



**Figure S1**  
**Alignment of the *ska* genes from the serotype M1, M2, M6, and M28 strains used in this study.**  
 The *ska* genes, and 100 nucleotides upstream of the ATG start codon, were aligned. Nucleotides that are identical across all four *ska* alleles are colored red. The start codon is colored green. The stop codon is colored blue. The nine nucleotides that base-pair to the Fas sRNA in serotype M1 GAS are highlighted in a purple box.

Strain Name	Information	Reference
MGAS2221	Serotype M1 clinical GAS isolate that has been genome sequenced	Sumby <i>et al.</i> , 2006
2221ΔFasX	MGAS2221 derivative in which the <i>fasX</i> gene has been replaced with a spectinomycin resistance cassette	Ramirez-Pena <i>et al.</i> , 2010
2221ΔFasX + vector	2221ΔFasX containing the empty shuttle vector pDCBB	Ramirez-Pena <i>et al.</i> , 2010
2221ΔFasX + pFasX	2221ΔFasX containing the pDCBB-derivative pFasX which contains a wild-type <i>fasX</i> allele downstream of the natural promoter	Ramirez-Pena <i>et al.</i> , 2010
MGAS10270	Serotype M2 clinical GAS isolate that has been genome sequenced	Beres <i>et al.</i> , 2007
M2 + vector	MGAS10270 containing the empty shuttle vector pDCBB	This work
M2 + pFasX	MGAS10270 containing the pDCBB-derivative pFasX which contains a wild-type <i>fasX</i> allele downstream of the natural promoter	This work
M2ΔFasX	MGAS10270 derivative in which the <i>fasX</i> gene has been replaced with a spectinomycin resistance cassette	This work
M2ΔFasX + vector	M2ΔFasX containing the empty shuttle vector pDCBB	This work
M2ΔFasX + pFasX	M2ΔFasX containing the pDCBB-derivative pFasX which contains a wild-type <i>fasX</i> allele downstream of the natural promoter	This work
M2ΔPIL	MGAS10270 derivative in which the pilus operon genes <i>10270_spy0109</i> to <i>0111</i> have been partially or fully deleted and a spectinomycin resistance cassette inserted	This work
MGAS10394	Serotype M6 clinical GAS isolate that has been genome sequenced	Banks <i>et al.</i> , 2004
M6ΔFasX	MGAS10394 derivative in which the <i>fasX</i> gene has been replaced with a spectinomycin resistance cassette	This work
M6ΔFasX + vector	M6ΔFasX containing the empty shuttle vector pDCBB	This work
M6ΔFasX + pFasX	M6ΔFasX containing the pDCBB-derivative pFasX which contains a wild-type <i>fasX</i> allele downstream of the natural promoter	This work
MGAS6180	Serotype M28 clinical GAS isolate that has been genome sequenced	Green <i>et al.</i> , 2005
M28ΔFasX	MGAS6180 derivative in which the <i>fasX</i> gene has been replaced with a spectinomycin resistance cassette	This work
M28ΔFasX + vector	M28ΔFasX containing the empty shuttle vector pDCBB	This work
M28ΔFasX + pFasX	M28ΔFasX containing the pDCBB-derivative pFasX which contains a wild-type <i>fasX</i> allele downstream of the natural promoter	This work

Table S1  
Table of GAS strains used in this study.

