

**ADDITIONAL FILE 1: Supplemental Information**

belonging to the manuscript

**A novel locus for mycelial aggregation forms a gateway to improved *Streptomyces* cell factories**

by

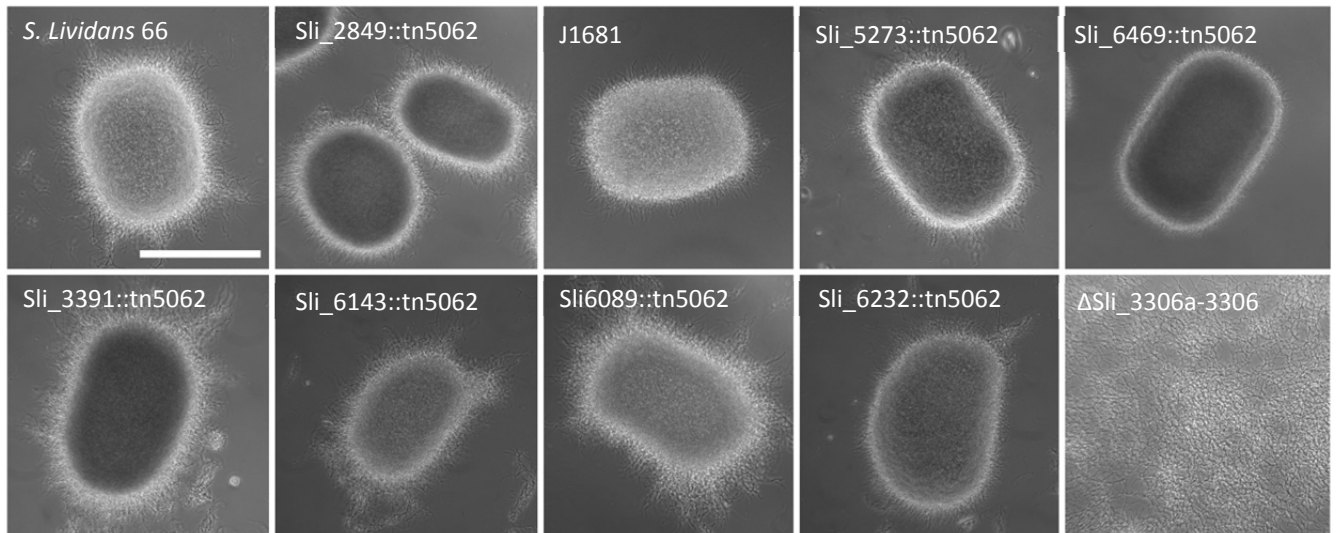
Dino van Dissel<sup>1</sup>, Dennis Claessen<sup>1</sup>, Martin Roth<sup>2</sup>, and Gilles P. van Wezel<sup>1,\*</sup>

<sup>1</sup>Molecular Biotechnology, Institute of Biology, Leiden University, PO Box 9505, 2300RA, Leiden, The Netherlands;

<sup>2</sup> Bio Pilot Plant, Leibniz Institute for Natural Product Research and Infection Biology, Hans Knöll Institute, Adolf-Reichwein-Str. 23, 07745 Jena, Germany

