8.5 Hz, 1H), 1.93 – 1.82 (m, 1H), 1.78 – 1.67 (m, 1H), 1.51 – 1.25 (m, 4H), 0.92 (t, *J* = 7.1 Hz, 3H). ¹³C NMR (151 MHz, CDCl₃) δ 142.98, 140.21, 129.66, 128.85, 126.71, 119.30 (q, *J* = 321.2 Hz), 63.15, 37.09, 27.92, 22.12, 13.82. HPLC chiralcel AS-H column (10 % isopropanol in hexanes, 0.5 mL/min) t_r= 10.893 min (major), 11.773 min (minor): 87 % ee. HRMS (ESI-TOF) *m/z* Calcd for C₁₂H₁₅F₃INaNO₂S⁺ [MNa]⁺ 443.9712, found 443.9711.



3m-di N-(1-(2,6-diiodophenyl)pentyl)-1,1,1-trifluoromethanesulfonamide

The compound 3m-di was prepared according to the general procedure and was purified by prep-TLC to

give a colorless oil in 24 % yield (26 mg). ¹H NMR (600 MHz, CDCl₃) δ 8.06 – 7.84 (m, 2H), 6.64 (t, *J* = 7.9 Hz, 1H), 6.32 (br s, 1H), 5.33 (dd, *J* = 9.6, 6.2 Hz, 1H), 2.17 – 2.06 (m, 1H), 2.00 – 1.87 (m, 1H), 1.59 – 1.31 (m, 4H), 0.94 (t, *J* = 7.2 Hz, 3H). ¹³C NMR (151 MHz, CDCl₃) δ 142.52, 141.25, 140.80, 130.87, 119.21 (q, *J* = 321.1 Hz), 102.69, 92.22, 65.79, 33.44, 28.05, 22.18, 13.84. HPLC chiralcel AS-H column (10 % isopropanol in hexanes, 0.5 mL/min) t_r= 10.400 min (major), 12.860 min (minor): 99 % ee. HRMS (ESI-TOF) *m/z* Calcd for C₁₂H₁₅F₃INaNO₂S⁺ [MNa]⁺ 569.8679, found 569.8680.



6a-mono methyl 3-(2-iodophenyl)-3-((trifluoromethyl)sulfonamido)propanoate

The compound **6a-mono** was prepared according to the general procedure and was purified by prep-TLC to give a colorless oil in 31 % yield (27 mg). ¹H NMR (400 MHz, CDCl₃) δ 7.85 (dd, *J* = 7.9, 1.1 Hz, 1H), 7.47 – 7.33 (m, 2H), 7.13 (d, *J* = 8.7 Hz, 1H), 7.03 (ddd, *J* = 7.9, 7.0, 2.1 Hz, 1H), 5.25 (dt, *J* = 9.5, 5.0 Hz, 1H), 3.66 (s, 3H), 2.98 (d, *J* = 5.0 Hz, 2H). ¹³C NMR (151 MHz, CDCl₃) δ 171.22, 140.77, 140.08, 130.04, 128.78, 127.30, 119.32 (q, *J* = 322.1, 321.4 Hz), 97.19, 58.89, 52.32, 39.04. HPLC chiralcel AD-H column (10 % isopropanol in hexanes, 0.2 mL/min) t_r= 29.827 min (minor), 32.413 min (major): 96 % ee. HRMS (ESI-TOF) *m/z* Calcd for C₁₁H₁₂F₃INO₄S⁺ [MH]⁺ 437.9478, found 437.9477.



6a-di methyl 3-(2,6-diiodophenyl)-3-((trifluoromethyl)sulfonamido)propanoate

The compound **6a-di** was prepared according to the general procedure and was purified by prep-TLC to give a colorless oil in 16 % yield (18 mg). ¹H NMR (600 MHz, CDCl₃) δ 7.95 (m, 2H), 6.67 (t, *J* = 7.9 Hz, 1H), 6.52 (d, *J* = 9.1 Hz, 1H), 5.88 (ddd, *J* = 10.3, 9.1, 5.0 Hz, 1H), 3.78 (s, 3H), 3.27 (dd, *J* = 15.4, 10.3 Hz, 1H), 2.85 (dd, *J* = 15.4, 5.0 Hz, 1H). ¹³C NMR (151 MHz, CDCl₃) δ 168.79, 142.54, 140.99, 139.67, 131.43, 119.13 (q, *J* = 321.4 Hz), 102.15, 92.52, 62.73, 52.45, 38.23. HRMS (ESI-TOF) *m*/*z* Calcd for C₁₁H₉F₃I₂NO₄S⁻ [M-H]⁻ 561.8299, found 561.8301.

6b methyl 3-(2-iodo-6-methylphenyl)-3-((trifluoromethyl)sulfonamido)propanoate

The compound **6b** was prepared according to the general procedure and was purified by prep-TLC to give

a colorless oil in 49 % yield (44 mg). ¹H NMR (600 MHz, CDCl₃) δ 7.76 (d, *J* = 7.6 Hz, 1H), 7.23 – 7.11 (m, 1H), 6.90 (dd, *J* = 7.6, 7.4 Hz, 1H), 5.75 (s, 0.2H), 5.69 – 5.62 (m, 0.8H), 3.77 (s, 0.6H), 3.71 (s, 2.4H), 3.33 (dd, *J* = 15.5, 8.8 Hz, 0.8H), 3.06 (dd, *J* = 15.5, 5.6 Hz, 0.8H), 2.91 (s, 0.2H), 2.80 (s, 0.2H), 2.51 (s, 3H). ¹³C NMR (151 MHz, CDCl₃) δ 176.61, 169.61, 140.04, 139.36, 138.49, 137.55, 133.14, 131.51, 130.05, 129.66, 119.20 (q, *J* = 321.2 Hz), 99.56, 93.97, 52.55, 52.26, 39.27, 38.79, 21.25, 20.70. HPLC chiralcel AS-H column (10 % isopropanol in hexanes, 0.3 mL/min) t_r= 29.640 min (minor), 31.907 min (major): 96 % ee. HRMS (ESI-TOF) *m/z* Calcd for C₁₂H₁₄F₃INO₄S⁺ [MH]⁺ 451.9635, found 451.9633.



6c methyl 3-(2-iodo-5-methylphenyl)-3-((trifluoromethyl)sulfonamido)propanoate

The compound **6c** was prepared according to the general procedure and was purified by prep-TLC to give a colorless oil in 44 % yield (40 mg). ¹H NMR (600 MHz, CDCl₃) δ 7.70 (d, *J* = 8.0 Hz, 1H), 7.21 (s, 1H), 7.05 (d, *J* = 9.1 Hz, 1H), 6.85 (d, *J* = 8.0 Hz, 1H), 5.21 (dt, *J* = 9.1, 5.1 Hz, 1H), 3.67 (s, 3H), 2.95 (d, *J* = 5.1 Hz, 2H), 2.32 (s, 3H). ¹³C NMR (151 MHz, CDCl₃) δ 171.18, 140.46, 139.76, 138.96, 131.06, 128.11, 119.35 (q, *J* = 320.7 Hz) 93.03, 58.77, 52.27, 39.14, 21.10. HPLC chiralcel AD-H column (5 % isopropanol in hexanes, 0.5 mL/min) t_r= 11.980 min (minor), 14.693 min (major): 94 % ee. HRMS (ESI-TOF) *m/z* Calcd for C₁₂H₁₄F₃INO₄S⁺ [MH]⁺ 451.9635, found 451.9629.

6d methyl 3-(2-iodo-4-methoxyphenyl)-3-((trifluoromethyl)sulfonamido)propanoate

The compound **6d** was prepared according to the general procedure and was purified by prep-TLC to give a colorless oil in 33 % yield (30 mg). ¹H NMR (500 MHz, CDCl₃) δ 7.37 (d, *J* = 2.6 Hz, 1H), 7.30 (d, *J* = 8.7 Hz, 1H), 6.92 (dd, *J* = 8.7, 2.6 Hz, 1H), 5.20 (t, *J* = 5.1 Hz, 1H), 3.79 (s, 3H), 3.66 (s, 3H), 2.94 (d, *J* = 5.1 Hz, 2H). ¹³C NMR (151 MHz, CDCl₃) δ 171.39, 159.49, 132.92, 127.62, 124.99, 119.35 (q, *J* = 321.1 Hz), 114.64, 97.27, 58.41, 55.52, 52.29, 39.53. HPLC chiralcel OJ column (10 % isopropanol in hexanes, 0.5 mL/min) t_r= 31.120 min (major), 39.807 min (minor): 98 % ee. HRMS (ESI-TOF) *m*/*z* Calcd for C₁₂H₁₃F₃INaNO₅S⁺ [MNa]⁺ 489.9403, found 489.9337.



6e methyl 3-(4-fluoro-2-iodophenyl)-3-((trifluoromethyl)sulfonamido)propanoate

The compound **6e** was prepared according to the general procedure and was purified by prep-TLC to give a colorless oil in 41 % yield (39 mg). ¹H NMR (600 MHz, CDCl₃) δ 7.58 (dd, *J* = 7.8, 2.6 Hz, 1H), 7.41 (dd, *J* = 8.8, 5.6 Hz, 1H), 7.12 (m, 1H), 5.21 (t, *J* = 4.9 Hz, 1H), 3.66 (s, 3H), 2.98 – 2.91 (m, 2H). ¹³C NMR (151 MHz, CDCl₃) δ 171.26 , 161.54 (d, *J* = 253.6 Hz), 136.84 (d, *J* = 3.5 Hz), 128.27 (d, *J* = 8.2 Hz), 126.93 (d, *J* = 23.7 Hz), 119.31 (q, *J* = 320.7 Hz), 115.93 (d, *J* = 21.2 Hz), 96.49 (d, *J* = 8.2 Hz), 58.15 , 52.39 , 38.86 . HPLC chiralcel OJ column (10 % isopropanol in hexanes, 0.3 mL/min) t_r= 37.327 min (major), 44.560 min (minor): 98 % ee. HRMS (ESI-TOF) *m/z* Calcd for C₁₁H₁₀F₄INaNO₄S⁺ [MNa]⁺ 477.9204, found 477.9201.



