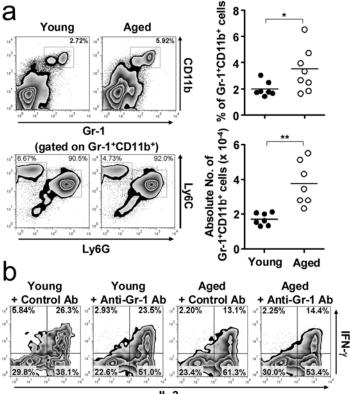
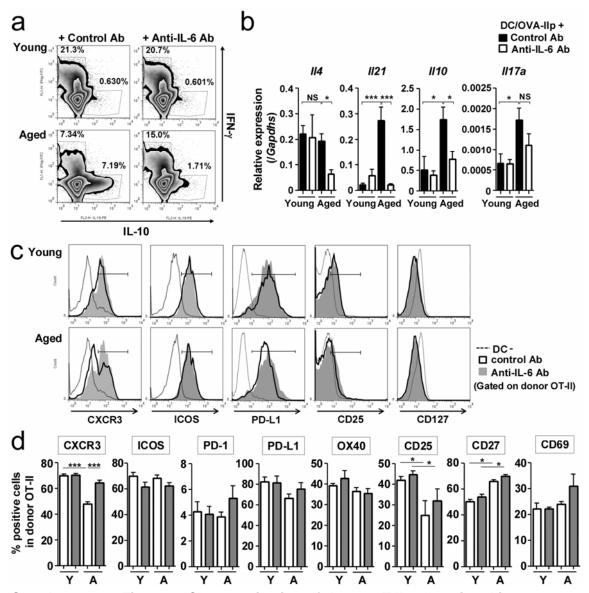


Supplementary Figure 1. Homeostatic expansion of young OT-II cells in young and aged environments. CFSE-labeled young OT-II cells were transferred into irradiated young or aged hosts. The indicated days after transfer, donor OT-II cells were harvested from spleen and LNs. CFSE intensity and CD44 expression (**a**) and the absolute number (**b**) of donor OT-II cells are shown. "Divided" and "Undivided" represent CFSE^{Io} and CFSE^{hi} population indicated in **a**, respectively. The data are representative of three independent experiments (mean \pm SEM with n = 4). Asterisks indicate the significance of the difference between young and aged group; * *p* < 0.05 (one-way ANOVA followed by Tukey's post hoc tests).

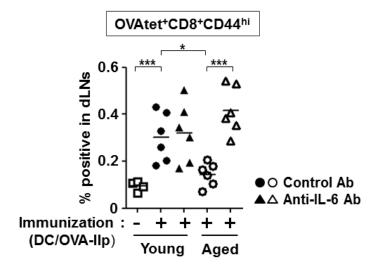


IL-2 (donor OT-II gated)

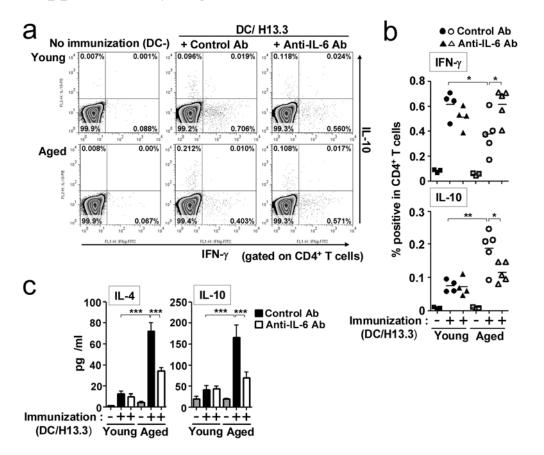
Supplementary Figure 2. Gr-1⁺CD11b⁺ cells increase in aged mice. (a) Spleen and lymph node cells were harvested from young or aged mice, and Gr-1⁺CD11b⁺ MDSC were analyzed. Representative plots of Gr-1 and CD11b expression (upper left) and Ly6C and Ly6G expression in the gated population (lower left), and total number of Gr-1⁺CD11b⁺MDSC (right) are shown (mean with n = 7-8). (b) Young naïve OT-II cells were primed *in vivo* as described in Figure 2. Mice were treated with control Ab or anti-Gr-1 Ab 4 days before immunization (n = 4/group). Seven days after immunization, donor OT-II cells were re-stimulated with PMA/ionomycin. Representative plots of IFN- γ and IL-2 production are shown. Representative data from three experiments are shown. Asterisks indicate the significance of the difference between young and aged group; * *p* < 0.05, ** *p* < 0.01 (unpaired Student's *t*-test).