# To whom correspondence should be addressed. Tel: +3171527430; email: g.wezel@biology.leidenuniv.nl

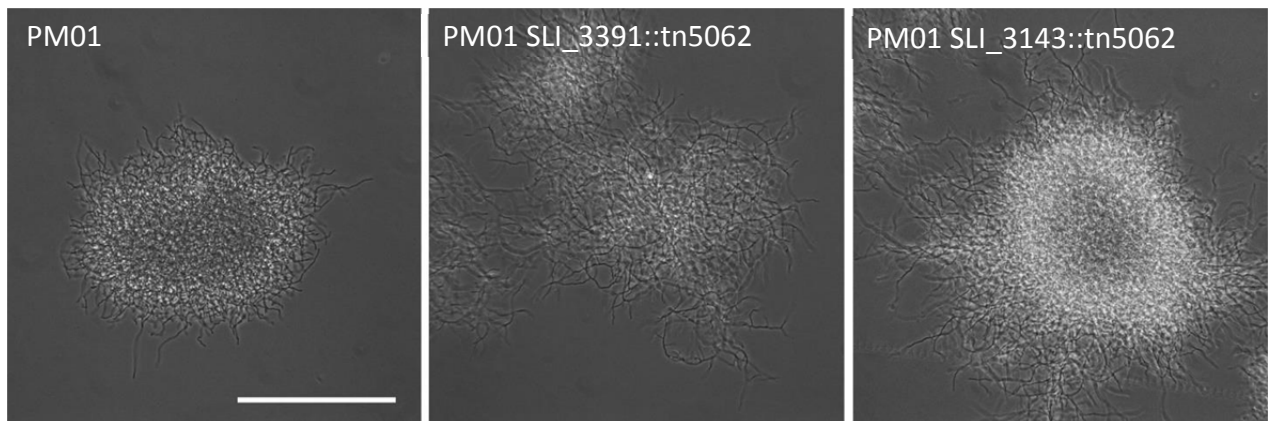
**Running title:** Morphological engineering of *Streptomyces*



**Figure S1. Phenotypes of disruption mutants of *S. lividans* in submerged cultures.**

Sli\_2849, Sli\_5273, Sli\_6469, Sli\_3391, Sli\_6143, Sli\_6089 and Sli\_6232 were disrupted by a transposon insertion. The region of Sli\_3306a-Sli\_3306 was removed by homologous recombination replacement. J1681 (*S. coelicolor*  $\Delta bldA$ ) was published previously [1].

Cultures were grown in baffled shake flasks in TSBS for 48 h. Scale bar, 200  $\mu$ m.



**Figure S2: Identification of suppressor mutations in *S. lividans* PM01 and PM02.**

SLI\_3391 and SLI\_6143 were identified by SNP analysis as the major changes during evolution of PM02 from PM01. Note that mutation of SLI\_3391 enhanced dispersed growth of PM01, giving a phenotype similar to that observed for PM02. Scale bar, 100  $\mu$ m.

**Table S1. Bacterial strains.**

Strain	Description and genotype	Reference
<i>Streptomyces lividans</i> 66 (1326)	SLP2+ SLP3+	[2]
PM01	Evolved from <i>S.lividans</i> 66	[3]
PM02	Evolved from PM01	[4]
J1681	J1501 $\Delta bldA$	[1]
GAD01	<i>S. lividans</i> 66 $\Delta SLI\_3306a::aacC4$ Apr <sup>R</sup>	This study
GAD02	<i>S. lividans</i> 66 $\Delta SLI\_3306a^{clean}$	This study
GAD03	<i>S. coelicolor</i> M145 $\Delta SCO2962::aacC4$ Apr <sup>R</sup>	This study
GAD04	<i>S. lividans</i> 66 $\Delta SLI\_3306a-Sli\_3306::aacC4$ Apr <sup>R</sup>	This study
GAD05	<i>S. lividans</i> 66 $\Delta SLI\_3306a-SLI3306^{clean}$	This study

IFD, in-frame deletion mutant;  
Apr<sup>R</sup> apramycin resistant.

**Table S2. Transposon-mediated gene-replacement cosmids.** Cosmid nomenclature refers to the *Streptomyces coelicolor* genome database (strepdb.streptomyces.org.uk). The genomic location of the insertion of the apramycin cassette is given for the *S. coelicolor* genome.

Cosmid name	target gene	Cosmids location in genome	Start gene	position relative to start
SC17.2.C04	SCO1907	2043368	2044163	795
C121.1.E05	SCO2513	2709849	2709485	364
E34.2.E04	SCO3043	3331409	3331178	231
2SCK36.1.F01	SCO4998	5437068	5437222	154
SC5B8.1.F05	SCO5821	6369817	6369367	450
SC2E9.1.F02	SCO5871	6426513	6426319	194
7H1.2.H01	SCO5952	6521053	6520547	506
SC9B1.2.C03	SCO6076	6670727	6670057	670