6f methyl 3-(4-chloro-2-iodophenyl)-3-((trifluoromethyl)sulfonamido)propanoate

The compound **6f** was prepared according to the general procedure and was purified by prep-TLC to give a colorless oil in 40 % yield (37 mg). ¹H NMR (500 MHz, CDCl₃) δ 7.93 (d, *J* = 1.4 Hz, 1H), 7.53 – 7.43 (m, 2H), 7.24 (d, *J* = 9.1 Hz, 1H), 5.28 (dt, *J* = 9.1, 4.8 Hz, 1H), 3.75 (s, 3H), 3.11 – 3.00 (m, 2H). ¹³C NMR (151 MHz, CDCl₃) δ 171.21, 139.46, 139.26, 134.90, 129.00, 127.99, 119.29 (q, *J* = 320.7 Hz), 97.06, 58.34, 52.44, 38.79. HPLC chiralcel OJ column (10 % isopropanol in hexanes, 0.5 mL/min) t_r= 19.533 min (major), 28.627 min (minor): 98 % ee. HRMS (ESI-TOF) *m/z* Calcd for C₁₁H₁₀ClF₃INaNO₄S⁺ [MH]⁺ 493.8908, found 493.8906.

6g methyl 3-(4-trifluoromethyl-2-iodophenyl)-3-((trifluoromethyl)sulfonamido)propanoate

The compound **6g** was prepared according to the general procedure and was purified by prep-TLC to give a colorless oil in 38 % yield (40 mg). ¹H NMR (600 MHz, CDCl₃) δ 8.14 – 8.08 (m, 1H), 7.66 (dd, *J* = 8.3, 1.7 Hz, 1H), 7.58 (d, *J* = 8.3 Hz, 1H), 5.30 – 5.23 (m, 1H), 3.68 (s, 3H), 3.04 – 2.95 (m, 2H). ¹³C NMR (151 MHz, CDCl₃) δ 171.10 , 144.80 , 136.89 (q, *J* = 3.6 Hz), 132.00 (q, *J* = 33.3 Hz), 127.72 , 125.66 (q, *J* = 3.4 Hz), 122.5 (q, *J* = 272.9 Hz), 119.31 (q, *J* = 320.9 Hz), 96.79 , 58.62 , 52.51 , 38.38 . HPLC chiralcel OJ column (10 % isopropanol in hexanes, 0.2 mL/min) t_r= 31.760 min (major), 40.273 min (minor): 99 % ee. HRMS (ESI-TOF) *m/z* Calcd for C₁₁H₁₀ClF₃INaNO₄S⁺ [MH]⁺ 505.9352, found 505.9355.



9a-mono *N*-(**2**-((*tert*-butyldimethylsilyl)oxy)-1-(**2**-iodophenyl)ethyl)-1,1,1-trifluoromethanesulfonamide The compound **9a-mono** was prepared according to the general procedure and was purified by prep-TLC to give a colorless oil in 20 % yield (20 mg). ¹H NMR (600 MHz, CDCl₃) δ 7.84 (d, *J* = 7.0 Hz, 1H), 7.38 (t, *J* = 7.5 Hz, 1H), 7.33 (d, *J* = 7.7 Hz, 1H), 7.02 (t, *J* = 7.6 Hz, 1H), 5.05 (dd, *J* = 4.5, 3.8 Hz, 1H), 3.95 (dd, *J* = 10.6, 3.8 Hz, 1H), 3.73 (dd, *J* = 10.6, 4.5 Hz, 1H), 0.84 (s, 9H), -0.06 (s, 3H), -0.09 (s, 3H). ¹³C NMR (151 MHz, CDCl₃) δ 139.77, 139.66, 129.89, 128.25, 128.14, 119.45 (q, *J* = 321.3 Hz), 97.39, 65.07, 63.28, 25.72, 18.24, -5.68, -5.79. The compound was deiodinated and then ee was determined. HPLC chiralcel AD-H column (5 % isopropanol in hexanes, 0.1 mL/min) t_r= 37.320 min (minor), 40.260 min (major): 87 % ee. HRMS (ESI-TOF) *m/z* Calcd for C₁₅H₂₂F₃INO₃SSi⁻ [M-H]⁻ 508.0092, found 508.0091.



9a-di *N*-(2-((*tert*-butyldimethylsilyl)oxy)-1-(2,6-diiodophenyl)ethyl)-1,1,1-trifluoromethanesulfonamide The compound **9a-di** was prepared according to the general procedure and was purified by prep-TLC to give a colorless oil in 20 % yield (25 mg). ¹H NMR (600 MHz, CDCl₃) δ 7.93 (m, 2H), 6.65 (t, *J* = 7.8 Hz, 1H), 6.24 (br, 1H), 5.49 (t, *J* = 7.6 Hz, 1H), 4.09 – 4.01 (m, 1H), 3.94 – 3.87 (m, 1H), 0.89 (s, 9H), 0.12 (s, 3H), 0.06(s, 3H). ¹³C NMR (151 MHz, CDCl₃) δ 142.51, 140.72, 138.24, 131.27, 119.33 (q, *J* = 321.7 Hz), 103.48, 92.85, 66.79, 62.04, 25.75, 18.22, -5.28, -5.31. HPLC chiralcel AD-H column (5 % isopropanol in hexanes, 0.1 mL/min) t_r= 32.867 min (minor), 39.473 min (major): 99 % ee. HRMS (ESI-TOF) *m/z* Calcd for C₁₅H₂₁F₃I₂NO₃SSi⁻ [M-H]⁻ 633.9058, found 633.9055.

9b

N-(2-((*tert*-butyldimethylsilyl)oxy)-1-(2-methyl-6-iodophenyl)ethyl)-1,1,1-trifluoromethanesulfonamide The compound **9b** was prepared according to the general procedure and was purified by prep-TLC to give a colorless oil in 49 % yield (51 mg). ¹H NMR (600 MHz, CDCl₃) δ 7.75 (d, *J* = 7.8 Hz, 1H), 7.19 (d, *J* = 7.4 Hz, 0.7H), 7.14 (d, *J* = 7.4 Hz, 0.3H), 6.94 – 6.82 (m, 1H), 5.48 – 5.40 (m, 0.3H), 5.27 – 5.19 (m, 0.7H), 4.16 – 4.08 (m, 0.7H), 4.06 – 3.98 (m, 0.7H), 3.88 (dd, *J* = 10.9, 5.0 Hz, 0.3H), 3.81 (t, *J* = 9.7 Hz, 0.3H), 2.53 (s, 1H), 2.43 (s, 2H), 0.91 (s, 3H), 0.83 (s, 6H), 0.12 – 0.06 (m, 2H), 0.04 (s, 2H), -0.04 (s, 2H). ¹³C NMR (151 MHz, CDCl₃) δ 140.28, 139.90, 138.40, 137.62, 136.24, 135.98, 133.04, 131.12, 129.87, 129.70, 119.32 (q, J = 321.1 Hz), 101.75, 93.97, 66.52, 63.10, 63.04, 56.92, 25.76, 25.67, 21.82, 20.98, 18.27, 18.11, -5.36, -5.43, -5.47, -5.51. The compound was deiodinated and the TBS ether was deprotected and then ee was determined. HPLC chiralcel AD-H column (10 % isopropanol in hexanes, 0.5 mL/min) t_r= 23.200 min (minor), 30.947 min (major): 91 % ee. HRMS (ESI-TOF) m/z Calcd for C₁₆H₂₄F₃INO₃SSi⁻ [M-H]⁻ 522.0248, found 522.0248.

9c *N*-(2-((*tert*-butyldimethylsilyl)oxy)-1-(2-fluoro-6-iodophenyl)ethyl)-1,1,1-trifluoromethanesulfonamide The compound **9**c was prepared according to the general procedure and was purified by prep-TLC to give a colorless oil in 41 % yield (43 mg). ¹H NMR (600 MHz, CDCl₃) δ 7.71 – 7.65 (m, 1H), 7.13 – 7.05 (m, 1H), 7.05 – 6.98 (m, 1H), 5.87 (s, 1H), 5.19 (s, 1H), 3.87 (s, 2H), 0.85 (s, 9H), 0.03 – -0.09 (m, 6H). ¹³C NMR (151 MHz, CDCl₃) δ 160.54 (d, *J* = 250.4 Hz), 136.02, 131.22 (d, *J* = 9.7 Hz), 127.74 (d, *J* = 9.7 Hz), 119.33 (d, *J* = 320.9 Hz), 116.73 (d, *J* = 23.8 Hz), 99.47, 64.27, 63.97, 25.66, 18.18, -5.59, -5.67. HPLC chiralcel AS-H column (5 % isopropanol in hexanes, 0.3 mL/min) t_r= 13.473 min (major), 14.520 min (minor): 99 % ee. HRMS (ESI-TOF) *m/z* Calcd for C₁₅H₂₃F₄INO₃SSi⁺ [MH]⁺ 528.0143, found 428.0141.



