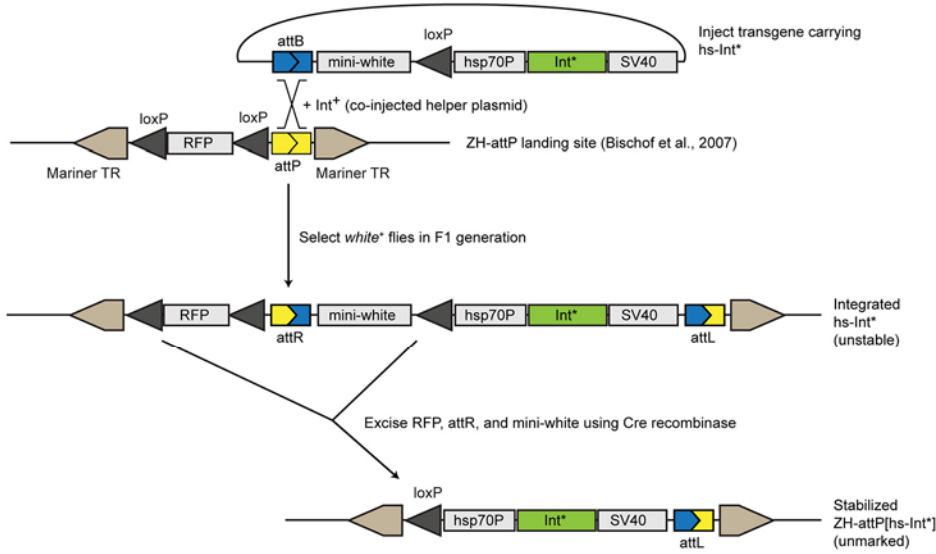


A. Strategy to stabilize hs-Int\* transgenes.



B. Crossing scheme to test excisionase activity of Int\* mutants.

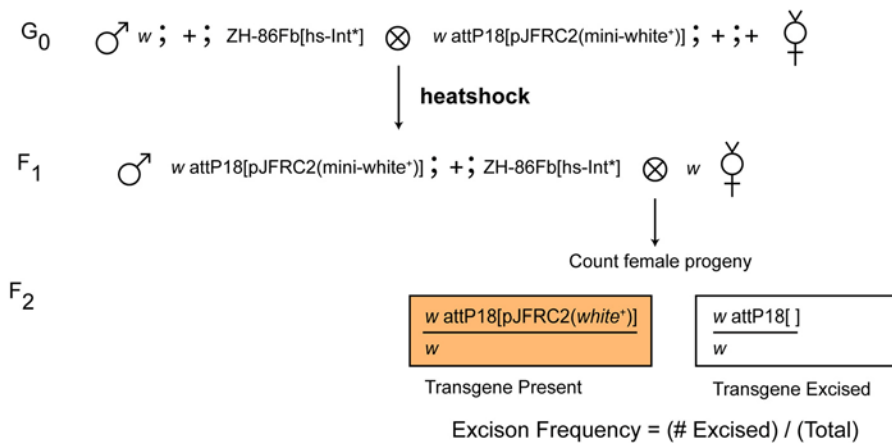


Figure S1. Testing Int\* variants for excisionase activity *in vivo*.

**Figure S1** Testing Int\* variants for excisionase activity *in vivo*. (A) Strategy to stably and site-specifically introduce Int\* into the genome: Int\* transgene constructs bearing a single loxP site (5' to hs-Int\*) were integrated into the landing sites *ZH-51C* or *ZH-86Fb* (Bischof *et al.* 2007), and integrants were identified by mini-white expression. To prevent Int\* from excising itself, sequences between the upstream-most loxP in the landing site and the loxP in the integrated transgene were eliminated by Cre recombinase. The removal of RFP and mini-white leaves the *hs-Int\** transgene unmarked, to avoid interference with downstream applications. (B) Crossing scheme to test Int\* variants for excisionase activity: Virgins bearing *attP18[JFRC2]* were crossed to males bearing *hs-Int\**, and progeny were heat shocked for one hour during the third larval instar. Following eclosion, individual males were crossed to *white* virgins, and the numbers of female progeny that were *mini-white*<sup>+</sup> (transgene present) and *white* (transgene excised) were counted. The frequency of transgene excision was determined by comparing the number of *white* female progeny to the total number of female progeny. Also see Figure 1D and Table S1.