Supplementary Figure 3. Characterization of donor OT-II cells primed in young or aged mice. Young naïve OT-II cells were transferred into young or aged mice, and then were primed *in vivo* as described in Fig. 1d. Mice were treated with control Ab or anti-IL-6 Ab 1 day before immunization. (a) Five days after immunization, isolated CD4⁺ T cells were re-stimulated with PMA/ionomycin. Representative plots of IFN- γ and IL-10 production in gated donor OT-II cells are shown. (b) Four days after immunization, isolated CD4⁺ T cells were analysed for the mRNA expression of the indicated genes by real-time qPCR. Each gene expression was normalized to the expression of mouse *Gapdh.* (c and d) Four or five days after immunization, the expression of indicated cell surface molecules on donor OT-II cells were assessed by flow cytometry. Representative histograms (c) and the frequencies of positive cells (d) in donor OT-II cells are shown are mean \pm SEM with n = 4-5 /group. Asterisks indicate the significance of the difference between indicated groups; * p < 0.05, ** p < 0.01, *** p < 0.001 (one-way ANOVA followed by Tukey's post hoc tests).



Supplementary Figure 4. Th1-dependent helper activity for tumour-specific CD8⁺ T cells is diminished by IL-6 in aged mice. CD45.1⁺ young naïve OT-II T cells were transferred into young or aged CD45.2⁺ mice. The mice were then immunized by transfer of DC pulsed with or without OVA-IIp. Mice were treated with control or anti-IL-6 Ab on day -1 and day 1. Five days after immunization, MCA-OVA were inoculated. Another 6 days later, OVA-specific CD8⁺ T cells in tumour draining LNs were analysed by flow cytometry. Representative data from three experiments are shown (mean with n = 4-6 /group). * p < 0.05, *** p < 0.001, ANOVA followed by Tukey's posttest.



Supplementary Figure 5. IL-6 controls the reciprocal regulation of IFN- γ and IL-4/10 responses in tumor-associated antigen-specific CD4⁺ T cells. (a and b) Young or aged mice were immunized by the transfer of DC pulsed MuLV EnvH13.3 peptide to prime cognate CD4⁺ T cells. Anti-IL-6 Ab or control Ab was injected 1 day before and after immunization. Five days after immunization, RMA tumor cells were inoculated in the mice. Further 5 days later, draining LNs were harvested and re-stimulated with H13.3 peptide-pulsed DC. Representative dot plots of intracellular cytokine staining (a) and the frequencies (b) of IFN- γ - or IL-10-positive cells in gated CD4⁺ T cells were shown. (c) Draining LN cells were stimulated with H13.3 peptide-pulsed DC for 36 h. The concentration of IL-4 or IL-10 in the culture supernatant was assessed by ELISA. The values are mean ± SEM with n = 4-5 mice/group. Data are representative from two independent experiments. Asterisks indicate the significance of the difference between indicated groups; * p < 0.05, ** p < 0.01, *** p < 0.001 (one-way ANOVA followed by Tukey's post hoc tests).

С а 🗆 WT CD8 SP CD4 SP WΤ c-Maf Mut 4 c-Maf Mut ³H incorporation 3.00% 80.7% 75.6% 3 47% Total No. (x 10⁻⁶) 2.5 8 3 10 10 (x 10⁻⁴ cpm) 2.0 6 500 1.5 2 4 1.0 9.95% 11.99 2 0.5 0 CD4 TCR-β⁺CD69⁺ in DP IL-6: -+ positive cells 20 10 10 14.1% 🗆 wт 13.0% CD69 15 c-Maf Mut 10 5 % 0 TCR-β (Gated on DP) b WΤ c-Maf Mut 11.0% 0.273% CD8 naive CD4 naive 0.313% 8.84% 10 NERGON Total No. (x 10⁻⁶) 5 15 4 **CD8** 10 3 2 1 57.1% 31.6% 10 58.5% 32.4% 0 CD4 CD4+CD62LhiCD44lo 10⁴ 10.1% 5.76% 4.02% 18.4% positive cells 30 103 10 D WT CD44 20 102 10 c-Maf Mut 10 10 2.64% .5% 6.79% 70.8% 100 % 0

CD62L (Gated on CD4)