9d

N-(2-((*tert*-butyldimethylsilyl)oxy)-1-(3-iodonaphthalen-2-yl)ethyl)-1,1,1-trifluoromethanesulfonamide The compound **9d** was prepared according to the general procedure and was purified by prep-TLC to give a colorless oil in 40 % yield (44 mg). ¹H NMR (400 MHz, cdcl₃) δ 8.40 (s, 1H), 7.83 – 7.75 (m, 2H), 7.72 (d, *J* = 8.9 Hz, 1H), 7.57 – 7.46 (m, 2H), 6.04 (s, 1H), 5.21 (ddd, *J* = 7.3, 4.1, 3.8 Hz, 1H), 4.04 (dd, *J* = 10.7, 3.8 Hz, 1H), 3.79 (dd, *J* = 10.7, 4.1 Hz, 1H), 0.82 (s, 9H), -0.10 (s, 3H), -0.18 (s, 3H). ¹³C NMR (151 MHz, CDCl₃) δ 139.28, 135.74, 134.37, 132.36, 127.86, 127.32, 127.20, 127.18, 126.47, 119.51 (q, *J* = 321.4 Hz), 94.13, 65.13, 63.02, 25.68, 18.20, -5.69, -5.80. HPLC chiralcel AS-H column (5 % isopropanol in hexanes, 0.1 mL/min) t_r= 48.873 min (minor), 53.613 min (major): 96 % ee. HRMS (ESI-TOF) *m*/*z* Calcd for C₁₉H₂₆F₃INO₃SSi⁺ [MH]⁺ 560.0394, found 560.0398.

Me NHTf

2a

HPLC chiralcel AS-H column (5 % isopropanol in hexanes, 0.5 mL/min) t_r = 11.200 min (minor), 30.613 min (major): 93 % ee.

2b

HPLC chiralcel AS-H column (5 % isopropanol in hexanes, 0.5 mL/min) t_r = 10.553 min (minor), 16.260 min (major): 89 % ee.



2c

HPLC chiralcel AS-H column (5 % isopropanol in hexanes, 0.5 mL/min) t_r = 9.733 min (minor), 12.880 min (major): 77 % ee.



2d

HPLC chiralcel AS-H column (5 % isopropanol in hexanes, 0.5 mL/min) t_r = 8.447 min (minor), 9.600 min (major): 83 % ee.



2e

HPLC chiralcel AS-H column (5 % isopropanol in hexanes, 0.5 mL/min) t_r = 14.440 min (minor), 18.167 min (major): 67 % ee.



2f

HPLC chiralcel AS-H column (5 % isopropanol in hexanes, 0.8 mL/min) t_r = 7.853 min (minor), 26.180 min (major): 77 % ee.



2g

HPLC chiralcel AS-H column (5 % isopropanol in hexanes, 0.8 mL/min) t_r = 9.687 min (minor), 32.247 min (major): 93 % ee.

2h

HPLC chiralcel AS-H column (5 % isopropanol in hexanes, 0.8 mL/min) t_r = 7.833 min (minor), 11.200 min (major): 89 % ee.





HPLC chiralcel AS-H column (5 % isopropanol in hexanes, 0.5 mL/min) t_r = 11.893 min (minor), 25.320 min (major): 91 % ee.



2j

HPLC chiralcel AS-H column (5 % isopropanol in hexanes, 0.8 mL/min) t_r = 7.100 min (minor), 12.013 min (major): 78 % ee.



2k

HPLC chiralcel AS-H column (10 % isopropanol in hexanes, 0.3 mL/min) t_r = 15.747 min (minor), 22.540 min (major): 78 % ee.



21

HPLC chiralcel AS-H column (5 % isopropanol in hexanes, 0.5 mL/min) t_r = 17.173 min (minor), 23.427 min (major): 67 % ee.



2m

HPLC chiralcel AS-H column (10 % isopropanol in hexanes, 0.5 mL/min) t_r = 9.827 min (minor), 10.853 min (major): 70 % ee.

2n

HPLC chiralcel AS-H column (10 % isopropanol in hexanes, 0.5 mL/min) t_r = 7.807 min (minor), 9.287 min (major): 45 % ee.

5a

HPLC chiralcel OJ column (10 % isopropanol in hexanes, 0.3 mL/min) t_r = 45.227 min (minor), 50.520 min (major): 85 % ee.



5b

HPLC chiralcel AS-H column (10 % isopropanol in hexanes, 0.5 mL/min) t_r = 17.280 min (minor), 18.913 min (major): 93 % ee.

5c

HPLC chiralcel OJ column (10 % isopropanol in hexanes, 0.5 mL/min) t_r = 16.933 min (major), 29.307 min (minor): 93 % ee.



HPLC chiralcel OJ column (10 % isopropanol in hexanes, 0.5 mL/min) t_r = 42.313 min (minor), 53.093 min (major): 82 % ee.

5e

HPLC chiralcel OJ column (10 % isopropanol in hexanes, 0.5 mL/min) t_r = 21.093 min (minor), 26.307 min (major): 90 % ee.

5f

HPLC chiralcel OJ column (10 % isopropanol in hexanes, 0.5 mL/min) t_r = 21.947 min (minor), 27.620 min (major): 92 % ee.

5g

HPLC chiralcel OJ column (10 % isopropanol in hexanes, 0.5 mL/min) t_r = 14.347 min (major), 17.787 min (minor): 65 % ee.



8a

HPLC chiralcel AD-H column (5 % isopropanol in hexanes, 0.1 mL/min) t_r = 37.700 min (major), 40.687 min (minor): 71 % ee.



8b

The TBS ether was deprotected and then ee was determined. HPLC chiralcel AS-H column (10 % isopropanol in hexanes, 0.3 mL/min) t_r = 20.167 min (minor), 23.213 min (major): 91 % ee.



8c

The TBS ether was deprotected and then ee was determined. HPLC chiralcel AS-H column (10 % isopropanol in hexanes, 0.2 mL/min) t_r = 39.120 min (major), 44.313 min (minor): 68 % ee.



8d

HPLC chiralcel AS-H column (5 % isopropanol in hexanes, 0.2 mL/min) t_r = 21.167 min (major), 24.160 min (minor): 72 % ee.

Deprotection of Trifluoromethanesulfonamides 31 (32)

To a 20 mL round bottle was added **3l** (1 equiv. 0.5 mmol), K_2CO_3 (2 equiv. 1.0 mmol), iodoacetonitrile (1.2 equiv. 0.6 mmol) and acetone (9mL). The mixture was stirred at room temperature for 12 h under nitrogen. Upon completion (monitored by TLC), the resulting solution was diluted by EtOAc, filtered and evaporated to afford the alkylated trifluoromethanesulfonamide, which could be used directly into the next step without purification.

To a 20 mL round bottle was added the alkylated trifluoromethanesulfonamide, Cs_2CO_3 (3 equiv.) and anhydrous THF. The mixture was stirred at 40 °C overnight under nitrogen. Upon completion, the resulting solution was cooled to 0 °C, at which time aq. HCl (3 N) was added. The mixture was stirred for another 2 h at room temperature and cooled to 0 °C. 1N NaOH was added to reach pH = 14, and BzCl (1.5 equiv.) was dissolved in Dioxane and added dropwise during 30 min at 0 °C. The mixture was warmed to room temperature and stirred for another 12 h. Upon completion, the mixture was cooled to 0 °C and 1N HCl was added to reach pH = 1. The mixture was extracted with EtOAc (20 mL × 3) and the organic layer was washed with brine and dried over MgSO₄. The solvent was evaporated under reduced pressure and the mixture was purified by column chromatography on silica gel (EtOAc/hexane= 1:10 as eluant) afforded **10** as a white solid (103 mg, 72 % yield). And the ee was determined by converting the product back to the triflate amide (95 % ee).

NHBz

10 N-(1-((2-iodo)-naphthalen-2-yl)ethyl)benzamide

¹H NMR (600 MHz, CDCl₃) δ 7.89 – 7.75 (m, 6H), 7.55 – 7.44 (m, 4H), 7.44 – 7.36 (m, 2H), 6.46 (d, *J* = 7.0 Hz, 1H), 5.56 – 5.44 (m, 1H), 1.69 (d, *J* = 6.9 Hz, 3H).

¹³C NMR (151 MHz, CDCl₃) δ 166.57, 140.44, 134.53, 133.32, 132.74, 131.47, 128.56, 128.54, 127.89, 127.59, 126.92, 126.23, 125.91, 124.77, 124.61, 49.25, 21.61.

Synthetic Transformations of Iodinated Products

Procedure for the preparation of compound 11

To a 10 mL schlenk tube was added **31** (1 equiv.), $Pd(PPh_4)_4$ (0.05 equiv.), HCO_2Na (2 equiv.) and DMF. The mixture was stirred under nitrogen at 110 °C for 5 h. After completion, the mixture was filtrated through a pad

of celite, concentrated and purified by prep-TLC to obtain a white solid (96 % yield).

Procedure for the preparation of compound 12

To a stirred solution of **3l** in THF was added iPrMgCl-LiCl (1.1 equiv.) dropwise at 0° C. The mixture was stirred at this temperature for 3 h and was quenched with D₂O. The resulting solution was extracted with EtOAc (3 times) and the organic layer was dried over MgSO₄, concentrated and purified by prep-TLC to obtain a white solid (99 % yield).



