Roberson et al., http://www.jcb.org/cgi/content/full/jcb.201411087/DC1



Figure S1. **Tmem231 and B9d1 are required for ciliary localization of membrane proteins.** *Tmem231* (A) and *B9d1* (B) wild-type and mutant MEFs stained for Tub^{Ac} (blue), γ -tubulin (red), DNA (gray), and either Adcy3, Arl13b, Inpp5e, or Ift88 (green). Bar, 10 µm. Adcy3, Arl13b, and Inpp5e fail to localize to cilia in *Tmem231* and *B9d1* mutant MEFs, whereas Ift88 remains unaffected. (C) Quantitation of the percentage of positive (green) and negative (purple) cilia from A and B. At least 10 cilia were scored per sample. (D) Cilia in the mutant MEFs are modestly longer than those of wild-type MEFs. (E) *Tmem231* and *B9d1* mutant MEFs ciliate less frequently than *Tmem231* and *B9d1* wild-type MEFs. Error bars represent the 95% confidence interval. *, P < 0.05, as measured by Student's *t* test with Welch's correction.

0 0

0

m

C

ы.



Figure S2. **B9d1 and Tmem231 are required for MKS complex assembly.** Ciliary TZ in B9d1 (A) and Tmem231 (B) wild-type and mutant MEFs stained for Tub^{Ac} (blue), γ -tubulin (red), DNA (gray), and either B9d1, Tmem231, Mks1, Tmem67, Rpgrip11, or Nphp1 (green). Bar, 10 µm. Tmem231 and B9d1 require each other for TZ localization. Mks1 and Tmem67 fail to localize to the ciliary TZ in the absence of *B9d1* or *Tmem231*. (C) Quantitation of the percentage of positive (green) and negative (purple) cilia from A and B. At least 10 cilia were scored per sample.

Human Mouse Turtle Chicken Zebrafish Ciona Trypanosoma Trypanosoma Laishmania Laishmania Laishmania Aurecocccus Thalassiosira Batrachochytrium Caenorhabditis Tetrahymena Drosophila Paramecium consensus		<pre>ALTELESHID_VERSURAACLESKIAMELULAAADTIIDLUVIERSHOGIMIRSSUESCOUNGOUNULLOPESDOF ANGTPPAFERLO</pre>
Human Mouse Turtle Chicken Xenopus Zebrafish Ciona Trypanosoma Leishmania Chlamydomonas Aurecococcus Thalassiosira Batrachochytrium Caenochabditis Tetrahymena Drosophila Paramecium consensus	117 118 118 116 116 116 116 114 121 97 1222 124 131	MLHEKLEL 105T
Human Mouse Turtle Chicken Zebrafish Ciona Trypanosoma Leishmania Chlamydomonas Aursococcus Aursococcus Tatrachobytrium Caenorhabditis Tetrahymena Drosophila Paramecium consensus	209 210 208 208 208 213 211 213 219 176 209 213 2250 261	DLEN UNAPGENUMETULNDPUTLUVGE - AAD DLEN UNAPGENUMETULNDPUTLUVGE - AAD DLEN UNAPGENUMETULNDPUTLUVGE - AAD DLEN UNAPGENUMETULDPUTLUVGE - AAD DLEN UNAPGENUMETULDPUTLUVGE - AAD DLEN UNAPGENUMETULUD GENERALKAPAN OVUS LLEP LUV BERK I TU DLEN UNAPGENUMETULUD GENERALKAPAN OVUS LLEP LUV BERK I TU ALDNU SSOCK DUT ULAPHEUTUG - AAN ALDNU SSOCK DUT ULAPHEUTUG - AAN ALDNU SSOCK DUT ULAPHEUTUG - AAN DEN UNAPGENUMETULUD GENERALKAPAN OVUS LLEP LUV GENERALKAPAN ALDNU SSOCK DUT ULAPHEUTUG - AAN ALDNU SSOCK DUT ULAPHEUTUG - AAS ALDNU SSOCK DUT ULAPHEUTUG - AAN DEN UNAPGENUMETULAPHEUTUG - AAN ALDNU SSOCK DUT ULAPHEUTUG - AAN ALDNU SSOCK DUT UNAPHEUTUG AAN ANNO SSOCK DUT UNAVER ALDNU SSOCK DUT UNAPHEUTUG - AAN ALDNU SSOCK DUT UNAPHEUTUG - AAN ALDNU SSOCK DUT UNAPHEUTUG AAN ANNO SSOCK DUT UNAVER ALDNU SSOCK DUT UNAPHEUTUG AAN ANNO SSOCK DUT UNAVER ALDNU SSOCK DUT UNAPHEUTUG ULAPTUK UNAPHEUTUG ULAPTUK UNA ALDNU SSOCK DUT UNAPHEUTUG ULAPTUK UNAPHEUTUG ULAPTUK UNAPHEUTUG ULAPTUK UNAPHEUTUG ULAPTUK UNA ALDNU SSOCK DUT UNAPHEUTUG ULAPTUK UNAPHEUTUG ULAPTUK UNA ALDNU SSOCK DUT UNAPHEUTUG ULAPTUK UNAPHEUTUG ULAPTUK UNAVER ANN AND AND
Human Mouse Turtle Chicken Xenopus Zebrafish Ciona Trypanosoma Leishmanla Guismydoous Thalassiosira Batrachochytrium Caenorhabditis Tetrahymena Drosophila Paramecium consensus	294 295 295 295 293 293 359 305 305 307 294 305 307 294 311 344 391	GVUTGIPUTVTPRGDLCKEHLS GVUTGIPUTVPGDLCKEHLS GVUTGIPUTVPSDLSPULSPVLSTK00