**Table S3. Plasmids and constructs.**

Plasmid or construct	Description	Reference
pWHM3	Cloning vector, <i>colE1</i> replicon, <i>pSG5</i> replicon, Thio <sup>R</sup> , Amp <sup>R</sup>	[5]
pSET152	Complementation vector, <i>oriT</i> RK2, pUC18 replicon, Apra <sup>R</sup>	[6]
pUWLcre	pUWLoriT derivative with <i>creA</i> gene under ermE* promoter, Thio <sup>R</sup>	[7]
pMAT1	pWHM3 containing flanking regions of <i>S. coelicolor</i> SCO2963 and SCO2962 with a <i>aac(3)IV-loxP</i> XbaI inserted between them in pWHM3 EcoRI-HindIII	this work
pMAT2	pWHM3 containing flanking regions of <i>S. coelicolor</i> SCO2963 with a <i>aac(3)IV-loxP</i> XbaI inserted between them in pWHM3 EcoRI-HindIII	this work
pMAT3	Cosmid StE59 derivative in which the <i>matB</i> coding sequence was replaced by the <i>aac(3)IV</i> resistance cassette	this work
pMAT4	pSET152 containing SCO2963 with the 500bp upstream (promoter) region	this work

**Table S4. Oligonucleotides.**

Name	Primer Sequence ^
matB_+2190	AGTCTCTAGAAAGCCGGTTCGGATGACCACC
matB_+3610	AGTCAAGCTTCCCTGTTCACCTCCCGCAACCG
matA_-1326	AGTCGAATTCCAGCCGGGCGGTGAGATTCC
matA_+43	ACTGTCTAGACGAGCACTCGTCGGCCGAAC
matA_2809	AGTCAAGCTTAGACGGTGTTCGCCGTCCATC
matA_+1466	ACTGTCTAGACCCCGGAGAACACCCTCTGATGG
matA_-54	TTCTTTGCCTGAGCACGGTGTGATAC
matB_+1528	TGGTACAGGACCACCCGGAAGAG
pmatA_-537	AGCTGAATTCGGCGCGTTACGAGAGCGGACTGAC
matA_1485	GATCTCTAGATCAGAGGGTGTTCCTCCGGGACAG
matB_FW_REDIRECT	CCGGGGTTCGGCCCGTTCGGTTCGTACGGCGGGGTGGTCATGTAGGCTGGAGCTGCTTC
matB_REV_REDIRECT	CCCCTCCCTCCCCCTGTCCCCGGAGAACACCCTCTGATGATTCCGGGGATCCGTTCGACC

^Restriction sites are underlined. TCTAGA, XbaI; AAGCTT, HindIII; GAATTC, EcoRI.

## References

1. Leskiw BK, Bibb MJ, Chater KF. **The use of a rare codon specifically during development?** *Mol Microbiol* 1991, **5**:2861-7.
2. Kieser T, Bibb MJ, Buttner MJ, Chater KF, Hopwood DA. **Practical *Streptomyces* genetics**. 2000.
3. Roth M, Noack D, Geuther R. **Maintenance of the recombinant plasmid pIJ2 in chemostat cultures of *Streptomyces lividans* 66 (pIJ2)**. *J Basic Microbiol* 1985, **25**:265-71
4. Roth M, Hoffmeier C, Geuther R, Muth G, Wohlleben W. **Segregational stability of pSG5-derived vector plasmids in continuous cultures of *Streptomyces lividans* 66**. *Biotechnol Lett* 1994, **16**:1225-30.
5. Vara J, Lewandowska-Skarbek M, Wang YG, Donadio S, Hutchinson CR. **Cloning of genes governing the deoxysugar portion of the erythromycin biosynthesis pathway in *Saccharopolyspora erythraea* (*Streptomyces erythreus*)**. *J Bacteriol* 1989, **171**:5872-81.
6. Bierman M, Logan R, O'Brien K, Seno ET, Rao RN, Schonher BE. **Plasmid cloning vectors for the conjugal transfer of DNA from *Escherichia coli* to *Streptomyces* spp**. *Gene* 1992, **116**:43-9.
7. Fedoryshyn M, Welle E, Bechthold A, Luzhetskyy A. **Functional expression of the Cre recombinase in actinomycetes**. *Appl Microbiol Biotechnol* 2008, **78**:1065-70.