12 1,1,1-trifluoro-N-(1-(naphthalen-2-yl-3-d)ethyl)methanesulfonamide

¹H NMR (600 MHz, CDCl₃) δ 7.91 – 7.80 (m, 3H), 7.75 (s, 1H), 7.56 – 7.47 (m, 2H), 5.28 (d, *J* = 7.9 Hz, 1H), 5.01 – 4.92 (m, 1H), 1.72 (d, *J* = 6.9 Hz, 3H).

¹³C NMR (151 MHz, CDCl₃) δ 138.05, 133.17, 133.01, 128.99, 128.02, 127.70, 126.66, 126.54, 124.79, 123.37 (t, *J* = 23.9 Hz), 119.52 (q, *J* = 320.8 Hz), 55.38, 23.37.

Procedure for the preparation of compound 13(33)

To a 10 mL schlenk tube was added **31** (0.1 mmol, 1 equiv.), 3-methoxyphenyl boronic acid (1.5 equiv.), $Pd(OAc)_2$ (0.025 equiv.), S-phos (0.05 equiv.), K_2CO_3 (3 equiv.) and CH_3CN/H_2O . The mixture was stirred under N₂ at 100 °C for 12 h. After completion, the mixture was filtrated through a pad of celite, concentrated and purified by prep-TLC to obtain the product as a colorless oil (93 % yield).



13 1,1,1-trifluoro-N-(1-(3-(3-methoxyphenyl)naphthalen-2-yl)ethyl)methanesulfonamide

¹H NMR (600 MHz, CDCl₃) δ 7.89 – 7.83 (m, 2H), 7.83 – 7.79 (m, 1H), 7.71 (s, 1H), 7.54 – 7.48 (m, 2H), 7.39 (t, *J* = 7.9 Hz, 1H), 7.02 – 6.90 (m, 3H), 5.39 (d, *J* = 7.9 Hz, 1H), 5.19 – 5.11 (m, 1H), 3.85 (s, 3H), 1.47 (d, *J* = 6.8 Hz, 3H).

¹³C NMR (151 MHz, CDCl₃) δ 159.52, 141.27, 138.29, 138.13, 132.69, 132.42, 129.59, 129.49, 127.63, 127.59, 126.79, 126.57, 124.19, 121.75, 119.40 (q, *J* = 321.2 Hz), 114.70, 113.67, 55.28, 52.50, 24.96.

Procedure for the preparation of compound 14 (34)

To a 10 mL schlenk tube was added 31 (0.1 mmol, 1 equiv.), CuCN (2 equiv.), L-proline (2 equiv.) and DMF

(0.5 mL). The mixture was stirred under N₂ at 120 $^{\circ}$ C for 48 h. After completion, the mixture was filtrated through a pad of celite, concentrated and purified by prep-TLC to obtain the product as a colorless oil (67 % yield).

14 N-(1-(3-cyanonaphthalen-2-yl)ethyl)-1,1,1-trifluoromethanesulfonamide

¹H NMR (400 MHz, CDCl₃) δ 8.30 (s, 1H), 7.94 – 7.81 (m, 3H), 7.73 – 7.56 (m, 2H), 6.04 (d, *J* = 6.9 Hz, 1H), 5.25 – 5.10 (m, 1H), 1.81 (d, *J* = 7.1 Hz, 3H).

¹³C NMR (151 MHz, CDCl₃) δ 138.81, 136.63, 134.58, 131.54, 129.93, 128.18, 126.83, 119.39 (q, *J* = 321.3 Hz), 118.33, 106.82, 54.71, 23.27.

Procedure for the preparation of compound 15 (35)

To a 10 mL schlenk tube was added **31** (0.1 mmol, 1 equiv.), CuI (0.2 equiv.), 1,10-phen (0.4 equiv.) Cs_2CO_3 and MeOH (0.5 mL). The mixture was stirred under air at 110 °C for 24 h. After completion, the mixture was filtrated through a pad of celite, concentrated and purified by prep-TLC to obtain the product as a colorless oil (77 % yield).

15 1,1,1-trifluoro-N-(1-(3-methoxynaphthalen-2-yl)ethyl)methanesulfonamide

¹H NMR (600 MHz, CDCl₃) δ 7.75 (t, *J* = 8.5 Hz, 2H), 7.61 (s, 1H), 7.47 (ddd, *J* = 8.1, 7.0, 1.2 Hz, 1H), 7.38 (ddd, *J* = 8.1, 7.0, 1.1 Hz, 1H), 7.26 (s, 1H), 6.14 (d, *J* = 9.4 Hz, 1H), 4.98 – 4.87 (m, 1H), 4.02 (s, 3H), 1.69 (d, *J* = 7.0 Hz, 3H).

¹³C NMR (151 MHz, CDCl₃) δ 154.90, 133.97, 129.82, 128.51, 127.71, 127.44, 126.93, 126.45, 124.53, 119.48 (q, *J* = 321.0 Hz), 106.69, 55.53, 23.21.

Procedure for the preparation of compound 16

To a stirred solution of **3l** in THF was added iPrMgCl-LiCl (1.1 equiv.) dropwise at 0 $^{\circ}$ C. The mixture was stirred at this temperature for 3 h and PhCHO was added. The resulting solution was warmed to room temperature slowly and stirred for another 2 h. After completion, the mixture was quenched with aq. NH₄Cl, extracted with EtOAc (3 times) and the organic layer was dried over MgSO₄, concentrated and purified by prep-TLC to obtain a white solid (98 % yield).



$16\ 1, 1, 1-trifluoro-\mathit{N-(1-(3-(hydroxy(phenyl))methyl)naphthalen-2-yl)ethyl)} methanesulfon a mide the state of the st$

¹H NMR (600 MHz, CDCl₃) δ 7.85 – 7.78 (m, 2H), 7.74 (d, *J* = 7.8 Hz, 1H), 7.58 (s, 1H), 7.53 – 7.45 (m, 2H), 7.43 – 7.36 (m, 4H), 7.37 – 7.30 (m, 1H), 6.27 (s, 1H), 5.96 (s, 1H), 5.42 – 5.32 (m, 1H), 2.64 (s, 1H), 1.49 (d, *J* = 6.8 Hz, 3H).

¹³C NMR (151 MHz, CDCl₃) δ 142.04, 138.92, 137.25, 132.97, 132.40, 128.86, 128.65, 127.94, 127.86, 127.40, 126.93, 126.76, 126.56, 126.53, 119.45 (q, *J* = 321.3 Hz), 74.45, 52.30, 24.08.



















































































































2a Area % Report

 Data File:
 1a-SM-C:\Documents
 and
 Settings\Yu
 lab
 hplc\Desktop\lingchu-kinetic

 resolution\CL07028-SM-ee-5%-0.5mL-40min-ASH



DAD-CH2 205 nm Results				
Retention Time	Area	Area %	Height	Height %
11.200	1145874	3.09	76299	10.17
30.613	35976428	96.91	674125	89.83
Totals				
	37122302	100.00	750424	100.00

3a Area % Report



DAD-CH2 205 nm Results				
Retention Time	Area	Area %	Height	Height %
7.533	50036964	96.61	3294342	97.02
10.133	1753692	3.39	101221	2.98
Totals				
	51790656	100.00	3395563	100.00

2b Area % Report

 Data File:
 1b-SM-C:\Documents
 and
 Settings\Yu
 lab
 hplc\Desktop\lingchu-kinetic

 resolution\CL07026-SM-ee-5%-0.5mL-20min-ASH

 Method:
 C:\EZChrom Elite\Enterprise\Projects\Default\Method\A 120 min without fc 0.5 ml per min.met

 Acquired:
 9/2/2013 12:48:50 PM

 Printed:
 1/9/2014 9:26:15 PM



DAD-CH2 205 nm Results				
Retention Time	Area	Area %	Height	Height %
10.553	3593116	4.51	226860	8.83
16.260	76022434	95.49	2341695	91.17
Totals				
	79615550	100.00	2568555	100.00

3b Area % Report



DAD-CH1 250 nm Results		• • • • •	TT = 1.	H • 1 · 0/
Retention Time	Area	Area %	Height	Height %
11.153	8587300	95.40	426316	95.51
13.567	414327	4.60	20051	4.49
Totals				
	9001627	100.00	446367	100.00

2c Area % Report



DAD-CH2 205 nm Results				
Retention Time	Area	Area %	Height	Height %
9.733	11711510	11.63	566571	17.06
12.880	89015576	88.37	2753515	82.94
Totals				
	100727086	100.00	3320086	100.00

3c Area % Report



DAD-CH2 205 nm Results Retention Time	Area	Area %	Height	Height %
17.227	38692545	93.36	1330248	93.49
19.080	2750934	6.64	92660	6.51
Totals				
Totulo	41443479	100.00	1422908	100.00

2d Area % Report

 Data File:
 1f-SM-C:\Documents
 and
 Settings\Yu
 lab
 hplc\Desktop\lingchu-kinetic

 resolution\CL07099-2-sm-ee-5%-0.5mL-20min-ASH



DAD-CH2 205 nm Results				
Retention Time	Area	Area %	Height	Height %
8.447	553170	8.29	45644	12.10
9.600	6121454	91.71	331511	87.90
Totals				
	6674624	100.00	377155	100.00