Figure S3. **TMEM231 is conserved in ciliated organisms.** ClustalW alignment of TMEM231 protein sequences reveals conservation of residues across diverse ciliated organisms. Residues highlighted in red indicate those mutated in OFD3 or MKS.



Figure S4. **Mutation of TMEM231 is associated with OFD3 and MKS.** Pedigree and chromatogram of OFD3- (A) and MKS (B–J)-affected individuals with recessive mutations in *TMEM231* (GenBank accession no. NM_001077418.2). (K) Functional consequences of c.664+4A>G. RT-PCR analysis conducted on primary skin fibroblasts from MKS-374 shows that the intronic change abolishes the canonical exon 5 splice donor sequence and results in retention of 47 bp of intron 5–6 to produce a premature frameshift p.Val222fsX21. H, homozygous; h, heterozygous; Pat, paternal; Mat, maternal.



Figure S5. *Tmem231* mutant alleles partially rescue Arl13b localization to the ciliary membrane. (A–E) Quantitation of various aspects of the rescue assays in *Tmem231* wild-type and mutant MEFs, including fluorescence intensity of Arl13b, Tub^{Ac}, and γ -tubulin, ciliary length, and γ -tubulin area. Tub^{Ac} intensity is decreased when mutant forms of Tmem231 are transfected, as compared with transfection of wild-type Tmem231. (F) Quantitation of fluorescence intensity of B9d1 at the TZ/basal body in *Tmem231* wild-type MEFs. Error bars represent the 95% confidence interval. *, P < 0.05, as measured by Student's *t* test with Welch's correction.