Supplementary Figure 6. Normal development and proportion of T cells in Maf^{Oft/Off} mutant mice. (a) Frequencies of CD4⁺CD8⁻, CD4⁻CD8⁺ single positive (SP), CD4⁺CD8⁺ double positive (DP) thymocytes in the thymus (upper left 2 panels), and TCR- β^{hi} CD69⁺ cells in gated CD4⁺CD8⁺ DP thymocytes (lower left panels) of homozygous c-Maf-mutant mice (c-Maf Mut) or littermate control mice (WT) were determined by flow cytometry. Absolute numbers of CD4⁺CD8⁻ or CD4⁻CD8⁺ thymocytes are also shown (right). (b) Frequencies of CD4⁺CD8⁻, CD4⁻CD8⁺ T cells in the spleen and lymph nodes (upper left 2 panels), and peripheral naïve (CD44^{lo}CD62L^{hi}) CD4⁺ T cells in the periphery are shown. Absolute numbers of CD4⁺CD4⁺ or CD44^{lo}CD8⁺ naïve T cells in the spleen and lymph nodes are also shown (right). The data are mean ± SEM with n = 3-4 mice/group. (c) Young naïve CD4⁺ T cells were isolated from homozygous c-Maf-mutant mice (c-Maf Mut) or littermate control mice (WT), and then were stimulated with plate-coated anti-CD3 and anti-CD28 Abs, and then their proliferation were determined by [³H] thymidine incorporation in triplicate assay. Data are one of two independent experiments and are mean ± SEM with n = 3-4 mice/group.

Supplementary Figure 6

IFN-γ II -4 IL-13 a 🖽 b + IL-12 40 20 IL-6 0+1L-6 + IL-6 + Control Ab ≁ 30 15 + IL-6 + Anti-IL-6R Ab ng / ml 150 Ocontrol Ab 20 Anti-IL-6R Ab 10 100 (bg / ml) 0 IL-21 IL-2 IL-5 40 0.4 0.3 50 30 0.3 ng / ml 0.2 0† 0 20 20 40 80 60 10 IL-21 0 150₁ -▲ + IL-6 + IL-12 n IL-10 11 -9 Foxp3 н. + + IL-12 Relative expression 0.8 1.0 2.5 0.25 + IL-6 (Gapdhs) 9.0 7 0 7 0 8.0 2.0 0.2 (m 100 (m / 6d) 50 🛧 No cytokine 0.6 Δ ng / ml 1.5 0.15 0.4 1.0 0.1 50 0.2 n 0 C CSTBILE Balbic Balbic CSTBILE MUS Ballol 0+ 0 AN CON z ঠ Not 20 40 60 80 Ma Time for stimulation (h) C3H/HeH C3H/HeH C3H/HeH C3H/HeH + IL-6 + IL-6 d + IL-6 · IL -6 C57BL/6 + anti-IL-4 Ab + anti-IL-21 Ab 2.17% 2.09% 8.21 ··· 23.29 16.19 1.78% - 10 2.88% 0.990% 7.29% 76.19 1.93% 4.96% IFN-y Balb/c 0.480% 0.882% 0.833% 0.832% 5 34 19.8 9 21

Supplementary Figure 7. IL-6 stimulation alters the early cytokine productions in activated CD4⁺ T cells through the c-Maf activity. (a) Young naïve OT-II cells were stimulated with anti-CD3 and anti-CD28 Abs *in vitro*. Indicated cytokines and Abs were also added at the initiation of culture. At the indicated time points after primary stimulation, the culture supernatants were collected, and then IL-4 and IL-21 concentrations were analysed by ELISA in triplicate assay. (b and c) Polyclonal naïve CD4⁺ T cells were isolated from young Balb/c, C57BL/6 or C3H/HeH (WT amd c-Maf^{Off/Off} mutant) background mice, and then were stimulated in the presence or absence of exogenous IL-6 under Th1-skewed condition (plus IL-12). Sixty hours later, indicated cytokine concentrations in the culture supernatant were determined. mRNA was also isolated to examine the gene expression of *Foxp3* by qPCR (c). The values are mean \pm SEM with n = 3 mice/group. Statistical significant differences between IL-6-sensitized and non-sensitized conditions in each strain are expressed as asterisks; * p < 0.05, ** p < 0.01, *** p < 0.001 (one-way ANOVA followed by Tukey's post hoc

0.860%

88.6%

5.20%

78.5%

IL-10

86.8%

3.16%

Supplementary Figure 7

tests). (d) Polyclonal naïve CD4⁺ T cells were isolated from young Balb/c, or C57BL/6 background mice, and then were stimulated in the presence or absence of IL-6 and indicated Abs under Th1-skewed condition (plus IL-12). Five days later, T cells were re-stimulated and analyzed for the ability to produce IFN- γ and IL-10. All data shown are representative of 2 or 3 independent experiments.