3d Area % Report

 Data File:
 1f-PR-C:\Documents
 and
 Settings\Yu
 lab
 hplc\Desktop\lingchu-kinetic

 resolution\CL07099-pr-ee-5%-0.5mL-20min-ASH



DAD-CH2 205 nm Results				
Retention Time	Area	Area %	Height	Height %
8.427	25936124	89.85	1657243	90.55
9.567	2928957	10.15	172898	9.45
Totals				
	28865081	100.00	1830141	100.00

2e Area % Report

 Data File:
 1e-SM-C:\Documents
 and
 Settings\Yu
 lab
 hplc\Desktop\lingchu-kinetic

 resolution\CL07058-sm-ee-5%-0.5mL-25min-ASH



DAD-CH2 205 nm Results				
Retention Time	Area	Area %	Height	Height %
14.440	14849463	16.78	501233	23.95
18.167	73659831	83.22	1591743	76.05
Totals				
	88509294	100.00	2092976	100.00

3e Area % Report

 Data File:
 1e-PR-C:\Documents
 and
 Settings\Yu
 lab
 hplc\Desktop\lingchu-kinetic

 resolution\CL07058-pr-re-ee-5%-0.5mL-25min-ASH



DAD-CH2 205 nm Results				
Retention Time	Area	Area %	Height	Height %
14.480	39925253	87.03	1157407	87.66
18.213	5948378	12.97	162980	12.34
Totals				
	45873631	100.00	1320387	100.00

2f Area % Report

 Data File:
 1d-SM-C:\Documents
 and
 Settings\Yu
 lab
 hplc\Desktop\lingchu-kinetic

 resolution\CL06105-SM-ee-5%-0.8mL-30min-ASH



DAD-CH2 205 nm Results				
Retention Time	Area	Area %	Height	Height %
7.853	3447949	11.88	268289	49.09
26.180	25587027	88.12	278199	50.91
Totals				
	29034976	100.00	546488	100.00

3f Area % Report

 Data File:
 1d-PR-C:\Documents
 and
 Settings\Yu
 lab
 hplc\Desktop\lingchu-kinetic

 resolution\CL06105-pr-ee-5%-0.5mL-40min-ASH



DAD-CH2 205 nm Results				
Retention Time	Area	Area %	Height	Height %
13.733	75586167	97.67	1605480	98.52
19.760	1803880	2.33	24080	1.48
Totals				
	77390047	100.00	1629560	100.00

2g Area % Report

Data File:3a-SM-C:\DocumentsandSettings\Yulabhplc\Desktop\lingchu-kineticresolution\CL06134-SM-ee-5%-0.8mL-40min-ASHMethod:C:\EZChrom Elite\Enterprise\Projects\Default\Method\A 120 min without fc 0.5 ml per min.metAcquired:8/22/2013 10:14:50 PMPrinted:1/10/2014 10:24:15 AM



DAD-CH2 205 nm Results		• • • • •	TT • 1.	H : 1. 0/
Retention Time	Area	Area %	Height	Height %
9.687	1759980	3.56	114474	14.75
32.247	47725381	96.44	661858	85.25
Totals				
	49485361	100.00	776332	100.00

3g Area % Report

 Data File:
 3a-PR-C:\Documents
 and
 Settings\Yu
 lab
 hplc\Desktop\lingchu-kinetic

 resolution\CL06134-PR-ee-5%-0.8mL-40min-ASH



DAD-CH2 205 nm Results				
Retention Time	Area	Area %	Height	Height %
9.387	81172696	98.49	4268645	99.06
21.127	1248480	1.51	40304	0.94
Totals				
	82421176	100.00	4308949	100.00

2h Area % Report

 Data File:
 3b-SM-C:\Documents
 and
 Settings\Yu
 lab
 hplc\Desktop\lingchu-kinetic

 resolution\CL06123-SM-ee-5%-0.8mL-13min-ASH



DAD-CH2 205 nm Results				
Retention Time	Area	Area %	Height	Height %
7.833	2750965	5.78	208061	9.22
11.200	44857038	94.22	2049113	90.78
Totals				
	47608003	100.00	2257174	100.00

3h Area % Report

 Data File:
 3b-PR-C:\Documents
 and
 Settings\Yu
 lab
 hplc\Desktop\lingchu-kinetic

 resolution\CL06123-PR-ee-5%-0.8mL-13min-ASH



DAD-CH2 205 nm Results				
Retention Time	Area	Area %	Height	Height %
8.447	84402874	98.06	4729336	98.18
12.593	1673606	1.94	87486	1.82
Totals				
	86076480	100.00	4816822	100.00

2i Area % Report

Data File:3c-SM-C:\DocumentsandSettings\Yulabhplc\Desktop\lingchu-kineticresolution\CL07029-sm-ee-5%-0.5mL-40min-ASHMethod:C:\EZChrom Elite\Enterprise\Projects\Default\Method\A 120 min without fc 0.5 ml per min.metAcquired:9/6/2013 12:56:41 AMPrinted:1/10/2014 10:32:03 AM



DAD-CH2 205 nm Results				
Retention Time	Area	Area %	Height	Height %
11.893	892586	1.46	30779	2.48
25.320	60213612	98.54	1208100	97.52
Totals				
	61106198	100.00	1238879	100.00

3i Area % Report



DAD-CH1 250 nm Results				
Retention Time	Area	Area %	Height	Height %
12.047	11338520	97.04	482224	97.52
15.640	346014	2.96	12270	2.48
Totals				
	11684534	100.00	494494	100.00

2j Area % Report

Data File:3d-SM-C:\DocumentsandSettings\Yulabhplc\Desktop\lingchu-kineticresolution\CL06120-SM-ee-5%-0.8mL-20min-ASHMethod:C:\EZChrom Elite\Enterprise\Projects\Default\Method\A 120 min without fc 0.5 ml per min.metAcquired:8/22/2013 8:38:28 PMPrinted:1/10/2014 10:37:29 AM



DAD-CH2 205 nm Results				
Retention Time	Area	Area %	Height	Height %
7.100	5722341	11.12	452938	18.91
12.013	45735849	88.88	1941802	81.09
Totals				
	51458190	100.00	2394740	100.00
3j Area % Report

 Data File:
 3d-PR-C:\Documents
 and
 Settings\Yu
 lab
 hplc\Desktop\lingchu-kinetic

 resolution\CL06120-PR-ee-5%-0.8mL-20min-ASH



DAD-CH2 205 nm Results				
Retention Time	Area	Area %	Height	Height %
7.967	45919187	96.11	2604305	97.69
15.753	1860675	3.89	61618	2.31
Totals				
	47779862	100.00	2665923	100.00

2k Area % Report

Data File:3e-SM-C:\EZChromElite\Enterprise\Projects\Default\Data\LingChu\CL08098-SM-ee-10%-0.3mL-40min-ASHMethod:C:\EZChrom Elite\Enterprise\Projects\Default\Method\A 120 min without fc 0.5 ml per min.metAcquired:12/26/2013 10:41:34 AMPrinted:1/10/2014 10:44:33 AM



DAD-CH2 205 nm Results				
Retention Time	Area	Area %	Height	Height %
15.747	4094157	10.89	185908	15.70
22.540	33507222	89.11	998394	84.30
Totals				
	37601379	100.00	1184302	100.00

3k Area % Report

Data File:3e-PR-C:\EZChromElite\Enterprise\Projects\Default\Data\LingChu\CL08098-PR-ee-10%-0.3mL-60min-ASHMethod:C:\EZChrom Elite\Enterprise\Projects\Default\Method\A 120 min without fc 0.5 ml per min.metAcquired:12/26/2013 11:09:20 AMPrinted:1/10/2014 10:47:38 AM



DAD-CH2 205 nm Results				
Retention Time	Area	Area %	Height	Height %
17.080	36972202	97.33	1420820	98.38
41.727	1015904	2.67	23411	1.62
Totals				
	37988106	100.00	1444231	100.00

2l Area % Report

 Data File:
 3f-SM-C:\Documents
 and
 Settings\Yu
 lab
 hplc\Desktop\lingchu-kinetic

 resolution\CL07084-sm-ee-5%-0.5mL-30min-ASH



DAD-CH2 205 nm Results				
Retention Time	Area	Area %	Height	Height %
17.173	39431375	16.58	1150810	25.70
23.427	198359465	83.42	3326859	74.30
Totals				
	237790840	100.00	4477669	100.00

31 Area % Report

 Data File:
 3f-PR-C:\Documents
 and
 Settings\Yu
 lab
 hplc\Desktop\lingchu-kinetic

 resolution\CL07084-pr-ee-5%-0.5mL-45min-ASH



DAD-CH2 205 nm Results		• • • • •	TT * 17	H : 14.04
Retention Time	Area	Area %	Height	Height %
17.880	93406456	97.10	2258366	98.39
36.760	2792665	2.90	36964	1.61
Totals				
	96199121	100.00	2295330	100.00

2m Area % Report

 Data File:
 C:\EZChrom

 Elite\Enterprise\Projects\Default\Data\LingChu\CL10022-1-sm-re-ee-10%-0.5mL-25min-ASH

 Method:
 C:\EZChrom Elite\Enterprise\Projects\Default\Method\A 25 min without frc 0.5 ml per min.met

 Acquired:
 7/3/2014 2:26:48 PM

 Printed:
 7/6/2014 3:10:04 PM



DAD-CH2 205 nm Results				
Retention Time	Area	Area %	Height	Height %
9.827	1111503	14.05	74561	16.76
10.853	6799405	85.95	370221	83.24
Totals				
	7910908	100.00	444782	100.00