	hnic N	Jucleotide	Protein	Exon/intron	AA conser-	Parental	PolvPhen 2-	Additional			Clinic	al featur	v	
$ \begin{array}{ c c c c c c c c c c c c c c c c c c c$		lteration"	change	(zygosity,	vation	consan-	Score/muta	mutations	-				3	
$ \begin{array}{ c c c c c c c c c c c c c c c c c c c$				segregation)		guinuity	tion taster		Ind. age	D	CNS	Kidney	Liver	Other
c.373C-G Pho12Abl 3 C. intentrolis 0.998/DC 24, XX, min(1) 2 min ENS ENS ENS Pho12Abl Pho1Abl	U	.241C>™	p.Leu81Phe	2 (het, m)	C. intestinalis	ои	0.976/DC		–21, 24 yr	1 upper limb	OMA; CVH	ESRD	_	lingual hamartomas, ID
Cu273C-G Pho125Abl 3 Currentoxic Anolu Circulation no 0998/DC 4 mints DWS; HC CK HF Creations Cu273C-G Ph0125Abl 3 Circulations 0.998/DC 23 wk 4 limbs DWS; CK HF Creations Cu273C-G Ph0125Abl 3 Circulations 0.998/DC 23 wk 4 limbs DWS; CK HF Creations Cu273C-G Ph0125Abl 3 Circulations 23 wk 4 limbs DWS; CK HF Creations Cu260A-1 Publications M	U	373C>G	p.Pro125Ala	3 (het, p)	C. intestinalis		0.998/DC		—24, 13 уг	2 upper limbs	OMA; CVH; DWS	ESRD		lingual hamartomas, ID
$ \begin{array}{c c c c c c c c c c c c c c c c c c c $	U	.373C>G	p.Pro125Ala	3 (hom)	C. intestinalis	оц	0.998/DC	46,XX,inv(10) (p11.1q21.1) (het, p)	40 wk	4 limbs	DWS; HC	сĸ	HPF	
$ \begin{array}{ c c c c c c c c c c c c c c c c c c c$	0	.373C>G	p.Pro125Ala	3 (het, m)	C. intestinalis	ou	0.998/DC		23 wk	4 limbs	DWS; CVA	СK	HPF	CP, epidymal cysts, micropenis, accessory
$ \begin{array}{ c c c c c c c c c c c c c c c c c c c$	U	269A>T	p.Asn90lle	2 (het, p)	C. elegans		0.987/DC							spleen/heterotaxia
$ \begin{array}{ c c c c c c c c c c c c c c c c c c c$	U	.373C>G	p.Pro125Ala	3 (het, p)	C. intestinalis	ou	0.998/DC	CC2D2A: c.834delG;	20 wk	4 limbs	DWS; CVA	Х	HPF	accessory spleen/heterotaxia,
$ \begin{array}{ c c c c c c c c c c c c c c c c c c c$	Ů	.544C>T	p.Gln182*	4 (het, m)			N/A	p.Leu279Cysf x* (het)						skeletal dysplasia (ulnar bowing)
$ \begin{array}{ c c c c c c c c c c c c c c c c c c c$	U	.363delC	p.Ile232Serfs*	6 (het, p)		ои	N/A		13 wk	2 upper limbs	AC	З		
$ \left[\begin{array}{cccccccccccccccccccccccccccccccccccc$	0	.664+4A>G		IVS5 (het, m)			N/A							
	о р	664+4A>G		IVS5 (hom)		yes	N/A		F1 14 wk	4 limbs	MEN	CK	HPF (CP, single umbilical artery, epididymal cysts
c664+4A-G IVS5 ves N/A 2 weet MEN CK HF pencentic fibrosis c.664G-x ^c p(m) 5 X.loevis yes N/A 2 weet EN:MC CK HF pencentic fibrosis c.664G-x ^c p.Vol222lle 5 X.loevis yes N/A F123 wk A limbs EN:MC CK M UGR, dbsent uvulo, micropenis, anomolous c.664G-x ^c p.Vol222lle 5 X.loevis P P D									F2 13 wk	4 limbs	BM	CK	_	IUGR
$ \begin{array}{c ccccccccccccccccccccccccccccccccccc$	υ	664+4A>G		IVS5 (hom)		yes	N/A		22 wk	2 upper limbs	MEN	CK	HPF	pancreatic fibrosis
F2 18 whF2 18	υ	664G>A°	p.Val222lle	5	X. laevis	yes	N/A		F1 23 wk	4 limbs	EN; MC	СК		IUGR, absent uvula, micropenis, anomalous pulmonary venous
$ \begin{array}{c c c c c c c c c c c c c c c c c c c $		_											Ū	connection
$ \begin{array}{c c c c c c c c c c c c c c c c c c c $				(hom)					F2 18 wk		Z	Х	<u> </u>	bowing of long bones, malposition of the feet, TGA
c.664+4A>GIVS5HPFmalposition of the hands(het, p)(het, p)CDCVH;CKHPFwith ulner deviation(het, p)No0.994/DC17 wk4 limbsPPFCP, short limbs;c.439-1G>CNS3no0.994/DC17 wk4 limbsPPFCP, short limbs;c.439-1G>CNet, m)0.994/DC17 wk4 limbsPPFCP, short limbs;c.439-1G>CP.Ala216Pro5G. gallusC.646G>CP.Ala216Pro5Pointion; bilderalc.646G>CP.Ala216Pro5G. gallusSingle unbilical artery, single unbilical artery, single unbilical artery,	υ	.373C>G	p.Pro125Ala	3 (het, m)	C. intestinalis	ou	0.998/DC		F1 25 wk	2 upper; 1 lower	HC; HPE	CK	HPF	splenopancreatic fusion
c.439-1G>C IVS3 no 0.994/DC 17 wk 4 limbs MEN CK HPF CP, short limbs; (het, m) (het, m) (het, m) CK HPF CP, short limbs; epididymal cysts; analposition of the left (het, m) CK HPF CP, short limbs; c.646G>C p.Ala216Pro 5 G. gallus evidition; bildteral c.646G>C p.Ala216Pro 5 G. gallus evidition; bildteral	0	.664+4A>G		IVS5 (het, p)					F2 16 wk		HC; CVH; CD	Х	HPF	malposition of the hands with ulnar deviation
c.646G>C p.Ala216Pro 5 Bindexitin bilateral deviations bilateral malposition of the feet, single unbilateral	0	.439-1G>C		IVS3 (het, m)		ou	0.994/DC		17 wk	4 limbs	MEN	сk	HPF	CP, short limbs; epididymal cysts; malnosition of the left
c.646G>C p.Ala216Pro ³ G. gallus G. gallus c.646G>C p.Ala216Pro ³ single umbilical artery,				ı										hand with ulnar
single unbilical artery,	υ	.646G>C	p.Ala216Pro	5 (het, p)	G. gallus								0 2	deviation; bilateral malposition of the feet,
														single umbilical artery,