Primer name	Sequence (5' - 3')	Role
M2PILA	gtcagatccAAATGTATGCCGCTTCCACGATAAAGCC	Used in construction of pilus operon mutant in M2 GAS
M2PILB	CTAAGAAAATGAGACGGAAAATCCAGCTGAATACAACTCTTTG	Used in construction of pilus operon mutant in M2 GAS
M2PILC	CAAAGAAGTGTGTATTCAGCTGGATTTCCGCTCATTTCTTAG	Used in construction of pilus operon mutant in M2 GAS
M2PILD	gcagatccAAGGTAATAACTACCTGGCTTAC	Used in construction of pilus operon mutant in M2 GAS
M2PILE	GGTGATATAAATTAATAGAGGTAG	Used in construction of pilus operon mutant in M2 GAS
M2PILF	TACTCTCAATCCTCTTTTTGGCTTAC	Used in construction of pilus operon mutant in M2 GAS
FASXA	TCGTGGATATAGCCAAACG	Used in construction of <i>fasX</i> mutant in M2 GAS
M2FASXB	gttattgattattataacatgattCATTAAATTTATTATAGCGAAAAAATCTTC	Used in construction of <i>fasX</i> mutant in M2 GAS
FASXC	ctatttaataaacgattaaaaaattatagGGGTTTTTGATAGGTAATAATC	Used in construction of <i>fasX</i> mutant in M2 GAS
M2FASXD	ATCTGAATTAAGAGGGGTAGAGG	Used in construction of <i>fasX</i> mutant in M2 GAS
M2FASXF	GAAGTATTTTCGCTATAAATAAATAAATGaatcatgattatataactataac	Used in construction of <i>fasX</i> mutant in M2 GAS
FASXSR	gtattaccatcaaaaaaacccCTATAAATTTTAACTCTGTTATTTAAATAG	Used in construction of <i>fasX</i> mutant in M2 GAS
FASXE	GAGCACCCCAAGGGCACTAGACG	Used in construction of <i>fasX</i> mutant in M2 GAS
M2FASXF	AAGTGGACAAAGAACTGAAGAC	Used in construction of <i>fasX</i> mutant in M2 GAS
M6fasXD	CGAATCATACTGGCATTTTGAATTAAG	Used to replace primer M2FASXD in construction of the M6 <i>fasX</i> mutant
M28fasXM	TTAACAAGGCAATGAGCATGAG	Used with FASXA to amplify the SPEC cassette from strain 2221Δ <i>fasX</i> for creation of M28 <i>fasX</i> mutant
107TMF	AAAAGGGAGTCAGAAAAGTAATCTTGGT	Taqman primer for <i>M1.cpa</i>
107TMR	CCGGAACCTGTGTTGGCATT	Taqman primer for <i>M1.cpa</i>
107TMP	CGTCAAGCTTTGAAGCACTGATTGATCC	Taqman probe for <i>M1.cpa</i>
109TMF	ACTACTGTCAAGCAAGACGGAAATAAG	Taqman primer for <i>M1.tee1</i>
109TMR	CGTATTGTATCCACCTTATCTGAATT	Taqman primer for <i>M1.tee1</i>
109TMP	TGACTTTAGTCACTGGTGTGTTCAAGCTACACC	Taqman probe for <i>M1.tee1</i>
111TMF	AAACCGAAAAATCGGAGCTTATT	Taqman primer for <i>M1.111</i>
111TMR	CGTAACCTACCATTTACACAGTTG	Taqman primer for <i>M1.111</i>
111TMP	AAACACCGGAACCTCATCAACCAGATACAA	Taqman probe for <i>M1.111</i>
M2.109 TMF	CGGTAGCGGACCAACTG	Taqman primer for <i>M2.fcaA</i>
M2.109 TMR	TCTGGTGTCTCAGCTCT-TACAAC	Taqman primer for <i>M2.fcaA</i>
M2.109 TMP	TAGCCCAAGTGTGACAGTTTGGCAGAA	Taqman probe for <i>M2.fcaA</i>
M2.110 TMF	GAAGGGACTCCAACCTGAAGG	Taqman primer for <i>M2.110</i>
M2.110 TMR	AATTCCAATGTGCCAACA	Taqman primer for <i>M2.110</i>
M2.110 TMP	CGCCATCTCAGTTGATTTCAATGG	Taqman probe for <i>M2.110</i>
M2.113 TMF	TAGGCAAGCTACTTTTG-TGTGAA	Taqman primer for <i>M2.113</i>
M2.113 TMR	TCCGGTTTTATGTTTTTC-AAAG	Taqman primer for <i>M2.113</i>
M2.113 TMP	TCTACTGTCTCGACTTATTTCTGACTTATCATGGTCA	Taqman probe for <i>M2.113</i>