3m-mono Area % Report

 Data File:
 C:\EZChrom

 Elite\Enterprise\Projects\Default\Data\LingChu\CL10022-1-PR-ee-10%-0.5mL-20min-ASH

 Method:
 C:\EZChrom Elite\Enterprise\Projects\Default\Method\A 20 min without fc 0.5 ml per min.met

 Acquired:
 7/2/2014 11:11:43 PM

 Printed:
 7/6/2014 3:15:48 PM



DAD-CH2 205 nm Results				
Retention Time	Area	Area %	Height	Height %
10.893	46737471	93.06	2215271	93.66
12.773	3487817	6.94	149915	6.34
Totals				
	50225288	100.00	2365186	100.00

3m-di Area % Report

 Data File:
 C:\EZChrom

 Elite\Enterprise\Projects\Default\Data\LingChu\CL10022-1-di-ee-10%-0.5mL-25min-ASH

 Method:
 C:\EZChrom Elite\Enterprise\Projects\Default\Method\A 25 min without frc 0.5 ml per min.met

 Acquired:
 7/3/2014 2:00:26 PM

 Printed:
 7/6/2014 3:19:30 PM



DAD-CH2 205 nm Results				
Retention Time	Area	Area %	Height	Height %
10.400	21413422	99.73	1252953	99.79
12.860	57268	0.27	2630	0.21
Totals				
	21470690	100.00	1255583	100.00

2n Area % Report

Data File:C:\EZChromElite\Enterprise\Projects\Default\Data\LingChu\CL10075-1-sm-ee-10%-0.5mL-25min-ASHMethod:C:\EZChrom Elite\Enterprise\Projects\Default\Method\A 25 min without frc 0.5 ml per min.metAcquired:8/19/2014 11:15:48 AMPrinted:8/24/2014 5:40:09 PM



DAD-CH1 250 nm Results				
Retention Time	Area	Area %	Height	Height %
7.807	548903	27.89	44676	32.78
9.287	1419192	72.11	91613	67.22
Totals				
	1968095	100.00	136289	100.00

Area % Report

 Data File:
 C:\EZChrom

 Elite\Enterprise\Projects\Default\Data\LingChu\CL10075-1-re-pr-ee-10%-0.5mL-25min-ASH

 Method:
 C:\EZChrom Elite\Enterprise\Projects\Default\Method\A 25 min without frc 0.5 ml per min.met

 Acquired:
 8/19/2014 11:42:10 AM

 Printed:
 8/24/2014 5:42:48 PM



DAD-CH1 250 nm Results				
Retention Time	Area	Area %	Height	Height %
8.273	5953359	94.65	427788	95.26
10.113	336756	5.35	21301	4.74
Totals				
	6290115	100.00	449089	100.00

 $\label{eq:2.1} Data \ File: \ 5a-SM-C: \ box{Documents} \ and \ Settings \ u \ lab \ hplc \ box{Desktop} \ ingchu-kinetic \ resolution \ CL08040-SM-ee-10\%-0.3mL-60min-OJ \ u \ box{Desktop} \ box{Desk$

Method:C:\EZChrom Elite\Enterprise\Projects\Default\Method\A 120 min without fc 0.5 ml per min.metAcquired:12/27/2013 12:39:27 PMPrinted:1/10/2014 11:03:52 AM



DAD-CH2 205 nm Results				
Retention Time	Area	Area %	Height	Height %
45.227	4338698	7.77	61188	10.81
50.520	51512114	92.23	505050	89.19
Totals				
	55850812	100.00	566238	100.00

6a Area % Report

Data File:5a-PR-C:\EZChromElite\Enterprise\Projects\Default\Data\LingChu\CL08040-PR-ee-10%-0.2mL-45min-ADHMethod:C:\EZChrom Elite\Enterprise\Projects\Default\Method\A 120 min without fc 0.5 ml per min.metAcquired:1/11/2014 11:54:45 AMPrinted:1/14/2014 10:05:51 AM



DAD-CH2 205 nm Results				
Retention Time	Area	Area %	Height	Height %
29.827	1888425	1.95	49058	2.55
32.413	95174677	98.05	1876398	97.45
Totals				
	97063102	100.00	1925456	100.00

5b Area % Report

 Data File:
 5b-SM-C:\Documents
 and
 Settings\Yu
 lab
 hplc\Desktop\lingchu-kinetic

 resolution\CL07116-sm-ee-10%-0.5mL-30min-ASH



DAD-CH2 205 nm Results				
Retention Time	Area	Area %	Height	Height %
17.280	1670982	3.59	59640	5.09
18.913	44850513	96.41	1111276	94.91
Totals				
	46521495	100.00	1170916	100.00

6b Area % Report

 Data File:
 5b-PR-C:\Documents
 and
 Settings\Yu
 lab
 hplc\Desktop\lingchu-kinetic

 resolution\CL07116-pr-ee-10%-0.3mL-40min-ASH



DAD-CH2 205 nm Results	A	A	II.:-h4	
Retention Time	Area	Area %	Height	Height %
29.640	2037367	2.13	48992	2.88
31.907	93423783	97.87	1649466	97.12
Totals				
	95461150	100.00	1698458	100.00

5c Area % Report

Data File:5c-SM-C:\EZChromElite\Enterprise\Projects\Default\Data\LingChu\CL08096-SM-ee-10%-0.5mL-60min-OJMethod:C:\EZChrom Elite\Enterprise\Projects\Default\Method\A 120 min without fc 0.5 ml per min.metAcquired:1/5/2014 2:21:19 PMPrinted:1/10/2014 11:17:15 AM



DAD-CH2 205 nm Results				
Retention Time	Area	Area %	Height	Height %
16.933	27482431	96.36	373223	96.65
29.307	1037923	3.64	12939	3.35
Totals				
	28520354	100.00	386162	100.00

6c Area % Report

Data File:5c-PR-C:\EZChromElite\Enterprise\Projects\Default\Data\LingChu\CL08096-PR-ee-5%-0.5mL-40min-ADHMethod:C:\EZChrom Elite\Enterprise\Projects\Default\Method\A 120 min without fc 0.5 ml per min.metAcquired:1/6/2014 9:35:51 PMPrinted:1/10/2014 11:25:25 AM



DAD-CH2 205 nm Results				
Retention Time	Area	Area %	Height	Height %
11.980	211866	3.09	11678	3.78
14.693	6650494	96.91	297508	96.22
Totals				
	6862360	100.00	309186	100.00

5d Area % Report

 Data File:
 5d-SM-C:\Documents
 and
 Settings\Yu
 lab
 hplc\Desktop\lingchu-kinetic

 resolution\CL08081-SM-ee-10%-0.5mL-60min-OJ



DAD-CH2 205 nm Results Retention Time	Area	Area %	Height	Height %
42 313	3405330	9.28	47562	16 55
52 002	22204192	00.72	220824	10.33 92.45
33.093	55504182	90.72	239824	05.45
Totals				
	36709512	100.00	287386	100.00

6d Area % Report

 Data File:
 5d-PR-C:\Documents
 and
 Settings\Yu
 lab
 hplc\Desktop\lingchu-kinetic

 resolution\CL08081-PR-ee-10%-0.5mL-60min-OJ



DAD-CH2 205 nm Results		• • • • •	TT • 17	H • 1/0/
Retention Time	Area	Area %	Height	Height %
31.120	39645952	99.00	358370	98.41
39.807	402004	1.00	5804	1.59
Totals				
	40047956	100.00	364174	100.00

5e Area % Report

Data File:C:\EZChromElite\Enterprise\Projects\Default\Data\LingChu\CL09065-2-SM-ee-10%-0.5mL-75min-OJMethod:C:\EZChrom Elite\Enterprise\Projects\Default\Method\A 60 min without fc 0.4 ml per min.metAcquired:4/2/2014 1:07:23 PMPrinted:4/4/2014 9:18:36 PM



DAD-CH2 205 nm Results				
Retention Time	Area	Area %	Height	Height %
19.987	4926216	4.99	111914	7.33
24.753	93889663	95.01	1414079	92.67
Totals				
	98815879	100.00	1525993	100.00

6e Area % Report

Data File:5f-PR-C:\EZChromElite\Enterprise\Projects\Default\Data\LingChu\CL08058-PR-ee-10%-0.3mL-75min-OJMethod:C:\EZChrom Elite\Enterprise\Projects\Default\Method\A 120 min without fc 0.5 ml per min.metAcquired:12/13/2013 10:18:54 PMPrinted:1/10/2014 2:42:08 PM



DAD-CH2 205 nm Results				
Retention Time	Area	Area %	Height	Height %
37.327	74377760	99.06	593490	98.15
44.560	707487	0.94	11171	1.85
Totals				
	75085247	100.00	604661	100.00

5f Area % Report

Data File:C:\EZChromElite\Enterprise\Projects\Default\Data\LingChu\CL08127-SM-ee-10%-0.5mL-40min-OJMethod:C:\EZChrom Elite\Enterprise\Projects\Default\Method\A 60 min without fc 0.4 ml per min.metAcquired:2/20/2014 10:03:16 AMPrinted:4/4/2014 9:16:47 PM



DAD-CH2 205 nm Results				
Retention Time	Area	Area %	Height	Height %
22.507	3389678	3.77	61198	5.54
27.940	86516469	96.23	1044197	94.46
Totals				
	89906147	100.00	1105395	100.00