exercise of the production of the great arteries. The proceeding of the production of the great arteries are arteries of the great arteries. The production of the great arteries are arteries of the great arteries of the great arteries. The production of the great arteries of the great arteries of the great arteries. The production of the great arteries of the great arteries of the great arteries of the great arteries. The production of the great arteries of the great arteries of the great arteries. The productions are numbered according to human CDNA reference sequence NM_001077418.2, isoform 1 (TMEM231), where +1 corresponds to the A of ATG start translation codon. The arterian of the great arteries are numbered according to human cDNA reference sequence NM_001077418.2, isoform 1 (TMEM231), where +1 corresponds to the A of ATG start translation codon. The arterian test is listed in Exome variant server database: TT = 0/TC = 1/CC = 6155.

Table S2.	С.	eleaans	strains	used
10010 02.	.	cicguiis	311 01113	0304

Strain	Genotype
PT709	nphp-4(tm925)
MX1855	tmem-231 (tm5963)
MX1251	mks-2(nx111)
MX754	mks-5(tm3100)
MX1415	N2; nxEx[<i>tmem-231</i> :gfp + Posm-5::xbx-1::tdTomato + rol-6(su1006)]
MX1450	nphp-4; nxEx[tmem-231::gfp + Posm-5::xbx-1::tdTomato + rol-6(su1006)]
MX1470	mks-2; nxEx[tmem-231::gfp + Posm-5::xbx-1::tdTomato + rol-6(su1006)]
MX1471	mks-5; nxEx[tmem-231::gfp + Posm-5::xbx-1::tdTomato + rol-6(su1006)]
YH930	N2; Ex[Posm-5::mks-5::tdTomato + Posm-5::dyf-11::gfp + rol-6(su1006)]
MX1811	tmem-231; nxEx[Posm-5::mks-5::tdTomato + Posm-5::dyf-11::gfp + rol-6(su1006)]
MX1420	N2; nxEx[Pbbs-8::tram-1::tdTomato + Pbbs-8::mks-2::gfp + rol-6(su1006)]
MX1813	tmem-231; nxEx[Pbbs-8::tram-1::tdTomato + Pbbs-8::mks-2::gfp + rol-6(su1006)]
YH237	N2; yhEx142[nphp-1::cfp + che-13::yfp + rol-6(su1006)]
MX1814	tmem-231; nxEx[nphp-1::cfp + che-13::yfp + rol-6(su1006)]
MX1065	N2; nxEx[<i>tmem-17</i> ::gfp + P <i>osm-5</i> ::xbx-1::tdTomato + <i>rol-6(su1006)</i>]
MX1815	<i>tmem-231</i> ; nxEx[<i>tmem-17</i> ::gfp + Pos <i>m-5</i> ::xbx-1::tdTomato + rol-6(su1006)]