M6.159 TMF	GGAGCAGCCCGCACTAA	Taqman primer for <i>M6.fexX</i>
M6.159 TMR	CCGCTTATGGTGTCTCAA-TT	Taqman primer for <i>M6.fexX</i>
M6.159 TMP	CCAGCTTCTTTTCGGTCTCTTAACCTCCA	Taqman probe for <i>M6.fexX</i>
M6.160 TMF	CGAATGACGGTTCAGGTA-CAGTATTATTAG	Taqman primer for <i>M6.tee6</i>
M6.160 TMR	GCTACTCTGTCGAAGGTA-AA-TTCACCTA	Taqman primer for <i>M6.tee6</i>
M6.160 TMP	ACTGACATCCCTAACCCAA	Taqman probe for <i>M6.tee6</i>
M6.srB TMF	AAGAAGCCCTCAGAAATGGA	Taqman primer for <i>M6.srB</i>
M6.srB TMR	TGAATGCCATCCCTAAATGA	Taqman primer for <i>M6.srB</i>
M6.srB TMP	TTGGGATTCAGCCGGTTCCA	Taqman probe for <i>M6.srB</i>
M28.cpa TMF	TTGGGACAGAATACCATCCA	Taqman primer for <i>M28.cpa</i>
M28.cpa TMR	TCCCAACTCACCAGTACTG	Taqman primer for <i>M28.cpa</i>
M28.cpa TMP	TCCATACGAATCAGGTCAACCATACTA	Taqman probe for <i>M28.cpa</i>
M28.lepA TMF	TTGAAAGTTGGTCAATTTGC	Taqman primer for <i>M28.lepA</i>
M28.lepA TMR	AACAGTCTCTCTCTGCGT	Taqman primer for <i>M28.lepA</i>
M28.lepA TMP	CCTCATCGCAGCTTGAGGG	Taqman probe for <i>M28.lepA</i>
M28.109 TMF	AATAACCGTACACTCAA-GTTCFA	Taqman primer for <i>M28.fcaA</i>
M28.109 TMR	AGCCACAATGTAAAGAACTGCAA	Taqman primer for <i>M28.fcaA</i>
M28.109 TMP	CTGGTGTGTAGGCACCTTGCTCCA	Taqman probe for <i>M28.fcaA</i>
TUFA TMF	AACTACTTTAACAGTGCATCACAACT	Taqman primer for <i>tufA</i>
TUFA TMR	AGAAGCGTAATCTTTTGGTTGGTT	Taqman primer for <i>tufA</i>
TUFA TMP	TATTTGGCACGTGCGCTTGCCCTCATC	Taqman probe for <i>tufA</i>
SKATM2 TMF	CGGCTACTTTGAGGTCAATTGATT	Taqman primer for <i>M1.ska / M2.ska</i>
SKATM2 TMR	CCGAACCTCTTTGTCAGCAA	Taqman primer for <i>M1.ska / M2.ska</i>
SKATM2 TMP	CAAGCGATGCAACCACTACTGATTGCAAA	Taqman probe for <i>M1.ska / M2.ska</i>
SKATM2 TMF	CGGCTACTTTGAGGTCAATTGATT	Taqman primer for <i>M6.ska</i>
M6SKATMR	CCGAATCATCTTTGTCAGCAA	Taqman primer for <i>M6.ska</i>
SKATM2 TMP	CAAGCGATGCAACCACTACTGATTGCAAA	Taqman probe for <i>M6.ska</i>
SKATM2 TMF	CGGCTACTTTGAGGTCAATTGATT	Taqman primer for <i>M28.ska</i>
M28SKATMR	CCGAATCATCTCTGCTCAGCAA	Taqman primer for <i>M28.ska</i>
SKATM2 TMP	CAAGCGATGCAACCACTACTGATTGCAAA	Taqman probe for <i>M28.ska</i>
UNR218	CTGActtaaacgactacatagAGAAAGGGAGATAAATGAAAAAGAAAC	Used with UNR219 to amplify the 5' end of gene 113 from M2 GAS and place it downstream of a T7 promoter for use in <i>in vitro</i> transcription
UNR219	ATTGGGATAGGATAGTAAAG	Used with UNR218 to amplify the 5' end of gene 113 from M2 GAS and place it downstream of a T7 promoter for use in <i>in vitro</i> transcription
UNR123	cttaaacgactacatagTAAATAAAGATTTACGAAGTC	Used with FASEND9 to amplify <i>fasX</i> or the <i>fasX</i> derivative <i>fasX172-77</i> and allow their use in <i>in vitro</i> transcription assays by placing a T7 promoter upstream
FASEND9	aaaaaacggcaaacggcagct	Used with UNR123 to amplify <i>fasX</i> or the <i>fasX</i> derivative <i>fasX172-77</i> and allow their use in <i>in vitro</i> transcription assays by placing a T7 promoter upstream