6f Area % Report

 Data File:
 5e-PR-C:\Documents
 and
 Settings\Yu
 lab
 hplc\Desktop\lingchu-kinetic

 resolution\CL08027-pr-ee-10%-0.5mL-40min-OJ



DAD-CH2 205 nm Results				
Retention Time	Area	Area %	Height	Height %
19.533	145655911	99.37	1723908	98.87
28.627	923417	0.63	19697	1.13
Totals				
	146579328	100.00	1743605	100.00

5g Area % Report

Data File:C:\EZChromElite\Enterprise\Projects\Default\Data\LingChu\CL09066-2-SM-ee-10%-0.5mL-75min-OJMethod:C:\EZChrom Elite\Enterprise\Projects\Default\Method\A 60 min without fc 0.4 ml per min.metAcquired:4/2/2014 1:38:43 PMPrinted:4/4/2014 9:22:34 PM



DAD-CH2 205 nm Results				
Retention Time	Area	Area %	Height	Height %
14.347	19794564	17.88	443985	23.24
17.787	90889220	82.12	1466167	76.76
Totals				
	110683784	100.00	1910152	100.00

6g Area % Report

Data File:C:\EZChromElite\Enterprise\Projects\Default\Data\LingChu\CL09066-1-PR-ee-10%-0.2mL-75min-OJMethod:C:\EZChrom Elite\Enterprise\Projects\Default\Method\A 60 min without fc 0.4 ml per min.metAcquired:4/2/2014 3:06:55 PMPrinted:4/4/2014 9:20:20 PM



DAD-CH2 205 nm Results				
Retention Time	Area	Area %	Height	Height %
31.760	525971055	99.66	4319443	99.64
40.273	1776529	0.34	15579	0.36
Totals				
	527747584	100.00	4335022	100.00

8a Area % Report

 Data File:
 5g-SM-C:\Documents
 and
 Settings\Yu
 lab
 hplc\Desktop\lingchu-kinetic

 resolution\CL08032-sm-ee-5%-0.1mL-60min-ADH



DAD-CH2 205 nm Results				
Retention Time	Area	Area %	Height	Height %
37.700	25074572	85.99	404499	86.26
40.687	4086691	14.01	64419	13.74
Totals				
	29161263	100.00	468918	100.00

9a-mono Area % Report

 Data File:
 5g-mono-C:\Documents
 and
 Settings\Yu
 lab
 hplc\Desktop\lingchu-kinetic

 resolution\CL08032-mono-ee-5%-0.1mL-60min-ADH

 Method:
 C:\EZChrom Elite\Enterprise\Projects\Default\Method\A 120 min without fc 0.5 ml per min.met

 Acquired:
 11/16/2013 3:15:12 PM

 Printed:
 1/10/2014 2:49:37 PM



DAD-CH2 205 nm Results				
Retention Time	Area	Area %	Height	Height %
37.320	5302078	6.34	83864	7.34
40.260	78360330	93.66	1058741	92.66
Totals				
	83662408	100.00	1142605	100.00

9a-di Area % Report

 Data File:
 5g-di-C:\Documents
 and
 Settings\Yu
 lab
 hplc\Desktop\lingchu-kinetic

 resolution\CL08032-di-ee-5%-0.1mL-60min-ADH



DAD-CH2 205 nm Results				
Retention Time	Area	Area %	Height	Height %
32.867	608676	0.25	11685	0.36
39.473	238492352	99.75	3190222	99.64
Totals				
	239101028	100.00	3201907	100.00

8b Area % Report

 Data File:
 5h-SM-C:\Documents
 and
 Settings\Yu
 lab
 hplc\Desktop\lingchu-kinetic

 resolution\CL07119-3-sm-ee-10%-0.3mL-40min-ASH



DAD-CH2 205 nm Results				
Retention Time	Area	Area %	Height	Height %
20.167	5106688	4.33	146380	5.76
23.213	112728212	95.67	2393278	94.24
Totals				
	117834900	100.00	2539658	100.00

9b Area % Report

 Data File:
 5h-PR-C:\Documents
 and
 Settings\Yu
 lab
 hplc\Desktop\lingchu-kinetic

 resolution\CL07119-pr-ee-10%-0.3mL-40min-ASH



DAD-CH2 205 nm Results				
Retention Time	Area	Area %	Height	Height %
23.200	1435500	4.21	45548	6.80
30.947	32677521	95.79	624377	93.20
Totals				
	34113021	100.00	669925	100.00

8c Area % Report

Data File:5i-SM-C:\EZChromElite\Enterprise\Projects\Default\Data\LingChu\CL08079-SM-ee-re-10%-0.2mL-45min-ASHMethod:C:\EZChrom Elite\Enterprise\Projects\Default\Method\A 120 min without fc 0.5 ml per min.metAcquired:1/12/2014 10:28:40 PMPrinted:1/14/2014 10:09:32 AM



DAD-CH1 250 nm Results				
Retention Time	Area	Area %	Height	Height %
39.120	4841826	83.98	81094	83.65
44.313	923718	16.02	15851	16.35
Totals				
	5765544	100.00	96945	100.00

9c Area % Report

Data File:5i-PR-C:\EZChromElite\Enterprise\Projects\Default\Data\LingChu\CL08079-PR-ee-5%-0.3mL-60min-ASHMethod:C:\EZChrom Elite\Enterprise\Projects\Default\Method\A 120 min without fc 0.5 ml per min.metAcquired:1/6/2014 1:19:09 PMPrinted:1/14/2014 10:16:46 AM



DAD-CH2 205 nm Results				
Retention Time	Area	Area %	Height	Height %
13.473	69496817	99.62	2861660	99.54
14.520	266465	0.38	13167	0.46
Totals				
	69763476	100.00	2874827	100.00

8d Area % Report

Data File:5j-SM-C:\EZChromElite\Enterprise\Projects\Default\Data\LingChu\CL08077-SM-ee-5%-0.2mL-45min-ASHMethod:C:\EZChrom Elite\Enterprise\Projects\Default\Method\A 120 min without fc 0.5 ml per min.metAcquired:1/13/2014 10:07:05 AMPrinted:1/14/2014 10:19:31 AM



DAD-CH2 205 nm Results				
Retention Time	Area	Area %	Height	Height %
21.167	66528912	85.95	1780549	86.69
24.160	10879612	14.05	273294	13.31
Totals				
	77408524	100.00	2053843	100.00

9d Area % Report

Data File:5j-PR-C:\EZChromElite\Enterprise\Projects\Default\Data\LingChu\CL08077-PR-ee-5%-0.1mL-90min-ASHMethod:C:\EZChrom Elite\Enterprise\Projects\Default\Method\A 120 min without fc 0.5 ml per min.metAcquired:1/13/2014 12:47:39 PMPrinted:1/14/2014 10:22:56 AM



DAD-CH2 205 nm Results				
Retention Time	Area	Area %	Height	Height %
48.873	1635782	1.81	24985	3.08
53.613	88890110	98.19	785945	96.92
Totals				
	90525892	100.00	810930	100.00

X-ray crystallographic data for 31

The single crystal X-ray diffraction studies were carried out on a Bruker Kappa APEX CCD diffractometer equipped with Mo K_{α} radiation ($\lambda = 0.71073$ Å). A 0.217 x 0.153 x 0.055 mm colorless block was mounted on a Cryoloop with Paratone oil. Data were collected in a nitrogen gas stream at 100(2) K using ϕ and ϖ scans. Crystal-to-detector distance was 40 mm and exposure time was 5 seconds per frame using a scan width of 2.0°. Data collection was 99.6% complete to 25.00° in θ . A total of 8740 reflections were collected covering the indices, -11 <=h<=11, -9 <=k<=9, -13 <=l<=13. 2957 reflections were found to be symmetry independent, with a R_{int} of 0.0307. Indexing and unit cell refinement indicated a primitive, monoclinic lattice. The space group was found to be *P*2₁. The data were integrated using the Bruker SAINT software program and scaled using the SADABS software program. Solution by direct methods (SHELXM) produced a complete phasing model consistent with the proposed structure.

All nonhydrogen atoms were refined anisotropically by full-matrix least-squares (SHELXL-2014). All hydrogen atoms were placed using a riding model. Their positions were constrained relative to their parent atom using the appropriate HFIX command in SHELXL-2014. Crystallographic data are summarized in Table S2.

Figure S1. X-ray Crystallographic Data of 31



Table S2. Crystal data and structure refinement for 31

Report date	2014-03-27	2014-03-27		
Identification code	Yu34	Yu34		
Empirical formula	C13 H11 F3 I N O2 S	C13 H11 F3 I N O2 S		
Molecular formula	C13 H11 F3 I N O2 S	C13 H11 F3 I N O2 S		
Formula weight	429.19	429.19		
Temperature	100 K	100 K		
Wavelength	0.71073 Å			
Crystal system	Monoclinic			
Space group	P 1 21 1			
Unit cell dimensions	a = 9.653(2) Å	$\alpha = 90^{\circ}$.		
	b = 7.5500(16) Å	$\beta = 106.626(6)^{\circ}.$		
	c = 10.651(2) Å	$\gamma = 90^{\circ}$.		
Volume	743.7(3) Å ³			
Z	2			
Density (calculated)	1.916 Mg/m ³			
Absorption coefficient	2.328 mm ⁻¹			
F(000)	416			
Crystal size	0.217 x 0.153 x 0.055 m	m ³		
Crystal color, habit	Colorless Block			
Theta range for data collection	3.357 to 26.358°.			
Index ranges	-11<=h<=11, -9<=k<=9,	-13<=l<=13		
Reflections collected	8740			
Independent reflections	2957 [R(int) = 0.0307]			
Completeness to theta = 25.000°	99.6 %			
Absorption correction	Semi-empirical from equ	uvalents		
Max. and min. transmission	0.0932 and 0.0585			
Refinement method	Full-matrix least-squares	s on F ²		
Data / restraints / parameters	2957 / 2 / 195			
Goodness-of-fit on F ²	1.043			
Final R indices [I>2sigma(I)]	R1 = 0.0235, wR2 = 0.04	R1 = 0.0235, $wR2 = 0.0449$		
R indices (all data)	R1 = 0.0254, wR2 = 0.04	R1 = 0.0254, $wR2 = 0.0460$		
Absolute structure parameter	0.007(13)	0.007(13)		
Extinction coefficient	n/a			
Largest diff. peak and hole	0.662 and -0.445 e.Å ⁻³	0.662 and -0.445 e.Å ⁻³		

References:

- 1. R. Giri, B.-F. Shi, K. M. Engle, N. Maugel, J.-Q. Yu, Transition metal-catalyzed C–H activation reactions: diastereoselectivity and enantioselectivity. *Chem. Soc. Rev.* **38**, 3242–3272 (2009).
- M. P. Doyle, R. Duffy, M. Ratnikov, L. Zhou, Catalytic carbene insertion into C–H bonds. *Chem. Rev.* 110, 704–724 (2010).
- 3. R. P. Reddy, H. M. L. Davies, Dirhodium tetracarboxylates derived from adamantylglycine as chiral catalyst for enantioselective C–H aminations. *Org. Lett.* **8**, 5013–5016 (2006).
- C. Liang, F. Robert-Peillard, C. Fruit, P. Muller, R. H. Dodd, P. Dauban, Efficient diastereoselective intermolecular rhodium-catalyzed C–H amination. *Angew. Chem. Int. Ed.* 45, 4641–4644 (2006).
- 5. D. N. Zalatan, J. Du Bois, A chiral rhodium carboxamidate catalyst for enantioselective C–H amination. *J. Am. Chem. Soc.* **130**, 9220–9221 (2008).
- E. Milczek, N. Boudet, S. Blakey, Enantioselective C–H amination using cationic ruthenium (II)–pybox catalysts. *Angew. Chem. Int. Ed.* 47, 6825–6828 (2008).
- 7. T. K. Hyster, L. Knorr, T. R. Ward, T. Rovis, Biotinylated Rh(III) complex in engineered streptavidin for accelerated asymmetric C–H activation. *Science* **338**, 500–503 (2012).
- B. Ye, N. Cramer, Chiral cyclopentadienyl ligand as stereocontrolling element in asymmetric C–H functionalization. *Science* 338, 504–506 (2012).
- F. Kakiuchi, P. L. Gendre, A. Yamada, H. Ohtaki, S. Murai, Atropselective alkylation of biaryl compounds by means of transition metal-catalyzed C–H/olefin coupling. *Tetrahedron: Asymmetry* 11, 2647–2651 (2000).
- B.-F. Shi, N. Maugel, Y.-H. Zhang, J.-Q. Yu, Pd^{II}-catalyzed enantioselective activation of C(sp²)–H and C(sp³)–H bonds using monoprotected amino acids as chiral ligands. *Angew. Chem. Int. Ed.* 47, 4882–4886 (2008).
- L. Chu, X.-C. Wang, C. E. Moore, A. L. Rheingold, J.-Q. Yu, Pd-catalyzed enantioselective C–H iodination: asymmetric synthesis of chiral diarylmethylamines. J. Am. Chem. Soc. 135, 16344–16347 (2013).
- M. Nakanishi, D. Katayev, C. Besnard, E. P. Kündig, Fused indolines by palladium-catalyzed asymmetric C–C coupling involving an unactivated methylene group. *Angew. Chem. Int. Ed.* 50, 7438–7441 (2011).
- 13. S. Anas, A. Cordi, H. B. Kagan, Enantioselective synthesis of 2-methyl indolines by palladium catalyzed asymmetric C(sp³)–H activation/cyclisation. *Chem. Comm.* **47**, 11483–11485 (2011).
- T. Saget, S. Lemouzy, N. Cramer, Chiral monodentate phosphines and bulky carboxylic acids: cooperative effects in palladium-catalyzed enantioselective C(sp³)–H functionalization. *Angew. Chem. Int. Ed.* 51, 2238–2242 (2012).
- D. Katayev, M. Nakanishi, T. Bürgi, E. P. Kündig, Asymmetric C(sp³)-H/C(Ar) coupling reactions. Highly enantio-enriched indolines via regiodivergent reaction of a racemic mixture. *Chem. Sci.* 3, 1422–1425 (2012).
- F. Schmidt, R. T. Stemmler, J. Rudolph, C. Bolm, Catalytic asymmetric approaches towards enantiomerically enriched diarylmethanols and diarylmethylamines. *Chem. Soc. Rev.* 35, 454–470 (2006).
- M. T. Robak, M. A. Herbage, J. A. Ellman, Synthesis and application of *tert*-butanesulfinamide. *Chem. Rev.* **110**, 3600–3740 (2010).

- J. Paetzold, J. E. Bäckvall, Chemoenzymatic dynamic resolution of primary amines. J. Am. Chem. Soc. 127, 17620–17621 (2005).
- M. Breuer, K. Ditrich, T. Habicher, B. Hauer, M. Keβeler, R. Stürmer, T. Zelinski, Industrial methods for the production of optically active intermediates. *Angew. Chem. Int. Ed.* 43, 788–824 (2004).
- 20. S. Arai, S. Bellemin-Laponnaz, G. C. Fu, Kinetic resolution of amines by a nonenzymatic acylation catalyst. *Angew. Chem. Int. Ed.* **40**, 234–236 (2001).
- C. K. De, E. G. Klauber, D. Seidel, Merging nucleophilic and hydrogen bonding catalysis: an anion binding approach to the kinetic resolution of amines. *J. Am. Chem. Soc.* 131, 17060–17061 (2009).
- 22. B. S. Fowler, P. J. Mikochik, S. J. Miller, Peptide-catalyzed kinetic resolution of formamides and thioformamides as an entry to nonracemic amines. *J. Am. Chem. Soc.* **132**, 2870–2871 (2011).
- D. -W. Gao, Q. Gu, S. -L. You, Pd(II)-catalyzed intermolecular direct C–H bond Iodination: an efficient approach toward the synthesis of axially chiral compounds via kinetic resolution. ACS *Catal.* 4, 2741–2745 (2014).
- 24. D. R. Jensen, J. S. Pugsley, M. S. Sigman, Palladium-catalyzed enantioselective oxidations of alcohols using molecular oxygen. *J. Am. Chem. Soc.* **123**, 7475–7476 (2001).
- 25. E. M. Ferreira, B. M. Stoltz, The palladium-catalyzed oxidative kinetic resolution of secondary alcohols with molecular oxygen. *J. Am. Chem. Soc.* **123**, 7725–7726 (2001).
- M. Tokunaga, J. F. Larrow, F. Kakiuchi, E. N. Jacobsen, Asymmetric catalysis with water: efficient kinetic resolution of terminal epoxides by means of catalytic hydrolysis. *Science* 277, 936–938 (1997).

- J. Song, Y. Wang, L. Deng, The mannich reaction of malonates with simple imines catalyzed by bifunctional cinchona alkaloids: enantioselective synthesis of β-amino acids. *J. Am. Chem. Soc.* 128, 6048–6049 (2006).
- A. Berkessel, F. Cleemann, S. Mukherjee, Kinetic resolution of oxazinones: an organocatalytic approach to enantiomerically pure β-amino acids. *Angew. Chem. Int. Ed.* 44, 7466–7469 (2005).
- V. D. Bumbu, V. B. Birman, Kinetic resolution of *N*-acyl-β-lactams via benzotetramisole-catalyzed enantioselective alcoholysis. *J. Am. Chem. Soc.* 133, 13902–13905 (2011).
- S. Zhou, J. Wang, X. Chen, J. L. Acena, V. A. Soloshonok, H. Liu, Chemical kinetic resolution of unprotected β-substituted-β-amino acids using recyclable chiral ligands. *Angew. Chem. Int. Ed.* 53, 7883–7886 (2014).
- Y. Zhang, Z. Lu, A. Desai, W. D. Wulff, Mapping the active site in a chemzyme: diversity in the *N*-substituent in the catalytic asymmetric aziridination of imines. *Org. Lett.* 10, 5429–5432 (2008).
- K. S. L., Chan, M. Wasa, L. Chu, B. N., Laforteza, M. Miura, J.-Q. Yu, Ligand–enabled cross-coupling of C(sp³)–H bonds with arylboron reagents via Pd(II)/Pd(0) catalysis. *Nature Chem.* 6, 146–150 (2014).
- 33. K. L. Billingsley, K. W., Anderson, S. L. Buchwald, A highly active catalyst for Suzuki–Miyaura cross-coupling reactions of heteroaryl compounds. *Angew. Chem. Int. Ed.* **45**, 3484–3488 (2006).
- 34. T.-S. Mei, D.-H. Wang, J.-Q. Yu, Expedient drug synthesis and diversification via *ortho*-C–H iodination using recyclable PdI₂ as the precatalyst. *Org. Lett.* **12**, 3140–3143 (2010).

35. M. Wolter, G. Nordmann, G. E. Job, S. L. Buchwald, Copper-catalyzed coupling of aryl iodides with aliphatic alcohols. *Org. Lett.* **4**, 973–976 (2002).