UNR291	acgttaaacgactacatagGAGAGAAACAGTGGAGAGAGAAGATTAATAAC	Used in association with UNR292 to amplify the 5' UTR and initial coding region of <i>M6.tee6</i> and allow its use in <i>in vitro</i> transcription assays by placing a T7 promoter upstream
UNR292	GTGAGTATCATCTTTGATAAAGCTG	Used in association with UNR291 to amplify the 5' UTR and initial coding region of <i>M6.tee6</i> and allow its use in <i>in vitro</i> transcription assays by placing a T7 promoter upstream
UNR212	CTGActtaaacgactacatagGAGAGAGAGAGAAATGAAAAAATAAATTAATCTG	Used in association with UNR214 to amplify the 5' UTR and initial coding region of <i>M28.fcaA</i> and allow its use in <i>in vitro</i> transcription assays by placing a T7 promoter upstream
UNR214	GATTTTACGTTTGTATTTAAAGAAG	Used in association with UNR212 to amplify the 5' UTR and initial coding region of <i>M28.fcaA</i> and allow its use in <i>in vitro</i> transcription assays by placing a T7 promoter upstream
UNR217	ATATGAGTAAACTTGGTCTGACGctattatcatgctcttataatcTCTACCTTAACATGAGATAACTTTC	Used in association with UNR218 to amplify the 5' UTR and first 158 codons of <i>M2.113</i> for use in <i>in vitro</i> transcription and translation reactions
UNR218	CTGActtaaacgactacatagAGAAAGGGAGATAAATGAAAAAGAAAC	Used in association with UNR217 to amplify the 5' UTR and first 158 codons of <i>M2.113</i> for use in <i>in vitro</i> transcription and translation reactions
UNR291	acgttaaacgactacatagGAGAGAAACAGTGGAGAGAGAAGATTAATAAC	Used in association with UNR293 to amplify the 5' UTR and first 158 codons of <i>M6.tee6</i> for use in <i>in vitro</i> transcription and translation reactions
UNR293	ATATGAGTAAACTTGGTCTGACGctattatcatgctcttataatcTAATGATGATTTGCAACACTGTTTGTCC	Used in association with UNR292 to amplify the 5' UTR and first 158 codons of <i>M6.tee6</i> for use in <i>in vitro</i> transcription and translation reactions
UNR220	ATATGAGTAAACTTGGTCTGACGctattatcatgctcttataatcTCCCTCAATACCACATTAACAAC	Used in association with UNR356 to amplify the 5' UTR and first 158 codons of <i>M28.fcaA</i> for use in <i>in vitro</i> transcription and translation reactions
UNR356	CTGActtaaacgactacatagGAAAAATCTCAACTTATTAAAGAGTG	Used in association with UNR220 to amplify the 5' UTR and first 158 codons of <i>M28.fcaA</i> for use in <i>in vitro</i> transcription and translation reactions
SPDTRN	ATATGAGTAAACTTGGTCTGACGctattatcatgctcttataatcTCCGACATAAGATAGACACTTTAAAC	Used in association with T7SPD to amplify the 5' UTR and first 158 codons of <i>spd</i> for use in <i>in vitro</i> transcription and translation reactions
T7SPD	cttaaacgactacatagGGTATGAGCGAAATAGAAAAAGG	Used in association with SPDTRN to amplify the 5' UTR and first 158 codons of <i>spd</i> for use in <i>in vitro</i> transcription and translation reactions

Table S2  
Table of primers and probes used in this study.